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(54) Titre : ANTICORPS THERAPEUTIQUES CONTRE LA PROTEINE ROR-1 ET LEURS METHODES D'UTILISATION
(54) Title: THERAPEUTIC ANTIBODIES AGAINST ROR-1 PROTEIN AND METHODS FOR USE OF SAME

(57) **Abrégé/Abstract:**

Therapeutic antibodies having binding specificity for ROR-1 (receptor tyrosine kinase like orphan receptor one) expressed on cancer cells (particularly leukemic and lymphomic cells) and pharmaceutical compositions containing one or more such antibodies for use in treating cancer. Methods for diagnosing such cancers through in vitro detection of binding to ROR-1 protein expressed on putative cancer cells are also provided.



ABSTRACT

Therapeutic antibodies having binding specificity for ROR-1 (receptor tyrosine kinase like orphan receptor one) expressed on cancer cells (particularly leukemic and lymphomic cells) and pharmaceutical compositions containing one or more such antibodies for use in treating cancer. Methods for diagnosing such cancers through in vitro detection of binding to ROR-1 protein expressed on putative cancer cells are also provided.

**THERAPEUTIC ANTIBODIES AGAINST ROR-1 PROTEIN
AND METHODS FOR USE OF SAME**

BACKGROUND

[0001] Tyrosine kinases are important mediators of the signaling cascade, determining key roles in diverse biological processes like growth, differentiation, metabolism and apoptosis in response to external and internal stimuli. Studies have implicated the role of tyrosine kinases in the pathophysiology of cancer. Schlessinger J. (2000) *Cell*, 103:211-225; and Robinson et al. (2000) *Oncogene*, 19:5548-5557. MacKeigan and colleagues used a large-scale RNAi approach to identify kinases that might regulate survival and apoptosis of a human tumor cell line (HeLa), RNAi to ROR1 was found as one of the most potent in inducing apoptosis among the set of RNAi targeting each of 73 different kinase-encoding genes. MacKeigan et al. (2005) *Nat Cell Biol.*, 7:591-600. However, these investigators did not examine the expression or function of ROR1 protein in these cells.

[0002] ROR1, receptor tyrosine kinase like orphan receptor one, is a molecule expressed at high levels during embryogenesis that plays a major role in the development of the skeleton, lungs and nervous system. ROR1 expression is greatly decreased in postpartum mammalian cells to levels that are barely detectable. ROR1 is a membrane-receptor with an intracellular kinase-like domain and extracellular Frizzled-like cysteine-rich domain, which is common to receptors of members of the Wnt-family. ROR1 is member of the ROR family that is evolutionarily conserved among *Caenorhabditis elegans*, *Drosophila*, mice and humans. Wilson C, Goberdhan DC, Steller H. Dror, a potential neurotrophic receptor gene, encodes a *Drosophila* homolog of the vertebrate Ror family of Trk-related receptor tyrosine kinases. *Proc Natl Acad Sci U S A.* 1993;90:7109-7113; Oishi et al. (1997) *J Biol Chem.*, 272:11916-11923; Masiakowski et al. (1992) *J Biol Chem.*, 267:26181-26190; Forrester et al. (2002) *Cell Mol Life Sci.*, 59:83-96; and Oishi et al. (1999) *Genes Cells*, 4:41-56. The actual functional role of the ROR1 protein during embryogenesis is unknown, although it is believed to be a receptor for Wnt proteins that regulate cellular polarity and cell-to-cell interactions.

[0003] Although principally an embryonic protein, ROR1 is expressed uniquely on certain cancer cells, including in CLL, small lymphocytic lymphoma, marginal cell B-Cell

lymphoma, Burkett's Lymphoma, and other cancers (e.g., breast cancers), but not on normal adult tissues and cells. In a recent study, it was found that ROR1, at both mRNA and protein level, was highly expressed in CLL B cells but not normal B cells. Moreover, it was found that ROR1 is a receptor for Wnt5a, which could induce activation of NF- κ B when co-expressed with ROR1 in HEK293 cells and enhance survival of CLL cells *in vitro*. This indicates that ROR1 is a CLL survival-signaling receptor for Wnt5a. Another study found that ROR1 was expressed in acute lymphocytic leukemia (ALL) as well. Shabani et al. (2007) *Tumour Biol.*, 28:318-326; and Baskar et al. (2008) *Clin Cancer Res.*, 14:396-404. Expression of ROR1 protein has now been demonstrated on a variety of hematologic and solid tumor cancers.

[0004] Therapeutic control of ROR1 expression is necessary. However, although polyclonal anti-ROR1 antibodies raised against ROR1 peptide are commercially available. The inventors developed a monoclonal anti-ROR1 antibody, terms 4A5, which reacts with the native ROR1 protein and is capable of detecting cell-surface expression of ROR1 for flow cytometric analysis. However, robustly therapeutic antibodies with demonstrable ability to inhibit ROR-1 mediated cancer cell proliferation to a degree that is therapeutically significant for slowing or preventing growth and metastasis have not been available.

SUMMARY OF THE INVENTION

[0005] The invention provides antibodies and combination of antibodies for *in vivo* and *in vitro* inhibition of ROR-1 cell mediated proliferation of cells from subjects with cancer, including lymphomas, CLL, small lymphocytic lymphoma, marginal cell B-Cell lymphoma, Burkett's Lymphoma, renal cell carcinoma, colon cancer, colorectal cancer, breast cancer, epithelial squamous cell cancer, melanoma, myeloma, stomach cancer, brain cancer, lung cancer, pancreatic cancer, cervical cancer, ovarian cancer, liver cancer, bladder cancer, prostate cancer, testicular cancer, thyroid cancer, and head and neck cancer, but not in blood or splenic lymphocytes of nonleukemic patients or normal adults.

[0006] The antibodies of the invention are also useful for differentiation between ROR1 expressing cancer cells ("ROR1 cancer") and normal cells. For example, an immunoassay that detects ROR1 in a sample from a subject by contacting the sample with a ROR1-specific

antibody of the invention and detecting immunoreactivity between the antibody and ROR1 in the sample is provided.

[0007] In accordance with a further aspect of the invention, a ROR1 cancer is diagnosed in a subject by detecting the presence or quantity of ROR1 protein in a sample.

[0008] The present invention includes compositions that include purified, isolated monoclonal antibodies and combinations thereof that bind specifically to ROR1 receptor protein.

BRIEF DESCRIPTION OF THE DRAWINGS

[0009] Figure 1 is a series of graphs illustrating the results of flow cytometric analysis of the expansion of CD5⁺B220^{low} leukemia B cells in ROR1 Tg mice following the adoptive transfer of 1×10^7 splenocytes from a ROR1 xTCL1 Tg mouse. Upper panel depicts the expansion from 2 to 4 weeks following adoptive transfer. Percentage of leukemic cells on the contour plot of mCD5 (x-axis) vs mB220 (y-axis) is indicated on above the gate on CD5⁺B220^{low} lymphocytes. Bottom panel depicts the relative ROR1 expression (x axis) using the mouse anti-ROR1 4A5 mAb.

[0010] Figure 2 is a diagram outlining the analysis of anti-ROR1 mAb on the adoptive transfer and engraftment of ROR1 XTCL1 leukemic splenocytes. ROR1 Tg mice (4 mice /group) were given 250ug of 4A5, D10 or control mIgG i.v. on day 0. The following day, 1×10^7 splenocytes from a ROR1 x TCL1 Tg mouse were adoptively transferred i.v. All mice were subsequently monitor weekly for expansion of CD5⁺B220^{low} leukemic B cells by flow cytometry beginning at 2 weeks post transfer.

[0011] Figure 3 is a series of graphs illustrating the results of a flow cytometric analysis which demonstrate that anti-ROR1 antibodies of the invention inhibited the development of CLL-like leukemia in ROR1 Tg mice. 2 weeks after adoptive transfer, the PBMC facs analysis were performed. The data showed the anti-ROR1 antibody D10 but not anti-ROR1 antibody 4A5 could markedly inhibit the CD5^{dull}B220⁺ and ROR1^{bright}B220⁺ leukemic B cell expansion.

[0012] Figure 4A is a series of graphs illustrating the results of *in vivo* testing in a murine model of human breast cancer. The anti-ROR1 antibodies inhibited breast cancer metastasis

in rag-/- g-/- deficiency mice. 5E5 MDA-MB-231 breast cancer cell were transferred by i.v. injection to rag-/- g-/- mice on day 1. The rag-/- g-/- deficiency mice were also i.v. injected isotype control or anti-ROR1 antibody (4A5, D10, and 4A5 plus D10) on day 1,3, 7 and 14 at 100 mg per mice. Figure 4A (center) also provides images from IVIS in vivo imaging procedures on the above mice, which were performed every week. 5 weeks later, the mice were sacrificed and histology analysis were performed (Figure 4B). The anti-ROR1 antibody D10 and the antibody combination (4A5 plus D10) both significantly inhibited metastasis of the breast cancer, with inhibition by D10 alone being greater than inhibition by 4a5 alone.

