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(54) **Enzymatic reductions with dihydrogen via metal catalyzed cofactor regeneration**

Enzymatische Reduktion mit Hydrogen mittels metallkatalysierter Kofaktorregenerierung

Réductions enzymatiques avec de l'hydrogène par régénération du cofacteur catalysée par un métal

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EP 1 621 613 B1

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Description

CROSS REFERENCE TO RELATED APPLICATIONS

- 5 **[0001]** This is a divisional application of EP-A-1 181 356 published on 14 September 2000 as WO 00/53731
[0002] This application claims priority to U.S. Provisional Patent Application Serial Number 60/123,833, filed March 11, 1999.

FIELD OF THE INVENTION

- 10 **[0003]** This invention relates generally to methods of employing H₂ as a reducing agent for an unsaturated organic compound, to form a reduced organic product, in the presence of a catalyst comprising a metal salt or complex, a nicotinamide cofactor; and a nicotinamide cofactor dependent enzyme.

BACKGROUND OF THE INVENTION

- 15 **[0004]** The present invention relates to catalytic processes for the use of H₂ as a reducing agent for organic compounds, in the presence of catalysts containing enzymes. Many man-made catalysts are known for reduction and/or hydrogenation reactions, but there are many limitations in the ability of the known catalysts to selectively reduce or hydrogenate one unsaturated functional group in the presence of another functional group. Moreover, most prior art catalysts and processes cannot selectively produce optically active products, as is highly desirable in the production of compositions for human or animal consumption, such as food or pharmaceuticals.

- 20 **[0005]** In contrast, many enzymes are capable of highly selective reduction of their natural substrates. In some cases enzymes catalyze unique transformations that require multiple steps by traditional synthetic methods. Moreover, a wide range of unnatural substrates, including a wide variety of unsaturated organic compounds, can be enzymatically reduced, with high chemo-, regio- and/or enantioselectivity, under mild reaction conditions. It is therefore highly desirable to employ enzymatic processes for reductions of unsaturated compounds of commercial interest, and especially for the preparation of chiral molecules, as taught by Simon et al. (Ang. Chem. Int. Ed. Engl. 1985, 24, 539-53). Unfortunately, most enzymes capable of catalyzing such reduction reactions require the presence of cofactors, which function as biological reducing agents. One broad class of enzymes capable of selective reduction and/or hydrogenation of unsaturated organic compounds are the nicotinamide dependent oxidoreductases, as discussed by Walsh (Enzymatic Reaction Mechanisms Freeman & Co., N.Y., 1979; pages 311-521).

- 25 **[0006]** Nicotinamide dependent oxidoreductases require the presence of nicotinamide cofactors. The structures of the naturally occurring nicotinamide cofactors, (NAD⁺, NADP⁺, NADH and NADPH) are shown below.

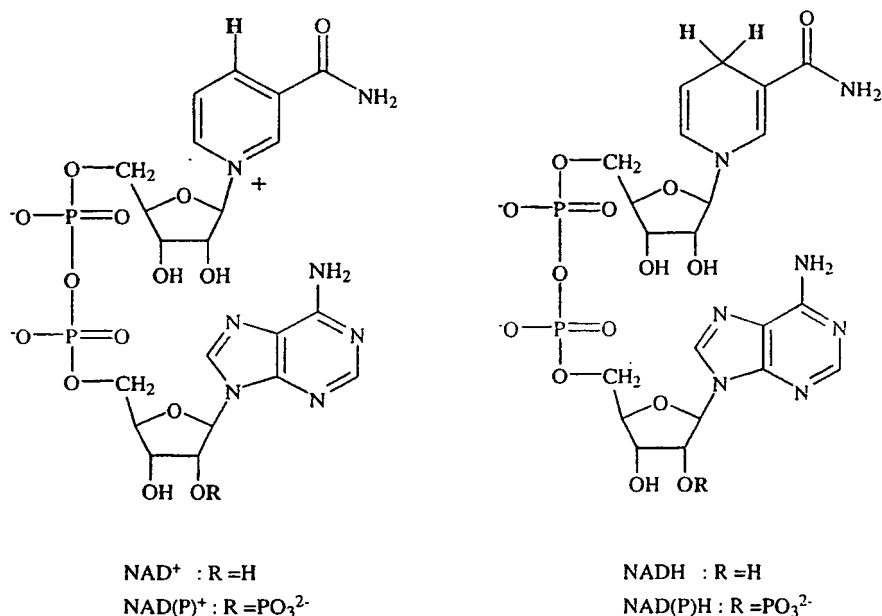
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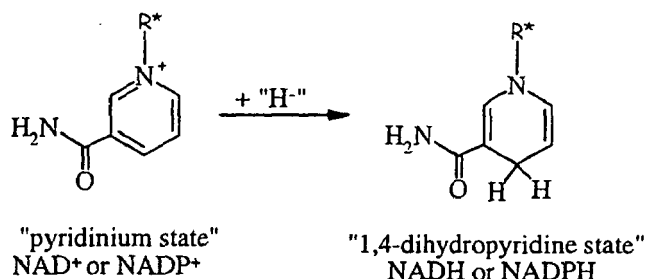
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- [0007]** A reduced nicotinamide cofactor (NADH or NADPH) binds to the nicotinamide cofactor dependent enzyme,

and transfers a "hydride" (two electrons and one hydrogen nucleus) to reduce a substrate that also binds to the enzyme. After the substrate is reduced, the enzyme releases the oxidized form of the nicotinamide cofactor (NAD⁺ or NADP⁺). Biological systems typically recycle the oxidized nicotinamide cofactors, by employing an external reducing agent, in combination with other enzymes, to regenerate the reduced form of the nicotinamide cofactors. In these nicotinamide cofactors the nicotinamide ring (shown schematically immediately below) is the reactive group. To regenerate the reduced cofactor, an external reducing agent must transfer the equivalent of a "hydride" to the oxidized (pyridinium) form of the cofactor, regioselectively to form the reduced (1,4-dihydropyridine) form of the cofactor.



[0008] Although nicotinamide cofactor dependent enzymes and nicotinamide cofactors are present in all living organisms at low concentrations, they tend to be chemically unstable under non-biological conditions, and are extremely expensive in purified form. Because of their high cost, most industrial processes that seek to employ a combination of enzymes and nicotinamide cofactors must supply a method to regenerate the nicotinamide cofactors.

[0009] A number of methods for cofactor regeneration are known, as discussed by Chenault and Whitesides (Appl. Biochem. Biotechnol. 1987, 14, 147-97), and in *Enzymes in Organic Synthesis* K. Drauz, H. Waldeman, Eds.; VCH: Weinheim, 1995; pages. 596-665. The most widely used methods for cofactor regeneration employ a chemical reducing agent and second enzyme to regenerate the nicotinamide cofactors. For example, using glucose as a reducing agent, glucose oxidase has been shown to successfully regenerate NADP⁺/NADPH through up to 4 x 10⁴ turnovers (see Wong, and Whitesides J. Org. Chem. 1982, 47, 2816-18; Wong et al., J. Am. Chem. Soc. 1985, 107, 4028-31; Obon et al., Biotech. Bioeng. 1998, 57, 510-17). Hummel et al., (Appl. Microbiol. Biotechnol. 1987, 26, 409-416) have shown that a combination of formate dehydrogenase and formate salts regenerates NADH from NAD⁺ with turnover numbers for the reduced cofactor as high as 6 x 10⁵. In these methods, a second enzyme couples the regenerated NADH to substrate reduction. In cases where activity of two separate enzyme systems can be accomplished *in vitro* without undue complexity or expense, reduction of substrates with a chemical reducing agent and two enzymes can be a viable cofactor regeneration method.

[0010] Prior art attempts to electrochemically regenerate the nicotinamide cofactors avoid the need for a second enzyme, but direct electrochemical methods have typically not achieved adequate cofactor regeneration, primarily due to formation of inactive nicotinamide-dimers. The addition of certain types of electron transfer catalysts or "mediators" to electrochemical methods can greatly improve electrochemical regeneration, as disclosed by Steckhan (Topics in Current Chemistry, 1994, 170, 83-111). The most successful mediators are the rhodium complexes disclosed by Steckhan et al (*Ang. Chem.*, 1982, 94, 786; U.S. Patent 4,526,661 and *Organometallics* 1991 10, 1568-77). Although these electrochemically-based systems have been successfully coupled to enzymatic reduction reactions, thus far cofactor turnover numbers remain too low to be commercially viable.

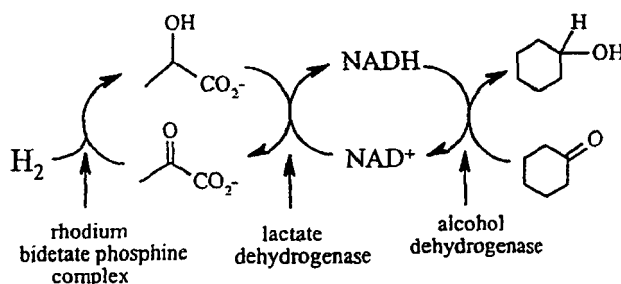
[0011] Photochemically assisted methods for chemical reduction of NAD(P)⁺ to NAD(P)H in the presence of similar rhodium electron transfer catalysts, and successful coupling to enzymes has been reported (Willner, et al., in J. Am. Chem. Soc., 1984, 106, 5352-53, and J. Chem. Soc., Perkin Trans., 2 1990, 559-64; Franke and Steckhan in *Angew. Chem. Intl. Ed. Engl.*, 1988, 27, 265; and Aono and Okura in *Inorg. Chim. Acta*, 1988, 152, 55-59). Nevertheless, an economically competitive and long-lived photochemical cofactor regeneration system which achieves cofactor regeneration at rates and efficiencies competitive with enzymatic methods has remained an elusive goal.

[0012] Cofactor regeneration with non-biological chemical reducing agents is a simple approach, but most chemical reducing agents are not desirably selective for production of 1,4-dihydro isomers of the cofactor nicotinamide ring, as discussed by Ohnishi and Tanimoto (*Tetrahedron Lett.* 1977, 1909-12). Dithionite salts are preferred reducing agents in this regard, providing up to about 10² turnovers of the nicotinamide cofactor, as described by Jones, et al. (*J. Chem. Soc., Chem. Commun.*, 1972, 856-57). Nevertheless, dithionite salts are incompatible with many enzymes and react directly with many substrates, are expensive, and generate undesirable sulfur-containing wastes. (such as isopropanol) and an alcohol dehydrogenase from *T. brockii*, in a "coupled substrate" method to reduce certain organic substrates. In the "coupled substrate" method one enzyme catalyzes both (a) reduction of NADP⁺ to NADPH by the hydride donor alcohol, and (b) reduction of ketone substrates such as 2-heptanone by NADPH.

[0013] Dihydrogen (H_2), is a highly desirable chemical reducing agent. H_2 is a strong reducing agent, and can be inexpensively produced and stored in high purity on a large scale. H_2 is typically innocuous towards enzymes and cofactors, and because it is completely consumed in most reduction reactions, it leaves no residues to complicate purification or create chemical waste. Many examples are known of the use of H_2 as a reducing agent in the presence

of transition metal catalysts (in the absence of enzymes or cofactors).
 [0014] Nevertheless, the use of hydrogen as a reducing agent in conjunction with enzymes has only been possible with complex multi-step or multi-component catalyst systems that employ indirect coupling of the of H_2 to cofactor regeneration. Wong et al (J. Am. Chem. Soc. 1981, 103, 6227-8), and Otsuka, et al. (J. Mol. Catal. 1989, 51, 35-9), have reported the use H_2 and hydrogenase enzymes, which are air-sensitive and not readily available, to reduce electron receptors such as methylviologen to produce radicals. The radicals can be coupled via a second enzyme (ferredoxin reductase) to NADH regeneration. The NADH is coupled to substrate reduction via a third enzyme.

[0015] Abril and Whitesides (J. Am. Chem. Soc. 1982, 104, 1552-54) reported a multi-component approach in which a water soluble rhodium complex of a bidentate phosphine ligand was employed to activate H_2 , but two other enzymes and high concentrations of a lactate/pyruvate hydrogen carrier intermediate were required for substrate reduction.



[0016] A recent report by Bhaduri, et al. (J. Am. Chem. Soc. 1998, 120, 12127-28) describes the use of H_2 to reduce NAD^+ to NADH via a "secondary coupling" system. H_2 reacts with a platinum carbonyl cluster in a methylene chloride phase and reduces a redox-active dye (Safranin O). The reduced dye then diffuses to an aqueous phase where it reduces NAD^+ to produce NADH. The NADH then combines with lactate dehydrogenase to reduce pyruvic acid to lactic acid. Nevertheless, the platinum carbonyl clusters are insoluble and unstable in water, necessitating the use of the redox-active dye and a two phase solvent system. No enzymes other than lactate dehydrogenase, or substrates other than pyruvic acid were reported.

