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(54) Title: IMMUNITY-INDUCING AGENT AND METHOD FOR DETECTION OF CANCER (54) 発明の名称: 免疫誘導剤及び癌の検出方法

WO 2010/005069 A1 [🖾3] 50 40 細胞障害活性(%) 30 20 AA 10 0 16 17 19 20 12 13 14 15 18 AA CYTOTOXIC ACTIVITY (%) (57) 要約:

(57) Abstract: Disclosed is an immunity-inducing agent comprising a recombinant vector as an active ingredient, wherein the recombinant vector comprises at least one polypeptide having an immunity-inducing activity or a polynucleotide encoding the polypeptide and enables the expression of the polypeptide in vivo, and wherein the polypeptide is selected from the following polypeptides (a) to (c): (a) a polypeptide which comprises at least seven contiguous amino acid residues contained in an amino acid sequence depicted in any odd SEQ ID number selected from SEQ ID NO:3 to SEQ ID NO:95 listed in the Sequence Listing; (b) a polypeptide which has a 90% or more sequence identity with the polypeptide (a) and comprises at least seven amino acid residues; and (c) a polypeptide which contains the polypeptide (a) or (b) as a partial sequence thereof. The immunity-inducing agent can be used for the treatment and/or prevention of cancer. Each of the above-mentioned polypeptides can react with an antibody occurring specifically in the serum collected from a cancer patient. Therefore, cancer in a living body can be detected by measuring the antibody in a sample.

CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, ____ TD, TG).

明細書の別個の部分として表した配列リスト (規則 5.2(a))

添付公開書類:

— 国際調査報告(条約第 21 条(3))

⁽a)配列表の配列番号3~95のうち奇数の配列番号に示されるアミノ酸配列中の連続する7個以上の アミノ酸からなるポリペプチド(b)前記(a)のポリペプチドと90%以上の配列同一性を有し、かつ7個以 上のアミノ酸からなるポリペプチド、及び(c)前記(a)又は(b)のポリペプチドを部分配列として含むポリペ プチドのポリペプチド類から選択されかつ免疫誘導活性を有する少なくとも1つのポリペプチド、又は 該ポリペプチドをコードするポリヌクレオチドを含み生体内で該ポリペプチドを発現可能な組換えベク ター、を有効成分として含有する免疫誘導剤は、癌の治療用及び/又は予防用として使用される。ま た、上記ポリペプチドは、癌患者の血清中に特異的に存在する抗体と反応するので、試料中の該抗体を 測定すれば、生体内の癌を検出することができる。

DESCRIPTION

Immunity-inducing Agent and Method for Detection of Cancer

TECHNICAL FIELD

5 [0001]

The present invention relates to a novel immunity-inducing agent useful as a therapeutic and/or prophylactic agent for cancer. Further, the present invention relates to a novel method for detection of cancer.

BACKGROUND ART

10 [0002]

Cancer is the commonest cause for death among all of the causes for death, and therapies carried out therefor at present are mainly surgical treatment in combination with radiotherapy and chemotherapy. In spite of the developments of new surgical methods and discovery of new anti-cancer agents in recent years,

15 treatment results of cancers are not improved very much at present except for some cancers. In recent years, by virtue of the development in molecular biology and cancer immunology, cancer antigens recognized by cytotoxic T cells reactive with cancers, as well as the genes encoding the cancer antigens, were identified, and expectations for antigen-specific immunotherapies have been raised (Non-patent Literature 1).

In immunotherapy, to reduce side effects, it is necessary that the peptide,

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[0003]

polypeptide or protein recognized as the antigen exist hardly in normal cells and exist specifically in cancer cells. In 1991, Boon et al. of Ludwig Institute in Belgium
isolated a human melanoma antigen MAGE 1, which is recognized by CD8-positive T cells, by a cDNA-expression cloning method using an autologous cancer cell line and cancer-reactive T cells (Non-patent Literature 2). Thereafter, the SEREX

(serological identifications of antigens by recombinant expression cloning) method, wherein tumor antigens recognized by antibodies produced in the living body of a cancer patient in response to the cancer of the patient himself are identified by application of a gene expression cloning method, was reported (Non-patent Literature 3; Patent Literature 1), and several cancer antigens have been isolated by this method (Non-patent Literatures 4 to 9). Using a part thereof as targets, clinical tests for cancer immunotherapy have started.

[0004]

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On the other hand, as in human, a number of tumors such as mammary gland cancer, leukemia and lymphoma are known in dogs and cats, and they rank high also in the statistics of diseases in dogs and cats. However, at present, no therapeutic agent and prophylactic agent exist which are effective for cancers in dogs and cats. Most of tumors in dogs and cats are realized by owners only after they advanced to grow bigger, and in many cases, it is already too late to visit a hospital to receive surgical excision of the tumor or administration of a human drug (an anticancer preparation or the like), so that those dogs and cats often die shortly after the treatment. Under such circumstances, if therapeutic agents and prophylactic agents for cancer effective for dogs and cats become available, their uses for canine cancers are expected to be developed.

20 [0005]

Since early detection of cancer leads to good treatment results, a method for detecting cancer which can be easily carried out by testing serum, urine or the like without physical and economical burden to cancer patients is demanded. Recently, methods wherein tumor products such as tumor markers are measured have been widely used as diagnostic methods using blood or urine. Examples of the tumor products include tumor-related antigens, enzymes, specific proteins, metabolites, tumor genes, products of tumor genes, and tumor-suppressor genes, and, in some

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cancers, a carcinoembryonic antigen CEA, glycoproteins CA19-9 and CA125, a prostate-specific antigen PSA, calcitonin which is a peptide hormone produced in thyroid, and the like are utilized as tumor markers in cancer diagnosis (Non-patent Literature 10). However, in most types of cancers, there are no tumor markers useful for cancer diagnosis. Further, since most of the tumor markers currently known exist only in very small amounts (e.g., in the order of pg/mL) in body fluid, their detection requires a highly sensitive measurement method or a special technique. Under such circumstances, if a novel cancer detection method by which various cancers can be detected by simple operations is provided, its use for diagnosis of various cancers are expected to be developed.

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[0006]

CD179b is known to be a part of the surrogate light chain of immunoglobulin and to be expressed on the membrane surfaces of precursor cells of B cells (pre-B cells and pro-B cells). It disappears upon differentiation of B cells and is not expressed in mature B cells. However, CD179b is known to be expressed in leukemia (pre-B cell leukemia) cells produced by cancerization of pre-B cells (Nonpatent Literatures 10 and 11). Further, CD179b is known to be expressed also in lymphoma (pre-B cell lymphoma) cells produced by cancerization of pre-B cells, and able to be used as a diagnostic marker for pre-B cell lymphoma (Non-patent

20 Literature 12). However, its specific expression has not been reported for leukemia cells other than pre-B cell leukemia cells, lymphomas other than pre-B cell lymphoma, breast cancer cells and the like. Further, there has been no report suggesting that enhancement of immunity against CD179b is useful for therapy and/or prophylaxis of cancer.

25 PRIOR ART LITERATURES Patent Literature

[0007]

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Patent Literature 1: US 5698396 B

Non-patent Literatures

[0008]

Non-patent Literature 1: Tsuyoshi Akiyoshi, "Cancer and Chemotherapy",

5 1997, Vol. 24, pp. 551-519

Non-patent Literature 2: Bruggen P. et al., Science, 254:1643-1647 (1991)
Non-patent Literature 3: Proc. Natl. Acad. Sci. USA, 92:11810-11813 (1995)
Non-patent Literature 4: Int. J. Cancer, 72:965-971 (1997)
Non-patent Literature 5: Cancer Res., 58:1034-1041 (1998)
Non-patent Literature 6: Int. J. Cancer, 29:652-658 (1998)
Non-patent Literature 7: Int. J. Oncol., 14:703-708 (1999)
Non-patent Literature 8: Cancer Res., 56:4766-4772 (1996)
Non-patent Literature 9: Hum. Mol. Genet 6:33-39 (1997)
Non-patent Literature 10: Adv. Immunol., 63:1-41 (1996)
Non-patent Literature 11: Blood, 92:4317-4324 (1998)
Non-patent Literature 12: Modern Pathology, 17:423-429 (2004)
DISCLOSURE OF THE INVENTION

PROBLEMS TO BE SOLVED BY THE INVENTION

[0009]

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The present invention aims to discover a novel polypeptide useful as an agent for therapy and/or prophylaxis and/or the like of cancer, thereby providing use of the polypeptide for an immunity-inducing agent. The present invention also aims to provide a method for detection of cancer, which is useful for diagnosis of cancer. MEANS FOR SOLVING THE PROBLEMS

25 [0010]

The present inventors intensively studied to obtain, by the SEREX method using serum from a canine patient from which a canine breast cancer tissue-derived cDNA library was prepared, cDNA encoding a protein which binds to antibodies existing in the serum derived from the tumor-bearing living body, and, based on a the cDNA, canine CD179b polypeptides having the amino acid sequences shown in the odd number IDs of SEQ ID NOs:5 to 95 (that is, SEQ ID NOs:5, 7, 9, 11, 13, 15, ...,

5 91 and 93) in SEQUENCE LISTING were prepared. Further, based on a human homologous gene of the obtained genes, a human CD179b polypeptide having the amino acid sequence shown in SEQ ID NO:3 was prepared, and, similarly, based on a bovine homologous gene, a bovine CD179b polypeptide having the amino acid sequence shown in SEQ ID NO:95 was prepared. The present inventors then 10 discovered that these CD179b polypeptides are specifically expressed in breast cancer, leukemia and lymphoma cells. Further, the present inventors discovered that, by administration of these CD179b to a living body, immunocytes against CD179b can be induced in the living body, and a tumor in the living body expressing CD179b can be regressed. Further, the present inventors discovered that a 15 recombinant vector comprising a polynucleotide encoding a CD179b polypeptide or a fragment thereof such that it can be expressed induces an anti-tumor effect against cancer expressing CD179b in the living body.

[0011]

Further, the present inventors discovered that a partial polypeptide in a
CD179b protein has a capacity to be presented by antigen-presenting cells, thereby allowing activation and growth of cytotoxic T cells specific to the peptide (immunity-inducing activity), and therefore that the peptide is useful for therapy and/or prophylaxis of cancer, and, further, that antigen-presenting cells contacted with the peptide and T cells contacted with the antigen-presenting cells are useful for the
therapy and/or prophylaxis of cancer. Further, the present inventors discovered that, since a recombinant polypeptide prepared based on the amino acid sequence of the above CD179b protein specifically reacts only with serum of a tumor-bearing living

body, cancer can be detected therewith. Based on the above discoveries, the present inventors completed the present invention.

[0012]

Thus, the present invention has the following characteristics.

5 [0013]

(1) An immunity-inducing agent comprising as an effective ingredient(s) at least one polypeptide selected from the polypeptides (a) to (c) below, the polypeptide(s) having an immunity-inducing activity/activities, or as an effective ingredient(s) a recombinant vector(s) which comprise(s) a polynucleotide(s) encoding the polypeptide(s) and is/are capable of expressing the polypeptide(s) in vivo:

(a) a polypeptide consisting essentially of not less than 7 consecutive amino acids in any one of the amino acid sequences shown in the odd number IDs of SEQ ID NOs:3 to 95 in SEQUENCE LISTING;

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(b) a polypeptide having a sequence identity of not less than 90% with the polypeptide (a) and consisting essentially of not less than 7 amino acids; and

(c) a polypeptide comprising the polypeptide (a) or (b) as a partial sequence thereof.

[0014]

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(2) The immunity-inducing agent according to (1) above, wherein the polypeptide (b) has a sequence identity of not less than 95% with the polypeptide (a).[0015]

(3) The immunity-inducing agent according to (1) above, wherein each of the polypeptide(s) having an immunity-inducing activity/activities is a polypeptide consisting essentially of not less than 7 consecutive amino acids in any one of the amino acid sequences shown in the odd number IDs of SEQ ID NOs:3 to 95 in SEQUENCE LISTING, or a polypeptide comprising the polypeptide as a partial

sequence thereof.

[0016]

(4) The immunity-inducing agent according to (3) above, wherein each of the polypeptide(s) having an immunity-inducing activity/activities is a polypeptide having any one of the amino acid sequences shown in the odd number IDs of SEQ ID NOs:3 to 95 in SEQUENCE LISTING.

[0017]

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(5) The immunity-inducing agent according to (3) above, wherein each of the polypeptide(s) having an immunity-inducing activity/activities is a polypeptide consisting essentially of not less than 7 consecutive amino acids in the region of aa l-34 or aa52-75 in any one of the amino acid sequences shown in the odd number IDs of SEQ ID NOs:3 to 95 in SEQUENCE LISTING, or a polypeptide comprising the polypeptide as a partial sequence thereof.

[0018]

(6) The immunity-inducing agent according to (5) above, wherein each of the polypeptide(s) having an immunity-inducing activity/activities is a polypeptide consisting essentially of the amino acid sequence shown in SEQ ID NO:108, SEQ ID NO:109, SEQ ID NO:110, SEQ ID NO:111, SEQ ID NO:112, SEQ ID NO:113, SEQ ID NO:114, SEQ ID NO:115, SEQ ID NO:116 or SEQ ID NO:117 in SEQUENCE
LISTING, or a polypeptide comprising as a partial sequence thereof the amino acid sequence shown in SEQ ID NO:108, SEQ ID NO:109, SEQ ID NO:110, SEQ ID NO:109, SEQ ID NO:110, SEQ ID NO:111, SEQ ID NO:112, SEQ ID NO:109, SEQ ID NO:110, SEQ ID NO:111, SEQ ID NO:111, SEQ ID NO:112, SEQ ID NO:113, SEQ ID NO:114, SEQ ID NO:115, SEQ ID NO:113, SEQ ID NO:114, SEQ ID NO:115, SEQ ID NO:113, SEQ ID NO:114, SEQ ID NO:115, SEQ ID NO:113, SEQ ID NO:114, SEQ ID NO:115, SEQ ID NO:111, SEQ ID NO:117 in SEQUENCE LISTING, the polypeptide having 8 to 12 amino acid residues.

25 **[0019]**

(7) The immunity-inducing agent according to any one of (1) to (6) above, comprising one or more of the polypeptides as an effective ingredient(s).

[0020]

(8) The immunity-inducing agent according to (7) above, wherein the polypeptide(s) is/are an agent(s) for treating antigen-presenting cells.[0021]

(9) The immunity-inducing agent according to any one of (1) to (8) above, which is for therapy and/or prophylaxis of an animal cancer(s).

[0022]

(10) The immunity-inducing agent according to (9) above, wherein the cancer(s) is/are a cancer(s) expressing the CD179b gene.

10 [0023]

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(11) The immunity-inducing agent according to (10), wherein the cancer(s) is/are breast cancer, leukemia and/or lymphoma.

[0024]

(12) The immunity-inducing agent according to any one of (1) to (11) above,further comprising an immunoenhancer.

[0025]

(13) An isolated antigen-presenting cell comprising a complex between the polypeptide having an immunity-inducing activity and an HLA molecule.
[0026]

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(14) An isolated T cell which selectively binds to a complex between the polypeptide having an immunity-inducing activity and an HLA molecule.
[0027]

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(15) A method for inducing immunity, the method comprising administering to an individual at least one polypeptide selected from the polypeptides (a) to (c) below, the polypeptide(s) having an immunity-inducing activity/activities, or a recombinant vector(s) which comprise(s) a polynucleotide(s) encoding the polypeptide(s) and is/are capable of expressing the polypeptide(s) in vivo: (a) a polypeptide consisting essentially of not less than 7 consecutive amino acids in any one of the amino acid sequences shown in the odd number IDs of SEQ ID NOs:3 to 95 in SEQUENCE LISTING;

(b) a polypeptide having a sequence identity of not less than 90% with the polypeptide (a) and consisting essentially of not less than 7 amino acids; and

(c) a polypeptide comprising the polypeptide (a) or (b) as a partial sequence thereof.

[0028]

(16) A method for detecting a cancer(s), which method is applied to a sample
 separated from a living body and comprises measuring expression of at least one of
 the polypeptides (a) to (c) below:

(a) a polypeptide consisting essentially of not less than 7 consecutive amino acids in any one of the amino acid sequences shown in the odd number IDs of SEQ ID NOs:3 to 95 in SEQUENCE LISTING;

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(b) a polypeptide having a sequence identity of not less than 90% with the polypeptide (a) and consisting essentially of not less than 7 amino acids.

(c) a polypeptide comprising the polypeptide (a) or (b) as a partial sequence thereof.

[0029]

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(17) The method according to (16) above, wherein the measurement of expression of the polypeptide(s) is carried out by measuring an antibody/antibodies which may be contained in the sample by immunoassay, which antibody/antibodies was/were induced in the living body against the polypeptide(s) to be measured. [0030]

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(18) A method for detecting a cancer(s), which is applied to a sample separated from a living body and comprises investigation of expression of the CD179b gene having a coding region having any one of the base sequences shown in SEQ ID NO:1 and the even number IDs of SEQ ID NOs:4 to 94 in SEQUENCE LISTING in a sample derived from a cancer patient, and comparison thereof with the expression level of the gene in a sample derived from a healthy individual. [0031]

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(19) A reagent for detecting a cancer(s), the reagent comprising a polypeptide which undergoes antigen-antibody reaction with an antibody induced in a living body against the polypeptide of any one of (a) to (c) below:

(a) a polypeptide consisting essentially of not less than 7 consecutive amino acids in any one of the amino acid sequences shown in the odd number IDs of SEQ ID NOs:3 to 95 in SEQUENCE LISTING;

(b) a polypeptide having a sequence identity of not less than 90% with the polypeptide (a) and consisting essentially of not less than 7 amino acids; and

(c) a polypeptide comprising the polypeptide (a) or (b) as a partial sequence thereof.

15 EFFECT OF THE INVENTION

[0032]

By the present invention, a novel immunity-inducing agent useful for therapy and/or prophylaxis and/or the like of cancer is provided. As particularly described in later-mentioned Examples, by administering the polypeptide used in the present invention to a tumor-bearing animal, immunocytes can be induced in the body of the tumor-bearing animal, and a cancer which has already occurred can be reduced or regressed.

[0033]

Further, by the present invention, a novel method for detection of cancer is provided. Since measurement of expression of the polypeptide in a sample by the method of the present invention enables detection of invisible small cancers and cancers which exist in deep parts of a body, the method is also useful for early detection of cancers in medical examinations and the like. If the method of the present invention is used in following-up of patients after cancer therapy, recurrence of the cancer can be detected in its early stage. Moreover, the method of the present invention makes it possible to assess the stage of cancer progression such as growth of the tumor, invasion of the tumor to the surrounding tissues, and metastasis of the

cancer to lymph nodes and distant organs.

BRIEF DESCRIPTION OF THE DRAWINGS

[0034]

Fig. 1 is a diagram showing the expression patterns of the gene encoding the
CD179b protein in normal tissues and tumor cell lines. Reference numeral 1
represents the expression pattern of the gene encoding the CD179b protein; and
reference numeral 2 represents the expression pattern of the GAPDH gene. In Fig.
1, reference numeral 1 in the ordinate represents the expression pattern of the gene
identified as described above, and reference numeral 2 represents the expression
pattern of the GAPDH gene as the control for comparison.

In Fig. 2, reference numerals 3, 4, 5, 6, 7, 8, 9 and 10 in the abscissa indicate the IFN-γ-producing abilities of HLA-A0201-positive CD8-positive T cells due to stimulation from T2 cells pulsed with the peptides of SEQ ID NOs:108, 109, 110, 113, 114, 115, 116 and 117, respectively. Reference numeral 11 indicates the result for the peptide of SEQ ID NO:118 used as the negative control (peptide having a sequence outside the scope of the present invention).

In Fig. 3, reference numerals 12, 13, 14, 15, 16, 17, 18 and 19 in the abscissa indicate the cytotoxic activities of HLA-A0201-positive CD8-positive T cells against Namalwa cells, which cells were stimulated using SEQ ID NOs:108, 109, 110, 113, 114, 115, 116 and 117, respectively. Reference numeral 20 indicates the cytotoxic activity of CD8-positive T cells induced using the peptide of the negative control (SEQ ID NO:118).

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In Fig. 4, reference numerals 21, 22, 23, 24 and 25 in the abscissa indicate the IFN-γ-producing abilities of HLA-A24-positive CD8-positive T cells due to stimulation from JTK-LCL cells pulsed with the peptides of SEQ ID NOs:110, 111, 112, 115 and 116, respectively. Reference numeral 26 indicates the result for the peptide of SEQ ID NO:118 used as the negative control.