[0013] Figure 5 provides a nucleotide coding sequence comparison of 4A5 Ig heavy chain (VH) to the closest germline mouse and human immunoglobulin (Ig) VH.

[0014] Figure 6 provides a nucleotide coding sequence comparison of G6 Ig heavy chain (VH) to the closest germline mouse and human immunoglobulin (Ig) VH.

[0015] Figure 7 provides a nucleotide coding sequence comparison of G3 Ig heavy chain (VH) to the closest germline mouse and human immunoglobulin (Ig) VH.

[0016] Figure 8 provides a nucleotide coding sequence comparison of H10 Ig heavy chain (VH) to the closest germline mouse and human immunoglobulin (Ig) VH.

[0017] Figure 9 provides a nucleotide coding sequence comparison of D10 Ig heavy chain (VH) to the closest germline mouse and human immunoglobulin (Ig) VH.

[0018] Figure 10 is a diagram and chart depicting the highly conserved nature of human and murine ROR1.

[0019] Figure 11 is a nucleotide comparison depicting the domain structure and sequence homology of human and murine ROR1 extracellular protein.

[0020] Figure 12 is a chart indicating the extracellular domain which the anti-ROR1 mAbs bind the ROR1 protein.

[0021] Figure 13 is a diagram depicting the chimeric ROR1 proteins generated to determine the binding domain of each of the anti-ROR1 mAbs.

[0022] Figure 14 is a diagram depicting the truncated ROR1 proteins generated to determine the sub-regions which each of the anti-ROR1 mAbs binds.

[0023] Figure 15 is a diagram depicting the amino acids which were murinized to determine residues critical for mAb binding to human ROR1 and a western blot showing that the 138 glutamic acid residue is critical for antibody D10 binding to human ROR1.

[0024] Figure 16 is a graph indicating the K_D values for antibody D10 (Figure 16a) and 4A5 (Figure 16b).

[0025] Figure 17 is a series of graphs illustrating the anti-ROR1 antibody D10 is highly active in *in vivo* assays.

[0026] Figure 18 is a diagram outlining the analysis of anti-ROR1 mAb on the adoptive transfer and engraftment of ROR1xTCL1 leukemic splenocytes. ROR1 Tg mice (5 mice/group) were given 250 ug of 4A5, D10 or control mIgG i.v. on day 0. The following day, 5×10^5 splenocytes from a ROR1 X TCL1 Tg mouse were adoptively transferred i.v. All mice were subsequently monitored weekly for expansion of $CD5^{\text{dull}}B200^+$ leukemic B cells by flow cytometry beginning at 2 weeks post transfer.

[0027] Figure 19 a series of graphs illustrating the results of flow cytometric analysis of the anti-ROR1 antibodies inhibiting the development of CLL-like leukemia in ROR1 Tg mice. 2 weeks after adoptive transfer, the PBMC facs analysis were performed. The data showed the anti-ROR1 antibody D10 but not anti-ROR1 antibody 4A5 could markedly inhibit the $CD5^{\text{dull}}B220^+$ and $ROR1^{\text{bright}}B220^+$ leukemic B cell expansion.

[0028] Figure 20 is a graph illustrating that anti-ROR1 antibody D10 inhibits the development and expansion of ROR1xTCL1 leukemic B cells in the blood of recipient animals until two weeks after receiving the last infusion of the mAb.

[0029] Figure 21 is a depiction of the rapid internalization of the anti-ROR1 antibody D10 into CLL cells.

[0030] Figure 22 is a series of graphs illustrating the results of flow cytometric analysis showing that anti-ROR1 antibodies D10 and 4A5 are both internalized into CLL cells. CLL cells were incubated with mouse anti-hROR1 Ab-Alex647 for 30 min at 4°C. Subsequently

the cells were washed and either left at 4°C or incubated for 4 hours at 37°C, followed by flow cytometry. The background signal with non-staining is also shown.

[0031] Figure 23 is a graph illustrating the kinetics of the internalization of anti-ROR1 antibodies D10 and 4A5.

[0032] Figure 24 is a diagram depicting the amino acids which were murinized to determine residues critical for mAb binding to human ROR1 and a western blot showing that the 111 isoleucine residue is critical for antibody 4A5 binding to human ROR1.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

[0033] The presently disclosed subject matter are described more fully below. However, the presently disclosed subject matter may be embodied in many different forms and should not be construed as limited to the embodiments set forth herein; rather, these embodiments are provided so that this disclosure will satisfy applicable legal requirements. Indeed, many modifications and other embodiments of the presently disclosed subject matter set forth herein will come to mind to one skilled in the art to which the presently disclosed subject matter pertains having the benefit of the teachings presented in the foregoing descriptions and the associated Figures. Therefore, it is to be understood that the presently disclosed subject matter is not to be limited to the specific embodiments disclosed and that modifications and other embodiments are intended to be included within the scope of the appended claims.

[0034] Antibodies of the invention were produced monoclonally using techniques as previously described. Briefly, Naturally occurring antibodies are generally tetramers containing two light chains and two heavy chains. Experimentally, antibodies can be cleaved with the proteolytic enzyme papain, which causes each of the heavy chains to break, producing three separate subunits. The two units that consist of a light chain and a fragment of the heavy chain approximately equal in mass to the light chain are called the Fab fragments (*i.e.*, the antigen binding fragments). The third unit, consisting of two equal segments of the heavy chain, is called the Fc fragment. The Fc fragment is typically not involved in antigen-antibody binding, but is important in later processes involved in ridding the body of the antigen.

[0035] Because Fab and F(ab')₂ fragments are smaller than intact antibody molecules, more antigen-binding domains are available than when whole antibody molecules are used. Proteolytic cleavage of a typical IgG molecule with papain is known to produce two separate antigen binding fragments called Fab fragments which contain an intact light chain linked to an amino terminal portion of the contiguous heavy chain via by disulfide linkage. The remaining portion of the papain-digested immunoglobulin molecule is known as the Fc fragment and consists of the carboxy terminal portions of the antibody left intact and linked together via disulfide bonds. If an antibody is digested with pepsin, a fragment known as an F(ab')₂ fragment is produced which lacks the Fc region but contains both antigen-binding domains held together by disulfide bonds between contiguous light and heavy chains (as Fab fragments) and also disulfide linkages between the remaining portions of the contiguous heavy chains (Handbook of Experimental Immunology. Vol 1: Immunochemistry, Weir, D. M., Editor, Blackwell Scientific Publications, Oxford (1986)).

[0036] As readily recognized by those of skill in the art, altered antibodies (*e.g.*, chimeric, humanized, CDR-grafted, bifunctional, antibody polypeptide dimers (*i.e.*, an association of two polypeptide chain components of an antibody, *e.g.*, one arm of an antibody including a heavy chain and a light chain, or an Fab fragment including VL, VH, CL and CH antibody domains, or an Fv fragment comprising a VL domain and a VH domain), single chain antibodies (*e.g.*, an scFv (*i.e.*, single chain Fv) fragment including a VL domain linked to a VH domain by a linker, and the like) can also be produced by methods well known in the art.

[0037] Monoclonal antibody (mAb) technology can be used to obtain mAbs to ROR1. Briefly, hybridomas are produced using spleen cells from mice immunized with ROR1 antigens. The spleen cells of each immunized mouse are fused with mouse myeloma Sp 2/0 cells, for example using the polyethylene glycol fusion method of Galfre, G. and Milstein, C., Methods Enzymol., 73:3-46 (1981). Growth of hybridomas, selection in HAT medium, cloning and screening of clones against antigens are carried out using standard methodology (Galfre, G. and Milstein, C., Methods Enzymol., 73:3-46 (1981)).

[0038] HAT-selected clones are injected into mice to produce large quantities of mAb in ascites as described by Galfre, G. and Milstein, C., Methods Enzymol., 73:3-46 (1981), which can be purified using protein A column chromatography (BioRad, Hercules, Calif.).

mAbs are selected on the basis of their (a) specificity for ROR1, (b) high binding affinity, (c) isotype, and (d) stability.

[0039] mAbs can be screened or tested for ROR1 specificity using any of a variety of standard techniques, including Western Blotting (Koren, E. et al., *Biochim. Biophys. Acta* 876:91-100 (1986)) and enzyme-linked immunosorbent assay (ELISA) (Koren, E. et al., *Biochim. Biophys. Acta* 876:91-100 (1986)).

[0040] Humanized forms of mouse antibodies can be generated by linking the CDR regions of non-human antibodies to human constant regions by recombinant DNA techniques (see, e.g., Queen et al., *Proc. Natl. Acad. Sci. USA* 86:10029-10033, 1989 and WO 90/07861).

Human antibodies can be obtained using phage-display methods (see, e.g., Dower et al., WO 91/17271; McCafferty et al., WO 92/01047). In these methods, libraries of phage are produced in which members display different antibodies on their outer surfaces. Antibodies are usually displayed as Fv or Fab fragments. Phage displaying antibodies with a desired specificity may be selected by affinity enrichment.