[0017] Despite the potential advantages of the use of H_2 as a reducing agent for regenerating nicotinamide cofactors, simple, effective, and economically attractive methods for doing so have not been achieved. There is an unmet need in the art for simple and effective methods for combining the low cost and environmental desirability of H_2 as a reducing agent with the exquisite selectivity of enzyme catalysis. It is to such a desirable object that the present invention is primarily directed.

SUMMARY OF THE INVENTION

[0018] The present invention meets the unmet needs in the art, by providing processes and catalyst compositions, which employ H_2 as a reducing agent for unsaturated organic compounds in the presence of catalysts comprising enzymes.

[0019] In accordance with the purpose(s) of this invention, as embodied and broadly described herein, the invention therefore relates to, in one aspect, a process for reducing an unsaturated organic compound, comprising mixing the unsaturated organic compound and H_2 in the presence of a catalyst to form a reduced organic product, wherein the catalyst comprises:

- a) at least one metal salt or complex,
 - b) at least one nicotinamide cofactor; and
 - c) a nicotinamide cofactor dependent enzyme,
- wherein:

- i) when the metal salt or complex is a platinum carbonyl cluster complex, the catalyst does not comprise a redox active dye; and
- ii) when the metal salt or complex is a rhodium phosphine complex, the nicotinamide cofactor dependent enzyme is not a mixture of horse liver alcohol dehydrogenase and lactate dehydrogenase.

EP 1 621 613 B1

[0020] In another aspect, the invention provides a process for reducing an unsaturated organic compound, comprising mixing the unsaturated organic compound and H₂ in the presence of a catalyst to form a reduced organic product, wherein the catalyst comprises:

- 5 a) a substantially aqueous buffer solution having a pH from about 6.5 to about 9.0,
b) a water-soluble metal salt or complex comprising iron, ruthenium, osmium, cobalt, rhodium, iridium, nickel, palladium, or copper,
c) a nicotinamide cofactor comprising NAD⁺, NADH, NADP⁺, NADPH, or a mixture thereof; and
10 d) one nicotinamide cofactor dependent enzyme classified under the EC system as an 1.x.1.y. class enzyme, wherein x is 1,3,4, 5 or 10.

[0021] In a different aspect, the invention provides a process for reducing an unsaturated organic compound, comprising:

- 15 a) contacting H₂ and a catalyst, and
b) contacting an unsaturated organic compound with the catalyst to form a reduced organic product, wherein the catalyst comprises:
- 20 i) at least one metal salt or complex,
ii) at least one nicotinamide cofactor; and
iii) a nicotinamide cofactor dependent enzyme, and wherein
iv) when the metal salt or complex is a platinum carbonyl cluster complex, the catalyst does not comprise a redox active dye; and
v) when the metal salt or complex is a rhodium phosphine complex, the nicotinamide cofactor dependent enzyme
25 is not a mixture of horse liver alcohol dehydrogenase and lactate dehydrogenase.

[0022] In yet another aspect, the invention provides a process for reducing an unsaturated organic compound, comprising:

- 30 a) contacting H₂, at least one metal salt or complex, and at least one nicotinamide cofactor to form at least some reduced nicotinamide cofactor, and
b) contacting the reduced nicotinamide cofactor, a nicotinamide cofactor dependent enzyme, and an unsaturated organic compound under conditions effective to form at least some of a reduced organic product, wherein
35 i) when the metal salt or complex is a platinum carbonyl cluster complex, the catalyst does not comprise a redox active dye; and
ii) when the metal salt or complex is a rhodium phosphine complex, the nicotinamide cofactor dependent enzyme is not a mixture of horse liver alcohol dehydrogenase and lactate dehydrogenase.

[0023] The invention further relates to a process comprising adding dihydrogen, H₂, to an unsaturated organic substrate using as a catalyst a mixture comprising an enzyme, a nicotinamide cofactor, a metal salt or complex and optionally ligands, wherein the metal of the metal salt or complex is selected from iron, cobalt, nickel, copper, ruthenium, palladium, osmium, and iridium.

[0024] The invention also relates to a composition for reducing unsaturated organic compounds comprising H₂ and a catalyst, the catalyst comprising:

- 45 a) at least one metal salt or complex,
b) at least one nicotinamide cofactor; and
50 c) a nicotinamide cofactor dependent enzyme, wherein
i) when the metal salt or complex is a platinum carbonyl cluster complex, the catalyst does not comprise a redox active dye; and
55 ii) when the metal salt or complex is a rhodium phosphine complex, the nicotinamide cofactor dependent enzyme is not a mixture of horse liver alcohol dehydrogenase and lactate dehydrogenase.

[0025] In another embodiment, the invention relates to a process for stabilizing the activity of an oxidoreductase

enzyme, comprising mixing a phosphine or phosphite with an oxidoreductase enzyme.

[0026] Additional advantages of the invention will be set forth in part in the description that follows, and in part will be obvious from the description, or may be learned by practice of the invention. The advantages of the invention will be realized and attained by means of the elements and combinations particularly pointed out in the appended claims. It is to be understood that both the foregoing general description and the following detailed description are exemplary and explanatory only and are not restrictive of the invention, as claimed.

DESCRIPTION OF THE PREFERRED EMBODIMENTS

[0027] The present invention may be understood more readily by reference to the following detailed description of preferred embodiments of the invention and the Examples included therein and to the Figures and their previous and following description. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only and is not intended to be limiting.

[0028] It must be noted that, as used in the specification and the appended claims, the singular forms "a," "an" and "the" include plural referents unless the context clearly dictates otherwise. Thus, for example, reference to "an aromatic compound" includes mixtures of aromatic compounds.

[0029] Often, ranges are expressed herein as from "about" one particular value, and/or to "about" another particular value. When such a range is expressed, another embodiment includes from the one particular value and/or to the other particular value. Similarly, when values are expressed as approximations, by use of the antecedent "about," it will be understood that the particular value forms another embodiment. It will be further understood that the endpoints of each of the ranges are significant both in relation to the other endpoint, and independently of the other endpoint.

[0030] In this specification and in the claims that follow, reference will be made to a number of terms that shall be defined to have the following meanings:

[0031] A weight percent of a component, unless specifically stated to the contrary, is based on the total weight of the formulation or composition in which the component is included.

[0032] A residue of a chemical species, as used in the specification and concluding claims, refers to the moiety that is the resulting product of the chemical species in a particular reaction scheme or subsequent formulation or chemical product, regardless of whether the moiety is actually obtained from the chemical species. Thus, an ethylene glycol residue in a polyester refers to one or more $-OCH_2CH_2O-$ repeat units in the polyester, regardless of whether ethylene glycol is used to prepare the polyester. Similarly, a sebacic acid residue in a polyester refers to one or more $-CO(CH_2)_8CO-$ moieties in the polyester, regardless of whether the residue is obtained by reacting sebacic acid or an ester thereof to obtain the polyester.

[0033] "Optional" or "optionally" means that the subsequently described event or circumstance may or may not occur, and that the description includes instances where said event or circumstance occurs and instances where it does not. For example, the phrase "optionally substituted lower alkyl" means that the lower alkyl group may or may not be substituted and that the description includes both unsubstituted lower alkyl and lower alkyl where there is substitution.

[0034] By the term "effective amount" of a compound or property as provided herein is meant such amount as is capable of performing the function of the compound or property for which an effective amount is expressed. As will be pointed out below, the exact amount required will vary from process to process, depending on recognized variables such as the compounds employed and the processing conditions observed. Thus, it is not possible to specify an exact "effective amount." However, an appropriate effective amount may be determined by one of ordinary skill in the art using only routine experimentation.

[0035] In one aspect, the invention provides a process for reducing an unsaturated organic compound, comprising mixing the unsaturated organic compound and H_2 in the presence of a catalyst to form a reduced organic product, wherein the catalyst comprises:

- a) at least one metal salt or complex,
- b) at least one nicotinamide cofactor; and
- c) a nicotinamide cofactor dependent enzyme.

[0036] In additional embodiments of the invention, one or more of the following further exclusions and/or limitations to the scope may apply:

- a) the catalyst comprises one and only one enzyme,
- b) the metal salt or complex does not comprise a platinum carbonyl cluster complex,
- c) the metal salt or complex does not comprise a rhodium phosphine complex, and/or
- d) the formation of the reduced organic product is capable of occurring in the absence of electrochemical or photochemical sources of external energy.

EP 1 621 613 B1

[0037] It is intended that when it is stated that the catalyst comprises "one" enzyme, one, and only one, enzyme is present in the catalyst.

[0038] Platinum carbonyl cluster complexes are metal salts or complexes having three or more platinum atoms, wherein each platinum atom is bonded to at least one carbon monoxide ligand. Examples of common platinum carbonyl complexes include but are not limited to $[\text{Pt}_9(\text{CO})_{18}]^{2-}$, and $[\text{Pt}_{12}(\text{CO})_{24}]^{2-}$.

[0039] When the metal salt or complex is a platinum carbonyl cluster complex; one or more of the following further exclusions and/or limitations may apply:

- a) the catalyst does not comprise a redox active dye, redox active dyes including but not being limited to Safranin O, methyl viologen, methylene blue, or the like,
- b) the unsaturated organic substrate is not a pyruvate,
- c) the reduced organic product is not a lactate,
- d) the enzyme does not comprise lactate dehydrogenase, or
- e) the process does not comprise water-immiscible solvents, which include but are not limited to methylene chloride, chloroform, and the like.

[0040] Rhodium phosphine complexes are metal salts or complexes having a rhodium atom bonded to a phosphine residue, the phosphine residue having the formula PR_3 , wherein the three R groups independently comprise hydrocarbyl groups or residues. In other embodiments, the metal salt or complex does not comprise a rhodium bis(phosphine) complex. Rhodium bis(phosphine) complexes have a rhodium atom bonded to two phosphine residues, that may or may not be bonded to each other through a bridging group, to form a bidentate phosphine ligand. In some embodiments, the metal salts or complexes do not comprise a water soluble rhodium phosphine complex.

[0041] When the metal salt or complex is a rhodium phosphine complex, one or more of the following further exclusions and/or limitations to the scope of certain embodiments of the present invention may apply:

- a) the nicotinamide cofactor dependent enzyme is not lactate dehydrogenase,
- b) the nicotinamide cofactor dependent enzyme is not horse liver alcohol dehydrogenase,
- c) the process does not comprise lactate or pyruvate, and/or
- d) the unsaturated organic substrate does not comprise cyclohexanone or 2-norbornanone.

[0042] In preferred embodiments of the processes of the current invention, the formation of the reduced organic product is capable of occurring in the absence of electrochemical or photochemical sources of external energy. In other words, the preferred processes of the present invention only require thermal energy to successfully reduce unsaturated organic compounds with H_2 , and they do not require the presence or supply of visible or ultraviolet light, or the presence or supply of surfaces or electrodes supplied with electromotive force differentials in order to induce the formation of reduced organic products. More particularly, preferred embodiments of the present invention do not require the presence or supply of light, or the presence or supply of surfaces or electrodes supplied with electromotive force differentials to initiate the reaction of H_2 with the metal salt or complex component of the catalyst.

[0043] The order of mixing of the H_2 , the unsaturated organic compound, the metal salt or complex, the nicotinamide cofactor, and the nicotinamide cofactor dependent enzyme may occur in any order, or simultaneously.

[0044] In a preferred embodiment, the invention provides a process for reducing an unsaturated organic compound, comprising mixing the unsaturated organic compound and H_2 in the presence of a catalyst to form a reduced organic product, wherein the catalyst comprises:

- a) at least one metal salt or complex,
 - b) at least one nicotinamide cofactor; and
 - c) a nicotinamide cofactor dependent enzyme,
- wherein:

- i) when the metal salt or complex is a platinum carbonyl cluster complex, the catalyst does not comprise a redox active dye; and
- ii) when the metal salt or complex is a rhodium phosphine complex, the nicotinamide cofactor dependent enzyme is not a mixture of horse liver alcohol dehydrogenase and lactate dehydrogenase.