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In Fig. 5, reference numerals 27, 28, 29, 30 and 31 indicate the cytotoxic activities of HLA-A24-positive CD8-positive T cells stimulated with the peptides of SEQ ID NO:110, 111, 112, 115 and 116, respectively, against JTK-LCL cells. Reference numeral 32 indicates the cytotoxic activity of CD8-positive T cells

10 induced using the peptide of the negative control (SEQ ID NO:118).

BEST MODE FOR CARRYING OUT THE INVENTION

[0035]

<Polypeptide>

Examples of the polypeptide contained in the immunity-inducing agent of the present invention as an effective ingredient include one or more polypeptide(s) selected from the polypeptides of (a), (b) and (c) below:

(a) a polypeptide which consists essentially of not less than 7 consecutive amino acids in a polypeptide having any one of the amino acid sequences shown in the odd number IDs of SEQ ID NOs:3 to 95 in SEQUENCE LISTING (that is, SEQ ID NO:3, 5, 7, 9, 11, 13, 15, ..., 93 and 95) and has an immunity-inducing activity;

(b) a polypeptide having a sequence identity of not less than 90% with the polypeptide (a), consisting essentially of not less than 7 amino acids, and having an immunity-inducing activity; and

(c) a polypeptide comprising the polypeptide (a) or (b) as a partial sequencethereof and having an immunity-inducing activity.

[0036]

As used herein, the term "polypeptide" means a molecule formed by a

plurality of amino acids linked together by peptide bonds, and includes not only polypeptide molecules having large numbers of amino acids constituting them, but also low-molecular-weight molecules having small numbers of amino acids (oligopeptides), and full-length molecules. In the present invention, proteins constituted by the total lengths of SEQ ID NO:3, 5, 7, 9, 11, 13, 15, ..., 93 and 95 are also included therein.

[0037]

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As used herein, the term "having an amino acid sequence" means that amino acid residues are arrayed in a specific order. Therefore, for example, "a polypeptide having the amino acid sequence shown in SEQ ID NO:3" means a polypeptide having the amino acid sequence of Leu Leu Arg Pro ... (snip) ... Ala Glu Cys Ser shown in SEQ ID NO:3, which polypeptide has a size of 176 amino acid residues. Further, for example, "polypeptide having the amino acid sequence shown in SEQ ID NO:3" may also be abbreviated as "polypeptide of SEQ ID NO:3". This also applies to the term "having a base sequence".

[0038]

As used herein, the term "immunity-inducing activity" means an ability to induce immunocytes which secrete cytokines such as interferon in a living body. [0039]

Whether or not a polypeptide has an immunity-inducing activity can be confirmed using, for example, the known ELISPOT assay. More particularly, for example, as described in the Examples below, cells such as peripheral blood mononuclear cells are obtained from a living body to which a polypeptide whose immunity-inducing activity is to be evaluated was administered, which cells are then cocultivated with the polypeptide, followed by measuring the amount(s) of a cytokine(s) and/or a chemokine(s) such as IFN- γ and/or interleukin (IL) produced by the cells using a specific antibody/antibodies, thereby measuring the number of immunocytes in the cells, which enables evaluation of the immunity-inducing activity. [0040]

Alternatively, as described in the later-mentioned Examples, when a recombinant polypeptide prepared based on the amino acid sequence of SEQ ID
NO:3, 5, 7, 9, 11, 13, 15, ..., 93 or 95 is administered to a tumor-bearing animal, the tumor can be reduced or regressed by its immunity-inducing activity. Thus, the above immunity-inducing activity can be evaluated also as an ability to suppress the growth of cancer cells expressing the polypeptide of SEQ ID NO:3, 5, 7, 9, 11, 13, 15, ..., 93 or 95, or to cause reduction or disappearance of a cancer tissue (tumor)
(hereinafter referred to as "anti-tumor activity"). The anti-tumor activity of a polypeptide can be confirmed by, for example, observation of whether or not the tumor is reduced or regressed when the polypeptide was administered to a tumor-bearing living body, as more particularly described in the Examples below. [0041]

Alternatively, the anti-tumor activity of a polypeptide can be evaluated also
 by observation of whether or not T cells stimulated with the polypeptide (that is, T
 cells brought into contact with antigen-presenting cells presenting the polypeptide)
 show a cytotoxic activity against tumor cells in vitro. The contact between the T
 cells and the antigen-presenting cells can be carried out by cocultivation of the both
 in a liquid medium, as mentioned below. Measurement of the cytotoxic activity can
 be carried out by, for example, a known method called ⁵¹Cr release assay described in
 Int. J. Cancer, 58: p317, 1994.

[0042]

In cases where the polypeptide is used for therapy and/or prophylaxis of cancer, the evaluation of the immunity-inducing activity is preferably carried out using the anti-tumor activity as an index, although the index is not restricted. [0043]

The amino acid sequence shown in each of SEQ ID NOs:3, 5, 7, 9, 11, 13, 15, ... 93 and 95 in SEQUENCE LISTING is the amino acid sequence of a polypeptide which binds to an antibody specifically existing in serum derived from a tumorbearing dog in the SEREX method using serum of the canine patient from which a 5 canine mammary gland cancer-derived cDNA library was prepared, or the amino acid sequence of CD179b isolated as a human homologous factor (homologue) of the polypeptide (see Example 1 below). The polypeptide (a) is a polypeptide which consists essentially of not less than 7 consecutive amino acids, preferably 8, 9 or not less than 10 consecutive amino acids in a polypeptide having any one of the amino 10 acid sequences shown in SEQ ID NO:3, 5, 7, 9, 11, 13, 15, ..., 93 and 95 in SEQUENCE LISTING and has an immunity-inducing activity. As known in the art, a polypeptide having not less than about 7 amino acid residues can exert its antigenicity and immunogenicity. Thus, a polypeptide having not less than 7 consecutive amino acid residues in the amino acid sequence shown in SEQ ID NO:3, 15 5, 7, 9, 11, 13, 15, ..., 93 or 95 can have an immunity-inducing activity, so that it can be used for preparation of the immunity-inducing agent of the present invention. [0044]

As a principle of immune induction by administration of a cancer antigenic polypeptide, the following process is known: the polypeptide is incorporated into an antigen-presenting cell and then degraded into smaller fragments by peptidases in the cell, followed by presentation of the fragments on the surface of the cell. The fragments are then recognized by a cytotoxic T cell or the like, which selectively kills cells presenting the antigen. The size of the polypeptide presented on the surface of the antigen-presenting cell is relatively small and about 7 to 30 amino acids.

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Therefore, from the view point of presenting thereof on the surface of the antigenpresenting cell, one preferred mode of the polypeptide (a) is a polypeptide composed of about 7 to 30 consecutive amino acids in the amino acid sequence shown in SEQ

ID NO:3, 5, 7, 9, 11, 13, 15, ..., 93 or 95, and more preferably, a polypeptide composed of 8 to 30 or 9 to 30 amino acids is sufficient as the polypeptide (a). In some cases, these relatively small polypeptides are presented directly on the surface of the antigen-presenting cells without incorporation thereof into the antigen-

5 presenting cells.

[0045]

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Further, since a polypeptide incorporated into an antigen-presenting cell is cleaved at random sites by peptidases in the cell to yield various polypeptide fragments, which are then presented on the surface of the antigen-presenting cell, administration of a large polypeptide such as the entire region of SEQ ID NO:3, 5, 7, 9, 11, 13, 15, ..., 93 or 95 inevitably causes production of polypeptide fragments by degradation thereof in the antigen-presenting cell, which fragments are effective for immune induction via the antigen-presenting cell. Therefore, also for immune induction via antigen-presenting cells, a large polypeptide can be used, and the polypeptide may be composed of not less than 30, preferably not less than 100, more preferably not less than 200 amino acids, which polypeptide may be still more preferably composed of the entire region of SEQ ID NO:3, 5, 7, 9, 11, 13, 15, ..., 93 or 95.

[0046]

Further, the polypeptides of the present invention can be checked with a checking medium by which epitope peptides having binding motifs of various types of HLA and composed of 8 to 12, preferably 9 to 10 amino acids can be searched, for example, HLA Peptide Binding Predictions

(http://bimas.dcrt.nih.gov/molbio/hla_bind/index.html) in Bioinformatics &

25 Molecular Analysis Selection (BIMAS), to screen peptides which may be epitope peptides. More particularly, a polypeptide composed of not less than 7 consecutive amino acids in the region of aa1-34 or aa52-75 in the amino acid sequence shown in

SEQ ID NO:3, 5, 7, 9, 11, 13, 15, ..., 93 or 95 is preferred, and, in the polypeptide of SEQ ID NO:3, the polypeptides shown in SEQ ID NOs:108 to 117 are more preferred.

[0047]

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The polypeptide (b) is the same polypeptide as the polypeptide (a) except that a small number of amino acid residues are substituted, deleted, added and/or inserted, which has a sequence identity of not less than 80%, preferably not less than 90%, more preferably not less than 95%, still more preferably not less than 98%, not less than 99% or not less than 99.5% to the original sequence, and has an immunity-

10 inducing activity. It is well known in the art that, in general, there are cases where a protein antigen retains substantially the same antigenicity or immunogenicity as the original even if the amino acid sequence of the protein is modified such that a small number of amino acids are substituted, deleted, added and/or inserted. Therefore, since the polypeptide (b) may also exert an immunity-inducing activity, it can be used 15 for preparation of the immunity-inducing agent of the present invention. Further, the polypeptide (b) is also preferably the same polypeptide as one having the amino acid sequence shown in SEQ ID NO:3, 5, 7, 9, 11, 13, 15, ..., 93 or 95 except that

[0048]

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As used herein, the term "sequence identity" in relation to amino acid sequences or base sequences means the value calculated by aligning two amino acid sequences (or base sequences) to be compared such that the number of matched amino acid residues (or bases) is maximum between the amino acid sequences (or base sequences), and dividing the number of matched amino acid residues (or the number of matched bases) by the total number of amino acid residues (or the total number of bases), which value is represented as a percentage (%). When the alignment is carried out, a gap(s) is/are inserted into one or both of the two sequences

one or several amino acid residues are substituted, deleted, added and/or inserted.

to be compared as required. Such alignment of sequences can be carried out using a
well-known program such as BLAST, FASTA or CLUSTAL W (Karlin and Altschul,
Proc. Natl. Acad. Sci. U.S.A., 87:2264-2268, 1993; Altschul et al., Nucleic Acids
Res., 25:3389-3402, 1997). When a gap(s) is/are inserted, the above-described
number of the total amino acid residues (or the total bases) is the number of residues
(or bases) calculated by counting one gap as one amino acid residue (or base).
When the thus counted numbers of the total amino acid residues (or bases) are
different between the two sequences to be compared, the identity (%) is calculated by
dividing the number of matched amino acid residues (or bases) by the number of the
total amino acid residues (or the total bases) in the longer sequence.

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[0049]

Among substitutions of amino acid residues, conservative amino acid substitutions are preferred. The 20 types of amino acids constituting the naturally occurring proteins may be classified into groups each of which has similar properties, for example, into neutral amino acids with side chains having low polarity (Gly, Ile, Val, Leu, Ala, Met, Pro), neutral amino acids having hydrophilic side chains (Asn, Gln, Thr, Ser, Tyr, Cys), acidic amino acids (Asp, Glu), basic amino acids (Arg, Lys, His) and aromatic amino acids (Phe, Tyr, Trp, His). It is known that, in most cases, substitutions of amino acids within the same group, that is, conservative substitutions, do not change the properties of the polypeptide. Therefore, in cases where an amino acid residue(s) in the polypeptide (a) of the present invention is/are substituted, the probability that the immunity-inducing activity can be maintained may be made high by conducting the substitution(s) within the same group.

[0050]

The polypeptide (c) comprises the polypeptide (a) or (b) as a partial sequence thereof and has an immunity-inducing activity. That is, the polypeptide (c) has another/other amino acid(s) or polypeptide(s) added at one end or the both ends of

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the polypeptide (a) or (b), and has an immunity-inducing activity. Such a polypeptide can also be used for preparation of the immunity-inducing agent of the present invention.

The above-described polypeptides can be synthesized by, for example, a chemical synthesis method such as the Fmoc method (fluorenylmethyloxycarbonyl method) or the tBoc method (t-butyloxycarbonyl method). Further, they can be synthesized by conventional methods using various types of commercially available peptide synthesizers. Further, the polypeptide of interest can be obtained using known genetic engineering techniques, by preparing a polynucleotide encoding the above polypeptide and incorporating the polynucleotide into an expression vector, which is then transfected into a host cell, followed by allowing the polypeptide to be produced in the host cell.

[0051]

The polynucleotide encoding the above polypeptide can be easily prepared by 15 a known genetic engineering technique or a conventional method using a commercially available nucleic acid synthesizer. For example, DNA having the base sequence shown in SEQ ID NO:4 can be prepared by carrying out PCR using a canine chromosomal DNA or cDNA library as a template, and a pair of primers designed such that the base sequence shown in SEQ ID NO:4 can be amplified 20 therewith. In the case of DNA having the base sequence of SEQ ID NO:1, this can be similarly prepared by using a human chromosomal DNA or cDNA library as the template. The reaction conditions for the PCR can be set appropriately, and examples thereof include, but are not limited to, repeating the reaction process of 94°C for 30 seconds (denaturation), 55°C for 30 seconds to 1 minute (annealing) and 25 72°C for 2 minutes (extension) for, for example, 30 cycles, followed by the reaction at 72°C for 7 minutes. Methods, conditions and the like of PCR are described in, for example, Ausubel et al., Short Protocols in Molecular Biology, 3rd ed., A

compendium of Methods from Current Protocols in Molecular Biology (1995), John Wiley & Sons (in particular, Chapter 15). Further, the desired DNA can be isolated by preparing an appropriate probe(s) or primer(s) based on the information of the base sequences and the amino acid sequences shown in SEQ ID NO:1 to 95 in

5 SEQUENCE LISTING in the present specification, and screening a cDNA library of human, dog, bovine or the like using the probe(s) or primer(s). The cDNA library is preferably prepared from a cell, organ or tissue expressing the protein of SEO ID NO:3, 5, 7, 9, 11, 13, 15, ..., 93 or 95. The above-described operations such as preparation of the probe(s) or primer(s), construction of a cDNA library, screening of 10 the cDNA library and cloning of the gene of interest are known to those skilled in the art, and can be carried out according to the methods described in, for example, Sambrook et al., Molecular Cloning, Second Edition, Current Protocols in Molecular Biology (1989); and Ausubel et al. (described above). From the thus obtained DNA, DNA encoding the polypeptide (a) can be obtained. Further, since codons encoding 15 each amino acid are known, a base sequence of a polynucleotide encoding a specific amino acid sequence can be easily specified. Therefore, the base sequences of polynucleotides encoding the polypeptide (b) and polypeptide (c) can also be easily specified, so that such polynucleotides can also be easily synthesized using a commercially available nucleic acid synthesizer according to a conventional method. [0052]

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The host cells are not restricted as long as they can express the abovedescribed polypeptide, and examples thereof include, but are not limited to, prokaryotic cells such as E. coli; and eukaryotic cells such as mammalian cultured cells including monkey kidney cells COS 1, Chinese hamster ovary cells CHO, a human embryonic kidney cell line HEK293 and a mouse embryonic skin cell line NIH3T3; budding yeast; fission yeast; silkworm cells; and Xenopus laevis egg cells. [0053]

In cases where prokaryotic cells are used as the host cells, an expression vector having the origin that enables its replication in a prokaryotic cell, promoter, ribosome binding site, multicloning site, terminator, drug resistant gene, nutrient complementary gene and/or the like is used. Examples of the expression vector for *E. coli* include the pUC system, pBluescriptII, pET expression system and pGEX expression system. By incorporating a DNA encoding the above polypeptide into such an expression vector and transforming prokaryotic host cells with the vector, followed by culturing the resulting transformants, the polypeptide encoded by the DNA can be expressed in the prokaryotic host cells. In this process, the polypeptide can also be expressed as a fusion protein with another protein (e.g., green fluorescent protein (GFP) or glutathione S-transferase (GST)).

[0054]

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In cases where eukaryotic cells are used as the host cells, an expression vector for eukaryotic cells having a promoter, splicing site, poly(A) addition site and/or the like is used as the expression vector. Examples of such an expression vector include pKA1, pCDM8, pSVK3, pMSG, pSVL, pBK-CMV, pBK-RSV, EBV vector, pRS, pcDNA3, pMSG and pYES2. In the same manner as described above, by incorporating a DNA encoding the above polypeptide into such an expression vector and transforming eukaryotic host cells with the vector, followed by culturing the resulting transformants, the polypeptide encoded by the DNA can be expressed in the eukaryotic host cells. In cases where pIND/V5-His, pFLAG-CMV-2, pEGFP-N1, pEGFP-C1 or the like is used as the expression vector, the above polypeptide can be expressed as a fusion protein to which a tag such as His tag (e.g., (His)₆ to (His)₁₀), FLAG tag, myc tag, HA tag or GFP was added.

25 [0055]

For the introduction of the expression vector into the host cells, well-known methods such as electroporation, the calcium phosphate method, the liposome

method, the DEAE dextran method and microinjection can be used. [0056]

Isolation and purification of the polypeptide of interest from the host cells can be carried out by a combination of known separation operations. Examples of the known separation operations include, but are not limited to, treatment with a denaturant such as urea, or a surfactant; ultrasonication treatment; enzyme digestion; salting-out or solvent fractional precipitation; dialysis; centrifugation; ultrafiltration; gel filtration; SDS-PAGE; isoelectric focusing; ion-exchange chromatography; hydrophobic chromatography; affinity chromatography; and reversed-phase chromatography.

[0057]

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The polypeptides obtained by the above method include, as mentioned above, those in the form of a fusion protein with another arbitrary protein. Examples thereof include fusion proteins with glutathion S-transferase (GST) and with a His tag. Such a polypeptide in the form of a fusion protein is also included within the scope of the present invention as the polypeptide (c). Further, in some cases, a polypeptide expressed in a transformed cell is modified in various ways in the cell after translation thereof. Such a polypeptide modified after translation thereof is also included within the scope of the present invention as long as it has an immunityinducing activity. Examples of such a post-translational modification include elimination of N-terminus methionine, N-terminus acetylation, glycosylation, limited degradation by an intracellular protease, myristoylation, isoprenylation and phosphorylation.

[0058]

25 <Immunity-inducing Agent>

As described concretely in the following Examples, the above-described polypeptide having an immunity-inducing activity can cause regression of an already

occurred tumor when administered to a tumor-bearing animal. Therefore, the immunity-inducing agent of the present invention can be used as a therapeutic and/or prophylactic agent for cancer.

[0059]

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The terms "cancer" and "tumor" used in the present specification mean a malignant neoplasm, and are used interchangeably.

[0060]

In this case, cancers to be treated are those expressing the CD179b gene, such as cancers expressing the gene encoding the polypeptide of SEQ ID NO: 3, 5, 7, 9, 11, 13, 15, ..., 93 or 95, preferably breast cancer, leukemia and lymphoma. Examples of these particular cancers include, but are not limited to, breast cancers (mammary gland cancer, combined mammary gland cancer, mammary gland malignant mixed tumor, intraductal papillary adenocarcinoma and the like), leukemias (chronic lymphocytic leukemia and the like), lymphomas (gastrointestinal lymphoma, digestive organ lymphoma, small/medium cell lymphoma and the like).

[0061]

The above-described polypeptide, or a recombinant vector comprising a polynucleotide encoding the polypeptide and capable of expressing the polypeptide in vivo can be used as a therapeutic method for immune induction. Further, it can be used as a therapeutic method for the purpose(s) of therapy and/or prophylaxis of animal cancer, and can also be used as a therapeutic method further comprising an immunoenhancer.

[0062]

The subject animal is a mammal such as a primate, pet animal, domestic animal or sport animal, preferably human, dog or cat.

[0063]

The administration route of the immunity-inducing agent of the present

invention to a living body may be either oral administration or parenteral administration, and is preferably parenteral administration such as intramuscular administration, subcutaneous administration, intravenous administration or intraarterial administration. In cases where the immunity-inducing agent is used for therapy of cancer, it may be administered to a regional lymph node in the vicinity of the tumor to be treated, as described in the Examples below, in order to enhance its anticancer activity. The dose may be any dose as long as the dose is effective for immune induction, and, for example, in cases where the agent is used for therapy and/or prophylaxis of cancer, the dose may be one effective for therapy and/or prophylaxis of the cancer. Further, the dose may vary depending on the body weight, sex (male or female), symptoms and the like. The dose effective for therapy and/or prophylaxis of cancer is appropriately selected depending on the size of the tumor, the symptom and the like, and usually, $0.0001 \ \mu g$ to $1000 \ \mu g$, preferably 0.001 μ g to 1000 μ g per subject animal per day, which may be administered once or in several times. The agent is preferably administered in several times, every several days to several months.