[0041] Human antibodies may be selected by competitive binding experiments, or otherwise, to have the same epitope specificity as a particular mouse antibody. Using these techniques, a humanized ROR1 antibody having the human IgG1 constant region domain and the human kappa light chain constant region domain with the mouse heavy and light chain variable regions. The humanized antibody has the binding specificity of a mouse ROR1 mAb, specifically the 4A5 mAb described in Examples 4 and 5.

[0042] It may be desirable to produce and use functional fragments of a mAb for a particular application. The well-known basic structure of a typical IgG molecule is a symmetrical tetrameric Y-shaped molecule of approximately 150,000 to 200,000 daltons consisting of two identical light polypeptide chains (containing about 220 amino acids) and two identical heavy polypeptide chains (containing about 440 amino acids). Heavy chains are linked to one another through at least one disulfide bond. Each light chain is linked to a contiguous heavy chain by a disulfide linkage. An antigen-binding site or domain is located in each arm of the Y-shaped antibody molecule and is formed between the amino terminal regions of each pair of disulfide linked light and heavy chains. These amino terminal regions of the light and heavy chains consist of approximately their first 110 amino terminal amino

acids and are known as the variable regions of the light and heavy chains. In addition, within the variable regions of the light and heavy chains there are hypervariable regions which contain stretches of amino acid sequences, known as complementarity determining regions (CDRs). CDRs are responsible for the antibody's specificity for one particular site on an antigen molecule called an epitope. Thus, the typical IgG molecule is divalent in that it can bind two antigen molecules because each antigen-binding site is able to bind the specific epitope of each antigen molecule. The carboxy terminal regions of light and heavy chains are similar or identical to those of other antibody molecules and are called constant regions. The amino acid sequence of the constant region of the heavy chains of a particular antibody defines what class of antibody it is, for example, IgG, IgD, IgE, IgA or IgM. Some classes of antibodies contain two or more identical antibodies associated with each other in multivalent antigen-binding arrangements.

[0043] Fab and F(ab')₂ fragments of mAbs that bind ROR1 can be used in place of whole mAbs. Because Fab and F(ab')₂ fragments are smaller than intact antibody molecules, more antigen-binding domains are available than when whole antibody molecules are used. Proteolytic cleavage of a typical IgG molecule with papain is known to produce two separate antigen binding fragments called Fab fragments which contain an intact light chain linked to an amino terminal portion of the contiguous heavy chain via by disulfide linkage. The remaining portion of the papain-digested immunoglobulin molecule is known as the Fc fragment and consists of the carboxy terminal portions of the antibody left intact and linked together via disulfide bonds. If an antibody is digested with pepsin, a fragment known as an F(ab')₂ fragment is produced which lacks the Fc region but contains both antigen-binding domains held together by disulfide bonds between contiguous light and heavy chains (as Fab fragments) and also disulfide linkages between the remaining portions of the contiguous heavy chains (Handbook of Experimental Immunology. Vol 1: Immunochemistry, Weir, D. M., Editor, Blackwell Scientific Publications, Oxford (1986)).

[0044] With respect to particular antibodies, "specific binding" refers to antibody binding to a predetermined antigen. Typically, the antibody binds with an affinity corresponding to a K_D of about 10^{-8} M or less, and binds to the predetermined antigen with an affinity (as expressed by K_D) that is at least 10 fold less, and preferably at least 100 fold less than its affinity for binding to a non-specific antigen (*e.g.*, BSA, casein) other than the predetermined

antigen or a closely-related antigen. Alternatively, the antibody can bind with an affinity corresponding to a K_A of about $10^6 M^{-1}$, or about $10^7 M^{-1}$, or about $10^8 M^{-1}$, or $10^9 M^{-1}$ or higher, and binds to the predetermined antigen with an affinity (as expressed by K_A) that is at least 10 fold higher, and preferably at least 100 fold higher than its affinity for binding to a non-specific antigen (e.g., BSA, casein) other than the predetermined antigen or a closely-related antigen.

[0045] Also, reference to “an antibody having binding specificity for ROR-1 protein” includes antibody fragments having at least 90% or 95% sequence identity to any of the polypeptide sequences disclosed in SEQ ID NOs: 2, 4, 6, 8, 12, 14, 16, 18 and 20, including variants modified by mutation to improve the utility thereof (e.g., improved ability to target specific cell types and the like). Such variants include those wherein one or more conservative substitutions are introduced into the heavy chain and/or the light chain of the antibody.

[0046] Such variants include those wherein one or more substitutions are introduced into the heavy chain nucleotide sequence and/or the light chain nucleotide sequence of the antibody. In some embodiments the variant has a light chain and/or heavy chain having a nucleotide sequence at least 80% or at least 90% or at least 95% identical to any of the nucleotide sequences set forth in SEQ ID NOs: 1, 3, 5, 7, 11, 13, 15, 17 and 19.

[0047] Polynucleotide sequences which code structural features of the antibodies of the invention include those whose sequences are set forth below. Each polynucleotide sequence is followed by the amino acid sequence of the encoded polypeptide. The light chain sequences which are considered to be “corresponding” to heavy chain sequences are those listed as being for the same antibody; i.e., the F2 heavy chain sequences correspond to the F2 light chain sequences, the D10 heavy chain sequences correspond to the D10 light chain sequences, and so forth.

SEQ ID NO: 1 4A5 Mouse Anti-ROR1 mAb Heavy Chain Variable Region Coding Sequence:

GAAGTGAAACTGGTGGAGTCTGGGGGAGGCTTAGTGAAGCCTGGAGGGTCCCTGAAACTCTC
CTGTGCAGCCTCTGGATT

CACTTTCAGTAGCTATGCCATGTCTTGGGTTCCGAGATTCCAGAGAAGAGGCTGGAGTGGG
TCGCATCCATTAGTCGTG

GTGGTACCACCTACTATCCAGACAGTGTGAAGGGCCGATTCACCATCTCCAGAGATAATGTC
AGGAACATCCTGTACCTG

CAAATGAGCAGTCTGAGGTCTGAGGACACGGCCATGTATTACTGTGGAAGATATGATTACGA
CGGGTACTATGCAATGGA

CTACTGGGGTCAAGGAACCTCAGTCACCGTCTCCTCA

SEQ ID NO: 2 4A5 Mouse Anti-ROR1 mAb Heavy Chain Variable Region Polypeptide
Sequence:

EVKLVESGGGLVKPGGSLKLSCAASGFTFSSYAMSWVRQIPEKRLEWVASISRGGTTYYPDS
VKGRFTISRDNVRNILYL

QMSSLRSEDAMYYCGRYDYGYYAMDYWGQTSVTVSS

SEQ ID NO: 3 4A5 Mouse Anti-ROR1 mAb Light Chain Variable Region Coding
Sequence:

GACATCAAGATGACCCAGTCTCCATCTTCCATGTATGCATCTCTAGGAGAGAGAGTCACTAT
CACTTGCAAGGCGAGTCC

GGACATTAATAGCTATTTAAGCTGGTTCCAGCAGAAACCAGGGAAATCTCCTAAGACCCTGA
TCTATCGTGCAAACAGAT

TGGTTGATGGGGTCCCATCAAGGTTTCAGTGGCGGTGGATCTGGGCAAGATTATTCTCTCACC
ATCAACAGCCTGGAGTAT

GAAGATATGGGAATTTATTATTGTCTACAGTATGATGAATTTCCGTACACGTTCCGGAGGGGG
GACCAAGCTGGAAATGAA

AC

SEQ ID NO: 4 4A5 Mouse Anti-ROR1 mAb Light Chain Variable Region Polypeptide

Sequence:

DIKMTQSPSSMYASLGERTITCKASPDINSYLSWFQQKPGKSPKTLIYRANRLVDGVPSRF
SGGGSGQDYSLTINSLEY

EDMGIYYCLOQYDEFPYTFGGGTKLEMK

SEQ ID NO: 5 F2, F12 and G6 Mouse Anti-ROR1 mAb Heavy Chain Variable Region

Coding Sequence:

GAGGTCCAGCTACAGCAGTCTGGACCTGAGCTGGAGAAGCCTGGCGCTTCAGTGAAGATATC
CTGCAAGGCTTCTGGTTT

CGCATTCACTGGCTACAACATGAACTGGGTGAAACAGACCAATGGAAAGAGCCTTGAGTGGA
TTGGAAGTATTGATCCTT

ACTATGGTGGTTCTACCTACAACCAGAAGTTCAAGGACAAGGCCACATTGACTGTAGACAAA
TCCTCCAGCACAGCCTAC

ATGCAACTCAAGAGCCTCACATCTGATGACTCTGCAGTCTATTACTGTGCAAGATCCCCGGG
GGGGACTATGCTATGGA

CTACTGGGGTCAAGGAACCTCAGTCACCGTCTCCTCA

SEQ ID NO: 6 F2, F12 and G6 Mouse Anti-ROR1 mAb Heavy Chain Variable Region

Polypeptide Sequence:

EVQLQQSGPELEKPGASVKISCKASGFAFTGYNMNWKQTNGKSLEWIGSIDPYYGGSTYNQ
KFKDKATLTVDKSSSTAY

MQLKSLTSDDSAVYYCARSPGGDYAMDYWGQGTSVTVSS

SEQ ID NO: 7 F2, F12 and G6 Mouse Anti-ROR1 mAb Light Chain Variable Region

Coding Sequence:

GACATCAAGATGACCCAGTCTCCATCTTCCATGTATGCATCTGTAGGAGAGAGAGTCACTAT
CACTTGTAAGGCGAGTCA

GGGCATTAATAGCTATTCAGGCTGGTTCAGCAGAAACCAGGGAAATCTCCTAAGACCCTGA
TTTATCGTGGAATAGAT

TGGTGGATGGGGTCCCATCAAGGTTCAAGTGGCAGTGGATCTGGGCAAGATTATTCTCTCACC
ATCAGCAGCCTGGAGTAT

GAAGATATGGGAATTTATTATTGTCTACAGTATGATGAGTTTCCGTACACGTTCCGGAGGGGG
GACCAAGCTGGAAATAAA

AC

SEQ ID NOs: 8 F2, F12 and G6 Mouse Anti-ROR1 mAb Light Chain Variable Region
Polypeptide Sequence:

DIKMTQSPSSMYASVGERVTITCKASQGINSYSGWFQQKPGKSPKTLIYRGNRLVDGVPSRF
SGSGSGQDYSLTISSLEY

EDMGIYYCLQYDEFPTYTFGGGTKLEIK

SEQ ID NO: 9 G3 Mouse Anti-ROR1 mAb Heavy Chain Variable Region Coding
Sequence:

CAGGTCCAACCTGCAGCAGCCTGGGGCTGAGCTTGTGAAGCCTGGGACTTCAGTGAAGCTGTC
CTGCAAGGCTTCTGGCTA

CAACTTCACCAACTACTGGATAAACTGGGTGAAGCTGAGGCCTGGACAAGGCCTTGAGTGGA
TTGGAGAAATTTATCCTG

GTAGTGGTAGTACTAATTACAATGAGAAGTTCAAGAGCAAGGCCACACTGACTGCAGACACA
TCCTCCAGCACAGCCTAC

ATGCAACTCAGCAGCCTGGCATCTGAAGACTCTGCTCTCTATTACTGTGCAAGAGATGGTAA
CTACTATGCTATGGACTA

CTGGGGTCAAGGAACCTCAGTCACCGTCTCCTCA

SEQ ID NO: 10 G3 Mouse Anti-ROR1 mAb Heavy Chain Variable Region Polypeptide

Sequence:

QVQLQQPGAELVKPGTSVKLSCKASGYNFTNYWINWVKLRPGQGLEWIGEIYPGSGSTNYNE
KFKSKATLTADTSSSTAY

MLSSLASEDSALYYCARDGNYYAMDYWGQGTSVTVSS

SEQ ID NO: 11 G3 Mouse Anti-ROR1 mAb Light Chain Variable Region Coding

Sequence:

GATATCCAGATGACACAGACTACATCCTCCCTGTCTGCCTCTCTGGGAGACAGAGTCACCAT
CACTTGCAGGGCAAGTCA

GGACATTAACAATTATTTAAACTGGTATCAACAGAAACCAGATGGAAGTGTAAACTCCTGA
TCTACTACACATCAGCAT

TACACTCAGGAGTCCCATCAAGGTTTCAGTGGCAGTGGGTCTGGAACAGATTATTCTCTCACC
ATTAGCAACCTGGAACAA

GAAGATATTGCCACTTACTTTTGCCAACAGGGTAATACGCTTCCTCCGTACACGTTCCGGAGG
GGGGACCAAGCTGGAAAT

AAAAC

SEQ ID NO: 12 G3 Mouse Anti-ROR1 mAb Light Chain Variable Region Polypeptide

Sequence:

DIQMTQTTSSLSASLGDRVTITCRASQDINNYLNWYQQKPDGTVKLLIYYTSALHSGVPSRF
SGSGSGTDYSLTISNLEQ

EDIATYFCQQGNTLPPYTFGGGTKLEIK

SEQ ID NO: 13 D10 Mouse Anti-ROR1 mAb Heavy Chain Variable Region Coding

Sequence:

CAGGTGCAGCTGAAGGAGTCAGGACCTGGCCTGGTGGCGCCCTCACAGACTCTGTCCATCAC
TTGCACTGTCTCTGGGTT

TTCATTAACCAGTTATGGTGTACACTGGGTTCGCCAGCCTCCAGGAAAGGGTCTGGAGTGGC
TGGGAGTAATATGGGCTG

GTGGATTCACAAATTATAATTCGGCTCTCAAGTCCAGACTGAGCATCAGCAAAGACAACCTCC
AAGAGCCAAGTTCTCTTA

AAAATGACCAGTCTGCAAACCTGATGACACAGCCATGTACTACTGTGCCAGGAGAGGTAGTTC
CTATTCTATGGACTATTG

GGGTCAAGGAACCTCAGTCACCGTCTCCTCA

SEQ ID NO: 14 D10 Mouse Anti-ROR1 mAb Heavy Chain Variable Region Polypeptide

Sequence

QVQLKESGPGLVAPSQTLSITCTVSGFSLTSYGVHWVRQPPGKLEWLGVIWAGGFNYNSA
LKSRLSISKDNSKSVLL

KMTSLQTDDTAMYICARRGSSYSMDYWGQGTSVTVSS

SEQ ID NO: 15 D10 Mouse Anti-ROR1 mAb Light Chain Variable Region Coding

Sequence:

GAAATTGTGCTCTCTCAGTCTCCAGCCATCACAGCTGCATCTCTGGGCCAAAAGGTCACCAT
CACCTGCAGTGCCAGTTC

AAATGTAAGTTACATCCACTGGTACCAGCAGAGGTCAGGCACCTCCCCAGACCATGGATTT
ATGAAATATCCAAACTGG

CTTCTGGAGTCCCAGTTCGCTTCAGTGGCAGTGGGTCTGGGACCTCTTACTCTCTCACAATC
AGCAGCATGGAGGCTGAA

GATGCTGCCATTTATTATTGTCAGCAGTGGAATTATCCTCTTATCACGTTCCGGCTCGGGGAC
AAAGTTGGAAATACAA

SEQ ID NO: 16 D10 Mouse Anti-ROR1 mAb Light Chain Variable Region Polypeptide
Sequence:

EIVLSQSPAITAASLGQKVTITCSASSNVSYIHWYQQRSGTSPRPWIYEISKLASGVPVRF
SGSGTSYSLTISSMEAE

DAAIYYCQQWNYPLITFGSGTKLEIQ

SEQ ID NO: 17 H10 and G11 Mouse Anti-ROR1 mAb Heavy Chain Variable Region
Coding Sequence:

GAAGTGAAGCTGGTGGAGTCTGGGGGAGGCTTAGTGAAGCCTGGAGGGTCCCTGAAACTCTC
CTGTGCAGCCTCTGGATT

CACTTTCAGTAGCTATGCCATGTCTTGGGTTCGCCAGACTCCAGAGAAGAGGCTGGAGTGGG
TCGCTTCCATTAGTACTG

GTGCTAGCGCCTACTTTCAGACAGTGTGAAGGGCCGATTCACCATCTCCAGAGATAATGCC
AGGAACATCCTGTACCTG

CAAATGAGCAGTCTGAGGTCTGAGGACACGGCCATGTATTATTGTGCAAGGATTACTACGTC
TACCTGGTACTTCGATGT

CTGGGGCGCAGGGACCACGGTCACCGTCTCCTCA

SEQ ID NO: 18 H10 and G11 Mouse Anti-ROR1 mAb Heavy Chain Variable Region
Polypeptide Sequence:

EVKLVESGGGLVKPGGSLKLSAASGFTFSSYAMSWVRQTPEKRLEWVASISTGASAYFPDS
VKGRFTISRDNARNILYL

QMSSLRSEDAMYYCARITTTSTWYFDVWGAGTTVTVSS

SEQ ID NO: 19 H10 and G11 Mouse Anti-ROR1 mAb Light Chain Variable Region Coding Sequence:

GACATCAAGATGACCCAGTCTCCATCTTCCATGTATGCATCTCTAGGAGAGAGAGTCACTAT
CACTTGCAAGGCGAGTCA

GGACATTAATAGTTATTTAAGCTGGTTCAGCAGAAACCAGGGAAATCTCCTAAGACCCTGA
TCTATCGTGCAAACAGAT

TGGTAGATGGGGTCCCATCAAGGTTCAAGTGGCAGTGGATCTGGGCAAGATTATTCTCTCACC
ATCAGCAGCCTGGAGTAT

GAAGATATGGGAATTTATTATTGTCTACAGTATGATGAGTTTCCGTACACGTTCCGGAGGGGG
GACCAAGCTGGAAATAAA

AC

SEQ ID NO: 20 H10 and G11 Mouse Anti-ROR1 mAb Light Chain Variable Region
Polypeptide Sequence:

DIKMTQSPSSMYASLGERVTITCKASQDINSYLSWFQQKPGKSPKTLIYRANRLVDGVPSRF
SGSGSGQDYSLTISLEY

EDMGIYYCLQYDEFPYTFGGGTKLEIK

[0048] In one aspect, antibodies are provided in which a heavy chain encoded by the polynucleotide sequence of SEQ ID NO:13 and a light chain encoded by the polynucleotide sequence of SEQ ID NO:15.