[0045] The unsaturated organic compounds of the invention are carbon containing molecules that have at least one multiple bond between two atoms of the compound, X and Y. The atoms X and Y may be any atom of the Periodic Table capable of forming multiple bonds. Preferably, the X and Y atoms are independently selected from carbon, nitrogen, oxygen, sulfur, and phosphorus atoms. Preferably, at least one of the X or Y atoms is a carbon atom. In many preferred

EP 1 621 613 B1

embodiments, one of the X or Y atoms is a carbon atom, and the other atom is carbon, oxygen or nitrogen atom. Preferably, the multiple bond between the atoms X and Y is a double bond, or a triple bond. The unsaturated organic compounds can have more than one multiple bond, and the multiple bonds may form conjugated combinations of multiple bonds.

5 **[0046]** Examples of classes of unsaturated organic compounds of the invention include but are not limited to

- a) a ketone, an aldehyde, a carboxylic acid, a carboxylic acid ester, or an amide,
- b) an α - β unsaturated derivative of a ketone, an aldehyde, a carboxylic acid, a carboxylic acid ester, or an amide,
- c) a fatty acid, a monoglyceride, a diglyceride, or a triglyceride, having an olefinic unsaturated group,
- 10 d) an olefin, an aromatic compound or a heteroaromatic compound,
- e) an imine, or an oxime,
- f) a sugar, an amino acid, a peptide, or a protein, wherein the sugar, amino acid, peptide, or protein has an unsaturated group.

15 **[0047]** Examples of unsaturated organic compounds include but are not limited to acetoin, 1,1-dimethoxyacetone, glycerone, acetophenone, 2-acetylfuran, hydroxyacetone, 6-methyl-5-heptene-2-one, 5-norbornene-2-one, 2-heptanone, 8-oxo-2-nonanone, cyclopropyl methyl ketone, L-sorbose, 2,4,6/3,5-pentahydroxycyclohexanone, aldose, glyoxylate, pyruvate, acetoacetate, ethyl 4-chloroacetoacetate, ethyl 4-oxo-hexanoate, oxaloacetate, 2,5-diketo-D-gluconic acid, D-glucono-1,5-lactone, 5 α -androstane-3,17-dione, androst-4-ene-3,17-dione, L lactaldehyde, fructose, D-glycer-20 aldehyde, orotate, 2-oxoglutarate, 3-hydroxypyruvate, glyoxylate, and L-lysine.

[0048] The reduced organic products of the invention typically correspond in structure to the starting unsaturated organic compound reduced, or a residue thereof, in which the multiple bond between the X and Y atoms is cleaved in a reduction process. Reduction is broadly defined for the purposes of this disclosure as the addition of one or more electrons to the multiple bond, the electrons having been donated by another compound, termed a reducing agent. Rearrangement and/or cleavage of functional groups may also occur during or subsequent to the reduction of the multiple bond. Often, reduction of the multiple bond is accompanied by the bonding of one or more hydrogen atoms to at least one of the X and Y atoms.

25 **[0049]** In preferred embodiments, the unsaturated organic compound comprises a carbon atom doubly bonded to another carbon, oxygen, or nitrogen atom, the reducing agent is H₂, and the reduced organic compound has a structure corresponding to the addition of two hydrogen atoms to the double bond, as illustrated below, in which case the reduction reaction may also be termed a hydrogenation reaction.



40 **[0050]** If the S1, S2 groups and/or Y groups illustrated in the equation above are different, reduction and formation of the new carbon-hydrogen bond can result in the formation of two enantiomeric isomers of the reduced organic product. The catalysts of the invention, which comprise enzymes, often selectively produce only one of the two possible enantiomeric isomers.

45 **[0051]** The processes of the invention occur in the presence of a catalyst comprising at least the following components:

- a) at least one metal salt or complex,
- b) at least one nicotinamide cofactor; and
- c) a nicotinamide cofactor dependent enzyme.

50 **[0052]** The components of the catalyst need not, but often do comprise a single phase or mixture, so long as the component can interact in a suitable manner. The components of the catalysts can be dissolved in one or more solvents, or can be dispersed upon or bonded to one or more support phases. In preferred embodiments of the invention, the three components of the catalyst comprise a mixture that is dispersed or substantially dissolved in a liquid medium. Preferably, the liquid medium comprises a homogeneous liquid phase. The liquid medium may comprise an organic solvent, water, or a mixture thereof. Preferably, the liquid medium comprises a substantially aqueous phase. In many preferred embodiments, the liquid medium comprises an aqueous buffer solution having a pH from about 6.5 to about 9.0, which typically maximizes the stability of the nicotinamide cofactor dependent enzymes and the nicotinamide cofactors. More preferably, the pH of the aqueous buffer solution is between about 7.0 to about 8.5. Aqueous buffer solutions

comprising phosphate salts are preferred aqueous buffer solutions.

[0053] The nicotinamide cofactors employed in the invention include the naturally occurring nicotinamide adenine dinucleotides described hereinabove, and further include any structural analogs thereof that are capable of effectively interacting with the nicotinamide cofactor dependent enzymes of the invention to reduce unsaturated organic compounds.

Structural analogs of the naturally occurring cofactors comprise compounds wherein the naturally occurring structure is modified by the addition of or removal of one or more functional groups. Preferably, the nicotinamide cofactors of the invention are the biologically preferred cofactors NAD⁺, NADH, NADP⁺, NADPH, or a mixture thereof. It is well known many nicotinamide cofactor dependent enzyme will preferentially bind and/or utilize only specifically phosphorylated cofactors (such as NADP⁺, and NADPH), while other nicotinamide cofactor dependent enzyme will preferentially bind and/or utilize only bind and/or utilize non-phosphorylated cofactors (such as NAD⁺, NADH).

[0054] The nicotinamide cofactor dependent enzymes employed in the invention comprise any naturally occurring or biotechnologically modified or engineered enzyme that requires the presence of a nicotinamide cofactor, or an analog thereof, in order to reduce or oxidize organic substrates. It is to be understood that while many nicotinamide cofactor dependent enzymes catalyze oxidation reactions in nature, they are nevertheless often useful for catalyzing the reduction processes of the invention.

[0055] The Enzyme Commission of International Union of Biochemistry and Molecular Biology ("EC") has devised a well-known four digit numerical system for classifying enzymes, in terms of the type of reaction that the enzymes catalyze. The EC classification system has been described by Dixon, Webb, Thorne, and Tipton ("Enzymes", Chapter 5, pages 207-230, Academic Press, 1979), for the purposes of describing the classification of enzymes and the relationship of the classifications to the substrates and products of the reactions catalyzed by the enzymes.

[0056] The first digit of an Enzyme Classification ("EC") corresponds to one of six classes of enzymes. The nicotinamide cofactor dependent enzymes of the present invention are all members of the class of oxidoreductases, which comprise enzymes that mediate the transfer of electrons, H atoms, or hydride atoms. Oxidoreductases all have a first digit EC classification of "1".

[0057] The second digit of an EC classification relates to subclasses of the enzymes specified by the first digit. For an oxidoreductase, the second digit pertains to a group of twenty subclasses of functional groups of the substrates and/or products for the enzyme, i.e. classes of the functional groups in the substrates or products which undergo oxidation or reduction. The third digit of an EC classification relates to another series of sub-subclasses. In the case of the oxidoreductases, a third digit of "1" indicates an oxidoreductase that requires the presence of nicotinamide cofactors. Therefore, the nicotinamide cofactor dependent enzymes of the present invention are all classified under the EC system as "1.x.1.y." class enzymes. The fourth digit of an EC classification specifies a serial number for the enzyme, which is often related to the particular identity of the natural substrate of the enzyme.

[0058] Preferred embodiments of the invention employ nicotinamide cofactor dependent enzymes, which may be further defined by the values of x and y in the EC classification of the enzyme. The second digit of an EC classification of an enzyme, i.e., the value of x, corresponds to the class of functional group oxidized or reduced by the enzyme. Preferred enzymes of the invention have x values of 1, 3, 4, 5, or 10, corresponding to ketone or aldehyde reductions (x=1); olefin reduction (x=3); imine reductions (x=4 or 5); and reduction of diphenols or ascorbate (x=10). Therefore in preferred embodiments of the present invention, the nicotinamide cofactor dependent enzyme is an enzyme classified under the EC system as an 1.x.1.y. class enzyme, wherein x is 1,3,4, 5 or 10. More than 340 enzymes are presently known to fall within those preferred classes of enzymes.

[0059] For enzymes within the class of 1.1.1.y enzymes, Table 1 below relates the value of y to the name of the enzyme and its natural substrates.

Table 1- 1.1.11.y Class Enzymes.

y	Name of Enzyme
1	alcohol dehydrogenase
2	alcohol dehydrogenase
4	butanediol dehydrogenase
5	diacetyl reductase
6	glycerol dehydrogenase
7	glycerol-3-phosphate dehydrogenase
14	L-Iditol 2-dehydrogenase
18	myo-inositol 2-dehydrogenase
21	aldose reductase
22	UDP glucose 6-dehydrogenase
26	glyoxalate dehydrogenase

EP 1 621 613 B1

(continued)

	y	Name of Enzyme
	27	L-lactate dehydrogenase
5	30	3-hydroxybutyrate dehydrogenase
	37	malate dehydrogenase
	40	malate dehydrogenase
	41	isocitrate dehydrogenase
	42	isocitrate dehydrogenase
10	44	phosphogluconate dehydrogenase
	47	glucose 1-dehydrogenase
	48	galactose 1-dehydrogenase
	49	glucose-6-phosphate 1-dehydrogenase
15	50	3 α -hydroxysteroid dehydrogenase
	51	3 (or 17) β -hydroxysteroid dehydrogenase
	53	3 α (or 20 β) hydroxysteroid dehydrogenase
	55	lactaldehyde reductase
	67	mannitol 2-dehydrogenase
20	72	glycerol dehydrogenase
	83	D-malate dehydrogenase
	95	glycerol dehydrogenase
	119	glucose 1-dehydrogenase
25	122	D-threo-aldose 1-dehydrogenase
	159	12 α -hydroxysteroid dehydrogenase
	176	12 α -hydroxysteroid dehydrogenase

30 **[0060]** For enzymes within the class of 1.3.1.y enzymes, enzymes of class 1.3.1.14 i.e. orotate reductases are preferred. (NADH)

[0061] For enzymes within the class of 1.4.1.y enzymes, Table 2 below relates the value of y to the name of the enzyme and/or its natural substrate.

Table 2: 1.4.1.y Class Enzymes.

	y	Name of Enzyme
35	1	alanine dehydrogenase
	2	glutamate dehydrogenase
	3	glutamate dehydrogenase
40	4	glutamate dehydrogenase
	5	L-amino-acid dehydrogenase
	7	serine dehydrogenase
	8	valine dehydrogenase
	9	leucine dehydrogenase
45	10	glycine dehydrogenase
	11	L-erythro-3,5-diaminohexanoate dehydrogenase
	12	2,4-diaminopentanoate dehydrogenase
	13	glutamate synthase
50	14	glutamate synthase
	15	lysine dehydrogenase
	16	diaminopimelate dehydrogenase
	17	N-methylalanine dehydrogenase
	18	lysine 6-dehydrogenase
55	19	tryptophane dehydrogenase
	20	phenylalanine dehydrogenase

[0062] In preferred embodiments of the invention, the nicotinamide cofactor dependent enzyme is an enzyme classified under the EC system as a 1.1.1.1, a 1.1.1.2, a 1.1.1.4, a 1.1.1.5, a 1.1.1.6, a 1.1.1.7, a 1.1.1.14, a 1.1.1.18, a 1.1.1.21, a 1.1.1.22, a 1.1.1.26, a 1.1.1.27, a 1.1.1.30, a 1.1.1.37, a 1.1.1.40, a 1.1.1.41, a 1.1.1.42, a 1.1.1.44, a 1.1.1.47, a 1.1.1.48, a 1.1.1.49, a 1.1.1.50, a 1.1.1.51, a 1.1.1.53, a 1.1.1.55, a 1.1.1.67, a 1.1.1.72, a 1.1.1.83, a 1.1.1.95, a 1.1.1.119, a 1.1.1.122, a 1.1.1.159, a 1.1.1.176, a 1.3.1.14, a 1.4.1.1, a 1.4.1.2, a 1.4.1.3, a 1.4.1.4, a 1.4.1.5, a 1.4.1.7, a 1.4.1.8, a 1.4.1.9, a 1.4.1.10, a 1.4.1.11, a 1.4.1.12, a 1.4.1.13, a 1.4.1.14, a 1.4.1.15, a 1.4.1.16, a 1.4.1.17, a 1.4.1.18, a 1.4.1.19 or a 1.4.1.20 class enzyme.