[0064]

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As concretely shown in the Examples below, the immunity-inducing agent of the present invention can cause reduction or regression of an already occurred tumor. Therefore, since the agent can exert its anticancer activity also against a small number of cancer cells in the early stage, development or recurrence of cancer can be prevented by using the agent before development of the cancer or after therapy for the cancer. That is, the immunity-inducing agent of the present invention is effective for both therapy and prophylaxis of cancer.

25 [0065]

The immunity-inducing agent of the present invention may contain only a polypeptide or may be formulated by mixing as appropriate with an additive such as a

pharmaceutically acceptable carrier, diluent or vehicle suitable for each administration mode. Formulation methods and additives which may be used are well-known in the field of formulation of pharmaceuticals, and any of the methods and additives may be used. Specific examples of the additives include, but are not

- limited to, diluents such as physiological buffer solutions; vehicles such as sucrose,
 lactose, corn starch, calcium phosphate, sorbitol and glycine; binders such as syrup,
 gelatin, gum arabic, sorbitol, polyvinyl chloride and tragacanth; and lubricants such
 as magnesium stearate, polyethylene glycol, talc and silica. Examples of the
 formulation include oral preparations such as tablets, capsules, granules, powders and
- 10 syrups; and parenteral preparations such as inhalants, injection solutions, suppositories and solutions. These formulations may be prepared by commonly known production methods.

[0066]

The immunity-inducing agent of the present invention may be used in combination with an immunoenhancer capable of enhancing the immune response in a living body. The immunoenhancer may be contained in the immunity-inducing agent of the present invention or administered as a separate composition to a patient in combination with the immunity-inducing agent of the present invention. [0067]

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Here, the patient is an animal, especially a mammal, preferably human, dog or cat.

Examples of the immunoenhancer include adjuvants. Adjuvants can

[0068]

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enhance the immune response by providing a reservoir of antigen (extracellularly or within macrophages), activating macrophages and stimulating specific sets of lymphocytes, thereby enhancing the immune response and hence the anticancer action. Therefore, especially in cases where the immunity-inducing agent of the present invention is used for therapy and/or prophylaxis of cancer, the immunityinducing agent preferably comprises an adjuvant, in addition to the above-described polypeptide as an effective ingredient. Many types of adjuvants are well-known in the art, and any of these adjuvants may be used. Specific examples of the adjuvants include MPL (SmithKline Beecham) and homologues of *Salmonella minnesota* Re 595 lipopolysaccharide obtained after purification and acid hydrolysis of the lipopolysaccharide; QS21 (SmithKline Beecham), pure QA-21 saponin purified from an extract of *Quillja saponaria*; DQS21 described in WO96/33739 (SmithKline Beecham); QS-7, QS-17, QS-18 and QS-L1 (So et al., "Molecules and cells", 1997, Vol. 7, p. 178-186); Freund's incomplete adjuvant; Freund's complete adjuvant; vitamin E; Montanide; alum; CpG oligonucleotides (for example, Kreig et al., Nature, Vol. 374, p. 546-549); poly-I:C and derivatives thereof (e.g., poly ICLC); and various

tocopherol. Among these, Freund's incomplete adjuvant; Montanide; poly-I:C and
derivatives thereof; and CpG oligonucleotides are preferred. The mixing ratio
between the above-described adjuvant and the polypeptide is typically about 1:10 to
10:1, preferably about 1:5 to 5:1, more preferably about 1:1. However, the adjuvant
is not limited to the above-described examples, and adjuvants known in the art other
than those described above (for example, Goding, "Monoclonal Antibodies:

water-in-oil emulsions prepared from biodegradable oils such as squalene and/or

20 Principles and Practice, 2nd edition", 1986) may be used when the immunityinducing agent of the present invention is administered. Preparation methods for mixtures or emulsions of a polypeptide and an adjuvant are well-known to those skilled in the art of vaccination.

[0069]

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Further, in addition to the above-described adjuvants, factors that stimulate the immune response of the subject may be used as the above-described immunoenhancer. For example, various cytokines having a property to stimulate

lymphocytes and/or antigen-presenting cells may be used as the immunoenhancer in combination with the immunity-inducing agent of the present invention. A number of such cytokines capable of enhancing the immune response are known to those skilled in the art, and examples thereof include, but are not limited to, interleukin-12 (IL-12), GM-CSF, IL-18, interferon- α , interferon- β , interferon- ω , interferon- γ , and Flt3 ligand, which have been shown to enhance the prophylactic action of vaccines. Such factors may also be used as the above-described immunoenhancer, and can be contained in the immunity-inducing agent of the present invention, or can be prepared as a separate composition to be used in combination with the immunityinducing agent of the present invention, to be administered to a patient

[0070]

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<Antigen-presenting Cells>

As concretely described in the Examples below, by bringing the abovedescribed polypeptide used in the present invention into contact with antigenpresenting cells in vitro, the antigen-presenting cells can be made to present the polypeptide. That is, the polypeptides (a) to (c) described above can be used as agents for treating antigen-presenting cells. Examples of the antigen-presenting cells include dendritic cells and B cells, and dendritic cells and B cells having MHC class I molecules are preferably employed. The agents for treating antigenpresenting cells mean agents for pulsing antigen-presenting cells, and, since pulsed

20 presenting cells mean agents for pulsing antigen-presenting cells, and, since pulsed antigen-presenting cells can have an ability to stimulate peripheral blood lymphocytes, the cells can be used as a vaccine.

[0071]

Various MHC class I molecules have been identified and well-known.
25 MHC molecules in human are called HLA. Examples of HLA class I molecules include HLA-A, HLA-B and HLA-C, more specifically, HLA-A1, HLA-A0201, HLA-A0204, HLA-A0205, HLA-A0206, HLA-A0207, HLA-A11, HLA-A24, HLA-

A31, HLA-A6801, HLA-B7, HLA-B8, HLA-B2705, HLA-B37, HLA-Cw0401 and HLA-Cw0602.

[0072]

The dendritic cells or B cells having MHC class I molecules can be prepared from peripheral blood by a well-known method. For example, tumor-specific dendritic cells can be induced by inducing dendritic cells from bone marrow, umbilical cord blood or patient's peripheral blood using granulocyte-macrophage colony-stimulating factor (GM-CSF) and IL-3 (or IL-4), and then adding a tumorrelated peptide to the culture system.

10 [0073]

By administering an effective amount of such dendritic cells, a response desired for therapy of a cancer can be induced. As the cells to be used, bone marrow or umbilical cord blood donated by a healthy individual, or bone marrow, peripheral blood or the like from the patient himself may be used. When autologous cells of the patient are used, high safety can be attained and serious side effects are expected to be avoided. The peripheral blood or bone marrow may be a fresh sample, cold-stored sample or frozen sample. As for the peripheral blood, whole blood may be cultured or the leukocyte components alone may be separated and cultured, and the latter is efficient and thus preferred. Further, among the leukocyte components, mononuclear cells may be separated. In cases where the cells are originated from bone marrow or umbilical cord blood, the whole cells constituting the bone marrow may be cultured, or mononuclear cells may be separated therefrom and cultured. Peripheral blood, the leukocyte components thereof and bone marrow cells contain mononuclear cells, hematopoietic stem cells and immature dendritic cells, from which dendritic cells are originated, and also CD4-positive cells and the like. As for the cytokine to be used, the production method thereof is not restricted and naturally-occurring or recombinant cytokine or the like may be employed as long

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as its safety and physiological activity have been confirmed. Preferably, a preparation with assured quality for medical use is used in a minimum necessary amount. The concentration of the cytokine(s) to be added is not restricted as long as the dendritic cells are induced, and usually, the total concentration of the cytokine(s) is preferably about 10 to 1000 ng/mL, more preferably about 20 to 500 ng/mL. 5 The culture may be carried out using a well-known medium usually used for culture of The culturing temperature is not restricted as long as the proliferation leukocytes. of the leukocytes is attained, and about 37°C which is the body temperature of human is most preferred. The atmospheric environment during the culturing is not 10 restricted as long as the proliferation of the leukocytes is attained, and 5% CO₂ is preferably allowed to flow. The culturing period is not restricted as long as a necessary number of the cells is induced therewith, and is usually 3 days to 2 weeks. As for the apparatuses used for separation and culturing of the cells, appropriate apparatuses, preferably those whose safety when applied to medical uses have been 15 confirmed, and whose operations are stable and simple, may be employed. In particular, as for the cell-culturing apparatus, not only the general vessels such as a Petri dish, flask and bottle, but also a layer type vessel, multistage vessel, roller bottle, spinner type bottle, bag type culturing vessel, hollow fiber column and the like may be used.

20 [0074]

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Bringing the above-described peptide into contact with the antigen presenting cells in vitro may be carried out by a well-known method. For example, it may be carried out by culturing the antigen-presenting cells in a culture medium containing the above-described polypeptide. The concentration of the peptide in the medium is not restricted, and usually about 1 μ g/ml to 100 μ g/ml, preferably about 5 μ g/ml to 20 μ g/ml. The cell density during the culturing is not restricted and usually about 10³ cells/ml to 10⁷ cells/ml, preferably about 5×10⁴ cells/ml to 5×10⁶ cells/ml. The

culturing may be carried out according to a conventional method, and is preferably carried out at 37°C under atmosphere of 5% CO₂. The maximum length of the peptide which can be presented on the surface of the antigen-presenting cells is usually about 30 amino acid residues. Therefore, in cases where the antigenpresenting cells are brought into contact with the polypeptide in vitro, the polypeptide may be prepared such that its length is not more than about 30 amino acid residues,

[0075]

although the length is not restricted.

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By culturing the antigen-presenting cells in the coexistence of the abovedescribed polypeptide, the polypeptide is incorporated into MHC molecules of the antigen-presenting cells and presented on the surface of the antigen-presenting cells. Therefore, using the above-described polypeptide, isolated antigen-presenting cells containing the complex between the polypeptide and the MHC molecules can be prepared. Such antigen-presenting cells can present the polypeptide against T cells in vivo or in vitro, and thereby induce, and allow proliferation of, cytotoxic T cells specific to the polypeptide.

[0076]

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By bringing the antigen-presenting cells prepared as described above having the complex between the above-described polypeptide and the MHC molecules into contact with T cells in vitro, cytotoxic T cells specific to the polypeptide can be induced and allowed to proliferate. This may be carried out by cocultivating the above-described antigen-presenting cells and T cells in a liquid medium. For example, it may be attained by suspending the antigen-presenting cells in a liquid medium, placing the suspension in vessels such as wells of a microplate, adding thereto T cells and then culturing the cells. The mixing ratio of the antigenpresenting cells to the T cells in the cocultivation is not restricted, and is usually about 1:1 to 1:100, preferably about 1:5 to 1:20 in terms of the ratio between the

numbers of cells. The density of the antigen-presenting cells to be suspended in the liquid medium is not restricted, and is usually about 100 to 10,000,000 cells/ml, preferably about 10,000 to 1,000,000 cells/ml. The cocultivation is preferably carried out at 37°C under atmosphere of 5% CO₂ in accordance with a conventional method. The culturing time is not restricted, and is usually 2 days to 3 weeks, preferably about 4 days to 2 weeks. The cocultivation is preferably carried out in the presence of one or more interleukins such as IL-2, IL-6, IL-7 and/or IL-12. In this case, the concentration of IL-2 and IL-7 is usually about 5 U/ml to 20 U/ml, the concentration of IL-6 is usually about 500 U/ml to 2000 U/ml, and the concentration of IL-12 is usually about 5 ng/ml to 20 ng/ml, but the concentrations of the interleukins are not restricted thereto. Here, "U" indicates the unit of activity. The above cocultivation may be repeated once to several times adding fresh antigenpresenting cells. For example, the operation of discarding the culture supernatant after the cocultivation and adding a fresh suspension of antigen-presenting cells to further conduct the cocultivation may be repeated once to several times. The

15 further conduct the cocultivation may be repeated once to several times. The conditions of the each cocultivation may be the same as described above.
[0077]

By the above-described cocultivation, cytotoxic T cells specific to the polypeptide are induced and allowed to proliferate. Thus, using the above-described polypeptide, isolated T cells can be prepared which selectively bind the complex between the polypeptide and the MHC molecule.

[0078]

As described in the Examples below, the genes encoding the polypeptides of SEQ ID NOs:3, 5, 7, 9, 11, 13, 15, ..., 93 and 95 are expressed specifically in breast cancer cells, leukemia cells and lymphoma cells. Therefore, it is thought that, in these cancer species, significantly higher numbers of the polypeptides of SEQ ID NOs:3, 5, 7, 9, 11, 13, 15, ..., 93 and 95 exist than in normal cells. When cytotoxic

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T cells prepared as described above are administered to a living body while a part of the polypeptides existing in cancer cells are presented by MHC molecules on the surfaces of the cancer cells, the cytotoxic T cells can damage the cancer cells using the presented polypeptides as markers. Since antigen-presenting cells presenting the above-described polypeptides can induce, and allow proliferation of, cytotoxic T cells specific to the polypeptides also in vivo, cancer cells can be damaged also by administering the antigen-presenting cells to a living body. That is, the cytotoxic T cells and the antigen-presenting cells prepared using the polypeptide are also effective as therapeutic and/or prophylactic agents for cancer, similarly to the immunity-inducing agent of the present invention.

[0079]

In cases where the above-described isolated antigen-presenting cells or isolated T cells are administered to a living body, these are preferably prepared by treating antigen presenting cells or T cells collected from the patient to be treated with the polypeptide (a) to (c) as described above in order to avoid the immune response in the living body that attacks these cells as foreign bodies. [0080]

The therapeutic and/or prophylactic agent for cancer comprising as an effective ingredient the antigen-presenting cells or T cells is preferably administered via a parenteral administration route such as intravenous or intraarterial administration. The dose is appropriately selected depending on the symptom, the purpose of administration and the like, and is usually 1 cell to 10,000,000,000,000 cells, preferably 1,000,000 cells to 1,000,000,000 cells, which dose is preferably administered once per several days to once per several months. The formulation may be, for example, the cells suspended in physiological buffered saline, and the formulation may be used in combination with another/other anticancer preparation(s) and/or cytokine(s). Further, one or more additives well-known in the field of

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formulation of pharmaceuticals may also be added.

[0081]

<Gene Vaccine>

Also by expression of the polynucleotide encoding the polypeptide (a) to (c) in the body of the subject animal, antibody production and cytotoxic T cells can be induced in the living body, and an effect comparable to that obtained in the case of administration of a polypeptide can be obtained. That is, the immunity-inducing agent of the present invention may be one comprising as an effective ingredient a recombinant vector having a polynucleotide encoding the polynucleotide (a) to (c), which recombinant vector is capable of expressing the polypeptide in a living body.

10 which recombinant vector is capable of expressing the polypeptide in a living body. Such a recombinant vector capable of expressing an antigenic polypeptide is also called gene vaccine.

[0082]

The vector used for production of a gene vaccine is not restricted as long as it 15 is a vector capable of expressing a polypeptide in a cell of the subject animal (preferably in a mammalian cell), and may be either a plasmid vector or a virus vector, and any known vector in the field of gene vaccines may be used. The polynucleotide such as DNA or RNA encoding the above-described polypeptide can be easily prepared, as mentioned above, by a conventional method. Incorporation of 20 the polynucleotide into the vector can be carried out using a method well-known to those skilled in the art.

[0083]

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The administration route of the gene vaccine is preferably a parenteral route such as intramuscular, subcutaneous, intravenous or intraarterial administration, and the dose may be appropriately selected depending on the type of the antigen and the like, and usually about 0.1 μ g to 100 mg, preferably about 1 μ g to 10 mg in terms of the weight of the gene vaccine per 1 kg of body weight.

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[0084]

Methods using a virus vector include those wherein a polynucleotide encoding the above-described polypeptide is incorporated into an RNA virus or DNA virus, such as a retrovirus, adenovirus, adeno-associated virus, herpes virus, vaccinia virus, pox virus, poliovirus or Sindbis virus, and then the subject animal is infected with the resulting virus. Among these methods, those using a retrovirus, adenovirus, adenoassociated virus, vaccinia virus or the like are especially preferred.

[0085]

Examples of other methods include a method wherein an expression plasmid is directly intramuscularly administered (DNA vaccine method), the liposome method, lipofectin method, microinjection method, calcium phosphate method and electroporation method, and the DNA vaccine method and liposome method are especially preferred.

Methods for actually making the gene encoding the above-described

polypeptide used in the present invention act as a pharmaceutical include the in vivo

method wherein the gene is directly transfected into the body, and the ex vivo

[0086]

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method wherein a kind of cells are collected from the subject animal and the gene is transfected into the cells ex vivo, followed by returning the cells to the body (Nikkei Science, 1994, April, p. 20-45; The Pharmaceutical Monthly, 1994, Vol. 36, No. 1, p. 23-48; Experimental Medicine, Extra Edition, 1994, Vol.12, No. 15; and references cited in these papers and the like). The in vivo method is more preferred.

[0087]

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In cases where the gene is administered by the in vivo method, the gene may be administered through an appropriate administration route depending on the disease to be treated, symptom and so on. It may be administered by, for example,

intravenous, intraarterial, subcutaneous, intramuscular administration or the like, or may be directly administered to the affected area in which a tumor exists. In cases where the gene is administered by the in vivo method, the gene may be formulated into a preparation such as a solution, and in general, it is formulated into an injection solution or the like containing DNA encoding the above-described peptide of the present invention as an effective ingredient. A commonly used carrier(s) may be added thereto as required. In the case of a liposome or membrane fusion liposome (Sendai virus (HVJ)-liposome or the like) containing the DNA, the liposome may be formulated into a liposome preparation such as a suspension, frozen preparation or centrifugally concentrated frozen preparation.

[0088]

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In the present invention, "the base sequence shown in SEQ ID NO:1" includes not only the base sequence expressly written in SEQ ID NO:1, but also the sequence complementary thereto. Thus, "a polynucleotide having the base sequence shown in SEQ ID NO:1" includes a single-stranded polynucleotide having the base sequence expressly written in SEQ ID NO:1, a single-stranded polynucleotide having the base sequence complementary thereto, and a double-stranded polynucleotide composed of these single-stranded polynucleotides. When the polynucleotide encoding the polypeptide used in the present invention is prepared, any one of these base sequences should be appropriately selected, and those skilled in the art can easily carry out the selection.

[0089]

<Detection of Cancer>

In a method of the present invention for detection of cancer, expression of the 25 polypeptide used in the present invention is measured using a sample separated from a living body. The method for measuring the expression of a polypeptide using the sample includes a method in which an antibody against the polypeptide, which

antibody is contained in the sample, is measured by immunoassay (Method 1); a method in which the polypeptide per se contained in the sample is measured by immunoassay (Method 2); and a method in which mRNA contained in the sample which encodes the polypeptide is measured (Method 3). In the method of the

5 present invention, the expression of the polypeptide may be measured by any of these three methods. In the present invention, the term "measurement" includes detection, quantification and semi-quantification.

[0090]

Here, CD179b was identified as a polypeptide which binds to an antibody 10 (cancer-specific antibody) specifically existing in serum derived from a tumorbearing dog, by the SEREX method using serum from a canine patient from which a canine breast cancer-derived cDNA library was prepared (see Example 1). That is, in the living body of a tumor-bearing dog, an antibody against CD179b is specifically induced. Thus, also by measuring an antibody against CD179b in a tumor-bearing 15 living body, a cancer expressing CD179b can be detected (see Example 7). Further, a canine cancer can be detected also by measuring CD179b as an antigen by the above Method 2. Further, since, as described in the Examples below, mRNA encoding the antigen polypeptide is significantly more highly expressed in cancer, especially in breast cancer and leukemia cells, than in normal tissues (see Example 1), 20 a canine cancer can be detected also by measuring the mRNA. As mentioned above, CD179b is known to be expressed on the membrane surfaces of precursor cells of B cells (pre-B cells), and therefore it is reported that CD179b is expressed in leukemia (pre-B cell leukemia) cells derived by cancerization of pre-B cells, but the fact that leukemia cells other than pre-B cell leukemia cells and breast cancer cells show 25 expression of CD179b was first discovered in the present invention. Accordingly, detection of leukemia other than pre-B cell leukemia cells, lymphoma and breast cancer became possible by investigating expression of CD179b.