[0049] In another aspect, an antibody of the present invention contains a heavy chain encoded by the polynucleotide sequence of SEQ ID NO:1 and a light chain encoded by the polynucleotide sequence of SEQ ID NO:3.

[0050] In further aspects, antibodies are provided which have a heavy chain encoded by the polynucleotide sequence of SEQ ID NO: 5 and a light chain encoded by the polynucleotide sequence of SEQ ID NO: 7; or by the polynucleotide sequence of SEQ ID NO: 9 and a light chain encoded by the polynucleotide sequence of SEQ ID NO: 11; or by the

polynucleotide sequence of SEQ ID NO: 15 and a light chain encoded by the polynucleotide sequence of SEQ ID NO: 17.

[0051] In another aspect, antibodies are provided which contain a heavy chain with the polypeptide sequence of SEQ ID NO:14 and a light chain with the polypeptide sequence of SEQ ID NO:16.

[0052] In another aspect, antibodies are provided which contain a heavy chain with the polypeptide sequence of SEQ ID NO:2 and a light chain with the polypeptide sequence of SEQ ID NO:4.

[0053] In one embodiment, isolated polynucleotides which encode an antibody that specifically binds ROR1 protein are provided which are (a) comprised of a heavy chain region coded by polynucleotides having at least 90% sequence identity with any of the sequences selected from the group consisting of SEQ ID NOs: 1, 5, 9, 13 or 17, (b) comprised of a corresponding light chain region encoded by polynucleotides having at least 90% sequence identity with any of the sequences selected from the group consisting of SEQ ID NOs: 3, 7, 11, 15 or 19, and (c) specifically binds either the 3' end or middle portion of the Ig-like region of the extracellular domain of human or murine ROR-1 protein.

[0054] Also provided are antibodies which bind residues within the middle of the Ig-like region of the extracellular domain of human or murine ROR-1 protein (amino acids 1-147 in the human molecule). In one aspect, the antibodies of the present invention bind to amino acids 70-130 of human ROR1. Examples of such antibodies include 4A5, G11, H10 and G3.

[0055] Alternatively or additionally, a residue corresponding to the one found in the extracellular domain of human ROR-1 protein at position 111 is critical to the binding activity of the antibodies.

[0056] Further provided are antibodies that bind residues within the 3' Ig-like region and the linker region between the Ig-like domain and the CRD domain of human or murine ROR-1 protein (amino acids 1-165 in the human molecule). In one aspect, the antibodies of the present invention bind to amino acids 130-165 of human ROR1. Examples of such antibodies include D10, F2, F12 and G6.

[0057] Alternatively or additionally, the antibodies bind a glutamic acid residue corresponding to the one found in the extracellular domain of human ROR-1 protein at position 138.

[0058] Alternatively or additionally, a residue corresponding to the one found in the extracellular domain of human ROR-1 protein at position 138 is critical to the binding activity of the antibodies.

[0059] Alternatively or additionally, the encoded antibody has *in vivo* activity in reducing leukemic or lymphomic cell burden in an art-accepted animal model at a rate of 2-8 times, or at least 2, 3, 4, 5, 6, 7, or 8 times, that of wild-type human anti-ROR1 antibody or monoclonal 4A5 antibody (disclosed herein).

[0060] Alternatively or additionally, the encoded antibody has *in vivo* activity in inhibiting CD5^{dull}B220⁺ and ROR1^{bright}B220⁺ leukemic B cell expansion.

[0061] Alternatively or additionally, the encoded antibody is internalized into leukemic or lymphomic cells at a rate of at least 2 times, or at least 2, 3, 4, 5, 6, 7, 8, 9 or 10 times that of monoclonal antibody 4A5. Such antibodies are particularly useful as carriers for drug delivery into a targeted cell.

[0062] An example of an antibody possessing all of the afore-mentioned functional characteristics is D10, which has a heavy chain region encoded by SEQ ID NO: 13 and a light chain region encoded by SEQ ID NO: 15.

[0063] In another aspect, polypeptides are provided which consist of or comprise antibodies which specifically bind ROR1 protein and are (a) comprised of a heavy chain region having at least 90% sequence identity with any of the sequences of SEQ. ID. NOs: 2, 6, 10, 14 or 18, (b) comprised of a corresponding light chain region having at least 90% sequence identity with any of the sequences of SEQ ID NOs: 4, 8, 12, 16 or 20, and (c) specifically binds either the 3' end or middle portion of the Ig-like region of the extracellular domain of human or murine ROR-1 protein. In one aspect, the isolated polypeptide is an antibody. In a further aspect, the polypeptide is a Fab or F(ab)'2.

[0064] In certain embodiments, an antibody of the present invention may further contain a detectable label. Such labels are known in the art and include radio-isotopes and fluorescent

labels. As such, internalization of a compound evidencing passage through transporters can be detected by detecting a signal from within a cell from any of a variety of reporters. The reporter can be a label such as a fluorophore, a chromophore, a radioisotope. Confocal imaging can also be used to detect internalization of a label as it provides sufficient spatial resolution to distinguish between fluorescence on a cell surface and fluorescence within a cell; alternatively, confocal imaging can be used to track the movement of compounds over time. In another approach, internalization of a compound is detected using a reporter that is a substrate for an enzyme expressed within a cell. Once the complex is internalized, the substrate is metabolized by the enzyme and generates an optical signal or radioactive decay that is indicative of uptake. Light emission can be monitored by commercial PMT-based instruments or by CCD-based imaging systems. In addition, assay methods utilizing LCMS detection of the transported compounds or electrophysiological signals indicative of transport activity are also employed.

[0065] In certain therapeutic embodiments, the selected antibody may be administered alone, in combination with another antibody of the invention, or with one or more combinatorial therapeutic agents to treat an ROR-1 cancer. When one or more the antibodies described herein are administered as therapeutic agents, they may exert a beneficial effect in the subject by a variety of mechanisms. For example, in certain embodiments, antibodies that specifically bind ROR1 are purified and administered to a patient to neutralize one or more forms of ROR1, to block one or more activities of ROR1, or to block or inhibit an interaction of one or more forms of ROR1 with another biomolecule; e.g., to treat CLL or other ROR1 cancers. All such therapeutic methods are practiced by delivery of a therapeutically effective dosage of a pharmaceutical composition containing the therapeutic antibodies and agents, which can be determined by a pharmacologist or clinician of ordinary skill in human cancer immunotherapy.

[0066] In one embodiment, the present invention provides for a method for of treating cancer by the administration to a human subject in need thereof of a therapeutically effective dose of an antibody according to the invention.

[0067] In another embodiment, the present invention provides a method for of treating cancer comprising administration to a human subject in need thereof of a therapeutically effective dose of an antibody according to the invention.

[0068] Advantageously, the methods of the invention provide for reduction of leukemic or lymphomic cell burden (as demonstrated in and equivalent to an art-accepted animal model) of 2-8 times, or at least 2, 3, 4, 5, 6, 7, or 8 times, that of wild-type human anti-ROR1 antibody or monoclonal 4A5 antibody (disclosed herein).

[0069] The methods of the invention further provide a therapeutic approach to inhibiting CD5^{dull}B220⁺ and ROR1^{bright}B220⁺ leukemic B cell expansion.

[0070] As discussed herein, the antibodies of the invention may include humanized antibodies, and can be combined for therapeutic use with additional active or inert ingredients, e.g., in conventional pharmaceutically acceptable carriers or diluents, e.g., immunogenic adjuvants, and optionally with adjunctive or combinatorially active molecules such as anti-inflammatory and anti-fibrinolytic drugs. Antibodies which readily internalize into cells as demonstrated herein with respect to the D10 antibody are also of particular use as carriers for drug delivery into target cells (for example, as shown in Figures 21-23). Those of ordinary skill in the art will be familiar with methods for producing antibody-drug conjugates useful in such drug delivery protocols.

[0071] In carrying out various assay, diagnostic, and therapeutic methods of the invention, it is desirable to prepare in advance kits comprises a combination of antibodies as described herein with other materials. For example, in the case of sandwich enzyme immunoassays, kits of the invention may contain an antibody that specifically binds ROR1 optionally linked to an appropriate carrier, a freeze-dried preparation or a solution of an enzyme-labeled monoclonal antibody which can bind to the same antigen together with the monoclonal antibody or of a polyclonal antibody labeled with the enzyme in the same manner, a standard solution of purified ROR1, a buffer solution, a washing solution, pipettes, a reaction container and the like. In addition, the kits optionally include labeling and/or instructional materials providing directions (i.e., protocols) for the practice of the methods described herein in an assay environment. While the instructional materials typically comprise written or printed materials, they are not limited to such. Any medium capable of storing such instructions and communicating them to an end user is contemplated. Such media include, but are not limited to electronic storage media (e.g., magnetic discs, tapes, cartridges, chips), optical media (e.g., CD ROM), and the like. Such media may include addresses to internet sites that provide such instructional materials.