[0063] In more preferred embodiments of the invention, the nicotinamide cofactor dependent enzyme is an enzyme classified under the EC system as 1.1.1.1, 1.1.1.2, 1.1.1.5, 1.1.1.6, 1.1.1.7, 1.1.1.14, 1.1.1.18, 1.1.1.21, 1.1.1.26, 1.1.1.27, 1.1.1.37, 1.1.1.40, 1.1.1.41, 1.1.1.42, 1.1.1.47, 1.1.1.48, 1.1.1.49, 1.1.1.72, 1.1.1.83, 1.1.1.95, 1.1.1.119, 14, 1.4.1.1, 1.4.1.3, 1.4.1.4, 1.4.1.9 or 1.4.1.20 class enzyme.

[0064] In other preferred embodiments of the invention the nicotinamide cofactor dependent enzyme is an enzyme classified under the EC system as a 1.1.1.1 or 1.1.1.2, class enzyme.

[0065] In the present invention, the specification of an enzyme by its EC classification does not necessarily strictly limit the classes or species of unsaturated organic compounds that may be reduced by the enzyme in the processes of the invention, or the reduced organic products produced. In many embodiments of the present invention, the methods of the invention will reduce unnatural unsaturated organic compounds, to produce unnatural reduced organic products, which will not be literally specified by the EC classification of an enzyme.

[0066] Nevertheless, it is to be understood that the 2nd and/or 4th digits of the EC classification of a particular oxidoreductase enzyme inherently identifies preferred classes of oxidation and/or reduction reactions that may be catalyzed by the enzyme, and/or the corresponding classes and/or species of substrates and products that may be involved. A listing of the names, EC classifications and the corresponding chemical reactions of the oxidoreductase enzymes relevant to the present invention may found in "Enzyme Nomenclature 1992 - Recommendations of the Nomenclature Committee of the International Union of Biochemistry and Molecular Biology on the Nomenclature and Classification of Enzymes", pages 1-154, Academic Press, 1992, for the purposes of describing the classification of enzymes and the relationship of the classifications to the substrates and products of the reactions catalyzed by the enzymes. Of particular relevance are pages 24-55 (1.1.1.y class enzymes), pages 65-67 (1.2.1.y class enzymes), pages 76-83 (1.3.1.y class enzymes), pages 87-89 (1.4.1.y class enzymes), pages 93-96 (1.5.1.y class enzymes), and 113 (1.10.1.y class enzymes).

[0067] For example, if the nicotinamide cofactor dependent enzyme is an alcohol dehydrogenase [1.1.1.1] or [1.1.1.2], a preferred reduced organic product is an alcohol.

[0068] In a similar manner, preferred classes or species of reduced organic products are hereinbelow identified for a number of particular EC enzyme classifications. For example, if the nicotinamide cofactor dependent enzyme is an aldose reductase [1.1.1.21], a preferred reduced organic product is an alditol. If the nicotinamide cofactor dependent enzyme is a glyoxalate reductase [1.1.1.26], a preferred reduced organic product is a glycolate or glycerate.

[0069] With respect to the identification of preferred species of reduced organic products for particular enzymes: If the nicotinamide cofactor dependent enzyme is a diacetyl reductase [1.1.1.5], a preferred reduced organic product is 3-hydroxy-2-butanone (acetoin). If the nicotinamide cofactor dependent enzyme is a glycerol dehydrogenase [1.1.1.6], a preferred reduced organic product is glycerol. If the nicotinamide cofactor dependent enzyme is an propanediol-phosphate dehydrogenase [1.1.1.7], a preferred reduced organic product is propane-1,2-diol 1-phosphate. If the nicotinamide cofactor dependent enzyme is an L-lactate dehydrogenase [1.1.1.27], a preferred reduced organic product is an (S)-lactate. If the nicotinamide cofactor dependent enzyme is a malate dehydrogenase [1.1.1.37], a preferred reduced organic product is (S)-malate. If the nicotinamide cofactor dependent enzyme is malate dehydrogenase [1.1.1.40], a preferred reduced organic product is (S)-malate. If the nicotinamide cofactor dependent enzyme is isocitrate dehydrogenase [1.1.1.41], a preferred reduced organic product is isocitrate. If the nicotinamide cofactor dependent enzyme is isocitrate dehydrogenase [1.1.1.42], a preferred reduced organic product is isocitrate.

[0070] Moreover, if the nicotinamide cofactor dependent enzyme is glucose 1-dehydrogenase [1.1.1.47], a preferred reduced organic product is β -D-glucose. If the nicotinamide cofactor dependent enzyme is galactose 1-dehydrogenase [1.1.1.48], a preferred reduced organic product is D-galactose. If the nicotinamide cofactor dependent enzyme is glucose 6-phosphate 1-dehydrogenase [1.1.1.49], a preferred reduced organic product is D-glucose 6-phosphate. If the nicotinamide cofactor dependent enzyme is glycerol dehydrogenase (NADP) [1.1.1.72], a preferred reduced organic product is glycerol. If the nicotinamide cofactor dependent enzyme is D-malate dehydrogenase [1.1.1.83], a preferred reduced organic product is (R)-malate. If the nicotinamide cofactor dependent enzyme is phosphoglycerate dehydrogenase [1.1.1.95], a preferred reduced organic product is 3-phosphoglycerate. If the nicotinamide cofactor dependent enzyme is glucose 1-dehydrogenase (NADP) [1.1.1.119], a preferred reduced organic product is D-glucose, D-mannose, 2-deoxy-D-glucose or 2-amino-2-deoxy-D-mannose.

[0071] Additionally, if the nicotinamide cofactor dependent enzyme is alanine dehydrogenase [1.4.1.1], a preferred reduced organic product is L-alanine. If the nicotinamide cofactor dependent enzyme is glutamate dehydrogenase [1.4.1.3], a preferred reduced organic product is L-glutamate. If the nicotinamide cofactor dependent enzyme is glutamate

dehydrogenase [1.4.1.4], a preferred reduced organic product is L-glutamate. If the nicotinamide cofactor dependent enzyme is leucine dehydrogenase [1.4.1.9], a preferred reduced organic product is L-leucine. If the nicotinamide cofactor dependent enzyme is phenylalanine dehydrogenase [1.4.1.20], a preferred reduced organic product is L-phenylalanine.

[0072] It is to be understood that in the preceding description relating the EC classifications of preferred enzymes to their substrates, the invention further provides methods for reducing man-made structural analogs of the natural substrates, to produce unnatural reduced organic products

[0073] It has been found that the maintenance of optimal activity of the enzymes is improved in the presence of certain stabilizers. Therefore, in preferred embodiments, the processes and compositions of the invention further comprise a stabilizer for the nicotinamide cofactor dependent enzyme. Preferably the stabilizers comprise sulfur or phosphorus compounds. Preferred sulfur containing stabilizers have sulfhydryl groups, and include compounds exemplified by dithiothreitol (DTT), mercaptoethanol, and the like. Preferred phosphorus containing stabilizers have phosphorus atoms with oxidizable pairs of unshared electrons, such as phosphines, or phosphites, which include compounds such as tris(m-sulfonatophenyl)phosphine trisodium salt (TPPTS), or 1,3,5-triaza-7-phosphaadamantane (PTA).

[0074] Most of the nicotinamide cofactor dependent enzymes of the invention have cysteine amino acid residues. Nevertheless, preferred nicotinamide cofactor dependent enzymes of the invention do not comprise a cysteine amino acid residue, because such enzymes may not require the presence of a stabilizer. Two cysteine-free enzymes which are preferred in the practice of the present invention are 2,5-diketo-D-gluconic acid reductase and glucose-6-phosphate and 1-dehydrogenase.

[0075] It is also to be understood that enzymes are often highly selective for the production of only one of two possible enantiomers of a reduced organic product. Therefore, in preferred embodiments, the invention provides processes wherein the reduced organic product is produced in a substantial enantiomeric excess.

[0076] The catalysts of the invention also comprise at least one metal salt or complex. The metal salt or complex may comprise any compound, composition, or phase containing at least one transition metal element, lanthanide metal element, or actinide metal element from the Periodic Table of the elements, with the proviso that the metal salt or complex is not nicotinamide cofactor dependent enzyme that contains a transition metal element, a lanthanide metal element, or an actinide metal element as part of its structure. Preferably, the metal salt or complex comprises a transition metal element selected from Groups 8, 9, 10, or 11 of the Periodic Table. Preferred elements from Groups 8, 9, 10, or 11 comprise, iron, ruthenium, osmium, cobalt, rhodium, iridium, nickel, palladium, platinum, or copper. In highly preferred embodiments of the invention, the metal salt or complex comprises ruthenium, rhodium, or palladium. The most preferred metal element is ruthenium.

[0077] The preferred oxidation state of the metal salt will vary with the identity of the metal, process variables such as the medium, the pressure of H₂, and the identity of any ligands bonded to the metal. It is to be understood that the oxidation state of the metal added to the mixture may be altered by the presence of the other components of the reaction. Examples of species of metal salts or complexes include but are not limited to iron trichloride, ferrocene, iron pentacarbonyl, ruthenium trichloride hydrate, [Cl₂Ru(TPPTS)₂]₂, osmium tetraoxide, dicobalt octacarbonyl, rhodium trichloride, [ClRh(P(C₆H₅)₃)₃], [ClRh(TPPTS)₃], nickel tetracarbonyl, nickel acetate, [Cl₂Pd(P(C₆H₅)₃)₂], Na₂[PtCl₆], CuBr₂, and the like.

[0078] In many embodiments of the invention, the metal salt or complex comprises one or more ligands. Ligands are any organic or inorganic compound that can coordinately, datively, or covalently bond to the metal atom of the metal salt or complex. Examples of suitable ligands include but are not limited to water, a hydroxide, an oxide, an amine, an amide, an imine, an oxime, an imide, a nitrogen containing heterocycle, a nitrogen containing macrocycle, a nitrile, a phosphine, a phosphide, a phosphite, an alcohol, a thiol, an alcoholate, a thiolate, a sulfur containing heterocycle, an oxygen containing heterocycle, an ether, a cyclic ether, a thioether; a phenol, a thiophenol, a phenolate, a thiophenolate; a halide, a hydride, a borohydride, a ketone, an aldehyde, a carboxylic acid, an ester, an amino acid, a carboxylate, an acetate, an iminate, an acetylacetonate, an iminoacetate, an iminoiminate, an alkene, an alkyne, a diene, an allyl residue, a dienyl residue a cyclopentadienyl residue, an indenyl residue, an arene, a polycyclic aromatic residue, a hydrocarbyl residue, carbon monoxide, a cyanide, nitric oxide, H₂, substituted silyl residues, a sulfate, a sulfoxide, a sulfone, a sulfonate, a phosphate, a phosphonate, or any ligand containing more than one of the above functional groups or residues.

[0079] In preferred embodiments of the invention, the metal salt or complex has a ligand comprising a phosphine residue, a phosphite residue, carbon monoxide, a cyclopentadienyl residue, an aromatic residue, a halide, or a hydride. A phosphine residue comprises a trivalent phosphorus residue having the formula PR₁R₂R₃. A phosphite residue comprises a trivalent phosphorus residue having at least one alkoxy residue in substitution for the R₁, R₂, or R₃ residues of a phosphine. Preferably, the R₁, R₂, and R₃ residues of the phosphines and/or phosphites are independently selected from hydrogen, alkyl, alkylene, aryl, or halide residues.

[0080] In other preferred embodiments of the invention, the metal salt or complex has a ligand comprising a phosphine residue, a phosphite residue, a cyclopentadienyl residue, or an aromatic residue, wherein the ligand has one or more polar functional groups. Preferred polar functional groups include one or more hydroxyl, carboxylic, amine, amide, ketone,

aldehyde, nitro, and other similar polar substituent groups for organic compounds. Preferred polar functional groups also include anionic groups, cationic groups, or poly(alkylene glycol) groups. Preferred anionic polar functional groups include carboxylates, sulfates, sulphonates, phosphates, phosphonates, and the like. Preferred cationic polar functional groups include ammonium groups, sulfonium groups, phosphonium groups, and the like. Preferred poly(alkylene glycol) functional groups include poly(ethylene glycol) groups, polypropylene glycol groups, polybutylene glycol groups, and the like.

[0081] Although the polar functional groups may serve various purposes, such as modification of the properties and reactivity of the metal complex, in many preferred embodiments, the polar functional group serves the purpose of increasing the water solubility of the ligand and/or the resulting metal salt or complex. A particularly preferred class of ligands are water soluble phosphine or phosphite ligands, which are useful as homogenous catalysts, as described by Kalck and Monteil (*Adv. Organomet. Chem.*, 1992, 34, 219-284). A preferred class of water soluble phosphine ligands comprises phosphine compounds having one or more anionic sulfonate groups. A well known example of such compounds comprises a salt of tris(m-sulfonatophenyl)phosphine. Another known water soluble phosphine ligand is 1,3,5-triaza-7-phosphaadamantane, as described by Daigle et al. (*Inorg. Synth.*, 1998, 32, 40-45).