[0091]

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In Method 1 above, measurement of the cancer-specific antibody which may exist in the sample can be easily carried out by immunoassay using an antigenic substance which immunologically reacts with the antibody. The immunoassay per se is a conventional well-known method as explained in detail below. As the antigenic substance which may be used in the immunoassay, the polypeptide (a) to (c) may be used. As antibodies have cross-reactivity, a molecule may be bound to an antibody which is induced against another immunogen, as long as the molecule has any structure thereon which is similar to the epitope of the immunogen. For example, polypeptides having high amino acid sequence homology to each other often have epitopes with similar structures, and in such cases the both polypeptides may have the same antigenicity. As concretely described in the Examples below, the human-derived polypeptide of SEQ ID NO:3 immunologically reacts with the antibody induced in the body of a tumor-bearing dog. Therefore, in Method 1 of the present invention, any mammalian homologous factor may be used as an antigen in the immunoassay.

[0092]

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structure, such as proteins, usually have a plurality of sites with different structures on their surface. Therefore, such an antigenic substance induces a plurality of kinds of antibodies which respectively recognize each of the sites in a living body. That is, an antibody induced in a living body against an antigenic substance such as a protein is a polyclonal antibody, which is a mixture of a plurality of kinds of antibodies. It should be noted that, in the present invention, the term "polyclonal antibody" means an antibody which exists in serum derived from a living body having an antigenic substance therein and is induced in the living body against the antigenic substance. [0093]

Antigenic substances having a large molecular weight and a complex

Measurement of the antibody in a sample may easily be carried out by immunoassay using the above-described polypeptide as an antigen. Immunoassays per se are well-known in the art, and includes, when classified based on the reaction mode, the sandwich method, competition method, agglutination method, Western blotting and the like. When classified based on the label, immunoassays include radioimmunoassay, fluorescence immunoassay, enzyme immunoassay, biotin immunoassay and the like, and the immunoassay of the above-described antibody

may be carried out by any of these immunoassays. Although not restricted, the sandwich ELISA and agglutination method may be preferably used as an

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- 10 immunoassay of the above antibody in the present invention, as these methods are simple and do not require a large-scale apparatus. In cases where an enzyme is used as a label of an antibody, the used enzyme is not particularly restricted as long as it satisfies such conditions that the turnover number is large, that the enzyme is stable even when it is bound to an antibody, that it specifically colors its substrate and the
- 15 like. For example, enzymes used in an ordinary enzyme immunoassay such as peroxidase, β -galactosidase, alkaline phosphatase, glucose oxidase, acetylcholinesterase, glucose-6-phosphate dehydrogenase, and malate dehydrogenase may be used. Enzyme inhibitors, coenzymes and the like may also be used. Binding of these enzymes with an antibody may be carried out by a known method 20 using a cross-linking agent such as a maleimide compound. As a substrate, known substances may be used depending on the kind of the used enzyme. For example, in cases where peroxidase is used as the enzyme, 3,3',5,5'-tetramethylbenzidine may be used; and in cases where alkaline phosphatase is used as the enzyme, paranitrophenol or the like may be used. As the radioisotope, those used in an ordinary 25 radioimmunoassay such as ¹²⁵I or ³H may be used. As the fluorescent dye, one used in an ordinary fluorescent antibody technique, such as fluorescein isothiocyanate (FITC), tetramethylrhodamine isothiocyanate (TRITC) or the like may be used.

[0094]

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These immunoassays per se are well-known in the art, and so it is not necessary to explain these immunoassays in the present specification. Briefly, in sandwich immunoassays, for example, the above-mentioned polypeptide used as an antigen is immobilized on a solid phase, and then reacted with a sample such as a serum. After washing the solid phase, the resultant is reacted with an appropriate secondary antibody. After washing the solid phase, the secondary antibody bound to the solid phase is measured. In the method for detecting cancer according to the present invention, it is preferred to immobilize an antigen polypeptide on a solid phase, because immobilization on a solid phase makes it possible to easily remove the unbound secondary antibody. As the secondary antibody, for example, anti-dog IgG antibody may be used in cases where the sample is obtained from dogs. The secondary antibody bound to the solid phase may be measured by labeling the secondary antibody with a labeling substance exemplified above. The thus measured amount of the secondary antibody corresponds to the amount of the abovementioned antibody in a serum sample. In cases where an enzyme is used as the labeling substance, the amount of the antibody may be measured by adding a substrate which is decomposed by the enzymatic activity to develop a color, and then optically measuring the amount of decomposed substrate. In cases where a radioisotope is used as the labeling substance, the amount of radiation from the radioisotope may be measured with a scintillation counter or the like.

[0095]

In Method 2 of the present invention, the polypeptide shown in SEQ ID NO:3, 5, 7, 9, 11, 13, 15, ..., 93 or 95 is measured, which polypeptide may be contained in the sample obtained from a living body. As explained above, the abundance of the cancer-specific antibody which immunologically reacts with the polypeptide shown in SEQ ID NO:3, 5, 7, 9, 11, 13, 15, ..., 93 or 95 or a homologous factor thereof is

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significantly high in cancer patients, which indicates that the production of the polypeptide or a homologous factor thereof, which is the antigen of the cancer-specific antibody, is significantly high in the cancer patients. Therefore, similarly to Method 1 above, cancers in a living body can be detected by measuring the polypeptide shown in SEQ ID NO:3, 5, 7, 9, 11, 13, 15, ..., 93 or 95 or a homologous factor thereof.

[0096]

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Measurement of the polypeptide in a sample may easily be carried out by a well-known immunoassay. Specifically, for example, the polypeptide having the amino acid sequence shown in the odd number ID of SEQ ID NOs:3 to 95 or a homologous factor thereof which may exist in a sample may be measured by preparing an antibody or antigen-binding fragment thereof which immunologically reacts with the polypeptide having the amino acid sequence shown in the odd number ID of SEQ ID NOs:3 to 95 or a homologous factor thereof, and then carrying out an immunoassay using the prepared antibody or fragment thereof. The immunoassay per se is a well-known conventional method as described above.

[0097]

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The term "antigen-binding fragment" herein means an antibody fragment such as the Fab fragment or the F(ab')₂ fragment contained in an antibody molecule, which has a binding capacity to an antigen. Although the antibody may be either a polyclonal antibody or monoclonal antibody, a monoclonal antibody is preferred for immunoassays and the like, because a high reproducibility is attained therewith. Methods for preparing a polyclonal or monoclonal antibody using a polypeptide as an immunogen are well-known, and the preparation may be easily carried out by a conventional method. For example, antibodies against the polypeptide may be induced by immunizing an animal with an immunogen, the polypeptide conjugated to a carrier protein such as keyhole limpet hemocyanin (KLH) or casein, together with

an adjuvant. Then antibody-producing cells such as spleen cells or lymphocytes are collected from the immunized animal and fused with myeloma cells to prepare hybridomas. Among the hybridomas, one producing an antibody which binds to the polypeptide shown in SEQ ID NO:3, 5, 7, 9, 11, 13, 15, ..., 93 or 95 or a

5 homologous factor thereof is selected and proliferated, and then the monoclonal antibody whose corresponding antigen is the above-mentioned protein may be collected from the culture supernatant. The above-described method is a conventional well-known method.

[0098]

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In Method 3 of the present invention, mRNA encoding CD179b, which may be contained in a sample obtained from a living body, is measured. As concretely described in the Examples below, the expression level of mRNA encoding CD179b is significantly high in cancer, especially, breast cancer and leukemia cells. Therefore, cancers in a living body can be detected by measuring the mRNA in a sample.

[0099]

In the detection method of the present invention, whether the subject living body suffers from cancer or not or the like is determined based on the expression level of the polypeptide measured as described above. Although the cancer detection may be attained simply by measuring the expression of the polypeptide in the subject living body, it is preferred to obtain the normal reference value by determining the expression level of the polypeptide (the amount of the antibody, polypeptide or mRNA) in one or more samples from healthy individuals to compare the measured value in the subject living body with the normal reference value, in view of increasing the detection accuracy. In order to further increase the detection accuracy, the cancer reference value may be obtained by determining the expression level of the polypeptide in samples obtained from many patients who have been revealed to suffer from cancer to compare the measured value of the subject living body with the both of the normal and cancer reference values. The above mentioned reference values may be determined by expressing the expression level of the polypeptide in each sample in values and calculating the average value thereof.

5 The normal and cancer reference values may be determined beforehand by measuring the expression level of the polypeptide in many healthy and cancer subjects. Thus, when the measured value is compared with the reference values in the method of the present invention, the reference values may be those predetermined.

[0100]

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The detection method of the present invention may be carried out in combination with detection using other cancer antigens and/or cancer markers so that the detection accuracy of cancers can be more improved.

[0101]

By the detection method of the present invention, cancers in a living body can be detected. The method of the present invention can detect even an invisible small tumor or a tumor which exists in a deep part of a body, and thus the method is useful for early detection of cancer. Further, by applying the detection method of the present invention to patients in the follow-up period after cancer therapy, the recurrent cancer, if any, can be detect in its early stage.

20 [0102]

If the more cancer cells expressing the prescribed polypeptide to be measured in the present invention proliferate in a tumor-bearing living body, the more the polypeptides and mRNAs encoding them accumulate in the body, which causes the increased amount of the antibodies against the above-mentioned polypeptides in the serum. On the other hand, the more cancer cells decrease, the more the accumulated polypeptides and mRNAs encoding them decrease in the living body, which causes the decreased amount of the antibodies against the above-mentioned

polypeptides in the serum. Thus, if the expression level of the prescribed polypeptide is high, it can be determined that tumor growth and/or metastasis of cancer occurred, i.e., the stage of progression of cancer is advanced. [0103]

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Further, as shown in the Example below, when compared between the same kinds of tumors, a malignant one produces significantly higher amount of the antibodies than a benign one. Therefore, if the expression level of the prescribed polypeptides is high, it can be determined that the grade of cancer malignancy is higher. That is, the grade of cancer malignancy can also be detected by the method of the present invention.

[0104]

Furthermore, the effect of the cancer therapy can be monitored based on the increase or decrease in the expression level of the prescribed polypeptides. Therefore, by observing the expression level of the above-mentioned polypeptides on an individual during or after cancer therapy, a clue to assess how much the administered anti-cancer agent was effective, or whether a portion of the tumor is left in the patient after extirpation of the tumor can be obtained, as well as a clue to find metastasis and/or recurrence as early as possible can be obtained during the follow-up. Appropriate treatment of cancer results in decrease in the expression level of the polypeptides compared to that in the tumor-bearing state before the therapy. In such a case, it can be judged that the effect of the therapy which was (is being) performed on the living body is/was good. In cases where the expression level of the polypeptides increases or is sustained, or once decreases and then increases, it can be judged that the effect of the therapy which was the useful basis for selection of a therapeutic method, such as decision to change the therapeutic method or to change the dose of an anti-cancer agent.

[0105]

Cancers to be detected by the method of the present invention are those expressing CD179b (excluding pre-B cell tumors), and examples thereof include, but are not limited to, mammary gland cancer, combined mammary gland cancer, mammary gland malignant mixed tumor, intraductal papillary adenocarcinoma,

leukemias (preferably, chronic lymphocytic leukemia excluding those of the pre-B
cell type) and lymphomas (preferably, gastrointestinal lymphoma, digestive organ
lymphoma, small/medium cell lymphoma, medium cell lymphoma and multicentric
lymphoma, excluding those of the pre-B cell type). The living bodies to which the
method of the present invention applies are mammals, preferably humans, dogs and
cats.

[0106]

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The sample to be subjected to the method of the present invention includes body fluids such as blood, serum, plasma, ascites and pleural effusion; tissues; and cells. In particular, serum, plasma, ascites and pleural effusion may be preferably used in Method 1 and Method 2 above. A tissue sample and cell sample are preferred in the case of Method 3 above in which mRNA is measured.

[0107]

The polypeptide used as an antigen for immunoassay in Method 1 may be provided as a reagent for detecting cancer. The reagent may consist only of the above-mentioned polypeptide, or may contain various additives useful for stabilizing the polypeptide, and the like. The reagent may also be provided in the form of being immobilized on a solid phase such as a plate or membrane.

[0108]

The antibody or an antigen-binding fragment thereof which immunologically reacts with the polypeptide of SEQ ID NO:3, 5, 7, 9, 11, 13, 15, ..., 93 or 95 or a homologous factor thereof, which is used for measuring the polypeptide or the homologous factor thereof by immunoassay in Method 2, may also be provided as a

reagent for detecting cancer. The reagent may also consist only of the abovementioned antibody or antigen-binding fragment thereof, or may contain various additives useful for stabilizing the antibody or antigen-binding fragment thereof, and the like. The antibody or antigen-binding fragment thereof may also be in the form of being conjugated with a metal such as manganese or iron. Since such a metalconjugated antibody or antigen-binding fragment thereof accumulates in a site in which a large amount of antigen protein exists when administered to a body, the existence of cancer cells which produce the antigen protein can be detected by measuring the metal by MRI or the like.

10 [0109]

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Furthermore, the above-described polynucleotide for cancer detection used for measuring mRNA in Method 3 may also be provided as a reagent for detecting cancer. The reagent for detecting cancer may also consist only of the polynucleotide, or may contain various additives useful for stabilizing the polynucleotide and the like.

15 The polynucleotide for cancer detection contained in the reagent is preferably a primer or a probe.

EXAMPLES

[0110]

The present invention will now be described more concretely by way of Examples, but the scope of the present invention is not limited to the particular examples below.

[0111]

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Example 1: Acquisition of Novel Cancer Antigen Protein by SEREX Method (1) Preparation of cDNA Library

From a canine mammary gland cancer tissue removed by surgery, total RNA was extracted by the Acid guanidium-Phenol-Chloroform method, and poly(A)⁺ RNA was purified using the Oligotex-dT30 mRNA purification Kit (manufactured by

Takara Shuzo Co., Ltd.) according to the protocol described in the attached instructions.

[0112]

Using the obtained mRNA (5 μ g), a canine mammary gland cancer-derived 5 cDNA phage library was synthesized. Preparation of the cDNA phage library was carried out using cDNA Synthesis Kit, ZAP-cDNA Synthesis Kit, and ZAP-cDNA Gigapack III Gold Cloning Kit (manufactured by STRATAGENE) in accordance with the protocols attached to the kits. The size of the prepared cDNA phage library was 2.99 × 10⁵ pfu/ml.

10 [0113]

(2) Screening of cDNA Library with Serum

Using the canine mammary gland cancer-derived cDNA phage library prepared as described above, immunoscreening was carried out. More particularly, host *E. coli* (XL1-Blue MRF') was infected with the library such that 2340 clones were included in a Φ90×15 mm NZY agarose plate, followed by culture at 42°C for 3 to 4 hours to allow formation of plaques. The plate was covered with a nitrocellulose membrane (Hybond C Extra; manufactured by GE Healthcare Bio-Science) impregnated with IPTG (isopropyl-β-D-thiogalactoside) at 37°C for 4 hours, to allow induction and expression of proteins, thereby transferring the proteins to the membrane. Thereafter, the membrane was recovered and soaked in TBS (10 mM Tris-HCl, 150 mM NaCl pH 7.5) supplemented with 0.5% non-fat dry milk, followed by being shaken at 4°C overnight to suppress nonspecific reactions. This filter was allowed to react with 500-fold diluted canine patient serum at room temperature for 2 to 3 hours.

25 [0114]

As the above-described canine patient serum, a total of 3 serum samples were used which were collected from each of the dog from which the above mammary

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gland cancer was removed and another mammary gland cancer canine patient. These sera were stored at -80°C and pretreated immediately before use. The pretreatment of the sera was carried out by the following method. That is, host E. *coli* (XL1-BLue MRF') was infected with λ ZAP Express phage into which no exogenous gene was inserted, and cultured on an NZY plate at 37°C overnight. Subsequently, 0.2 M NaHCO₃ buffer (pH 8.3) containing 0.5 M NaCl was added to the plate, and the plate was left to stand at 4°C for 15 hours, followed by recovering the supernatant as an E. coli/phage extract. Thereafter, the recovered E. coli/phage extract was passed through an NHS-column (manufactured by GE Healthcare Bio-Science) to immobilize the proteins derived from the *E. coli*/phage. The serum from the canine patient was passed through this protein-immobilized column and allowed to react with the proteins, thereby removing antibodies that adsorb to E. coli and the phage from the serum. The serum fraction passed through the column without being adsorbed was 500-fold diluted with TBS supplemented with 0.5% nonfat dry milk, and the resulting dilution was used as a material for the immunoscreening.

[0115]

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The membrane to which the thus treated serum and the above-described fusion proteins were blotted was washed with TBS-T (0.05% Tween 20/TBS) 4 times, and goat anti-dog IgG (Goat anti Dog IgG-h+I HRP conjugated; manufactured by BETHYL Laboratories, Inc.) which was 5000-fold diluted with TBS supplemented with 0.5% non-fat dry milk was allowed, as a secondary antibody, to react at room temperature for 1 hour. Detection was carried out by an enzymatic coloring reaction using the NBT/BCIP reaction solution (manufactured by Roche), and colonies whose positions were identical to those of positive sites of the coloring reaction were collected from the Φ90×15mm NZY agarose plate, and dissolved into 500 µl of SM buffer (100 mM NaCl, 10 mM MgCISO₄, 50 mM Tris-HCl, 0.01% gelatin, pH7.5).

The second and third screenings were carried out by repeating the same method as described above until the colonies positive in the coloring reaction became single colonies, thereby isolating 45 positive clones after screening of 92820 phage clones reactive with IgG in the serum.

5 [0116]

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(3) Homology Search of Isolated Antigen Genes

To subject the 45 positive clones isolated by the above method to sequence analysis, an operation to convert the phage vector to a plasmid vector was carried out. More particularly, 200 μ l of a solution prepared such that the host *E. coli* (XL1-Blue MRF') was contained to an absorbance OD₆₀₀ of 1.0, 250 μ l of the purified phage solution and 1 μ l of ExAssist helper phage (manufactured by STRATAGENE) were mixed together, and the resulting mixture was allowed to react at 37°C for 15 minutes, followed by adding 3 ml of LB broth thereto and culturing the resultant at 37°C for 2.5 to 3 hours. This was immediately followed by 20 minutes of incubation in a water bath at 70°C and centrifugation at 1000×g for 15 minutes, after

which the supernatant was collected as a phagemid solution. Subsequently, 200 μ l of a solution prepared such that the phagemid host *E. coli* (SOLR) was contained to an absorbance OD₆₀₀ of 1.0 and 10 μ l of the purified phagemid solution were mixed together, and the resulting mixture was allowed to react at 37°C for 15 minutes,

followed by plating a 50 µl aliquot of the resultant on LB agar medium supplemented with ampicillin (50 µg/ml final concentration) and culturing at 37°C overnight.
Single colonies of the transformed SOLR were picked up and cultured in LB medium supplemented with ampicillin (50 µg/ml final concentration) at 37°C, followed by purifying plasmid DNAs having inserts of interest using QIAGEN plasmid Miniprep Kit (manufactured by QIAGEN).

[0117]

Each purified plasmid was subjected to analysis of the full-length sequence of

the insert by the primer walking method using the T3 primer shown in SEQ ID NO:96 and the T7 primer shown in SEQ ID NO:97. By this sequence analysis, the gene sequences shown in the even number IDs of SEQ ID NOs:4 to 92 were obtained. Using the base sequences and the amino acid sequences (odd number IDs of SEQ ID NOs: 5 to 93) of these genes, homology search against known genes were carried out using a homology search program BLAST (http://www.ncbi.nlm.nih.gov/BLAST/), and, as a result, it was revealed that all the obtained 45 genes were those encoding CD179b. The homologies among the 45 genes were 94 to 99% in terms of the base sequences and 96 to 99% in terms of the amino acid sequences. The homologies between these genes and the gene encoding a human homologous factor were 62 to 82% in terms of the base sequences and 69 to 80% in terms of the amino acid sequences, in the region translated to a protein. The base sequence of the human homologous factor is shown in SEQ ID NO:1, and the amino acid sequences of the human homologous factor are shown in SEQ ID NOs:2 and 3. Further, the homologies between these genes and the gene encoding a bovine homologous factor

- 15 homologies between these genes and the gene encoding a bovine homologous factor were 68 to 82% in terms of the base sequences and 56 to 77% in terms of the amino acid sequences, in the region translated to a protein. The base sequence of the bovine homologous factor is shown in SEQ ID NO:94, and the amino acid sequence of the bovine homologous factor is shown in SEQ ID NO:95. The homology
- 20 between the gene encoding the human homologous factor and the gene encoding the bovine homologous factor was 62% in terms of the base sequences and 72% in terms of the amino acid sequences, in the region translated to a protein.