[0072] In general, an *in vitro* method of diagnosing a ROR-1 cancer will comprise contacting putative cancer cells from a human subject with an antibody according to the invention, and detecting binding with ROR-1 expressed on said cells as compared to expression on post-embryonic human non-cancer cells. All such diagnostic methods are practiced by delivery of a diagnostically effect quantity of antibodies according to the invention, which can be determined by a diagnostician or *in vitro* diagnostic engineer of ordinary skill in human cancer diagnosis.

[0073] The following examples are intended to illustrate but not limit the invention.

EXAMPLE 1: GENERATION OF MONOCLONAL ANTI-ROR1 ANTIBODIES

[0074] For the production of the hybridoma-generated mAbs, mice were inoculated with DNA, protein and adenoviral constructs that express the extracellular portion (AA 1-406) of the ROR1 protein that include the Ig-like, CRD and Kringle domains and adjacent linker regions (Figures 10-11). Because of the high degree of homology between the murine and human molecules, a variety of cytokines and immune stimulatory agents, such as Freund's Complete Adjuvant, were co-injected to maximize the generation of anti-human ROR1 antibodies. Hybridoma-generated mAbs were generated and screened for binding to human and murine ROR1. An example of hybridoma derived mAbs is D10.

EXAMPLE 2: GENERATION OF ANTI-ROR1 ANTIBODIES USING PHAGE DISPLAY

[0075] A second set of antibodies was generated through the use of a proprietary enhanced phage library (Alere, Inc. San Diego). These anti-human ROR1 antibodies bind epitopes that span the entire length of the extra-cellular domain of the ROR1 protein (Figure 12). An example of a phage display derived anti-ROR1 antibody is 4A5.

EXAMPLE 3: IN VITRO ANALYSIS OF ANTI-ROR1 ANTIBODIES

[0076] Antibodies generated through either hybridomas or phage display were screened for binding to human and murine ROR1. It was determined that the anti-ROR1 antibodies D10 and 4A5 bound only to human ROR1 and did not cross react with murine ROR1.

EXAMPLE 4: DETERMINATION OF BINDING SITES FOR ANTI-ROR1 ANTIBODIES

[0077] Because the anti-ROR1 mAbs are species specific, a series of chimeric proteins were generated that were used to determine the binding site for each of the anti-ROR1 mAbs (Figure 13). As a second level screen, a series of deletion constructs were generated to determine the actual extracellular ROR1 domain to which the mAbs bind. Once the binding domain was identified, truncated chimeric ROR1 molecules to identify specific sub-regions were generated that are recognized by the anti-human ROR1 mAbs (Figure 14). As a final step, the actual amino acids targeted by these antibodies were determined. For this final screen, murinized human amino acids in the sub-domain fragments were generated to determine critical residues required for mAb binding (Figure 15). From this screening paradigm, the binding sub-domains for the mAbs were determined (Figure 15). It was determined that the D10 anti-human ROR1 mAb required the glutamic acid residue at position 138 for binding to the Ig-like domain of the human ROR1 molecule. When this amino acid is replaced with the murine molecule's lysine residue, the D10 molecule no longer bound to the ROR1 protein.

[0078] In a similar manner, it was determined that 4A5 anti-human ROR1 mAb required the isoleucine residue at position 111 for binding to human ROR1 molecule (Figure 24). When this amino acid is replaced with the murine molecule's asparagine residue, the 4A5 molecule no longer bound to the ROR1 protein. It was also determined that the anti-ROR1 antibodies G11, H10 and G3 bind the same region as 4A5.

[0079] Using standard cross blocking techniques the binding sites for anti-ROR1 antibodies F2, F12 and G6 were determined. These experiments determined that antibodies F2, F12 and G6 cross block the anti-ROR1 antibody D10, indicating that they share a binding site.

EXAMPLE 5: DETERMINATION OF THE K_D VALUES FOR THE ANTI-ROR1 ANTIBODIES D10 AND 4A5

[0080] The K_D values for the anti-ROR1 antibodies was determined using standard techniques. It was determined that the K_D for the D10 antibody was 40 nM and for the antibody 4A5 was 4 nM (Figures 16A & B).

EXAMPLE 6: IN VIVO ANALYSIS OF ANTI-ROR1 ANTIBODIES

[0081] The D10 mAb was assessed in several in vivo models. In a murine *in vivo* xenograph, niche-dependent, activity model two doses of the mAb were administered at 10 mg/kg against 4 primary patient CLL cells in 76 mice. As shown in Figure 17, D10 mAb substantially eliminated patient CLL cells in a dose dependent manner. In contrast, the 4A5 mAb had minimal activity in these studies even though the kDa of this mAb is 10 fold greater (4 vs. 40) for the D10 mAb.

[0082] In addition to this activity model, the D10 mAb was also tested in an immune competent transgenic mouse model that spontaneously generates leukemic cells expressing the human ROR1 protein (Figures 18-20). The ROR1-specific mAbs D10 and 4A5 or control IgG antibodies (10 mg/kg) were administered before and after adoptive transfer of ROR1xTCL1 CLL B cells into Balb C mice. The D10 mAb, but not control IgG or 4A5, was able to inhibit the development and expansion of the ROR1xTCL1 leukemic B cells in the blood of recipient animals until two weeks after receiving the last infusion of MAb.

[0083] Along with the anti-leukemic activity of this mAb, it has also been shown that the D10 anti-ROR1 antibody is internalized into patient CLL cells and B cell leukemia and lymphoma cell lines at a greater rate and degree than other anti-ROR1 MAbs that bind other antigenic sites on the extracellular portion of the ROR1 protein (Figures 21-23). Because of the absence of the ROR1 protein on post-partum tissues and its rapid rate of internalization, the D10 mAb may serve as an excellent carrier protein for drugs; for example, for use in directed antibody-drug conjugate (ADC) mediated cytotoxicity. Based on these preclinical findings, the D10 mAb has potential to have therapeutic activity against ROR1 expressing leukemias, lymphomas and solid tumor cancers as a targeted therapy and/or conjugated drug carrier.

[0084] Although the foregoing subject matter has been described in some detail by way of illustration and example for purposes of clarity of understanding, it will be understood by those skilled in the art that certain changes and modifications can be practiced within the scope of the appended claims.

THE EMBODIMENTS OF THE INVENTION FOR WHICH AN EXCLUSIVE PROPERTY OR PRIVILEGE IS CLAIMED ARE DEFINED AS FOLLOWS:

1. An isolated antibody that specifically binds receptor tyrosine kinase like orphan receptor one (ROR1) protein and comprises a heavy chain variable region comprising SEQ ID NO:14 and a light chain region comprising SEQ ID NO:16.
2. The antibody according to claim 1, wherein it further specifically binds either the 3' or middle Ig-like region of the extracellular domain of human or murine ROR-1 protein.
3. The antibody according to claim 1, wherein it binds the 3' end of the Ig-like region of the extracellular domain of human or murine ROR-1 protein from position 1-147.
4. The antibody according to claim 3, wherein it further binds a glutamic acid residue corresponding to the one found in the extracellular domain of human ROR-1 protein at position 138.
5. The antibody according to claim 1, wherein it further reduces leukemic or lymphomic cell burden in an art-accepted animal model at a rate of 2-8 times, or at least 2, 3, 4, 5, 6, 7, or 8 times that of wild-type human anti-ROR1 antibody or monoclonal 4A5 antibody.
6. The antibody according to claim 1, wherein it further inhibits CD5^{dull}B220⁺ and ROR1^{bright} B220⁺ leukemic B cell expansion.
7. The antibody according to claim 1, wherein it further is internalized into leukemic or lymphomic cells at a rate of at least 2 times, or at least 2, 3, 4, 5, 6, 7, 8, 9 or 10 times that of monoclonal antibody 4A5.
8. A pharmaceutically acceptable anti-ROR1 antibody composition comprising the antibody according to claim 1 and a pharmaceutically acceptable carrier.
9. An isolated antibody which binds the same epitope as antibody D10, wherein the heavy chain variable region of D10 is encoded by SEQ ID NO:13 and the light chain variable region of D10 is encoded by SEQ ID NO:15.