[0082] Preferred metal salts or complexes are significantly soluble in water. Significant water solubility permits the metal salt or complex to react with and/or activate H₂ in the water phase, and rapid transfer of hydrogen to reduce the water soluble nicotinamide cofactors. Metal salts or complexes are significantly soluble in water if the metal of the metal salt or complex is solubilized to the extent of at least about 1 part per million in water. Preferably, the metal is soluble to the extent of greater than about 10 parts per million in water. Even more preferably, the metal is soluble to the extent of greater than about 100 parts per million in water.

[0083] H₂, i.e. dihydrogen, is the reducing agent supplied to the processes of the invention. H₂ has very significant practical and environmental advantages as compared with other chemical reducing agents, as discussed hereinabove. H₂ is also very economically attractive from a cost perspective. The table below illustrates the relative costs of a series of relevant reducing agents.

Table 3 - Cost of One Mole of Various "Hydride Reagents"

Reducing Agent	mol wt.	\$/mole
H ₂	2	0.01 ^a
glucose	180	0.24 ^b
Na[CO ₂ H]	68	3 ^b
NaBH ₄	38	32 ^b
NADH	709	26,781 ^c
NADPH	833	358,190 ^c

^aChemical Market Reporter ^bAldrich Catalog ^cSigma Catalog

[0084] The H₂ may be present at any pressure which is effective to produce at least some of the reduced organic product. The pressure of H₂ that will produce at least some reduced organic product will vary with the compositions of the catalyst, the reaction temperature, and other reaction conditions, and variables. Preferably the H₂ is present at a pressure less than about 100 atmospheres. More preferably, the H₂ is present at a pressure from about 0.1 atmospheres to about 50 atmospheres. Even more preferably, the H₂ is present at a pressure from about 1 atmosphere to about 20 atmospheres.

[0085] The processes of the invention can occur at any temperature that induces the reduction of the unsaturated organic compounds, while not substantially denaturing the activity of the nicotinamide enzyme, or substantially deactivating the nicotinamide cofactors. Preferably, the processes of the invention are conducted at a temperature from about 0°C to about 90 °C. More preferably, the processes of the invention are conducted at a temperature from about 0°C to about 45 °C.

[0086] In a preferred embodiment, the invention provides a process for reducing an unsaturated organic compound, comprising mixing the unsaturated organic compound and H₂ in the presence of a catalyst to form a reduced organic product, wherein the catalyst comprises:

- a) a substantially aqueous buffer solution having a pH from about 6.5 to about 9.0,
- b) a water-soluble metal salt or complex comprising iron, ruthenium, osmium, cobalt, rhodium, iridium, nickel, palladium, or copper,
- c) a nicotinamide cofactor comprising NAD⁺, NADH, NADP⁺, NADPH, or a mixture thereof; and
- d) one nicotinamide cofactor dependent enzyme classified under the EC system as an 1.x.1.y. class enzyme, wherein

x is 1,3,4, 5 or 10.

[0087] In another preferred embodiment, the invention provides a process comprising adding dihydrogen, H₂, to an unsaturated organic substrate using as a catalyst a mixture comprising an enzyme, a nicotinamide cofactor, a metal salt or complex and optionally ligands, wherein the metal of the metal salt or complex is selected from iron, cobalt, nickel, copper, ruthenium, palladium, osmium, iridium. In even more preferred embodiments, the metal of the metal salt or complex is selected from ruthenium and palladium.

[0088] It is clear that in the absence of a catalyst, the unsaturated organic compounds of the reaction typically do not react with H₂, and are not reduced by H₂ to form the reduced organic products of the invention. Therefore, the invention provides a preferred process for reducing an unsaturated organic compound, comprising:

- a) contacting H₂ and a catalyst, and
- b) contacting an unsaturated organic compound with the catalyst to form a reduced organic product, wherein the catalyst comprises:

- i) at least one metal salt or complex,
- ii) at least one nicotinamide cofactor; and
- iii) a nicotinamide cofactor dependent enzyme, and wherein
- iv) when the metal salt or complex is a platinum carbonyl cluster complex, the catalyst does not comprise a redox active dye; and
- v) when the metal salt or complex is a rhodium phosphine complex, the nicotinamide cofactor dependent enzyme is not a mixture of horse liver alcohol dehydrogenase and lactate dehydrogenase.

[0089] It is to be understood that in the embodiments of the invention described immediately above, H₂, the catalyst, and the unsaturated organic compound may all be present in a mixture or single phase, but it is not necessary that the unsaturated organic product is directly contacted with H₂. In some embodiments, the catalyst (or certain components thereof) might be contacted with H₂ in one reaction phase or reactor, while the unsaturated organic product was contacted with catalyst in a different phase or reactor. The steps of contacting H₂ and a catalyst, and contacting an unsaturated organic compound with the catalyst may occur simultaneously, or sequentially.

[0090] Furthermore, in an alternative but similar embodiment, the invention provides a process for reducing an unsaturated organic compound, comprising:

- a) contacting H₂, at least one metal salt or complex, and at least one nicotinamide cofactor to form at least some reduced nicotinamide cofactor, and
- b) contacting the reduced nicotinamide cofactor, a nicotinamide cofactor dependent enzyme, and an unsaturated organic compound under conditions effective to form at least some of the reduced organic product, wherein

- i) when the metal salt or complex is a platinum carbonyl cluster complex, the catalyst does not comprise a redox active dye; and
- ii) when the metal salt or complex is a rhodium phosphine complex, the nicotinamide cofactor dependent enzyme is not a mixture of horse liver alcohol dehydrogenase and lactate dehydrogenase.

[0091] In the embodiment described immediately above, the contacting steps may occur simultaneously, or sequentially; the reaction steps may also occur in the same phase or reactor. Alternatively, the metal salt or complex, and the nicotinamide cofactor might be contacted with H₂ in one reaction phase or reactor, while the unsaturated organic product was contacted with enzyme and the reduced organic cofactor in a different phase or reactor.

[0092] In preferred embodiments of step a) of the process described immediately above, the H₂ reacts with the metal salt or complex to form at least some of a metal hydrogen complex, and the metal hydrogen complex reacts directly with the nicotinamide cofactor to transfer hydrogen from the metal hydrogen complex to the nicotinamide cofactor.

[0093] In yet another aspect, the current invention provides a composition for reducing unsaturated organic compounds comprising H₂ and a catalyst, the catalyst comprising:

- a) at least one metal salt or complex,
- b) at least one nicotinamide cofactor; and
- c) a nicotinamide cofactor dependent enzyme, wherein

- i) when the metal salt or complex is a platinum carbonyl cluster complex, the catalyst does not comprise a redox active dye; and
 ii) when the metal salt or complex is a rhodium phosphine complex, the nicotinamide cofactor dependent enzyme is not a mixture of horse liver alcohol dehydrogenase and lactate dehydrogenase.

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 [0094] As previously described hereinabove, many embodiments of the above-described processes for reducing unsaturated organic compounds with H₂ employ oxidoreductase enzymes whose activity is enhanced or maintained by the presence of certain stabilizers. It is known in the art to employ certain sulfur containing stabilizers, and in particular it is known in the art to employ stabilizers having sulfhydryl groups, which are exemplified by compounds such as dithiothreitol (DTT), mercaptoethanol, and the like.

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 [0095] Unexpectedly, it has been discovered that certain phosphorus containing compounds can also stabilize oxidoreductase enzymes. The enzyme stabilization occurs independently of the presence or absence of the H₂, or unsaturated organic compounds employed in the processes described above. In particular, it has been unexpectedly discovered that phosphines or phosphites can stabilize oxidoreductase enzymes in a variety of processes.

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 [0096] Therefore, in one aspect the invention provides a process for stabilizing the activity of an oxidoreductase enzyme, comprising mixing a phosphine or phosphite with an oxidoreductase enzyme. In preferred embodiments of the invention, the mixing of the phosphine or phosphite with the oxidoreductase enzyme occurs in a liquid medium. The liquid medium may comprise water, an organic solvent, or a mixture thereof. Preferably, the liquid medium comprises a substantially aqueous buffer solution.

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 [0097] Preferably, the mixing of the phosphine or phosphite with the oxidoreductase enzyme is effective to slow the loss of activity of the oxidoreductase enzyme, as compared to the rate of loss of activity of the oxidoreductase enzyme in the absence of the phosphine or phosphite. While not wishing to be bound by theory, it is believed that the phosphine or phosphite compounds serve as reducing agents for certain oxidation sensitive components of the enzymes, including cysteine amino acid residues contained in the enzymes.

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 [0098] Preferably, the phosphine or the phosphite comprise phosphorus derivatives PR_n(O-R)_{3-n}, wherein n is an integer of from zero to three. Preferably, the R groups comprise from 1 to about 10 carbon atoms. The R groups (whether bonded to phosphorus or oxygen) are independently selected, and may be the same or different. Preferably, the R groups are hydrocarbyl, or substituted hydrocarbyl, groups, which include alkyl, cycloalkyl, aromatic and heteroaromatic groups.

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 [0099] Preferably, one or more of the R groups is substituted with one or more polar groups. Preferred polar groups include salts of anionic groups such as sulfonate, and carboxylate groups; salts of cationic groups such as alkylammonium groups, or polar but electrically neutral groups such as hydroxyl, amino, alcohol, or polyalkylene glycol groups. Polyethylene glycol groups are preferred polyalkylene glycol groups. The polar groups are believed to be beneficial in improving the effectiveness of the phosphine or phosphite stabilizers because the polar groups tend to increase the water solubility of the phosphines or phosphites, so that they can more effectively interact with the oxidoreductase enzymes, which are also water soluble.

Experimental

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 [0100] The following examples are put forth so as to provide those of ordinary skill in the art with a complete disclosure and description of how the compounds, and associated processes and methods are constructed, used, and evaluated, and are intended to be purely exemplary of the invention and are not intended to limit the scope of what the inventors regard as their invention. Efforts have been made to ensure accuracy with respect to numbers (e.g., amounts, temperature, etc.) but some errors and deviations should be accounted for. Unless indicated otherwise, parts are parts by weight, temperature is in °C (Celsius) or is at ambient temperature, and pressure is at or near atmospheric.

Materials and Methods

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 [0101] Tris(m-sulfonatophenyl)phosphine trisodium salt (TPPTS) was purchased from Strem Chemical and sodium diphenylphosphinobenzene-3-sulfonate (TPPMS) from TCI America. Alcohol dehydrogenase enzymes of *Thermanaerobium Brockii* (TBADH, EC 1.1.1.2), β-nicotinamide adenine dinucleotide (NAD⁺), β-nicotinamide adenine dinucleotide phosphate (NADP⁺) and their respective reduced forms were purchased from Sigma Chemical and stored in a refrigerator (-10°C). The preparation of TBADH is described in U.S. Patent No. 4,352,885. RuCl₃-hydrate (38.4 % Ru) was obtained from Colonial Metals, Inc. 1,3,5-Triaza-7-phosphaadamantane (PTA) was prepared by the procedure of Daigle, et al. (Inorg. Synth. 1998, 32, 40-45). [Cl₂Ru(TPPTS)₂]₂ was prepared by the procedure of Hernandez, and Kalck (J. Mol. Catal. A: Chem., 1993, 116, 117-130). Cl₂Ru(PTA)₄ was prepared by the procedure of Darensbourg, et al., (Inorg. Chem., 1994, 33, 200-08). Phosphate buffer was prepared using KHPO₄. Procedures for 2-heptanone reduction using isopropanol to regenerate NADP were adapted from Keinan, et al. (J. Am. Chem. Soc. 1986, 108, 162). ¹H and ¹³C NMR

EP 1 621 613 B1

spectra were recorded on a Varian Gemini-300 spectrometer. Reference samples of NAD⁺, NADH, NADP⁺ and NADPH were dissolved in D₂O and their chemical shifts (δ) are reported relative to sodium 3-(trimethylsilyl)propionate (TMSP). Chemical shift assignments have been made in accord with Ragg, et al. (J. Biochim. Biophys. Acta 1991, 1076, 49) and Oppenheimer (Proc. Natl. Acad. Sci. U. S. 1971, 68, 3200). GLC was conducted on a HP 6890 gas chromatograph with a 30' DB-FFAP capillary column and flame ionization detection. Samples of (R)-(-)-2-heptanol (95%) and (S)-(+)-2-heptanol (99%, 97% ee) were obtained from Aldrich Chemical. Samples were analyzed by conversion to their trifluoroacetate esters by a standardized treatment with trifluoroacetic anhydride. The enantiomeric excess of (S)-2-heptanol produced by enzymatic reduction was determined by chiral GC and comparison to (R)- and (S)-2-heptanol standards using a Cyclodex-B column (30 m x 0.25, 0.25 μ film) at 40°C. Under these conditions the retention times of the trifluoroacetates of (R)-2-heptanol and (S)-2-heptanol were 28.15 minutes and 28.77 minutes.