[0118]

(4) Analysis of Expression in Various Tissues

Expressions of the genes obtained by the above method in canine and human normal tissues and various cell lines were investigated by the RT-PCR (Reverse Transcription-PCR) method. The reverse transcription reaction was carried out as

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follows. That is, from 50 to 100 mg of each tissue or $5-10 \times 10^6$ cells of each cell line, total RNA was extracted using the TRIZOL reagent (manufactured by INVITROGEN) according to the protocol described in the attached instructions. Using this total RNA, cDNA was synthesized by the Superscript First-Strand

5 Synthesis System for RT-PCR (manufactured by INVITROGEN) according to the protocol described in the attached instructions. As the cDNAs of human normal tissues (brain, hippocampus, testis, colon and placenta), Gene Pool cDNA (manufactured by INVITROGEN), QUICK-Clone cDNA (manufactured by CLONETECH) and Large-Insert cDNA Library (manufactured by CLONETECH) 10 were used. The PCR reaction was carried out as follows, using primers specific to the obtained canine genes (shown in SEQ ID NOs:98 and 99) and their human homologous gene (shown in SEO ID NOs:100 and 101). That is, reagents and an attached buffer were mixed such that concentrations/amounts of $0.25 \,\mu$ of a sample prepared by the reverse transcription reaction, $2 \mu M$ each of the above primers, 0.2 15 mM each of dNTPs, and 0.65 U ExTaq polymerase (manufactured by Takara Shuzo Co., Ltd.) were attained in a total volume of 25 μ l, and the reaction was carried out with 30 cycles of 94°C for 30 seconds, 60°C for 30 seconds and 72°C for 30 seconds using a Thermal Cycler (manufactured by BIO RAD). The above-described primers specific to genes having the base sequences shown in SEQ ID NOs: 98 and 99 were 20 for amplification of the positions 32 to 341 in the base sequence shown in SEQ ID NO:4, and for amplification of the region common to all the canine CD179b genes shown in the even number IDs of SEQ ID NOs: 4 to 92. Further, the primers specific to genes having the base sequences shown in SEQ ID NOs:100 and 101 were for amplification of the positions 216 to 738 in the base sequence shown in SEQ ID 25 NO:1. As a control for comparison, primers specific to GAPDH (shown in SEO ID NOs:102 and 103) were used at the same time. As a result, as shown in Fig. 1, the obtained canine genes did not show expression in normal canine tissues at all, but

showed strong expression in canine breast cancer tissues. In terms of expression of the human homologous gene, bone marrow was the only human normal tissue wherein its expression was confirmed, but, in human cancer cells, its expression was detected in leukemia cell lines and breast cancer cell lines, so that specific expression of CD179b in the leukemia cell lines and the breast cancer cell lines was confirmed. [0119]

In Fig. 1, reference numeral 1 in the ordinate represents the expression pattern of the gene identified as described above, and reference numeral 2 represents the expression pattern of the GAPDH gene as the control for comparison.

10 [0120]

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Example 2: Preparation of Canine and Human Novel Cancer Antigen Protein

(1) Preparation of Recombinant Protein

Based on the gene of SEQ ID NO:4 obtained in Example 1, a recombinant protein was prepared by the following method. That is, reagents and an attached 15 buffer were mixed such that concentrations/amounts of 1 µl of the vector prepared from the phagemid solution obtained in Example 1 and subjected to the sequence analysis, 0.4 μ M each of two kinds of primers having Ndel and Kpnl restriction sites (described in SEQ ID NOs:104 and 105), 0.2 mM dNTP, and 1.25 U PrimeSTAR HS polymerase (manufactured by Takara Shuzo Co., Ltd.) were attained in a total 20 volume of 50 µl, and PCR was carried out with 30 cycles of 98°C for 10 seconds and 68°C for 40 seconds using a Thermal Cycler (manufactured by BIO RAD). The above-described two kinds of primers were those for amplification of the region encoding the 5th to 120th amino acids in the amino acid sequence shown in SEQ ID NO:5. After the PCR, the amplified DNA was subjected to electrophoresis using 25 2% agarose gel, and a DNA fragment of about 350 bp was purified using OIAquick Gel Extraction Kit (manufactured by QIAGEN). [0121]

The purified DNA fragment was ligated into a cloning vector pCR-Blunt (manufactured by Invitrogen). *E. coli* was transformed with the resulting ligation product, and plasmids were recovered thereafter, followed by confirming, by sequencing, that the sequence of the amplified gene fragment matches the sequence

of interest. The plasmid having the sequence that matched the sequence of interest was treated with restriction enzymes *Ndel* and *Kpnl* and purified using QIAquick Gel Extraction Kit, followed by inserting the gene sequence of interest into an expression vector for *E. coli*, pET30b (manufactured by Novagen) that had been treated with restriction enzymes *Ndel* and *Kpnl*. Usage of this vector enables production of a
His-tag fusion recombinant protein. *E. coli* for expression, BL21 (DE3), was transformed with this plasmid, and expression of the protein of interest was induced in *E. coli* with 1 mM IPTG.

[0122]

On the other hand, based on the gene of SEQ ID NO:1, a recombinant protein 15 of the human homologous gene was prepared by the following method. Reagents and an attached buffer were mixed such that concentrations/amounts 1 µl of the cDNA prepared in Example 1 whose expression could be confirmed by the RT-PCR method in cDNAs from various tissues/cells, 0.4 µM each of two kinds of primers having EcoRI and Sall restriction sites (described in SEQ ID NOs:106 and 107), 0.2 20 mM dNTP, and 1.25 U PrimeSTAR HS polymerase (manufactured by Takara Shuzo Co., Ltd.) were attained in a total volume of 50 μ l, and PCR was carried out with 30 cycles of 98°C for 10 seconds and 68°C for 40 seconds using a Thermal Cycler (manufactured by BIO RAD). The above-described two kinds of primers were those for amplification of the total length the amino acid sequence shown in SEQ ID 25 NO:3. After the PCR, the amplified DNA was subjected to electrophoresis using 2% agarose gel, and a DNA fragment of about 540 bp was purified using QIAquick Gel Extraction Kit (manufactured by QIAGEN).

[0123]

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The purified DNA fragment was ligated into a cloning vector pCR-Blunt (manufactured by Invitrogen). *E. coli* was transformed with the resulting ligation product, and plasmids were recovered thereafter, followed by confirming, by sequencing, that the sequence of the amplified gene fragment matches the sequence of interest. The plasmid having the sequence that matched the sequence of interest was treated with restriction enzymes *Eco*RI and *Sal*I and purified using QIAquick Gel Extraction Kit, followed by inserting the gene sequence of interest into an expression vector for *E. coli*, pET30a (manufactured by Novagen) that had been treated with restriction enzymes *Eco*RI and *Sal*I. Usage of this vector enables production of a His-tag fusion recombinant protein. *E. coli* for expression, BL21 (DE3), was transformed with this plasmid, and expression of the protein of interest was induced in *E. coli* with 1 mM IPTG.

[0124]

15 (2) Purification of Recombinant Protein

The above-obtained recombinant *E. coli* cells that expresse SEQ ID NO:1 and SEQ ID NO:4, respectively, were cultured in LB medium supplemented with 30 μ g/mL kanamycin at 37°C until the absorbance at 600 nm reached about 0.7, and then isopropyl- β -D-1-thiogalactopyranoside was added thereto such that its final concentration should be 1 mM, followed by culturing them at 30°C for 20 hours. Subsequently, the cells were collected by centrifugation at 4,800 rpm for 10 minutes. The pellet of the cells was suspended in phosphate-buffered saline and further subjected to centrifugation at 4,800 rpm for 10 minutes to wash the cells.

[0125]

The cells were suspended in phosphate-buffered saline and subjected to sonication on ice. The sonicated solution of *E. coli* was centrifuged at 7,000 rpm for 20 minutes to obtain the supernatant as the soluble fraction and the precipitate as

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the insoluble fraction.

[0126]

The insoluble fraction was suspended in 4% Triton-X100 solution and centrifuged at 7,000 rpm for 20 minutes. This operation was repeated twice and an operation of removal of proteases was carried out.

[0127]

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The residue was suspended in 20 mM phosphate buffer (pH 8.0) containing 6M guanidine hydrochloride, and the resulting suspension was left to stand at 4°C for 20 hours to denature proteins. Thereafter, the suspension was centrifuged at 7,000 rpm for 20 minutes, and the obtained soluble fraction was placed in a nickel chelate column prepared by a conventional method (carrier: Chelating Sepharose (trademark) Fast Flow (GE Health Care); column volume: 5 mL; equilibration buffer: 20 mM phosphate buffer (pH 8.0) containing 6M guanidine hydrochloride). The fraction that was not adsorbed to the column was washed away with 10 column volumes of 20 mM phosphate buffer (pH 8.0) containing 10 mM imidazole, and elution was immediately carried out with a four-step density gradient of 50 mM-500 mM imidazole, to obtain a purified fraction, which was used thereafter as a material for administration tests.

20 [0128]

To 1 ml of a reaction buffer (20 mM Tris-HCl, 50 mM NaCl, 2 mM CaCl₂; pH 7.4), 200 μ l of the purified preparation obtained by the above-described method was aliquoted, and 2 μ l of enterokinase (manufactured by Novagen) was then added thereto, followed by leaving it to stand at room temperature overnight to cleave His tag. The resulting product was purified using Enterokinase Cleavage Capture Kit (manufactured by Novagen) in accordance with the protocol attached to the kit. Subsequently, the buffer contained in 1.2 ml of the purified preparation obtained by

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the above-described method was replaced with physiological phosphate buffer (manufactured by Nissui Pharmaceutical) by ultrafiltration using NANOSEP 10K OMEGA (manufactured by PALL), and the resulting solution was filtered aseptically using HT Tuffryn Acrodisc 0.22 µm (manufactured by PALL) and used in the

5 following experiments.

[0129]

Example 3: Test of Administration of Recombinant Protein to Cancer-bearing Dog (1) Antitumor Assay

The anti-tumor effect of the recombinant protein which was purified as 10 described above was assessed in a tumor-bearing dog (breast cancer) having an epidermal tumor.

[0130]

An equal amount of Freund's incomplete adjuvant (manufactured by Wako Pure Chemicals) was mixed with 100 μ g (0.5 ml) of the recombinant polypeptide 15 purified as described above, to prepare a therapeutic agent for cancer. This was administered to a regional lymph node in the vicinity of the tumor a total of 3 times, by carrying out the subsequent administrations 3 days and 7 days after the first administration. As a result, the tumor with a size of about 55 mm³ at the time of administration of the therapeutic agent for cancer was reduced in size to 30 mm³ 10 days after the first administration; to 16 mm³ 20 days after the first administration; and to 10 mm³ 30 days after the first administration.

[0131]

Further, to another canine patient suffering from mammary gland cancer, a mixture of 100 µg (0.5 ml) of the above-described polypeptide derived from dog and 0.5 ml of Freund's incomplete adjuvant was administered in the same manner as described above a total of 3 times. Further, concurrently with the respective administrations, 100 µg of canine interleukin 12 was administered subcutaneously.

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As a result, the tumor with a size of about 155 mm³ at the time of administration of the therapeutic agent for cancer completely regressed 24 days after the first administration.

[0132]

5 (2) Immune Inducibility Assay

Blood of the canine patient in which the anti-tumor effect was obtained in the administration test in the above-described (1) was collected before the administration of the therapeutic agent for cancer, and 10 days and 30 days after the first administration. Peripheral blood mononuclear cells were isolated according to a conventional method, and by the ELISPOT assay for IFNy using them, the immune inducibility of each administered recombinant protein was assayed.

[0133]

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In a 96-well plate manufactured by Millipore (MultiScreen-IP, MAIPS 4510), 100 µL/well of 70% ethanol was placed and the plate was left to stand for 5 minutes, 15 followed by removal of the ethanol by aspiration. The plate was washed with sterile water and 300 µL/well of 200 mM Sodium Bicarbonate (pH8.2) was placed therein. After leaving it to stand for 5 minutes, Sodium Bicarbonate was removed by aspiration, and then the plate was washed. Subsequently, 0.5 μ /well of anti-canine interferon y monoclonal antibody (manufactured by R&D, clone 142529, MAB781) 20 mixed with 200 mM Sodium Bicarbonate was placed in wells, and the plate was incubated at 37°C overnight to immobilize the primary antibody. After removal of the primary antibody by aspiration, 300 µL/well of a blocking solution (1% BSA-5% sucrose-200 mM Sodium Bicarbonate (pH8.2)) was added to the wells, and the plate was incubated at 4°C overnight to block the plate. After removal of the blocking 25 solution by aspiration, 300 µL/well of 10% fetal calf serum-containing RPMI medium (manufactured by Invitrogen) was placed in the wells, and the plate was left to stand for 5 minutes, followed by removal of the medium by aspiration.

Subsequently, 5×10^5 cells/well of the canine peripheral blood mononuclear cells suspended in 10% fetal calf serum-containing RPMI medium were placed in the plate, and 10 µL/well of the canine-derived polypeptide or human-derived polypeptide used in each administration was added thereto, followed by culturing the cells under the conditions of 37°C and 5% CO₂ for 24 hours, to allow immunocytes that might exist in the peripheral blood mononuclear cells to produce interferon v. After the culture, the medium was removed, and the wells were washed 6 times with a washing solution (0.1% Tween 20-200mM Sodium Bicarbonate (pH8.2)). In each well, 100 μ L of rabbit anti-dog polyclonal antibody 1000-fold diluted with the above-described blocking solution was placed, and the plate was incubated at 4°C overnight. After washing the wells 3 times with the above-described washing solution, 100 μ L of HRP-labeled anti-rabbit antibody 1000-fold diluted with the above-described blocking solution was placed in each well, and the reaction was allowed to proceed at 37°C for 2 hours. After washing the wells 3 times with the above-described washing solution, the resultant was colored with Konica Immunostain (manufactured by Konica), and the wells were washed with water to stop the reaction. Thereafter, the membrane was dried, and the number of the appeared spots was counted using KS ELISPOT (manufactured by Carl Zeiss, Inc.). As a result, in peripheral blood mononuclear cells sampled before the administration of the polypeptide, no spot was detected. On the other hand, in the canine patient after the administration of the polypeptide, 18 and 87 spots were detected in the peripheral blood mononuclear cells sampled 10 days and 30 days, respectively, after the administration.

[0134]

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From the above results, it was confirmed that immunocytes which specifically react with the administered recombinant protein and produce interferon γ were induced in the canine patient to which the recombinant protein was administered, and it was thought that the anti-tumor effect described in the above-described (1) was

exerted by immunoreactions in which these immunocytes were mainly involved. [0135]

Example 4: Induction of CD8-positive T Cells Reactive with Epitopes of CD179bderived Peptide

(1) Prediction of Peptide Motifs Which Bind to HLA-A0201 and HLA-A24

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Information on the amino acid sequence of the human CD179b protein was obtained from GenBank. For prediction of HLA-A0201 and HLA-A24 binding motifs, the amino acid sequence of the human CD179b protein was analyzed employing a computer-based prediction program using a known BIMAS software (available at http://bimas.dcrt.nih.gov/molbio/hla_bind/). As a result, 8 kinds of peptides shown in SEQ ID NOs:108 to 110 and SEQ ID NOs:113 to 117, which were expected to be capable of binding to the HLA-A0201 molecule; and 5 kinds of peptides shown in SEQ ID NOs:110 to 112, SEQ ID NO:115 and SEQ ID NO:116, which were expected to be capable of binding to the HLA-A24 molecule; were selected.

[0136]

(2) Induction of Peptide Epitope-reactive CD8-positive T Cells

From an HLA-A0201-positive healthy individual, peripheral blood was isolated, and the peripheral blood was overlaid on Lymphocyte separation medium (OrganonpTeknika, Durham, NC), followed by centrifugation thereof at 1,500 rpm at room temperature for 20 minutes. A PBMC-containing fraction was recovered and washed 3 times (or more) with cold phosphate buffer to obtain peripheral blood mononuclear cells (PBMCs). The obtained PBMCs were suspended in 20 ml of AIM-V medium (manufactured by Life Technologies, Inc., Grand Island, NY), and allowed to adhere to a culturing flask (manufactured by Falcon) at 37°C under 5% CO₂ for 2 hours. The cells which were not adhered were used for the preparation of T cells, and the adhered cells were used for the preparation of dendritic cells.

[0137]

The adhered cells were cultured in AIM-V medium in the presence of IL-4 (1000 U/ml) and GM-CSF (1000 U/ml). Six days later, the medium was replaced with AIM-V medium supplemented with IL-4 (1000 U/ml), GM-CSF (1000 U/ml), IL-6 (1000 U/ml, Genzyme, Cambridge, MA), IL-1ß (10 ng/ml, Genzyme, Cambridge, MA) and TNF- α (10 ng/ml, Genzyme, Cambridge, MA), and the culturing was continued for another 2 days. The obtained population of cells which did not adhere was used as the dendritic cells.

[0138]

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The prepared dendritic cells were suspended in AIM-V medium at a cell density of 1×10⁶ cells/ml, and the peptide shown in SEQ ID NOs:108 to 110 or SEQ ID NOs:113 to 117, which are sequences selected in the above (1) and expected to be capable of binding to the HLA-A201 molecule, was added to the resulting suspension at a concentration of 10 μ g/ml, followed by culture using a 96-well plate under the conditions of 37°C, 5% CO₂ for 4 hours. Thereafter, the cells were irradiated with 15 X-ray (3000 rad), washed with AIM-V medium, suspended in AIM-V medium containing 10% human AB serum (Nabi, Miami, FL), IL-6 (1000 U/ml) and IL-12 (10 ng/ml, Genzyme, Cambridge, MA), and placed in wells of a 24-well plate at a population of 1×10^5 cells/well. The prepared T cell population was added to the wells at a population of 1×10^6 cells/well, and the cells were cultured at 37°C under 20 5% CO₂. Seven days later, each culture supernatant was discarded, and the cells were treated with each of the peptides obtained in the same manner as described above. After irradiation with X-ray, the dendritic cells were suspended in AIM-V medium containing 10% human AB serum (Nabi, Miami, FL), IL-7 (10 U/ml, 25 Genzyme, Cambridge, MA) and IL-2 (10 U/ml, Genzyme, Cambridge, MA) (cell

density: 1×10^{5} cells/ml), and the cells were placed in wells of a 24-well plate at a cell population of 1×10^5 cells/well and further cultured. The same operations were

repeated 4 to 6 times at intervals of 7 days, and the stimulated T cells were then recovered, after which induction of CD8-positive T cells were confirmed by flow cytometry.

[0139]

Also for the peptides shown in SEQ ID NOs:110, 111, 112, 115 and 116, which were expected to be capable of binding to the HLA-A24 molecule, induction of peptide epitope-reactive CD8-positive T cells was attempted using dendritic cells and a T cell population induced from peripheral blood of an HLA-A24-positive healthy individual.

10 [0140]

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As a negative control, a peptide outside the scope of the present invention (SEQ ID NO:118) was used.

[0141]

Example 5: Determination of CD179b-derived Cytotoxic T Cell Antigen Epitopes

15 Which Stimulate HLA-A0201-positive CD8-positive T Cells

(1) IFN-γ-Producing Ability

In order to examine the specificity of each of the T cells, whose growth was confirmed among the T cells induced as described above, to peptide epitopes, 5×10^3 T cells were added to 5×10^4 T2 cells (Salter RD et al., Immunogenetics, 21:235-246 (1985), purchased from ATCC) which were pulsed with each peptide and expresses the HLA-A0201 molecule (cultured in AIM-V medium supplemented with each peptide at a concentration of 10 µg/ml, at 37°C under 5% CO₂ for 4 hours), and the cells were cultured in AIM-V medium containing 10% human AB serum in a 96-well plate for 24 hours. The supernatant after the culturing was recovered and the production amount of IFN- γ was measured by ELISA. As a result, production of IFN- γ was confirmed in the culture supernatants in the wells of T2 cells pulsed with the peptides of SEQ ID NOs:108 to 110 and SEQ ID NOs:113 to 117, when

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compared with the culture supernatants in the wells of T2 cells which were not pulsed with a peptide (Fig. 2). From these results, it was revealed that the abovedescribed peptides are T cell epitope peptides having a capacity to specifically stimulate, and allow proliferation of, the HLA-A0201-positive CD8-positive T cells, thereby inducing production of IFN- γ .

[0142]

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In Fig. 2, reference numerals 3, 4, 5, 6, 7, 8, 9 and 10 in the abscissa indicate the IFN- γ -producing abilities of the HLA-A0201-positive CD8-positive T cells due to stimulation from the T2 cells pulsed with the peptides of SEQ ID NOs:108, 109,

10 110, 113, 114, 115, 116 and 117, respectively. Reference numeral 11 indicates the result for the peptide of SEQ ID NO:118 used as the negative control.