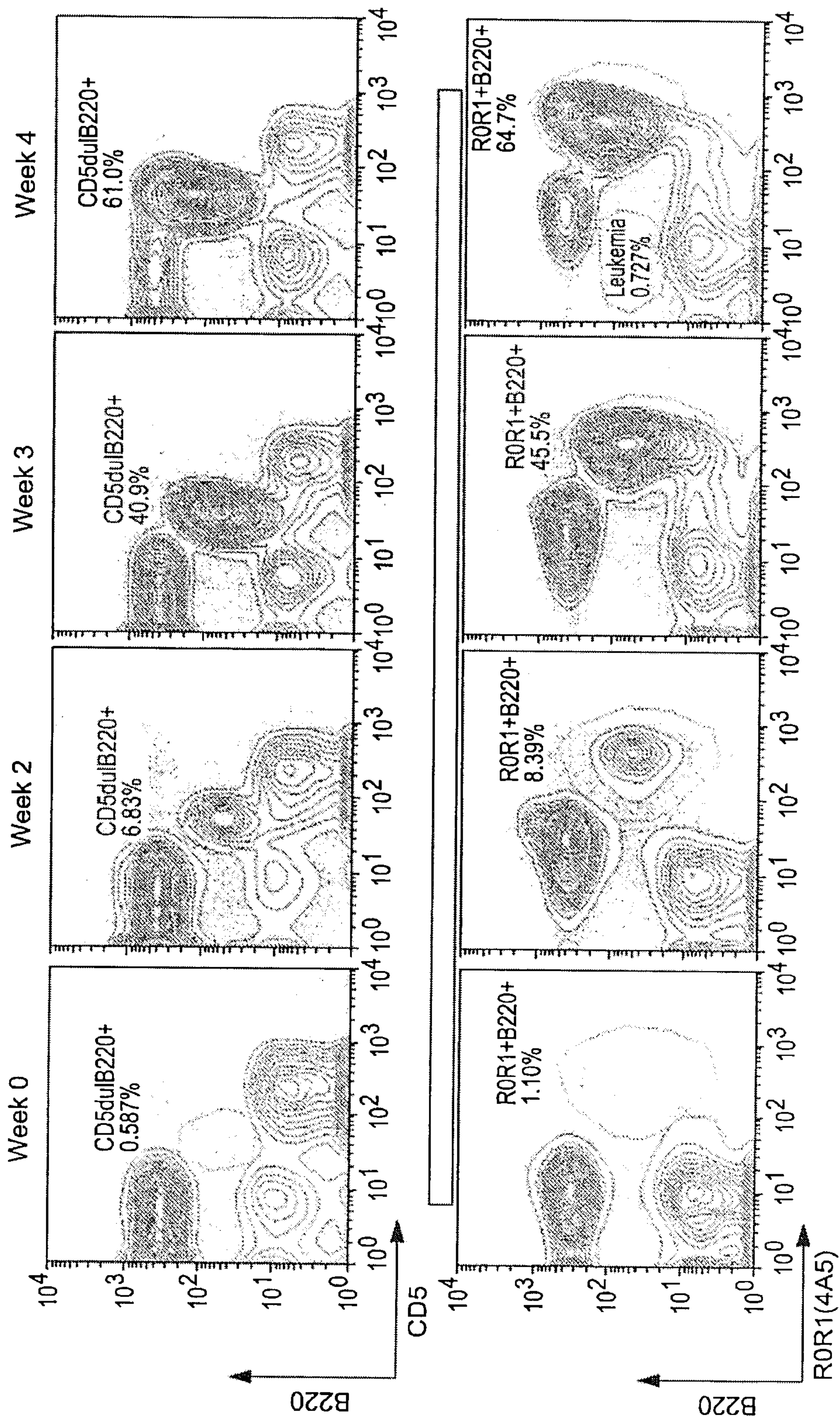


FIG. 1

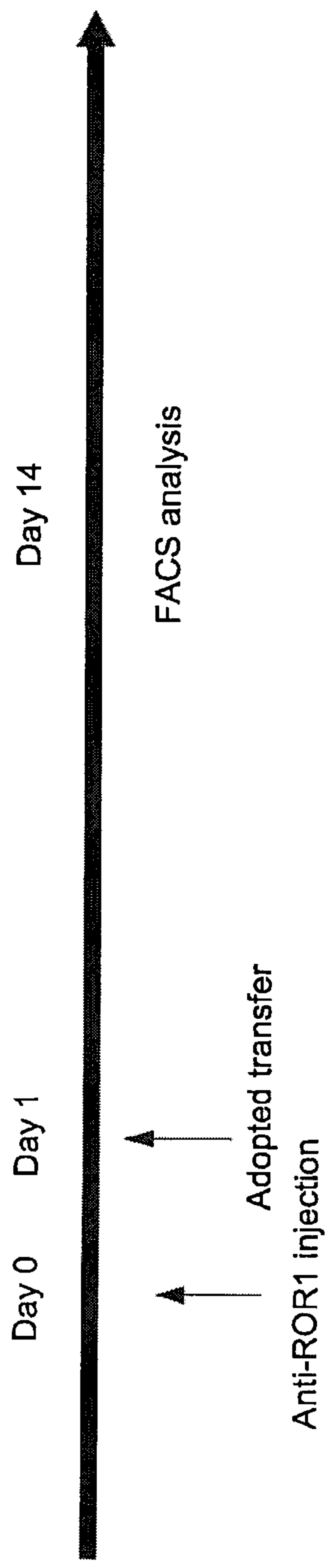


FIG. 2

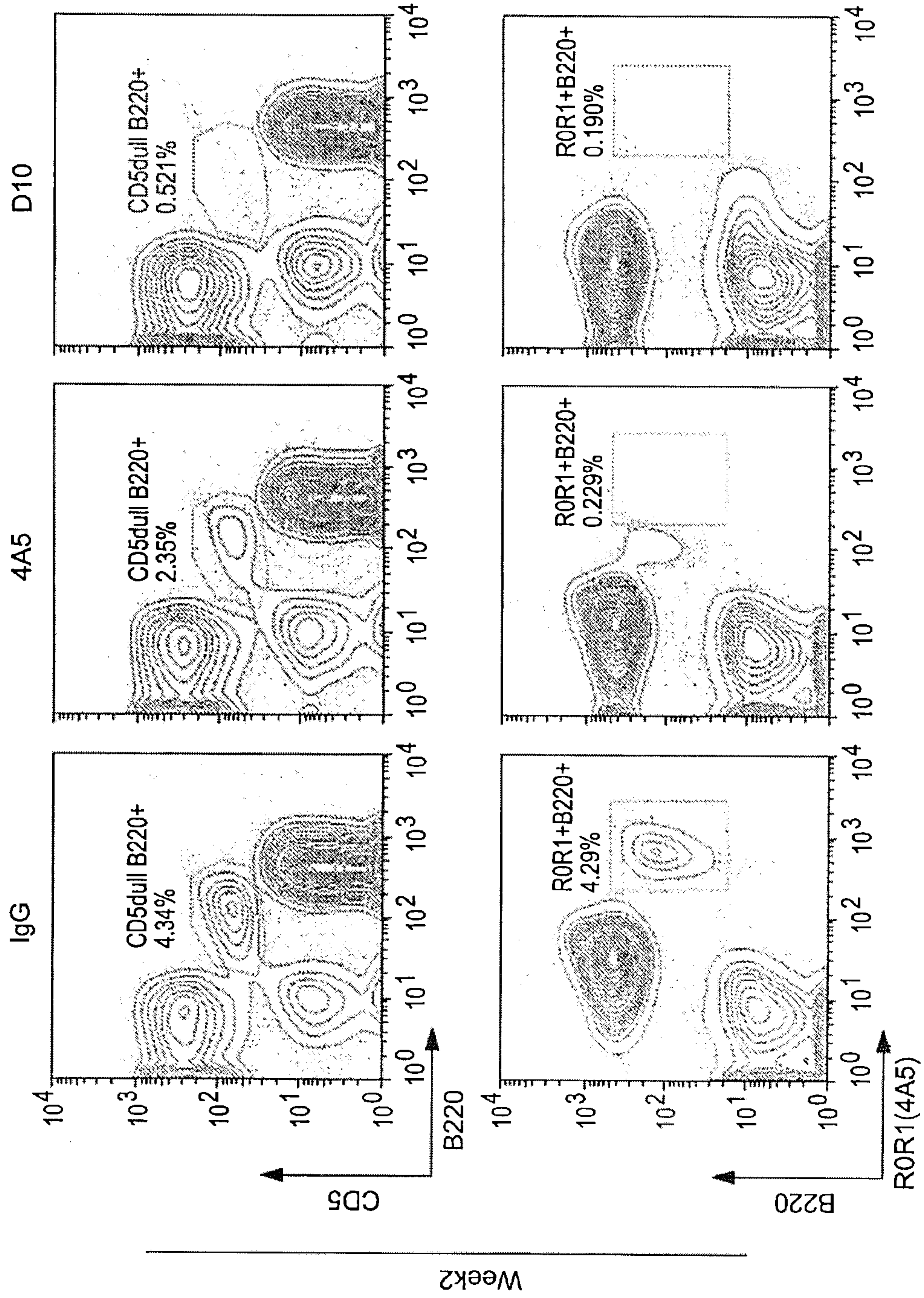


FIG. 3

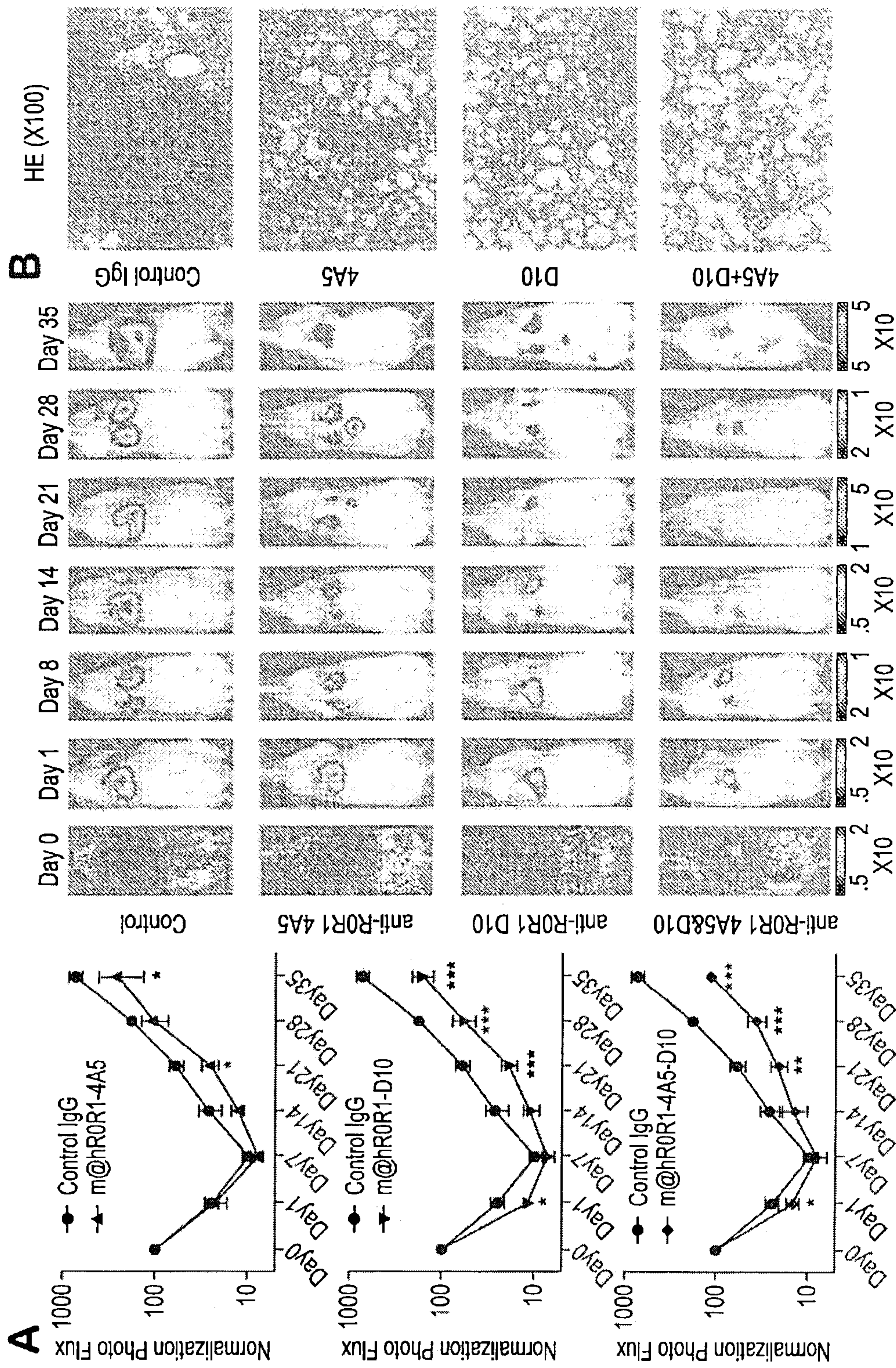


FIG. 4

Comparison of 4A5 Ig Heavy Chain to The Closest
 Germline Mouse and Human IGHV

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	FR1-IMGT (1-26)	CDR1-IMGT (27-38)	FR2-IMGT (39-55)
4A5_VH	1 10 20 30 40 50		
	EVKLVESGG.GLVKPGGSLKLSCAAS	GFTE...SSYA	MSWVRQIPEKRLWVAS
MuIGHV5-09*02A.....DT.....T
HuIGHV3-48*01	..Q.....Q.....R.....S	..N.....A.G.G.....SY

	CDR2-IMGT (56-65)	FR3-IMGT (66-104)
4A5_VH	60 70 80 90 100	
	ISRG...GTT	YYPDSVK.GRFTISRDNVRNIIYLQMSLRSED
MuIGHV5-09*02	..S.....SY.A.T.....L...A.
HuIGHV3-48*01	..SS..SS.I	..A.....AK.S.....N...A...V...A.

FIG. 5

Comparison of 2-G3 Ig Heavy Chain to The Closest
Germline Mouse and Human IGHV

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FR1-IMGT (1-26) CDR1-IMGT (27-38) FR2-IMGT (39-55)

	1	10	20	30	40	50
2-G3_VH	QVQLQQPGQ	.ELVKP	GTSVKLSCKAS	GYNF....	TNYW	INWVKLRPGQGLEWIGE
MuIGHV1-55*01	A...M.....	..T.....	S...T...	Q.....D
HuIGHV1-46*01V.S....	VK...A...V.....	..T.....	S.Y	MH..RQA.....	...M.I

CDR2-IMGT (56-65) FR3-IMGT (66-104)

	60	70	80	90	100
G3_VH	IYPG..	SGST	NYNEKFK.SKATLT	ADTSSSTAYMQLSS	LASEDSALYYCAR
MuIGHV1-55*01
HuIGHV1-46*01	.N.S..G...	S.AQ..Q.GRV.M.R...	T.V..E.....	R...T.V.....	

FIG. 7

Comparison of 3-H10 Ig Heavy Chain to The Closest
 Germline Mouse and Human IGHV

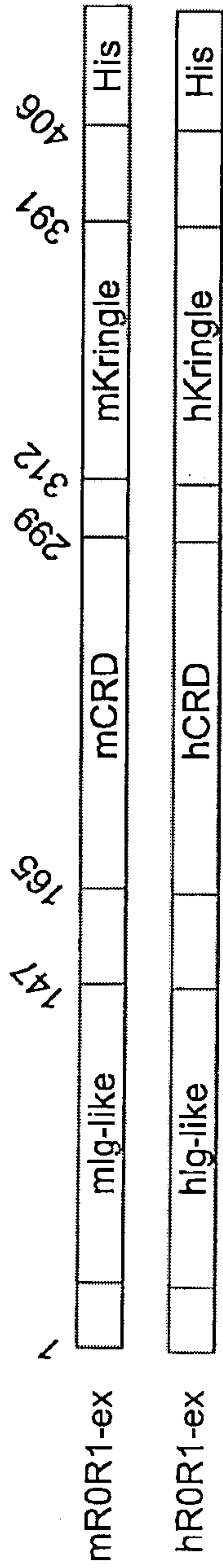
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	FR1-IMG (1-26)	CDR1-IMG (27-38)	FR2-IMG (39-55)