Example 1

[0102] Example 1 demonstrates that H₂ can be used to convert 2-butanone to 2-butanol in the presence of a catalyst comprising a ruthenium complex, TBADH and NADP⁺.

[0103] NADP⁺ (28.0 mg, 36.6 μ mol) was placed in 2 mL of a 0.1 M phosphate buffer and the pH adjusted to 8.1 with NaOH. TBADH (1.2 mg, 8.8 units) was added to this solution and it was placed in an 80 mL Fisher-Porter bottle. The Fisher-Porter bottle was evacuated and refilled with argon three times then [Cl₂Ru(TPPTS)₂]₂ (6.4 mg, 5.0 μ mol) and 2-butanone (5 μ l) were added under a flow of argon. The Fisher-Porter bottle was evacuated and filled with 70 psi of H₂ and heated to 40°C for 2.0 hr. The pressure was released from the bottle and a liquid sample analyzed by GLC showed the production of 9.6 μ mol of 2-butanol.

Comparative Example 1a

[0104] Comparative Example 1a demonstrates that without the enzyme (TBADH) the conditions applied in Example 1 produce no reaction.

[0105] NADP⁺ (39.0 mg, 51.5 μ mol) was placed in 2 mL of a 0.1 M phosphate buffer. The pH of this solution was adjusted to 8.1 with NaOH and then placed in an 80 mL Fisher-Porter bottle. The Fisher-Porter bottle was evacuated and refilled with argon three times then [Cl₂Ru(TPPTS)₂]₂ (8.8 mg, 6.9 μ mol) and 2-butanone (10 μ l) were added under a flow of argon. The Fisher-Porter bottle was evacuated and filled with 70 psi of H₂ and heated to 40°C for 2.0 hr. The pressure was released from the bottle and a liquid sample analyzed by GLC showed the production of no 2-butanol. The reaction solution was stripped to dryness under reduced pressure and the residue was dissolved in D₂O. ¹H NMR analysis showed that only a trace of NADP⁺ remained and that NADPH was the dominant species remaining.

Example 2

[0106] Example 2 demonstrates that H₂ can be used to convert 2-heptanone to 2-heptanol in significant enantiomeric in the presence of a catalyst comprising a ruthenium complex, TBADH and NADP⁺.

[0107] An 80 mL Fisher-Porter bottle with a micro-liquid sampling tube was charged with 20 mL of a 0.1 M phosphate buffer (pH = 7.0), 2-heptanone (100 μ l, 718 μ mol), TPPTS (74 mg, 130.7 μ mol), NADP⁺ (10 mg, 13.1 μ mol) and TBADH (2.7 mg, 19.8 units) under a flow of argon. Ruthenium catalyst, [Cl₂Ru(TPPTS)₂]₂ (17 mg, 6.5 μ mol), was finally added and the Fisher-Porter bottle was sealed, evacuated and filled with 70 psi of H₂. Evacuation and refilling with H₂ were repeated two times and the apparatus was then heated to 60°C with an oil bath. After 10 hours GC analysis of a microsample removed via the liquid sampler showed the production of 227.8 μ mol of 2-heptanone. After an additional four hours an additional 13.0 μ mol had accumulated (240.8 μ mol). The pressure was released from the bottle and a liquid sample analyzed by chiral GC showed the 2-heptanol to be 70.5 % (S) and 29.5% (R) (41.0 % ee). Turnovers of catalyst components: NADP⁺_{TO} = 18.4

Example 3

[0108] Example 3 demonstrates that the ruthenium catalyst [Cl₂Ru(TPPTS)₂]₂ is a catalyst precursor for the reduction of NAD(P)⁺ with H₂.

[0109] NADP⁺ (36.0 mg, 47.0 μ mol) was placed in 2 mL of a 0.1 M phosphate buffer. The pH of this solution was adjusted to 8.3 with NaOH and then placed in an 80 mL Fisher-Porter bottle. The Fisher-Porter bottle was evacuated and refilled with argon three times then [Cl₂Ru(TPPTS)₂]₂ (9.7 mg, 3.7 μ mol) was added under a flow of argon. The Fisher-Porter bottle was evacuated and filled with 70 psi of H₂ and heated to 40°C for 3.0 hr. The volatiles were removed under reduced pressure and a sample dissolved in D₂O. To this was added 20 μ l of trimethylsilylpropionate standard solution. ¹H NMR integration shows that 29.0 μ mole of NADPH have been produced under these conditions. NADP⁺

EP 1 621 613 B1

^1H (D_2O): δ 9.35 (s) N_2 , 9.17 (d, 0.6 Hz) N_6 , 8.84 (d, 1.0) N_4 , 8.42 (s) A_2 , 8.21 (t, 0.7) N_5 , 8.11 (s) A_8 , 6.10 (d, 5.2 Hz) N_1 , 6.02 (d,) A_1 , 4.77 (t,) A_2 , 4.77 (t,) A_2 , 4.56 (br s), 4.51 (br s), 4.45 (br s), 4.40 (br s), 4.26 (br s).

NADPH

^1H (D_2O): δ 8.48 (s) A_8 , 8.21 (s) A_2 , 6.94 (s) N_6 , 6.13 (d,) A_1 , 5.98 (d, 0.7) N_1 , 8.11 (s) A_8 , 6.10 (d, 5.2 Hz) N_1 , 6.02 (d,) A_1 , 4.78 (t,) A_2 , 4.71 (t,) A_2 , 4.51 (t,) , 4.39 (br s), 4.22 (br m), 4.09 (br s), 2.71 (dd,) P_4

Example 4

[0110] Example 4 demonstrates that the ruthenium catalyst $[\text{Cl}_2\text{Ru}(\text{TPPTS})_2]_2$ is a catalyst precursor for the reduction of NAD^+ with H_2 .

[0111] NAD^+ (34.0 mg, 38.0 μmol) was placed in 2 mL of a 0.1 M phosphate buffer. The pH of this solution was adjusted to 8.1 with NaOH and then placed in an 80 mL Fisher-Porter bottle. The Fisher-Porter bottle was evacuated and refilled with argon three times then $[\text{Cl}_2\text{Ru}(\text{TPPTS})_2]_2$ (6.9 mg, 2.6 μmol) was added under a flow of argon. The Fisher-Porter bottle was evacuated and filled with 70 psi of H_2 and stirred to 23 $^\circ\text{C}$ for 17.0 hr. The volatiles were removed under reduced pressure and a sample dissolved in D_2O . To this was added 20 μl of trimethylsilylpropionate standard solution. ^1H NMR integration shows that 9.2 μmole of NADH have been produced under these conditions.

NAD⁺

$^1\text{H}(\text{D}_2\text{O})$: δ 9.35 (s) N_2 , 9.17 (d, 0.6 Hz) N_6 , 8.84 (d, 1.0) N_4 , 8.42 (s) A_2 , 8.21 (t, 0.7) N_5 , 8.11 (s) A_8 , 6.10 (d, 5.2 Hz) N_1 , 6.02 (d,) A_1 , 4.77 (t,) A_2 , 4.56 (br s), 4.51 (br s), 4.45 (br s), 4.40 (br s), 4.26 (br s).

NADH

$^1\text{H}(\text{D}_2\text{O})$: δ 8.48 (s) A_8 , 8.21 (s) A_2 , 6.94 (s) N_6 , 6.13 (d,) A_1 , 5.98 (d, 0.7) N_1 , 8.11 (s) A_8 , 6.10 (d, 5.2 Hz) N_1 , 6.02 (d,) A_1 , 4.78 (t,) A_2 , 4.71 (t,) A_2 , 4.51 (t,) , 4.39 (br s), 4.22 (br m), 4.09 (br s), 2.71 (dd,) P_4 .

Example 5

[0112] Example 5 demonstrates that iodide is not a poison for the catalyst derived from ruthenium complex $[\text{Cl}_2\text{Ru}(\text{TPPTS})_2]_2$.

[0113] NAD^+ (34.0 mg, 38.0 μmol) was placed in 2 mL of a 0.1 M phosphate buffer containing 0.07 M sodium iodide. The pH of this solution was adjusted to 8.1 with NaOH and then placed in an 80 mL Fisher-Porter bottle. The Fisher-Porter bottle was evacuated and refilled with argon three times then $[\text{Cl}_2\text{Ru}(\text{TPPTS})_2]_2$ (7.7 mg, 2.9 μmol) was added under a flow of argon. The Fisher-Porter bottle was evacuated and filled with 70 psi of H_2 and stirred to 23 $^\circ\text{C}$ for 13.0 hr. The volatiles were removed under reduced pressure and a sample dissolved in D_2O . To this was added 20 μl of trimethylsilylpropionate standard solution. ^1H NMR integration shows that 7.4 μmole of NADH have been produced under these conditions.

Example 6

[0114] Example 6 demonstrates that other water-soluble ruthenium complexes are catalysts for cofactor regeneration with H_2 .

[0115] Using a procedure identical to that of Example 4 above $\text{Cl}_2\text{Ru}(\text{PTA})_4$ (4.8 mg, 6.0 μmol) and NAD^+ (33.0 mg, 50.0 μmol) were reacted at 70 psi and 40 $^\circ\text{C}$ for 15.5 hr. ^1H NMR assay shows that 11.0 μmole of NADH have been produced under these conditions.

Example 7

[0116] Example 7 demonstrates that the phosphine ligands used in the above examples in ruthenium complexes are necessary to generate catalysts for cofactor regeneration.

[0117] Using a procedure identical to that of Example 4 above RuCl_3 -hydrate (1.6 mg, 38.4 % Ru, 6.0 μmol) and NAD^+ (33.0 mg, 50.0 μmol) were reacted at 70 psi and 40 $^\circ\text{C}$ for 2.5 hr. ^1H NMR assay shows that no NADH has been produced under these conditions.

Example 8

[0118] Example 8 demonstrates that the ruthenium complexes used in the above examples are necessary for the regeneration of cofactor with hydrogen.

[0119] Following the procedure of Example 4, but excluding the ruthenium catalyst, NAD^+ (33.0 mg, 50.0 μmol) was placed 2 mL of a 0.1 M (aq) KHPO_4 with 0.07 M sodium iodide in an 80 mL Fisher-Porter bottle and the pH adjusted to 8.15 with NaOH. After establishing a H_2 -atmosphere (70 psig) and heating to 40 $^\circ\text{C}$ for 2.5 hr ^1H NMR analysis showed

no production of NADH.

Example 9

5 [0120] Example 9 demonstrates that the hydrogen is necessary for the regeneration of cofactors described in the above examples.

[0121] Following the procedure of Example 4, $[\text{Cl}_2\text{Ru}(\text{TPPTS})_2]_2$ (7.9 mg, 6.0 μmol) and NAD^+ (33.0 mg, 50.0 μmol) was placed 2 mL of 0.1 M (aq) KHPO_4 with 0.07 M sodium iodide and an atmosphere of argon rather than H_2 , was placed over the reaction solution. After heating to 40°C for 2.5 hr. ^1H NMR analysis showed no NADH. The argon atmosphere was then replaced by H_2 and NADH was produced without significant catalyst deactivation.

Example 10

[0122] Example 10 demonstrates that the TPPTS can be used as a stabilizer for TBADH.

15 [0123] Phosphate buffer (10 mL, 0.1 M, pH = 7.0) was placed in a 100 mL Schlenk tube under argon. 2-Heptanone (4.92 g, 43.1 mmol), isopropanol (1.57 g, 26.1 mmol), heptadecane (41.0 μl), TPPTS (23 mg, 40.0 μmol), NADP^+ (0.4 mg, 0.5 μmol) and TBADH (2.7 mg, 19.8 units) were then added under a flow of argon. After evacuation and refilling with argon three times the apparatus was heated to 38°C with an oil bath. GC analysis every five hours showed a continuous production of 2-heptanol for the initial 20 hours at a rate corresponding to - 210 turnovers of NADP^+ per hour (total of 2.17 mmoles of 2-heptanol in 21 hours). After and additional 24 hours the yield of 2-heptanol had not changed. The final yield of 2-heptanol corresponds to 4,343 turnovers of NADP^+ .

Comparative Example 10a

25 [0124] Comparative Example 10a shows TBADH productivity without a stabilizer.