[0143]

(2) Cytotoxicity Assay

Subsequently, whether or not the peptides of SEQ ID NOs:108 to 110 and 15 SEQ ID NOs:113 to 117 used in the present invention are presented on the HLA-A0201 molecules on tumor cells which are HLA-A0201-positive and express CD179b, and whether or not the CD8-positive T cells stimulated by these peptides can damage the tumor cells which are HLA-A0201-positive and express CD179b were examined. In a 50-ml centrifugal tube, 10^6 cells of a B cell leukemia cell line. 20 Namalwa cells (purchased from ATCC), whose expression of CD179b had been confirmed, were collected, and 100 µCi of chromium 51 was added thereto, followed by incubation at 37°C for 2 hours. Thereafter, the cells were washed 3 times with RPMI medium (manufactured by Gibco) containing 10% fetal calf serum (manufactured by Gibco), and placed in wells of a 96-well V-bottom plate in an amount of 10^3 cells/well. Further, to each well, 5×10^4 T cells suspended in RPMI 25 medium containing 10% fetal bovine serum, which cells were stimulated by each peptide, and HLA-A0201-positive, peptide epitope-reactive and CD8-positive, were

added, followed by culture at 37°C under 5% CO₂ for 4 hours. Thereafter, by measuring the amount of chromium 51 in the culture supernatant, which was released from the damaged tumor cells, the cytotoxic activity of the CD8-positive T cells stimulated by each peptide was calculated. As a result, it was revealed that the HLA-A0201-positive CD8-positive T cells stimulated by the peptide have a cytotoxic activity against Namalwa cells (Fig. 3). The CD8-positive T cells induced using the negative control peptide (SEQ ID NO:118) did not show a cytotoxic activity. Thus, it was proved that each of the peptides used in the present invention (SEQ ID NOs:108 to 110 and SEQ ID NOs:113 to 117) is presented on the HLA-A0201

10 molecules on tumor cells which are HLA-A0201-positive and express CD179b, and that the peptide has an ability to induce CD8-positive cytotoxic T cells which can damage such tumor cells.

[0144]

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The cytotoxic activity was determined by, as described above, mixing 10⁵ 15 CD8-positive T cells stimulated and induced with each of the peptides used in the present invention and 10³ cells of the B cell leukemia cell line Namalwa which were made to incorporate chromium 51; culturing the resulting mixture for 4 hours; measuring the amount of chromium 51 released to the culture medium after the culturing; and calculating the cytotoxic activity of the CD8-positive T cells against 20 the Namalwa cells according to the following equation*.

[0145]

*Equation: Cytotoxic activity (%) = the amount of chromium 51 released from Namalwa cells upon addition of CD8-positive T cells / the amount of chromium 51 released from the target cells upon addition of 1 N hydrochloric acid × 100.

25 [0146]

In Fig. 3, reference numerals 12, 13, 14, 15, 16, 17, 18 and 19 in the abscissa indicate the cytotoxic activities of the HLA-A0201-positive CD8-positive T cells

against the Namalwa cells, which T cells were stimulated using SEQ ID NOs:108, 109, 110, 113, 114, 115, 116 and 117, respectively. Reference numeral 20 indicates the cytotoxic activity of CD8-positive T cells induced using the peptide of the negative control (SEQ ID NO:118).

5 [0147]

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Example 6: Determination of CD179b-derived Cytotoxic T Cell Antigen Epitopes Which Stimulate HLA-A24-positive CD8-positive T Cells

(1) IFN-γ-Producing Ability

In order to examine the specificity of the peptide epitope-reactive CD8positive T cells induced in Example 3(2) to peptide epitopes in the same manner as in Example 5(1), 5×10^3 cells of the above-described T cells were added to 5×10^4 JTK-LCL cells expressing HLA-A24 molecules (purchased from RIKEN), which JTK-LCL cells were pulsed using the peptide of SEQ ID NOs:110, 111, 112, 115 or 116 (cultured in AIM-V medium supplemented with each peptide at a concentration of 10

µg/ml, at 37°C under 5% CO₂ for 4 hours), and the cells were cultured in AIM-V medium containing 10% human AB serum in a 96-well plate for 24 hours. The supernatant after the culturing was recovered and the production amount of IFN-γ was measured by ELISA. As a result, production of IFN-γ was confirmed in the culture supernatants in the wells of JTK-LCL cells pulsed with the peptides of SEQ
ID NOs:110, 111, 112, 115 and 116, when compared with the culture supernatants in the wells of JTK-LCL cells which were not pulsed with a peptide (Fig. 4). From these results, it was revealed that the above-described peptides are T cell epitope peptides having a capacity to specifically stimulate, and allow proliferation of, the HLA-A24-positive CD8-positive T cells, thereby inducing production of IFN-γ.

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[0148]

In Fig. 4, reference numerals 21, 22, 23, 24 and 25 in the abscissa indicate the IFN-γ-producing abilities of the HLA-A24-positive CD8-positive T cells due to

stimulation from the JTK-LCL cells pulsed with the peptides of SEQ ID NOs:110, 111, 112, 115 and 116, respectively. Reference numeral 26 indicates the result for the peptide of SEQ ID NO:118 used as the negative control.

[0149]

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5 (2) Cytotoxicity Assay

Subsequently, whether or not the peptides of SEQ ID NOs:110, 111, 112, 115 and 116 used in the present invention are presented on the HLA-A24 molecules on cells which are HLA-A24-positive and express CD179b, and whether or not the CD8-positive T cells stimulated by these peptides can damage the tumor cells which are HLA-A24-positive and express CD179b were examined in the same manner as in Example 5(2). In a 50-ml centrifugal tube, 10⁶ JTK-LCL cells, which are HLA-A24-positive and express CD179b, were collected, and 100 µCi of chromium 51 was added thereto, followed by incubation at 37°C for 2 hours. Thereafter, the cells were washed 3 times with RPMI medium containing 10% fetal calf serum, and placed in wells of a 96-well V-bottom plate in an amount of 10^3 cells/well. Further, to each well, 5×10^4 T cells suspended in RPMI medium containing 10% fetal calf serum, which cells were stimulated with each peptide, and HLA-A24-positive, peptide epitope-reactive and CD8-positive, were added, followed by culture at 37°C under 5% CO₂ for 4 hours. Thereafter, by measuring the amount of chromium 51 in the culture supernatant, which was released from the damaged cells, the cytotoxic activity of the CD8-positive T cells stimulated by each peptide was calculated. As a result, it was revealed that the HLA-A24-positive CD8-positive T cells stimulated by the peptide have a cytotoxic activity against JTK-LCL cells (Fig. 5). Thus, it was proved that each of the peptides used in the present invention (SEQ ID NOs:110, 111, 112, 115 and 116) is presented on the HLA-A24 molecules on cells which are HLA-A24-positive and express CD179b, and that the peptide has an ability to induce CD8positive cytotoxic T cells which can damage such cells. The CD8-positive T cells

induced using the negative control peptide (SEQ ID NO:118) did not show a cytotoxic activity.

[0150]

In Fig. 5, reference numerals 27, 28, 29, 30 and 31 indicate the cytotoxic activities of the HLA-A24-positive CD8-positive T cells stimulated with the peptides of SEQ ID NO:110, 111, 112, 115 and 116, respectively, against JTK-LCL cells. Reference numeral 32 indicates the cytotoxic activity of CD8-positive T cells induced using the peptide of the negative control (SEQ ID NO:118). [0151]

10 Example 7: Detection of Cancer Using Recombinant Protein

(1) Detection of Canine Cancer

From 153 canine patients whose malignant tumor was confirmed and 264 healthy dogs, blood was collected, and sera were separated therefrom. Using the dog-derived cancer antigen protein prepared in Example 2 (the 5th to 120th amino acids in the amino acid sequence shown in SEQ ID NO:5) and anti-dog IgG antibody, the titer of IgG antibody in the sera which specifically reacts with the polypeptide was measured by ELISA.

[0152]

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Immobilization of the prepared polypeptide on a solid phase was carried out
by placing 100 μL/well of the recombinant protein solution diluted to 100 μg/mL
with phosphate-buffered saline in a 96-well Immobilizer Amino plate (manufactured
by Nunc), followed by leaving the plate to stand at 4°C overnight. Blocking was
carried out by adding 100 μL/well of a solution, which was prepared by dissolving 4
g of Block Ace powder (manufactured by DS Pharma Biomedical Co., Ltd.) into 100
ml of purified water, into the wells, and shaking the plate at room temperature for 1
hour. The serum 1000-fold diluted with the blocking solution was added to the
wells in an amount of 100 μL/well, and the plate was shaken at room temperature for

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3 hours to allow the reaction to proceed. The wells were washed 3 times with phosphate-buffered saline containing 0.05% Tween 20 (manufactured by Wako Pure Chemical Industries, Ltd.)(hereinafter referred to as PBS-T), and 100 μ L/well of HRP-modified dog IgG antibody (Goat anti Dog IgG(-H+L) HRP conjugated: manufactured by BETHYL Laboratories) 3000-fold diluted with the blocking solution was added thereto, followed by shaking the plate at room temperature for 1 hour to allow the reaction to proceed. After washing the wells 3 times with PBS-T, 100 μ I/well of an HRP substrate TMB (1-Step Turbo TMB (tetramethylbenzidine), PIERCE) was added, and the enzyme-substrate reaction was allowed to proceed at room temperature for 30 minutes. Thereafter, 100 μ I/well of 0.5 M sulfuric acid solution (manufactured by Sigma-Aldrich Japan) was added to the wells to stop the reaction, and the absorbance at 450 nm was measured using a microplate reader. As

15 described above except that the prepared recombinant protein was not immobilized, or except that the tumor-bearing dog serum was not reacted, was designed for comparison.

a control, a case where the same operation was carried out in the same manner as

[0153]

As the cancer species to be used for the above detection of cancer, 112 samples of breast cancer, 31 samples of lymphoma and 10 samples of leukemia which had been definitely diagnosed as malignant by pathological diagnosis were used. [0154]

These sera derived from the living bodies of the tumor-bearing dogs showed significantly high antibody titers against the recombinant protein. It was revealed that, by diagnosing a sample showing twice the average value of healthy canine samples as malignant, 61 samples (54%) of breast cancer, 21 samples (71%) of lymphoma and 7 samples (70%) of leukemia could be successfully diagnosed as

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malignant. When the test was similarly carried out using sera from 30 canine patients having a mammary gland tumor which had been definitely diagnosed as benign, the number of samples showing twice the average value of healthy canine samples was 0.

5 [0155]

In the same manner, using the human-derived cancer antigen protein prepared in Example 2 (the amino acid sequence shown in SEQ ID NO:3) and anti-dog IgG antibody, the titer of IgG antibody which specifically reacts with the polypeptide in each of the above-described tumor-bearing dog serum samples was measured by

10 ELISA. As a result, it was revealed that 56 samples (50%) of breast cancer, 18 samples (58%) of lymphoma and 5 samples (50%) of leukemia could be judged as malignant.

[0156]

When the detection was carried out in the same manner as described above using pleural effusion and ascites collected from canine patients with terminal cancer, values similar to the results obtained by the detection method using serum could be detected, and diagnosis of the cancer was possible.

[0157]

(2) Detection of Human Cancer

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In the same manner, using the human-derived cancer antigen protein (the amino acid sequence shown in SEQ ID NO:3) used in the above detection and antihuman IgG antibody, the titer of IgG antibody in a healthy individual which specifically reacts with the polypeptide was measured. The secondary antibody to be used was an HRP-modified anti-human IgG antibody (manufactured by HRP-Goat Anti-Human IgG(H+L) Conjugate: manufactured by Zymed Laboratories) 10000diluted with the blocking solution. As a positive control, egg white albumin which was prepared to 50 µg/ml with phosphate-buffered saline and immobilized on the solid phase was used. As a result, in the case of the egg white albumin, seven healthy individuals showed an absorbance of 0.45 at 450 nm on average, which was high. On the other hand, in the case of the above-described polypeptide, the absorbance was 0, which means that the reaction was not detected at all.

5 [0158]

Further, in the same manner as described above, using 17 samples of sera derived from patients suffering from malignant breast cancer (purchased from Promeddx), the titer of IgG antibody in each serum which specifically reacts with the human-derived cancer antigen protein (amino acid sequence shown in SEQ ID NO:3) was similarly measured. As a result, in the case of the above-described polypeptide, the 17 breast cancer patients showed an absorbance of 0.28 at 450 nm on average, which was high. Thus, it was revealed that cancer can be detected by the present method also in human.

INDUSTRIAL APPLICABILITY

15 [0159]

The present invention is useful for therapy of cancer since it provides an immunity-inducing agent containing a polypeptide which exerts an anti-tumor activity against a cancer(s) (tumor(s)) such as breast cancer, leukemia and/or lymphoma. Further, the present invention is useful for diagnosis of cancer since it provides a novel detection method for cancer.

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[0160]

In the specification the term "comprising" shall be understood to have a broad meaning similar to the term "including" and will be understood to imply the inclusion of a stated integer or step or group of integers or steps but not the exclusion of any other integer or step or group of integers or steps. This definition also applies to variations on the term "comprising" such as "comprise" and "comprises."

The reference to any prior art in this specification is not, and should not be taken as an acknowledgement or any form of suggestion that the referenced prior art forms part of the common general knowledge in Australia.

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CLAIMS

1. A method for inducing immunity for therapy and/or prophylaxis of breast cancer, leukemia and/or lymphoma, said method comprising administering a therapeutically effective amount of as an effective ingredient(s) at least one polypeptide selected from the polypeptides (a) to (c) below, said polypeptide(s) having an immunity-inducing activity/activities, or as an effective ingredient(s) a recombinant vector(s) which comprise(s) a polynucleotide(s) encoding said polypeptide(s) and is/are capable of expressing said polypeptide(s) in vivo:

(a) a polypeptide consisting essentially of not less than 7 consecutive amino acids in any one of the amino acid sequences shown in the odd number IDs of SEQ ID NOs:3 to 95 in SEQUENCE LISTING;

(b) a polypeptide having a sequence identity of not less than 90% with said polypeptide (a) and consisting essentially of not less than 7 amino acids; and

(c) a polypeptide comprising said polypeptide (a) or (b) as a partial sequence thereof.

2. The method according to claim 1, wherein said polypeptide (b) has a sequence identity of not less than 95% with said polypeptide (a).

3. The method according to claim 1, wherein each of said polypeptide(s) having an immunity-inducing activity/activities is a polypeptide consisting essentially of not less than 7 consecutive amino acids in any one of the amino acid sequences shown in the odd number IDs of SEQ ID NOs:3 to 95 in SEQUENCE LISTING, or a polypeptide comprising said polypeptide as a partial sequence thereof.

4. The method according to claim 3, wherein each of said polypeptide(s) having 25 an immunity-inducing activity/activities is a polypeptide having any one of the amino acid sequences shown in the odd number IDs of SEQ ID NOs:3 to 95 in SEQUENCE LISTING.

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5. The method according to claim 3, wherein each of said polypeptide(s) having an immunity-inducing activity/activities is a polypeptide consisting essentially of not less than 7 consecutive amino acids in the region of aa1-34 or aa52-75 in any one of the amino acid sequences shown in the odd number IDs of SEQ ID NOs:3 to 95 in SEQUENCE LISTING, or a polypeptide comprising the polypeptide as a partial sequence thereof.

6. The method according to claim 5, wherein each of said polypeptide(s) having an immunity-inducing activity/activities is a polypeptide consisting essentially of the amino acid sequence shown in SEQ ID NO:108, SEQ ID NO:109, SEQ ID NO:110, SEQ ID NO:111, SEQ ID NO:112, SEQ ID NO:113, SEQ ID NO:114, SEQ ID NO:115, SEQ ID NO:116 or SEQ ID NO:117 in SEQUENCE LISTING, or a polypeptide comprising as a partial sequence thereof the amino acid sequence shown in SEQ ID NO:108, SEQ ID NO:109, SEQ ID NO:110, SEQ ID NO:111, SEQ ID NO:112, SEQ ID NO:113, SEQ ID NO:114, SEQ ID NO:115, SEQ ID NO:116 or SEQ ID NO:117 in SEQUENCE LISTING, said polypeptide having 8 to 12 amino acid residues.

7. The method according to any one of claims 1 to 6, comprising one or more of said polypeptides as an effective ingredient(s).

8. The method according to claim 7, wherein said polypeptide(s) is/are an agent(s) for treating antigen-presenting cells.

9. The method according to any one of claims 1 to 8, wherein said cancer(s) is/are a cancer(s) expressing the CD179b gene.

10. The method according to any one of claims 1 to 9, further comprising an immunoenhancer.

25 11. A method for inducing immunity for therapy and/or prophylaxis of breast cancer, leukemia and/or lymphoma, said method comprising administering to an individual at least one polypeptide selected from the polypeptides (a) to (c) below,

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said polypeptide(s) having an immunity-inducing activity/activities, or a recombinant vector(s) which comprise(s) a polynucleotide(s) encoding said polypeptide(s) and is/are capable of expressing said polypeptide(s) in vivo:

(a) a polypeptide consisting essentially of not less than 7 consecutive amino acids in any one of the amino acid sequences shown in the odd number IDs of SEQ ID NOs:3 to 95 in SEQUENCE LISTING;

(b) a polypeptide having a sequence identity of not less than 90% with said polypeptide (a) and consisting essentially of not less than 7 amino acids; and

(c) a polypeptide comprising said polypeptide (a) or (b) as a partial sequence thereof.

12. A method for detecting breast cancer or leukemia (excluding pre-B cell leukemia), which method is applied to a sample separated from a living body and comprises measuring expression of at least one of the polypeptides (a) to (c) below:

(a) a polypeptide produced in the living body and having a reactivity to bind, by antigen-antibody reaction, to an antibody against a polypeptide consisting of amino acids in any one of the amino acid sequences shown in the odd number IDs of SEO ID NOs:3 to 95 in SEQUENCE LISTING;

(b) a polypeptide produced in the living body and having a reactivity to bind, by antigen-antibody reaction, to an antibody against a polypeptide having a sequence identity of not less than 95% with said polypeptide (a); and

(c) a polypeptide produced in the living body and having a reactivity to bind, by antigen-antibody reaction, to an antibody against a polypeptide comprising said polypeptide (a) or (b) as a partial sequence thereof.

13. The method according to claim 12, wherein the measurement of expression of said polypeptide(s) is carried out by measuring an antibody/antibodies which may be contained in the sample by immunoassay, which antibody/antibodies was/were induced in the living body against said polypeptide(s) to be measured.

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14. A method for detecting breast cancer or leukemia (excluding pre-B cell leukemia), which is applied to a sample separated from a living body and comprises investigation of expression of the CD179b gene having a coding region having any one of the base sequences shown in SEQ ID NO:1 and the even number IDs of SEQ ID NO:4 to 94 in SEQUENCE LISTING in a sample derived from a cancer patient, and comparison thereof with the expression level of the gene in a sample derived from a healthy individual.

15. A reagent when used to detect breast cancer or leukemia (excluding pre-B cell leukemia), said reagent comprising a polypeptide which undergoes antigen-antibody reaction with an antibody induced in a living body against the polypeptide consisting of the amino acid sequence shown in SEQ ID NO:3.

16. A method for inducing immunity substantially as herein described with reference to the examples and/or drawings but excluding the comparative examples.

17. An isolated antigen-presenting cell substantially as herein described with reference to the examples and/or drawings but excluding the comparative examples.

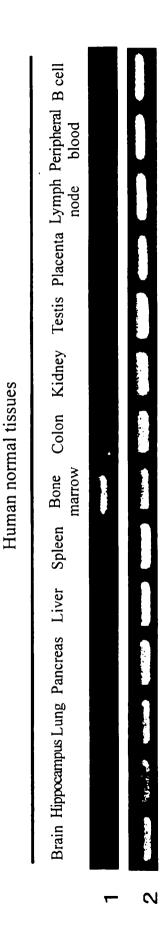
18. An isolated T cell substantially as herein described with reference to the examples and/or drawings but excluding the comparative examples.

19. A method for detecting breast cancer or leukemia (excluding pre-B cell leukemia) substantially as herein described with reference to the examples and/or drawings but excluding the comparative examples.

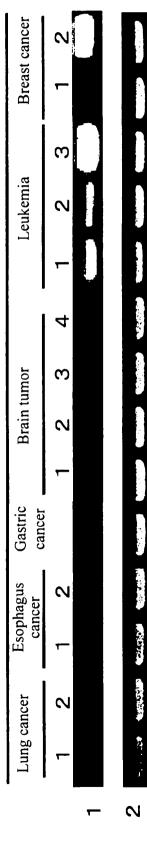
20. A reagent when used to detect breast cancer or leukemia (excluding pre-B cell leukemia) substantially as herein described with reference to the examples and/or drawings but excluding the comparative examples.