[0125] Identical amounts of all reagents in a 2-heptanone reduction with isopropanol (Example 10) except for the water-soluble phosphine (TPPTS) were heated to 38°C with an oil bath for 23 hours. The yield of 2-heptanol was 0.13 mmol, corresponding to a total of 271 turnovers of NADP^+ .

Comparative Example 10b

[0126] Comparative Example 10b shows TBADH productivity with dithiothreitol (DTT) as a stabilizer.

35 [0127] Using the procedure described in Comparative Example 10a, identical amounts of all reagents except twice the amount of 2-heptanone (9.84, 86.2 mmol) and substituting DTT as a stabilizer (6 mg, 40 μmol) in place of TPPTS, the reduction of 2-heptanone with isopropanol was carried out at 38°C for 19 hours yielding 2.51 mmol of 2-heptanol, corresponding to a total of 5,021 turnovers of NADP^+ .

Comparative Example 10c

40 [0128] Comparative Example 10c shows TBADH productivity with mercaptoethanol as a stabilizer.

[0129] Using the procedure described in Example 10 and identical amounts of all reagents but substituting mercaptoethanol as a stabilizer (2.8 μl , 40 μmol) in place of DTT, the reduction of 2-heptanone with isopropanol was carried out at 38°C for 22 hours yielding 2.56 mmol of 2-heptanol, corresponding to a total of 5,121 turnovers of NADP^+ .

Example 11

[0130] Example 11 demonstrates that triphenylphosphine (TPP) can be used as a stabilizer for TBADH.

50 [0131] Using the procedure described in Example 10 and identical amounts of all reagents but substituting triphenylphosphine as a stabilizer (10 mg, 40 μmol) in place of TPPTS, the reduction of 2-heptanone with isopropanol was carried out at 37°C for 19 hours yielding 2.15 mmol of 2-heptanol, corresponding to a total of 4,301 turnovers of NADP^+ .

Example 12.

[0132] Example 12 demonstrates that "monosulfonated triphenylphosphine" (TPPMS) can be used as a stabilizer for TBADH.

55 [0133] Using the procedure describe in Example 10 and identical amounts of all reagents but substituting triphenylphosphine as a stabilizer (15 mg, 40 μmol) in place of TPPTS, the reduction of 2-heptanone with isopropanol was carried out at 38°C for 16 hours yielding 1.93 mmol of 2-heptanol, corresponding to a total of 3,859 turnovers of NADP^+ .

[0134] It will be apparent to those skilled in the art that various modifications and variations can be made in the present invention.

[0135] Other embodiments of the invention will be apparent to those skilled in the art from consideration of the specification and practice of the invention disclosed herein. It is intended that the specification and examples be considered as exemplary only, with a true scope of the invention being indicated by the following claims.

Claims

1. A process for reducing an unsaturated organic compound, comprising mixing the unsaturated organic compound and H₂ in the presence of a catalyst to form a reduced organic product, wherein the catalyst comprises:

- a) at least one metal salt or complex,
- b) at least one nicotinamide cofactor ; and
- c) only one nicotinamide cofactor dependent enzyme classified under the EC system as an 1.x.1.y. class enzyme, wherein x is 1, 3, 4, 5, or 10,

wherein:

when the metal salt or complex is a platinum carbonyl cluster complex, the catalyst does not comprise a redox active dye.

2. The process of Claim 1, wherein the metal salt or complex comprises a metal of Groups 8, 9, 10, or 11 of the Periodic Table.

3. The process of Claim 1, wherein the metal salt or complex comprises iron, ruthenium, osmium, cobalt, rhodium, iridium, nickel, palladium, platinum, or copper.

4. The process of Claim 1, wherein the metal salt or complex comprises ruthenium, rhodium, or palladium.

5. The process of Claim 1, wherein the metal salt or complex comprises ruthenium.

6. The process of Claim 5, wherein the catalyst is dispersed in a liquid medium, wherein the liquid medium comprises an aqueous buffer solution having a pH from about 6.5 to about 9.0.

7. The process of Claim 1, wherein the metal salt or complex comprises one or more ligands.

8. The process of Claim 7, wherein the ligand comprises a phosphine residue, a phosphite residue, carbon monoxide, a cyclopentadienyl residue, an aromatic residue, a halide, or a hydride.

9. The process of Claim 7, wherein the ligand comprises a phosphine residue, a phosphite residue, a cyclopentadienyl residue, or an aromatic residue, wherein the ligand has a polar functional group.

10. The process of Claim 9, wherein the polar functional group is an anionic group, a cationic group, or a poly(alkylene glycol) group.

11. The process of Claim 7, wherein the ligand comprises 1,3,5-triaza-7-phosphaadamantane or a salt of tris(m-sulfonatophenyl)phosphine.

12. The process of Claim 1, wherein the nicotinamide cofactor comprises NAD⁺, NADH, NADP⁺, NADPH, or a mixture thereof.

13. The process of Claim 1, wherein the unsaturated organic compound comprises

- a) a ketone, an aldehyde, a carboxylic acid, a carboxylic acid ester, or an amide,
- b) an α-β unsaturated derivative of a ketone, an aldehyde, a carboxylic acid, a carboxylic acid ester, or an amide,
- c) a fatty acid, a monoglyceride, a diglyceride, or a triglyceride, having an olefinic unsaturated group,
- d) an olefin, an aromatic compound or a heteroaromatic compound,

e) an imine, or an oxime, or

f) a sugar, an amino acid, a peptide, or a protein, wherein the sugar, amino acid, peptide, or protein has an unsaturated group.

- 5 14. The process of Claim 1, wherein the unsaturated organic compound is a ketone, or an aldehyde.
15. The process of Claim 1, wherein the nicotinamide cofactor dependent enzyme is an enzyme classified under the EC system as a 1.1.1.1, a 1.1.1.2, a 1.1.1.4, a 1.1.1.5, a 1.1.1.6, a 1.1.1.7, a1.1.1.14, a1.1.1.18, a1.1.1.21; a1.1.1.22, a1.1.1.26, a1.1.1.27, a1.1.1.30, a 1.1.1.37, a 1.1.1.40, a 1.1.1.41, a 1.1.1.42, a 1.1.1.47, a 1.1.1.48, a 1.1.1.49, a 1.1.1.50, a 1.1.1.51, a 1.1.1.53, a 1.1.1.55, a 1.1.1.67, a 1.1.1.72, a 1.1.1.83, a 1.1.1.95, a 1.1.1.119, a 1.1.1.122, a1.1.1.159, a1.1.1.176, a1.3.1.14, a1.4.1.1, a1.4.1.2, a1.4.1.3, a1.4.1.4, a 1.4.1.5, a1.4.1.6, a1.4.1.7, a1.4.1.8, a1.4.1.9, a1.4.1.10, a1.4.1.11, a1.4.1.12, a1.4.1.13, a 1.4.1.14, a 1.4.1.15, a 1.4.1.16, a 1.4.1.17, a 1.4.1.18, a 1.4.1.19, or a 1.4.1.20 class enzyme.
- 15 16. The process of Claim 1, wherein the nicotinamide cofactor dependent enzyme is an enzyme classified under the EC system as a 1.1.1.1, 1.1.1.2, 1.1.1.5, 1.1.1.6, 1.1.1.7, 1.1.1.14, 1.1.1.18, 1.1.1.21, 1.1.1.26, 1.1.1.27, 1.1.1.37, 1.1.1.40, 1.1.1.41, 1.1.1.42, 1.1.1.47, 1.1.1.48, 1.1.1.49, 1.1.1.72, 1.1.1.83, 1.1.1.95, 1.1.1.119, 14, 1.4.1.1, 1.4.1.3, 1.4.1.4, 1.4.1.9 or 1.4.1.20 class enzyme.
- 20 17. The process of Claim 1, wherein the nicotinamide cofactor dependent enzyme is an enzyme classified under the EC system as a 1.1.1.1 or 1.1.1.2, class enzyme.
18. The process of Claim 1, wherein the nicotinamide cofactor dependent enzyme does not comprise a cysteine amino acid residue.
- 25 19. The process of Claim 18, wherein the nicotinamide cofactor dependent enzyme comprises 2,5-diketo-D-gluconic acid reductase or glucose-6-phosphate and 1-dehydrogenase.
20. The process of Claim 1, further comprising a stabilizer for the nicotinamide cofactor dependent enzyme.
- 30 21. The process of Claim 20, wherein the stabilizer comprises a phosphine or a phosphite.
22. The process of Claim 1, wherein the reduced organic product is produced in a substantial enantiomeric excess.
- 35 23. The process of claim 1, wherein the catalyst comprises:
- a) a substantially aqueous buffer solution having a pH from about 6.5 to about 9.0,
- b) a water-soluble metal salt or complex comprising iron, ruthenium, osmium, cobalt, rhodium, iridium, nickel, palladium, or copper,
- 40 c) a nicotinamide cofactor comprising NAD⁺, NADH, NADP⁺, NADPH, or a mixture thereof; and
- d) only one nicotinamide cofactor dependent enzyme classified under the EC system as an 1.x.1.y. class enzyme, wherein x is 1,3,4, 5 or 10.

45 **Patentansprüche**

1. Verfahren zur Reduzierung einer ungesättigten organischen Verbindung, umfassend Mischen der ungesättigten organischen Verbindung mit H₂ in Gegenwart eines Katalysators zur Bildung eines reduzierten organischen Produkts, wobei der Katalysator umfasst:
- 50 a) mindestens ein Metallsalz oder -komplex,
- b) mindestens ein Nikotinamid-Cofaktor; und
- c) nur ein Nikotinamid-Cofaktor-abhängiges Enzym, welches unter dem EC-System als Enzym der 1.x.1.y.-Klasse klassifiziert wird, wobei x 1, 3, 4, 5 oder 10 ist,
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wobei:

wenn das Metallsalz oder -komplex ein Platin-Carbonyl-Clusterkomplex ist, umfasst der Katalysator keinen

EP 1 621 613 B1

redoxaktiven Farbstoff.

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2. Verfahren nach Anspruch 1, wobei das/der Metallsalz oder -komplex ein Metall der Gruppen 8, 9, 10 oder 11 des Periodensystems umfasst.
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3. Verfahren nach Anspruch 1, wobei das/der Metallsalz oder -komplex Eisen, Ruthenium, Osmium, Kobalt, Rhodium, Iridium, Nickel, Palladium, Platin oder Kupfer umfasst.
4. Verfahren nach Anspruch 1, wobei das/der Metallsalz oder -komplex Ruthenium, Rhodium oder Palladium umfasst.
5. Verfahren nach Anspruch 1, wobei das/der Metallsalz oder -komplex Ruthenium umfasst.
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6. Verfahren nach Anspruch 5, wobei der Katalysator in einem flüssigen Medium dispergiert ist, wobei das flüssige Medium eine wässrige Pufferlösung umfasst, die einen pH-Wert von ungefähr 6,5 bis ungefähr 9,0 hat.
7. Verfahren nach Anspruch 1, wobei das/der Metallsalz oder -komplex einen oder mehrere Liganden umfasst.
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8. Verfahren nach Anspruch 7, wobei der Ligand einen Phosphin-Rest, einen Phosphit-Rest, Kohlenmonoxid, einen Cyclopentadienyl-Rest, einen aromatischen Rest, ein Halogenid oder ein Hydrid umfasst.
9. Verfahren nach Anspruch 7, wobei der Ligand einen Phosphin-Rest, einen Phosphit-Rest, einen Cyclopentadienyl-Rest oder einen aromatischen Rest umfasst, wobei der Ligand eine polare funktionelle Gruppe hat.
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10. Verfahren nach Anspruch 9, wobei die polare funktionelle Gruppe eine anionische Gruppe, eine kationische Gruppe oder eine Poly(Alkylenglycol)-Gruppe ist.
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11. Verfahren nach Anspruch 7, wobei der Ligand 1,3,5-Triaza-7-phosphaadamantan oder ein Tris(m-sulfonatophenyl)phosphin-Salz umfasst.
12. Verfahren nach Anspruch 1, wobei der Nikotinamid-Cofaktor NAD⁺, NADH, NADP⁺, NADPH oder eine Mischung davon umfasst.
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13. Verfahren nach Anspruch 1, wobei die ungesättigte organische Verbindung umfasst
- a) ein Keton, einen Aldehyd, eine Carbonsäure, einen Carbonsäureester oder ein Amid,
- b) ein α - β -ungesättigtes Derivat von Keton, Aldehyd, Carbonsäure, Carbonsäureester oder Amid,
- c) eine Fettsäure, ein Monoglycerid, ein Diglycerid oder ein Triglycerid, die eine olefinische ungesättigte Gruppe haben,
- d) ein Olefin, eine aromatische Verbindung oder eine heteroaromatische Verbindung,
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- e) ein Amin oder ein Oxim, oder
- f) ein Zucker, eine Aminosäure, ein Peptid oder ein Protein, wobei der Zucker, Aminosäure, Peptid oder Protein eine ungesättigte Gruppe haben.
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14. Verfahren nach Anspruch 1, wobei die ungesättigte organische Verbindung ein Keton oder ein Aldehyd ist.
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15. Verfahren nach Anspruch 1, wobei das Nikotinamid-Cofaktor-abhängiges Enzym ein Enzym ist, welches unter dem EC-System als ein Enzym einer 1.1.1.1-, einer 1.1.1.2-, einer 1.1.1.4-, einer 1.1.1.5-, einer 1.1.1.6-, einer 1.1.1.7-, einer 1.1.1.14-, einer 1.1.1.18-, einer 1.1.1.21-, einer 1.1.1.22-, einer 1.1.1.26-, einer 1.1.1.27-, einer 1.1.1.30-, einer 1.1.1.37-, einer 1.1.1.40-, einer 1.1.1.41-, einer 1.1.1.42-, einer 1.1.1.47-, einer 1.1.1.48-, einer 1.1.1.49- einer 1.1.1.50- einer 1.1.1.51-, einer 1.1.1.53-, einer 1.1.1.55-, einer 1.1.1.67-, einer 1.1.1.72-, einer 1.1.1.83-, einer 1.1.1.95-, einer 1.1.1.119-, einer 1.1.1.122-, einer 1.1.1.159-, einer 1.1.1.176-, einer 1.3.1.14-, einer 1.4.1.1-, einer 1.4.1.2-, einer 1.4.1.3-, einer 1.4.1.4-, einer 1.4.1.5-, einer 1.4.1.6-, einer 1.4.1.7-, einer 1.4.1.8-, einer 1.4.1.9-, einer 1.4.1.10-, einer 1.4.1.11-, einer 1.4.1.12-, einer 1.4.1.13-, einer 1.4.1.14-, einer 1.4.1.15-, einer 1.4.1.16-, einer 1.4.1.17-, einer 1.4.1.18-, einer 1.4.1.19- oder einer 1.4.1.20-Enzymklasse klassifiziert ist.
- 55
16. Verfahren nach Anspruch 1, wobei das Nikotinamid-Cofaktor-abhängiges Enzym ein Enzym ist, welches unter dem EC-System als ein Enzym einer 1.1.1.1-, 1.1.1.2-, 1.1.1.5-, 1.1.1.6-, 1.1.1.7-, 1.1.1.14-, 1.1.1.18-, 1.1.1.21-, 1.1.1.26-, 1.1.1.27-, 1.1.1.37-, 1.1.1.40-, 1.1.1.41-, 1.1.1.42-, 1.1.1.47-, 1.1.1.48-, 1.1.1.49-, 1.1.1.72-, 1.1.1.83-, 1.1.1.95-,

EP 1 621 613 B1

1.1.1.119-, 14-, 1.4.1.1-, 1.4.1.3-, 1.4.1.4-, 1.4.1.9- oder 1.4.1.20-Enzymklasse klassifiziert ist.

5 17. Verfahren nach Anspruch 1, wobei das Nikotinamid-Cofaktor-abhängiges Enzym ein Enzym ist, welches unter dem EC-System als ein Enzym einer 1.1.1.1- oder 1.1.1.2-Enzymklasse klassifiziert ist.

18. Verfahren nach Anspruch 1, wobei das Nikotinamid-cofactor-abhängiges Enzym keinen Aminosäurerest Cystein umfasst.

10 19. Verfahren nach Anspruch 18, wobei das Nikotinamid-Cofactor-abhängiges Enzym eine 2,5-diketo-D-Gluconsäure-Reduktase oder Glucose-6-phosphat und 1-Dehydrogenase umfasst.

20. Verfahren nach Anspruch 1, umfassend ferner einen Stabilisator für das Nikotinamid-Cofaktor-abhängiges Enzym.

15 21. Verfahren nach Anspruch 20, wobei der Stabilisator ein Phosphin oder ein Phosphit umfasst.

22. Verfahren nach Anspruch 1, wobei das reduzierte organische Produkt in einem substantiellen enantiomeren Überschuss produziert wird.

20 23. Verfahren nach Anspruch 1, wobei der Katalysator umfasst:

- 25 a) eine im Wesentlichen wässrige Pufferlösung, die ein pH von ungefähr 6,5 bis ungefähr 9,0 hat,
b) ein wasserlösliches Metallsalz oder -komplex umfassend Eisen, Ruthenium, Osmium, Kobalt, Rhodium, Iridium, Nickel, Palladium oder Kupfer,
c) einen Nikotinamid-Cofaktor umfassend NAD⁺, NADH, NADP⁺, NADPH oder eine Mischung davon; und
d) nur ein Nikotinamid-Cofaktor-abhängiges Enzym, welches unter dem EC-System als Enzym der 1.x.1.y.-Klasse klassifiziert wird, wobei x 1, 3, 4, 5 oder 10 ist.

Revendications

30 1. Procédé pour réduire un composé organique insaturé, comprenant le mélange du composé organique insaturé et de H₂ en présence d'un catalyseur pour former un produit organique réduit, dans lequel le catalyseur comprend :

- 35 a) au moins un sel ou complexe de métal,
b) au moins un cofacteur nicotinamide ; et
c) une seule enzyme dépendante du cofacteur nicotinamide classée aux termes du système EC comme une enzyme de classe 1.x.1.y., dans laquelle x est 1, 3, 4, 5 ou 10, dans lequel :

40 lorsque sel ou complexe de métal est un complexe d'agrégats platine carbonyle, le catalyseur ne comprend pas de colorant ayant une activité d'oxydoréduction.

45 2. Procédé selon la revendication 1, dans lequel le sel ou complexe de métal comprend un métal des Groupes 8, 9, 10 ou 11 du Tableau Périodique.

3. Procédé selon la revendication 1, dans lequel le sel ou complexe de métal comprend du fer, du ruthénium, de l'osmium, du cobalt, du rhodium, de l'iridium, du nickel, du palladium, du platine ou du cuivre.

50 4. Procédé selon la revendication 1, dans lequel le sel ou complexe de métal comprend du ruthénium, du rhodium ou du palladium.

5. Procédé selon la revendication 1, dans lequel le sel ou complexe de métal comprend du ruthénium.

55 6. Procédé selon la revendication 5, dans lequel le catalyseur est dispersé dans un milieu liquide, dans lequel le milieu liquide comprend une solution tampon aqueuse ayant un pH d'environ 6,5 à environ 9,0.

7. Procédé selon la revendication 1, dans lequel le sel ou complexe de métal comprend un ou plusieurs ligand(s).

EP 1 621 613 B1

8. Procédé selon la revendication 7, dans lequel le ligand comprend un résidu phosphine, un résidu phosphite, du monoxyde de carbone, un résidu cyclopentadiényle, un résidu aromatique, un halogénure ou un hydrure.
- 5 9. Procédé selon la revendication 7, dans lequel le ligand comprend un résidu phosphine, un résidu phosphite, un résidu cyclopentadiényle ou un résidu aromatique, dans lequel le ligand a un groupe fonctionnel polaire.
10. Procédé selon la revendication 9, dans lequel le groupe fonctionnel polaire est un groupe anionique, un groupe cationique ou un groupe poly(alkylène glycol).
- 10 11. Procédé selon la revendication 7, dans lequel le ligand comprend du 1,3,5-triaza-7-phosphaadamantane ou un sel de tris(m-sulfonatophényl)phosphine.
12. Procédé selon la revendication 1, dans lequel le cofacteur nicotinamide comprend NAD⁺, NADH, NADP⁺, NADPH, ou un mélange de ceux-ci.
- 15 13. Procédé selon la revendication 1, dans lequel le composé organique insaturé comprend
- a) une cétone, un aldéhyde, un acide carboxylique, un ester d'acide carboxylique ou un amide,
b) un dérivé α - β -insaturé d'une cétone, d'un aldéhyde, d'un acide carboxylique, d'un ester d'acide carboxylique ou d'un amide,
c) un acide gras, un monoglycéride, un diglycéride ou un triglycéride ayant un groupe insaturé oléfinique,
d) une oléfine, un composé aromatique ou un composé hétéroaromatique,
e) une imine ou un oxyme, ou
f) un sucre, un acide aminé, un peptide ou une protéine, dans lequel le sucre, l'acide aminé, le peptide ou la protéine a un groupe insaturé.
- 20 14. Procédé selon la revendication 1, dans lequel le composé organique insaturé est une cétone ou un aldéhyde.
15. Procédé selon la revendication 1, dans lequel l'enzyme dépendante du cofacteur nicotinamide est une enzyme classée aux termes du système EC comme une enzyme de classe 1.1.1.1, 1.1.1.2, 1.1.1.4, 1.1.1.5, 1.1.1.6, 1.1.1.7, 1.1.1.14, 1.1.1.18, 1.1.1.21, 1.1.1.22, 1.1.1.26, 1.1.1.27, 1.1.1.30, 1.1.1.37, 1.1.1.40, 1.1.1.41, 1.1.1.42, 1.1.1.47, 1.1.1.48, 1.1.1.49, 1.1.1.50, 1.1.1.51, 1.1.1.53, 1.1.1.55, 1.1.1.67, 1.1.1.72, 1.1.1.83, 1.1.1.95, 1.1.1.119, 1.1.1.122, 1.1.1.159, 1.1.1.176, 1.3.1.14, 1.4.1.1, 1.4.1.2, 1.4.1.3, 1.4.1.4, 1.4.1.5, 1.4.1.6, 1.4.1.7, 1.4.1.8, 1.4.1.9, 1.4.1.10, 1.4.1.11, 1.4.1.12, 1.4.1.13, 1.4.1.14, 1.4.1.15, 1.4.1.16, 1.4.1.17, 1.4.1.18, 1.4.1.19 ou 1.4.1.20.
- 30 16. Procédé selon la revendication 1, dans lequel l'enzyme dépendante du cofacteur nicotinamide est une enzyme classée aux termes du système EC comme une enzyme de classe 1.1.1.1, 1.1.1.2, 1.1.1.5, 1.1.1.6, 1.1.1.7, 1.1.1.14, 1.1.1.18, 1.1.1.21, 1.1.1.26, 1.1.1.27, 1.1.1.37, 1.1.1.40, 1.1.1.41, 1.1.1.42, 1.1.1.47, 1.1.1.48, 1.1.1.49, 1.1.1.72, 1.1.1.83, 1.1.1.95, 1.1.1.119, 14, 1.4.1.1, 1.4.1.3, 1.4.1.4, 1.4.1.9 ou 1.4.1.20.
- 35 17. Procédé selon la revendication 1, dans lequel l'enzyme dépendante du cofacteur nicotinamide est une enzyme classée aux termes du système EC comme une enzyme de classe 1.1.1.1 ou 1.1.1.2.
18. Procédé selon la revendication 1, dans lequel l'enzyme dépendante du cofacteur nicotinamide ne comprend pas un résidu acide aminé cystéine.
- 40 19. Procédé selon la revendication 18, dans lequel l'enzyme dépendante du cofacteur nicotinamide comprend de l'acide 2,5-dicéto-D-gluconique réductase ou du glucose-6-phosphate et de la 1-déshydrogénase.
- 50 20. Procédé selon la revendication 1, comprenant en outre un stabilisant pour l'enzyme dépendante du cofacteur nicotinamide.
21. Procédé selon la revendication 20, dans lequel le stabilisant comprend une phosphine ou un phosphite.
- 55 22. Procédé selon la revendication 1, dans lequel le produit organique réduit est produit dans un excès énantiomérique substantiel.
23. Procédé selon la revendication 1, dans lequel le catalyseur comprend :

EP 1 621 613 B1

- a) une solution tampon substantiellement aqueuse ayant un pH d'environ 6,5 à environ 9,0,
b) un sel ou complexe de métal soluble dans l'eau comprenant du fer, du ruthénium, de l'osmium, du cobalt, du rhodium, de l'iridium, du nickel, du palladium ou du cuivre,
c) un cofacteur nicotinamide comprenant NAD^+ , NADH , NADP^+ , NADPH ou un mélange de ceux-ci ; et
d) une seule enzyme dépendante du cofacteur nicotinamide classée aux termes du système EC comme une enzyme de classe 1.x.1.y., dans laquelle x est 1, 3, 4, 5 ou 10.

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REFERENCES CITED IN THE DESCRIPTION

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