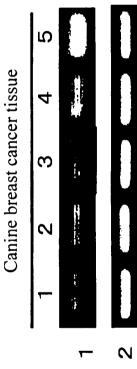
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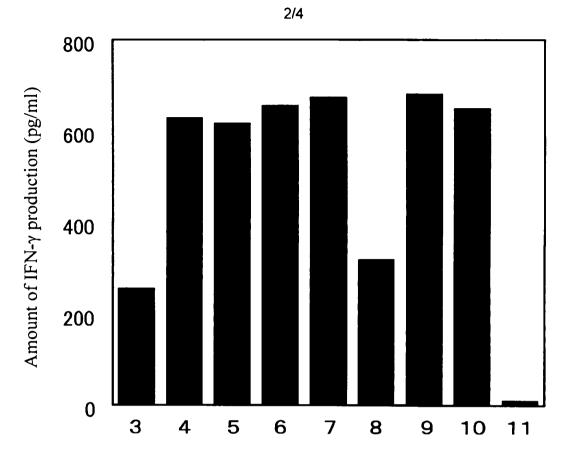




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Fig.1

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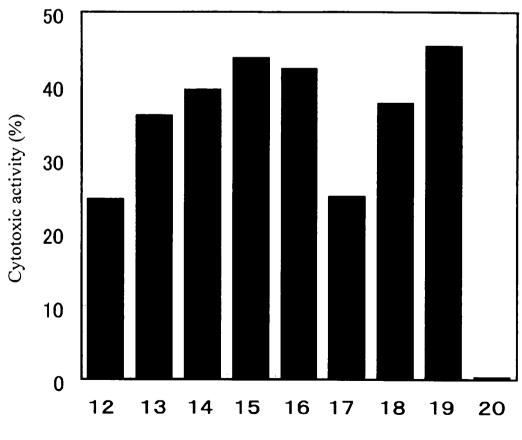
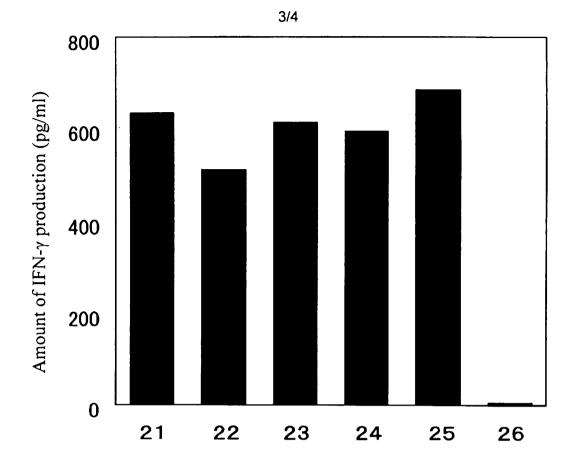
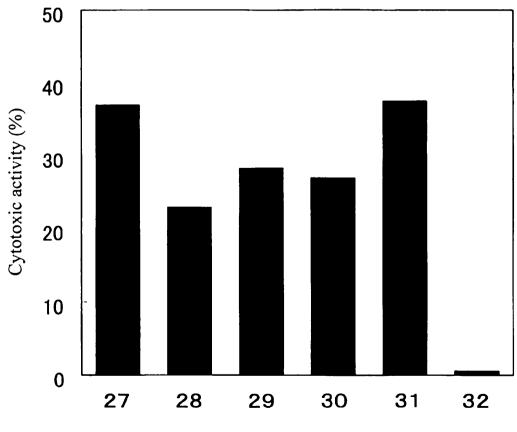


Fig.3







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Fig.5

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SEQUENCE LISTING(09062) Trp Tyr Gln Gln Leu Pro Gly Arg Gly Pro Arg Thr Val Ile Phe Thr 35 40 aca cat agt cga ccc tcg ggg gtg tcc gat cga ttc tct gcc tcc aag Thr His Ser Arg Pro Ser Gly Val Ser Asp Arg Phe Ser Ala Ser Lys 50 55 60 193 tct ggc agc aca gcc acc ctg acc atc tct ggg ctc cag gct gag gat Ser Gly Ser Thr Ala Thr Leu Thr Ile Ser Gly Leu Gln Ala Glu Asp 65 70 75 80 241 gag gct gat tat tac tgc tca acg tgg gat gat agt ctc agt gct gct Glu Ala Asp Tyr Tyr Cys Ser Thr Trp Asp Asp Ser Leu Ser Ala Ala 85 90 95 289 gtg ttc ggc gga ggc acc cac ctg acc gtc ctc ggt cag ccc aag gcc Val Phe Gly Gly Gly Thr His Leu Thr Val Leu Gly Gln Pro Lys Ala 337 100 105 110 tcc ccc tcg gtc aca ctc ttc ccg ccc tcc tct gag gag ctc ggc gcc Ser Pro Ser Val Thr Leu Phe Pro Pro Ser Ser Glu Glu Leu Gly Ala 385 120 115 aac aag gcc acc ctg gtg tgc ctc atc agc gac ttc tac ccc agc ggc Asn Lys Ala Thr Leu Val Cys Leu Ile Ser Asp Phe Tyr Pro Ser Gly 130 135 140 433 gtg acg gtg gcc tgg aag gca gac ggc agc ccc gtc acc cag ggc gtg Val Thr Val Ala Trp Lys Ala Asp Gly Ser Pro Val Thr Gln Gly Val 145 150 160 481 gag acc acc aag ccc tcc aag cag agc aac aac aag tac gcg gcc agc Glu Thr Thr Lys Pro Ser Lys Gln Ser Asn Asn Lys Tyr Ala Ala Ser 529 165 170 agc tac ctg agc ctg acg cct gac aag tgg aaa tct cac agc agc ttc Ser Tyr Leu Ser Leu Thr Pro Asp Lys Trp Lys Ser His Ser Ser Phe 577 180 190 agc tgc ctg gtc acg cac gag ggg agc acc gtg gag aag aag gtg gcc Ser Cys Leu Val Thr His Glu Gly Ser Thr Val Glu Lys Lys Val Ala 625 195 200 ccc gca gag tgc tct tag gttcccgacg gccccgccca ccgaaggggg 673 Pro Ala Glu Cys Ser 210 cccggagcct caggacctcc aggaggatct tgcctcccat ctgggtcatc ccgcccttct 733 793 aaa 796 <210> 15 <211> 213 <212> PRT Canis familiaris <213> <400> 15 Leu Thr Gln Pro Ala Ser Val Ser Gly Ser Leu Gly Gln Arg Ile Thr 1 5 10 15 Ile Ser Cys Thr Gly Ser Ser Asn Ile Gly Gly Asn Asn Val Gly 20 25 30 30

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838

859

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SEQUENCE LISTING(09062) 200 195 205 aag tgg aaa tct cac agc agc ttc agc tgc ctg gtc acg cac gag ggg Lys Trp Lys Ser His Ser Ser Phe Ser Cys Leu Val Thr His Glu Gly 210 215 220 673 acc gtg gag aag aag gtg gcc ccc gca gag tgc tct tag Thr val Glu Lys Lys val Ala Pro Ala Glu Cys Ser 230 235 715 agc Ser 225 775 gttcctgatg tcccccgccc accaaagggg gctcagagcc tcaggacctc caggaggatc 835 ttgcctccca tctgggtcat cccagccttt ccccttaaac ccaggcaaca ttcaataaag 875 tgttctttct tcaatcagaa aaaaaaaaa aaaaaaaaa <210> 21 <211> 237 <212> PRT <213> Canis familiaris <400> 21 Ser Asn Met Ala Trp Ser Pro Leu Leu Leu Thr Leu Leu Val Tyr Cys 1 5 10 15 Thr Gly Ser Trp Ala Gln Ser Val Leu Thr His Pro Thr Ser Val Ser 20 25 30 Gly Ser Leu Gly Gln Arg Val Thr Ile Ser Cys Ser Gly Ser Thr Asn 35 40 45 Asn Ile Gly Thr Val Gly Ala Gly Trp Tyr Gln Gln Phe Pro Gly Lys 50 55 60 Ala Pro Lys Leu Leu Ile Tyr Ser Asp Gly Asn Arg Pro Ser Gly Val 65 70 75 80 Pro Asp Arg Phe Ser Gly Ser Lys Ser Gly Asn Ser Ala Thr Leu Thr 85 90 95 Ile Ile Gly Leu Gln Ala Glu Asp Glu Ala Asp Tyr Tyr Cys Gln Ser 100 105 110 100 Val Asp Pro Thr Leu Gly Gly His Val Phe Gly Gly Gly Thr His Leu 115 120 125 Thr Val Leu Gly Gln Pro Lys Ala Ser Pro Ser Val Thr Leu Phe Pro 130 135 140 Pro Ser Ser Glu Glu Leu Gly Ala Asn Lys Ala Thr Leu Val Cys Leu 145 150 155 160 Ile Ser Asp Phe Tyr Pro Ser Gly Val Thr Val Ala Trp Lys Ala Asp 165 170 175

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SEQUENCE LISTING(09062) Cys Leu Ile Ser Asp Phe Tyr Pro Ser Gly Val Thr Val Ala Trp Lys 165 170 175 165 gca gac ggc agc ccc gtc acc cag ggc gtg gag acc acc aag ccc tcc Ala Asp Gly Ser Pro Val Thr Gln Gly Val Glu Thr Thr Lys Pro Ser 180 185 190 577 aag cag agc aac aac aag tac gcg gcc agc agc tac ctg agc ctg acg Lys Gln Ser Asn Asn Lys Tyr Ala Ala Ser Ser Tyr Leu Ser Leu Thr 625 200 cct gac aag tgg aaa tct cac agc agc ttc agc tgc ctg gtc acg cac Pro Asp Lys Trp Lys Ser His Ser Ser Phe Ser Cys Leu Val Thr His 210 215 220 673 gag ggg agc acc gtg gag aag aag gtg gcc ccc gca gag tgc tct tag Glu Gly Ser Thr Val Glu Lys Lys Val Ala Pro Ala Glu Cys Ser 721 230 235 gttcccgacg gccccgccca ccgaaggggg cccggagcct caggacctcc aggaggatct 781 tgcctcccat ctgggtcatc ccgctcttct ccccgcaccc aggcagcact caataaagtg 841 ttctttgttc aatcagaaaa a 862 23 <210> 239 <211> <212> PRT <213> Canis familiaris <400> 23 Met Ile Phe Thr Met Ala Trp Ser Pro Leu Leu Leu Gly Leu Leu Ala 1 5 10 15 His Cys Thr Gly Ser Trp Ala Gln Ser Met Leu Thr Gln Pro Ala Ser 20 25 30 Val Ser Gly Ser Leu Gly Gln Lys Val Thr Ile Ser Cys Thr Gly Ser 35 40 45 Ser Ser Asn Ile Gly Ala Tyr Tyr Val Ser Trp Tyr Gln Gln Ser Pro 50 55 60 Gly Lys Gly Pro Arg Thr Val Ile Tyr Gly Asp Asn Tyr Arg Pro Ser 65 70 75 80 80 Gly Val Pro Asp Arg Phe Ser Gly Ser Lys Ser Gly Ser Ser Ala Thr 85 90 95 Leu Thr Ile Ser Gly Leu Gln Ala Glu Asp Glu Ala Glu Tyr Tyr Cys 100 105 110 Leu Ser Trp Asp Asn Ser Leu Arg Gly Gly Val Phe Gly Gly Gly Thr 115 120 125 His Leu Thr Val Leu Gly Gln Pro Lys Ala Ser Pro Ser Val Thr Leu 130 135 140 130 140

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Ala Asp Gly	Ser Pro 180	Val Thr	Gln Gly 185	Val Glu	Thr Thr	Lys Pro 190	Ser
Lys Gln Ser 195	Asn Asn	Lys Tyr	Ala Ala 200	Ser Ser	Tyr Leu 205	Ser Leu	Thr
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Page 23

SEQUENCE LISTING(09062) Gly Thr Thr Ala Ile Leu Thr Ile Ser Gly Leu Gln Ala Glu Asp Glu 105 100 110 Ala Asp Tyr Tyr Cys Ser Ala Tyr Asp Ser Ser Leu Gly Gly Thr Ile 115 120 125 Phe Gly Gly Gly Thr Phe Leu Thr Val Leu Gly Gln Pro Lys Ala Ser 130 135 140 Pro Ser Val Thr Leu Phe Pro Pro Ser Ser Glu Glu Leu Gly Ala Asn 150 145 155 160 Lys Ala Thr Leu Val Cys Leu Ile Ser Asp Phe Tyr Pro Ser Gly Val 165 170 175 165 Thr Val Ala Trp Lys Ala Asp Gly Ser Pro Val Thr Gln Gly Val Glu 180 185 190 Thr Thr Lys Pro Ser Lys Gln Ser Asn Asn Lys Tyr Ala Ala Ser Ser 195 200 205 Tyr Leu Ser Leu Thr Pro Asp Lys Trp Lys Ser His Ser Ser Phe Ser 210 215 220 Cys Leu Val Thr His Glu Gly Ser Thr Val Glu Lys Lys Val Ala Pro 225 230 235 240 Ala Glu Cys Ser <210> 26 <211> 729 <212> DNA <213> Canis familiaris <220> <221> CDS <222> (2)..(574) <223> <400> 26 c tcc aac att gga ggt aat cat gta ggt tgg tac caa caa ttt cca gga Ser Asn Ile Gly Gly Asn His Val Gly Trp Tyr Gln Gln Phe Pro Gly 1 5 10 15 49 aga ggc ccc aga act gtc atc tat agc aca aat gtt cga ccc tcg ggg Arg Gly Pro Arg Thr Val Ile Tyr Ser Thr Asn Val Arg Pro Ser Gly 20 25 30 97 gtg ccc gat cga ttc tct ggc tcc aag tct gac aac aca ggc acc ctg Val Pro Asp Arg Phe Ser Gly Ser Lys Ser Asp Asn Thr Gly Thr Leu 35 40 45 145 acc atc tct gga ctc cag gct gag gat gag gct gat tat tat tgc gca Thr Ile Ser Gly Leu Gln Ala Glu Asp Glu Ala Asp Tyr Tyr Cys Ala 50 55 60 193 acg tgg gat gat agt ctc agt gtt tct ctg ttc ggc gga ggc acc cac 241 Page 24

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289

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385

529

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Gln Gln Leu Pro Gly Arg Gly Pro Arg Thr Leu Phe Tyr Arg Ala Thr 65 70 75 80 Gly Arg Pro Ser Gly Val Pro Asp Arg Phe Ser Ala Ser Arg Ser Gly 85 90 95 Thr Thr Ala Thr Leu Thr Ile Ser Gly Leu Gln Pro Glu Asp Glu Ala 100 105 110 Asp Tyr Tyr Cys Ser Ser Tyr Asp Ser Thr Leu Phe Ser Val Phe Gly 115 120 125 Gly Gly Thr Tyr Leu Thr Val Leu Gly Gln Pro Lys Ala Ser Pro Ser 130 135 140 Val Thr Leu Phe Pro Pro Ser Ser Glu Glu Leu Gly Ala Asn Lys Ala 145 150 155 160 Thr Leu Val Cys Leu Ile Ser Asp Phe Tyr Pro Ser Gly Val Thr Val 165 170 175 Ala Trp Lys Ala Asp Gly Ser Pro Val Thr Gln Gly Val Glu Thr Thr 180 185 190 Lys Pro Ser Lys Gln Ser Asn Asn Lys Tyr Ala Ala Ser Ser Tyr Leu 195 200 205 Ser Leu Thr Pro Asp Lys Trp Lys Ser His Ser Ser Phe Ser Cys Leu 210 215 220 Val Thr His Glu Gly Ser Thr Val Glu Lys Lys Val Ala Pro Ala Glu 225 230 235 240 Cys Ser <210> 30 <211> 762 <212> DNA <213> Canis familiaris <220> <221> <222> CDS (1)..(609)<223> <400> 30 ggc cag agg gtc acc atc tcc tgc act gga agc ccc aat gtt ggt tat Gly Gln Arg Val Thr Ile Ser Cys Thr Gly Ser Pro Asn Val Gly Tyr 1 5 10 15 ggc aat tac gtg ggc tgg tac cag cag ctc cca gga aca ggc ccc aga Gly Asn Tyr Val Gly Trp Tyr Gln Gln Leu Pro Gly Thr Gly Pro Arg 20 25 30 Page 28

48

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Phe Ser Gly Ser Thr Ser Gly Ser Ser Ala Thr Leu Thr Ile Ser Gly 50 55 60 60 Leu Gln Ala Glu Asp Glu Ala Asp Tyr Tyr Cys Ser Ser Tyr Asp Ile 65 70 75 80 Ser Leu Gly Gly Val Val Phe Gly Gly Gly Thr His Leu Thr Val Leu 85 90 95 Gly Gln Pro Lys Ala Ser Pro Ser Val Thr Leu Phe Pro Pro Ser Ser 105 Glu Glu Leu Gly Ala Asn Lys Ala Thr Leu Val Cys Leu Ile Ser Asp 120 115 125 Phe Tyr Pro Ser Gly Val Thr Val Ala Trp Lys Ala Asp Gly Ser Pro 130 135 140 Val Thr Gln Gly Val Glu Thr Thr Lys Pro Ser Lys Gln Ser Asn Asn 145 150 155 160 Lys Tyr Ala Ala Ser Ser Tyr Leu Ser Leu Thr Pro Asp Lys Trp Lys 165 170 175 165 170 Ser His Ser Ser Phe Ser Cys Leu Val Thr His Glu Gly Ser Thr Val 180 185 190 190 185 Glu Lys Lys Val Ala Pro Ala Glu Cys Ser 195 200 <210> 32 826 <211> <212> DNA <213> Canis familiaris <220> <221> CDS <222> (1)..(678)<223> <400> 32 ctt gtc agc ctc ctg gct ctc tgc aca ggt tct gtg gcc tcc tat gtg Leu Val Ser Leu Leu Ala Leu Cys Thr Gly Ser Val Ala Ser Tyr Val 1 5 10 15 48 ctg aca cag ccg ccg tcc atg agt gtg acc ctg agg cag acg gcc cgc Leu Thr Gln Pro Pro Ser Met Ser Val Thr Leu Arg Gln Thr Ala Arg 20 25 30 96 atc acc tgt gag gga gac agc att gga gat aaa aga gtt tac tgg tac Ile Thr Cys Glu Gly Asp Ser Ile Gly Asp Lys Arg Val Tyr Trp Tyr 35 40 45 144 cag cag aaa ctg ggc cgg ggc ccg atg ttg att att tat gat ggt acc Gln Gln Lys Leu Gly Arg Gly Pro Met Leu Ile Ile Tyr Asp Gly Thr 192 Page 30

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Asp Tyr Tyr Cys Gln Val Trp Asp Asn Gly Glu Ile Ile Phe Gly Gly 100 105 110
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Thr Leu Phe Pro Pro Ser Ser Glu Glu Leu Gly Ala Asn Lys Ala Thr 130 135 140
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Pro Ser Lys Gln Ser Asn Asn Lys Tyr Ala Ala Ser Ser Tyr Leu Ser 180 185 190
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49

97

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Ile Ser Cys Thr Gly Ser Ser Ser Asn Ile Gly Ser Asn Asp Val Gly Page 33	

SEQUENCE LISTING(09062) 25 30 20 Trp Tyr Gln Gln Leu Pro Gly Arg Gly Pro Lys Thr Val Val Ser Asn 35 40 45 Thr Asn Ile Arg Pro Ser Gly Val Pro Asp Arg Phe Ser Ala Ser Lys 50 55 60 Ser Gly Ser Thr Ala Thr Leu Thr Ile Ser Gly Leu Gln Ala Glu Asp 65 70 75 80 Glu Ala Asp Tyr Tyr Cys Ser Thr Trp Asp Asn Ser Leu Ser Thr Tyr 85 90 95 Met Phe Gly Ser Gly Thr Gln Leu Thr Val Leu Gly Gln Pro Lys Ala 100 105 110 Ser Pro Ser Val Thr Leu Phe Pro Pro Ser Ser Glu Glu Leu Gly Ala 115 120 125 Asn Lys Ala Thr Leu Val Cys Leu Ile Ser Asp Phe Tyr Pro Ser Gly 130 135 140 Val Thr Val Ala Trp Lys Ala Asp Gly Ser Pro Ile Thr Gln Gly Val 145 150 155 160 160 Glu Thr Thr Lys Pro Ser Lys Gln Ser Asn Asn Lys Tyr Ala Ala Ser 165 170 175 Ser Tyr Leu Ser Leu Thr Pro Asp Lys Trp Lys Ser His Ser Ser Phe 180 185 190 Ser Cys Leu Val Thr His Glu Gly Ser Thr Val Glu Lys Lys Val Ala 195 200 205 Pro Ala Glu Cys Ser 210 <210> 36 930 <211> <212> DNA <213> Canis familiaris <220> <221> <222> CDS (1)..(774)<223> <400> 48 gtg atg acc tcc acc atg ggc tgg ttc cct ctc atc ctc acc ctc ctc Val Met Thr Ser Thr Met Gly Trp Phe Pro Leu Ile Leu Thr Leu Leu 96 Page 34

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tca gtg tct Ser val Ser 50	ggg tcc Gly Ser	ctg ggo Leu Gly 55	cag Gln	agg Arg	gtc val	acc Thr	atc Ile 60	tcc Ser	tgc Cys	act Thr	gga Gly	192
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ctc cca gga Leu Pro Gly												288
ccc tcg ggg Pro Ser Gly												336
gca acc ctg Ala Thr Leu 115	Thr Ile	tct gg <u>o</u> Ser Gly	ctc Leu 120	cag Gln	gct Ala	gag Glu	gat Asp	gaa Glu 125	gcc Ala	gat Asp	tat Tyr	384
tac tgc tca Tyr Cys Ser 130			Ser									432
ggc acc cac Gly Thr His 145												480
aca ctc ttc Thr Leu Phe	ccg ccc Pro Pro 165	tcc tct Ser Ser	gag Glu	gag Glu	ctc Leu 170	ggc Gly	gcc Ala	aac Asn	aag Lys	gcc Ala 175	acc Thr	528
ctg gtg tgc Leu Val Cys	ctc atc Leu Ile 180	agc gac Ser Asp	ttc Phe	tac Tyr 185	ccc Pro	agc Ser	ggc Gly	gtg Val	acg Thr 190	gtg Val	gcc Ala	576
tgg aag gca Trp Lys Ala 195	Āsp Ğly	agc cco Ser Pro	gtc Val 200	acc Thr	cag Gln	ggc Gly	gtg Val	gag Glu 205	acc Thr	acc Thr	aag Lys	624
ccc tcc aag Pro Ser Lys 210	cag agc Gln Ser	aac aac Asn Asr 215	Lys	tac Tyr	gcg Ala	gcc Ala	agc Ser 220	agc Ser	tac Tyr	ctg Leu	agc Ser	672
ctg acg cct Leu Thr Pro 225												720
acg cac gag Thr His Glu	ggg agc Gly Ser 245	acc gto Thr Val	gag Glu	aag Lys	aag Lys 250	gtg Val	gcc Ala	ccc Pro	gca Ala	gag Glu 255	tgc Cys	768
tct tag gtt Ser	cccgacg	gccccgcc	ca c	cgaag	99999	g cco	cgga	gcct	cago	gacci	tcc	824
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Ser

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agc tgc ctg gtc aca cac gag ggg agc acc gtg gag aag aag gtg gcc Ser Cys Leu Val Thr His Glu Gly Ser Thr Val Glu Lys Lys Val Ala 210 215 220 674 ccc gca gag tgc tct tag gttcccgacg ccccgccca cctaaggggg 722 Pro Āla Ģļu Cys Ser 225 782 cccggagcct caggacctcc aggaggatct tgcctcctat ctgggtcatc ccgcccttct 842 ccccacaccc aggcagcact caataaattg ttctttgttc aatcagaaaa aaggggggcc 843 С 39 <210> <211> 227 <212> PRT <213> Canis familiaris <400> 39 Met Tyr Lys Ile Leu Glu Ser Thr Tyr Ile Val Lys Arg Ser Ile Thr 1 5 10 15 Val Pro Gln Pro Pro Phe Val Ser Val Thr Leu Arg Asp Thr Ala His 20 25 30 Ile Thr Cys Gly Gly Asp Asn Ile Gly Ser Lys Tyr Val Gln Trp Ile 35 40 45 Gln Gln Asn Pro Gly Gln Ala Pro Val Val Ile Ile Tyr Arg Asp Thr 50 55 60 Lys Arg Pro Thr Trp Ile Pro Glu Arg Phe Ser Gly Ala Asn Ser Gly 65 70 75 80 Asn Thr Ala Thr Leu Thr Ile Ser Gly Val Leu Ala Glu Asp Glu Ala 85 90 95 Asp Tyr Tyr Cys Gln Val Thr Asp Ser Gly Pro Gln Thr Asn Val Phe 100 105 110 Gly Gly Gly Thr His Leu Thr Val Leu Ser Gln Pro Lys Ala Ser Pro 115 120 125 Ser Val Thr Leu Phe Pro Pro Ser Ser Glu Glu Leu Gly Ala Asn Lys 140 130 135 Ala Thr Leu Val Cys Leu Ile Ser Asp Phe Tyr Pro Ser Gly Val Thr 145 150 155 160 160 Val Ala Trp Lys Ala Asp Gly Ser Pro Val Thr Gln Gly Val Glu Thr 165 170 175 Thr Lys Pro Ser Lys Gln Ser Asn Asn Lys Tyr Ala Ala Ser Ser Tyr

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SEQUENCE LISTING(09062) 165 175 170 agc ccc gtc acc cag ggc gtg gag acc acc aag ccc tcc aag cag agc Ser Pro Val Thr Gln Gly Val Glu Thr Thr Lys Pro Ser Lys Gln Ser 577 185 190 180 aac aac aag tac gcg gcc agc agc tac ctg agc ctg acg cct gac aag Asn Asn Lys Tyr Ala Ala Ser Ser Tyr Leu Ser Leu Thr Pro Asp Lys 195 200 205 625 tgg aaa tct cac agc agc ttc agc tgc ctg gtc acg cac gag ggg agc Trp Lys Ser His Ser Ser Phe Ser Cys Leu Val Thr His Glu Gly Ser 210 215 220 673 acc gtg gag aag aag gtg gcc ccc gca gag tgc tct tag gttcccgacg Thr Val Glu Lys Lys Val Ala Pro Ala Glu Cys Ser 225 230 235 722 gccccgccca ccgaaggggg cccggagcct caggacctcc aggaggatct tgcctcccat 782 ctgggtcatc ccgcccttct ccccgcaccc aggcagcact caataaagtg ttctttgttc 842 aatcagaaaa aaaaaa 858 <210> 41 <211> 236 <212> PRT <213> Canis familiaris <400> 41 Ser Asn Met Ala Trp Ser Pro Leu Leu Leu Thr Leu Leu Ala Tyr Cys 1 5 10 15 10 Thr Gly Ser Trp Ala Gln Ser Val Leu Thr Gln Pro Thr Ser Val Ser 20 25 30 Gly Ser Leu Gly Gln Arg Val Thr Ile Ser Cys Ser Gly Ser Thr Asn 35 40 45 Asn Ile Gly Ile Val Gly Ala Ser Trp Tyr Gln Gln Leu Pro Gly Lys 50 55 60 Ala Pro Lys Leu Leu Val Tyr Ser Val Gly Asp Arg Pro Ser Gly Val 65 70 75 80 65 Pro Asp Arg Phe Ser Gly Ser Asn Ser Gly Asn Ser Ala Thr Leu Thr 85 90 95 Ile Thr Gly Leu Gln Ala Glu Asp Glu Ala Asp Tyr Tyr Cys Gln Ser 100 105 110 Phe Asp Thr Thr Leu Gly Ala Val Phe Gly Gly Gly Thr His Leu Thr 115 Val Leu Gly Gln Pro Lys Ala Ser Pro Ser Val Thr Leu Phe Pro Pro 130 135 140

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Leu Ile Ser Asp Phe Tyr Pro Ser Gly Val Thr Val Ala Trp Lys Ala 50 55 60	
Asp Gly Ser Pro Val Thr Gln Gly Val Glu Thr Thr Lys Pro Ser Lys 65 70 75 80	
Gln Ser Asn Asn Lys Tyr Ala Ala Ser Ser Tyr Leu Ser Leu Thr Pro 85 90 95	
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Gly Ser Thr Val Glu Lys Lys Val Ala Pro Ala Glu Cys Ser 115 120 125	
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ccg ccc tcc tct gag gag ctc ggc gcc aac aag gcc acc ctg gtg tgc Pro Pro Ser Ser Glu Glu Leu Gly Ala Asn Lys Ala Thr Leu Val Cys 35 40 45	144

SEQUENCE LISTING(09062) ctc atc agc gac ttc tac ccc agc ggc gtg acg gtg gcc tgg aag gca Leu Ile Ser Asp Phe Tyr Pro Ser Gly Val Thr Val Ala Trp Lys Ala 192 55 50 60 gac ggc agc ccc gtc acc cag ggc gtg gag acc acc aag ccc tcc aag Asp Gly Ser Pro Val Thr Gln Gly Val Glu Thr Thr Lys Pro Ser Lys 240 65 70 75 80 cag agc aac aac aag tac gcg gcc agc agc tac ctg agc ctg acg cct Gln Ser Asn Asn Lys Tyr Ala Ala Ser Ser Tyr Leu Ser Leu Thr Pro 85 90 95 288 gac aag tgg aaa tct cac agc agc ttc agc tgc ctg gtc acg cac gag Asp Lys Trp Lys Ser His Ser Ser Phe Ser Cys Leu Val Thr His Glu 336 100 105 110 ggg agc acc gtg gag aag aag gtg gcc ccc gca gag tgc tct tag Gly Ser Thr Val Glu Lys Lys Val Ala Pro Ala Glu Cys Ser 115 120 381 115 120 gttcccgacg gccccgccca ccgaaggggg cccggagcct caggacctcc aggaggatct 441 tgcctcccat ctgggtcatc ccgctcttct ccccgcaccc aggcagcact caataaagtg 501 ttctttgttc aat 514 <210> 45 <211> 126 <212> PRT Canis familiaris <213> <400> 45 Met Phe Glu Ala Val Ser Gln Cys Ala Val Phe Gly Gly Gly Thr His Leu Thr Val Leu Gly Gln Pro Lys Ala Ser Pro Ser Val Thr Leu Phe 20 25 30 Pro Pro Ser Ser Glu Glu Leu Gly Ala Asn Lys Ala Thr Leu Val Cys 35 40 45 Leu Ile Ser Asp Phe Tyr Pro Ser Gly Val Thr Val Ala Trp Lys Ala 50 55 60 Asp Gly Ser Pro Val Thr Gln Gly Val Glu Thr Thr Lys Pro Ser Lys 65 70 75 80 Gln Ser Asn Asn Lys Tyr Ala Ala Ser Ser Tyr Leu Ser Leu Thr Pro 85 90 95 Asp Lys Trp Lys Ser His Ser Ser Phe Ser Cys Leu Val Thr His Glu 100 105 110 Gly Ser Thr Val Glu Lys Lys Val Ala Pro Ala Glu Cys Ser 115 120

Page 43

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46

561

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Pro Ser Ser Glu Glu Leu Gly Ala Asn Lys Ala Thr Leu Val Cys Leu 50 60 Ile Ser Asp Phe Tyr Pro Ser Gly Val Thr Val Ala Trp Lys Ala Asp 65 70 75 80 Gly Ser Pro Ile Thr Gln Gly Val Glu Thr Thr Lys Pro Ser Lys Gln 85 90 Ser Asn Asn Lys Tyr Ala Ala Ser Ser Tyr Leu Ser Leu Thr Pro Asp 100 105 110 Lys Trp Lys Ser His Ser Ser Phe Ser Cys Leu Val Thr His Glu Gly 120 125 Ser Thr Val Glu Lys Lys Val Ala Pro Ala Glu Cys Ser 130 135 140 <210> 48 <211> 514 <212> DNA Canis familiaris <213> <220> <221> CDS <222> (1)..(381)<223> <400> 48 atg ttc gag gct gtg tca cag tgt gct gtg ttc ggc gga ggc acc cac Met Phe Glu Ala Val Ser Gln Cys Ala Val Phe Gly Gly Gly Thr His 1 5 10 15 48 ctg acc gtc ctc ggt cag ccc aag gcc tcc ccc tcg gtc aca ctc ttc Leu Thr Val Leu Gly Gln Pro Lys Ala Ser Pro Ser Val Thr Leu Phe 96 20 25 30 ccg ccc tcc tct gag gag ctc ggc gcc aac aag gcc acc ctg gtg tgc Pro Pro Ser Ser Glu Glu Leu Gly Ala Asn Lys Ala Thr Leu Val Cys 144 35 40 ctc atc agc gac ttc tac ccc agc ggc gtg acg gtg gcc tgg aag gca Leu Ile Ser Asp Phe Tyr Pro Ser Gly Val Thr Val Ala Trp Lys Ala 50 55 60 192 gac ggc agc ccc gtc acc cag ggc gtg gag acc acc aag ccc tcc aag Asp Gly Ser Pro Val Thr Gln Gly Val Glu Thr Thr Lys Pro Ser Lys 240 70 cag agc aac aag tac gcg gcc agc agc tac ctg agc ctg acg cct Gln Ser Asn Asn Lys Tyr Ala Ala Ser Ser Tyr Leu Ser Leu Thr Pro 85 90 95 288 95 gac aag tgg aaa tct cac agc agc ttc agc tgc ctg gtc acg cac gag Asp Lys Trp Lys Ser His Ser Ser Phe Ser Cys Leu Val Thr His Glu 100 105 110 336 105 ggg agc acc gtg gag aag aag gtg gcc ccc gca gag tgc tct tag Gly Ser Thr Val Glu Lys Lys Val Ala Pro Ala Glu Cys Ser 381

SEQUENCE LISTING(09062)

45

40

35

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SEQUENCE LISTING(09062) Pro Pro Ser Ser Glu Glu Leu Gly Ala Asn Lys Ala Thr Leu Val Cys 35 40 45 ctc atc agc gac ttc tac ccc agc ggc gtg acg gtg gcc tgg aag gca Leu Ile Ser Asp Phe Tyr Pro Ser Gly Val Thr Val Ala Trp Lys Ala 50 55 60 192 gac ggc agc ccc gtc acc cag ggc gtg gag acc acc aag ccc tcc aag Asp Gly Ser Pro Val Thr Gln Gly Val Glu Thr Thr Lys Pro Ser Lys 240 70 cag agc aac aac aag tac gcg gcc agc agc tac ctg agc ctg acg cct Gln Ser Asn Asn Lys Tyr Ala Ala Ser Ser Tyr Leu Ser Leu Thr Pro 85 90 95 288 95 gac aag tgg aaa tct cac agc agc ttc agc tgc ctg gtc acg cac gag Asp Lys Trp Lys Ser His Ser Ser Phe Ser Cys Leu Val Thr His Glu 336 100 105 110 ggg agc acc gtg gag aag aag gtg gcc ccc gca gag tgc tct tag Gly Ser Thr Val Glu Lys Lys Val Ala Pro Ala Glu Cys Ser 115 120 125 381 441 gttcccgacg gccccgccca ccgaaggggg cccggagcct caggacctcc aggaggatct tgcctcccat ctgggtcatc ccgctcttct ccccgcaccc aggcagcact caataaagtg 501 514 ttctttgttc aat <210> 51 126 <211> <212> PRT Canis familiaris <213> <400> 51 Met Phe Glu Ala Val Ser Gln Cys Ala Val Phe Gly Gly Gly Thr His 1 5 10 15 Leu Thr Val Leu Gly Gln Pro Lys Ala Ser Pro Ser Val Thr Leu Phe 20 25 30 Pro Pro Ser Ser Glu Glu Leu Gly Ala Asn Lys Ala Thr Leu Val Cys 35 40 45 Leu Ile Ser Asp Phe Tyr Pro Ser Gly Val Thr Val Ala Trp Lys Ala 50 55 60 Asp Gly Ser Pro Val Thr Gln Gly Val Glu Thr Thr Lys Pro Ser Lys 70
75
80 Gln Ser Asn Asn Lys Tyr Ala Ala Ser Ser Tyr Leu Ser Leu Thr Pro 85 90 95 Asp Lys Trp Lys Ser His Ser Ser Phe Ser Cys Leu Val Thr His Glu 100 105 110 105 Gly Ser Thr Val Glu Lys Lys Val Ala Pro Ala Glu Cys Ser 115 120 125

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48

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gcc tcc cga gca gct gca ggg ttt gtg ctc gag gct gtg tca cag tgt Ala Ser Arg Ala Ala Ala Gly Phe Val Leu Glu Ala Val Ser Gln Cys 35 40 45	144
att gtg ttc ggc gga ggc acc cat ctg acc gtc ctc ggt cag ccc aag Ile Val Phe Gly Gly Gly Thr His Leu Thr Val Leu Gly Gln Pro Lys 50 55 60	192
gcc tcc cct tcg gtc aca ctc ttc ccg ccc tcc tct gag gag ctt ggc Ala Ser Pro Ser Val Thr Leu Phe Pro Pro Ser Ser Glu Glu Leu Gly 65 70 75 80	240
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Ile val Phe Gly Gly Gly Thr His Leu Thr Val Leu Gly Gln Pro Lys 50 55 60	

Ala Ser Pro Ser Val Thr Leu Phe Pro Pro Ser Ser Glu Glu Leu Gly 65 70 75 80	
Ala Asn Lys Ala Thr Leu Val Cys Leu Ile Ser Asp Phe Tyr Pro Ser 85 90 95	
Gly Val Thr Val Ala Trp Lys Ala Asp Gly Ser Pro Ile Thr Gln Gly 100 105 110	
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SEQUENCE LISTING(09062) Ser Val Ala Ser Tyr Val Leu Thr Gln Ser Pro Ser Val Ser Val Thr 20 25 30 ctg gga cag acg gcc agc atc acc tgt agg gga aac agc att gga agg Leu Gly Gln Thr Ala Ser Ile Thr Cys Arg Gly Asn Ser Ile Gly Arg 144 40 aaa gat gtt cat tgg tac cag cag aag ccg ggc caa gcc ccc ctg ctg Lys Asp Val His Trp Tyr Gln Gln Lys Pro Gly Gln Ala Pro Leu Leu 192 55 60 att atc tat aat gat aac agc cag ccc tca ggg atc cct gag cga ttc Ile Ile Tyr Asn Asp Asn Ser Gln Pro Ser Gly Ile Pro Glu Arg Phe 65 70 75 80 240 tct ggg acc aac tca ggg agc acg gcc acc ctg acc atc agt gag gcc Ser Gly Thr Asn Ser Gly Ser Thr Ala Thr Leu Thr Ile Ser Glu Ala 288 85 90 caa acc aac gat gag gct gac tat tac tgc cag gtg tgg gaa agt agc Gln Thr Asn Asp Glu Ala Asp Tyr Tyr Cys Gln Val Trp Glu Ser Ser 100 105 110 336 gct gat tgt tgg gta ttc ggt gaa ggg acc cag ctg acc gtc ctc ggt Ala Asp Cys Trp Val Phe Gly Glu Gly Thr Gln Leu Thr Val Leu Gly 115 120 125 384 cag ccc aag tcc tcc ccc ttg gtc aca ctc ttc ccg ccc tcc tct gag Gln Pro Lys Ser Ser Pro Leu Val Thr Leu Phe Pro Pro Ser Ser Glu 432 gag ctc ggc gcc aac aag gct acc ctg gtg tgc ctc atc agc gac ttc Glu Leu Gly Ala Asn Lys Ala Thr Leu Val Cys Leu Ile Ser Asp Phe 145 150 155 160 480 160 tac ccc agt ggc ctg aaa gtg gct tgg aag gca gat ggc agc acc atc Tyr Pro Ser Gly Leu Lys Val Ala Trp Lys Ala Asp Gly Ser Thr Ile 165 170 175 528 atc cag ggc gtg gaa acc acc aag ccc tcc aag cag agc aac aag Ile Gln Gly Val Glu Thr Thr Lys Pro Ser Lys Gln Ser Asn Asn Lys 576 180 185 190 tac acg gcc agc agc tac ctg agc ctg acg cct gac aag tgg aaa tct Tyr Thr Ala Ser Ser Tyr Leu Ser Leu Thr Pro Asp Lys Trp Lys Ser 195 200 205 624 cac agc agc ttc agc tgc ctg gtc acg cac cag ggg agc acc gtg gag His Ser Ser Phe Ser Cys Leu Val Thr His Gln Gly Ser Thr Val Glu 672 215 aag aag gtg gcc cct gca gag tgc tct tag gtccctgaga attcctgaga Lys Lys Val Ala Pro Ala Glu Cys Ser 225 230 722 tggagcette etcaeccaga cacceettee ceagtteace ttgtgeecet gaaaaeceae 782 cctggaccag ctcagaccag gcaggtcact catcctccct gtttctactt gtgctcaata 842 aagactttat catttatcac tg 864 <210> 59

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864

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I.

I

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130 135 140
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Gln Thr Asn Asp Glu Ala Asp Tyr Tyr Cys Gln Glu Met His Thr Pro 100 105 110
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Lys Asp Val His Trp Tyr Gln Gln Lys Pro Gly Gln Ala Pro Leu Leu Page 66										

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