3-H10_VH	1 10 20 30 40 50		
	EVKLVESGG.GLVKPGGSLKLSCAAS	GFAF...TGYN	MNWVKQTNGKSLEWIGS
MuIGHV5-9*02A.....DT
HuIGHV3-23*04	..Q.....Q.....R.....A.G.G.....SA

	CDR2-IMG (56-65)	FR3-IMG (66-104)
H10_VH	60 70 80 90 100	
	ISTG...AST YFPDSVK.GRFTISRDNARNILYLQMSLRSED	TAMYCAR
MuIGHV5-9*02	..S...G... Y.....T.....L.....
HuIGHV3-23*04	..GS..GG.. YA.....	SK.T.....N...A.....V.....K

FIG. 8

Human and Murine ROR1 Proteins are Highly Conserved



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Region	Position	# of different aa
Ig-like	1---147aa	12
Linker between Ig-like and CRD	148---165aa	1
CRD	166---299aa	1
Linker between CRD and Kringle	300---312aa	0
Kringle	313---391aa	1
Linker between Kringle and TM	392---406aa	0

FIG. 10

Domain Structure and Sequence Homology
of Human and Murine ROR1 Extracellular Protein

hROR1	MHRPRRRGTRPPLLLAALLAARGAAQAQETELSVSAELVPTSSWNISSELNPKDSYLTLDPEPMNNITSLGQTAELHCK	80
mROR1P.....D.....T.....ID.G.....	
hROR1	<u>VSGNPPPTIRWFKNDAPVVQEPRLSFRSTIYGSRLRIRNLDTTDTGYFCVAATNGKEVSSIGVLFVKFGPPPTASPGY</u>	160
mROR1S.....I...A.N.....K...T.....	
	Ig Domain	
hROR1	<u>SDEYEEDGFCQPYRGIACARFIGNRTVYMESLHMQGEIENQITAAFTMIGTSSHLSDKCSQFAIPSLCHYAFPYCDETSS</u>	240
mROR1V.....	
hROR1	<u>VPKPRDLCRDECEILENVLCQTEYIFARSNPMLMRLKLPNCEDLPQPESPEAANCIRIGIPMADPINKNHKCYNSTGVD</u>	320
mROR1	
hROR1	<u>YRGTVSVTKSGRQCQPWNSQYPHTHTFTALRFPPELNGGHSYCRNPGNQKEAPWCFTLDENFKSDLCDIPACDSKDSKEKN</u>	400
mROR1S.....	
hROR1	KMEILY	
mROR1	

FIG. 11

Anti-ROR1 Mabs Generated Across Extracellular Domain

Binding sites of antibodies					
No.	5'-Ig-like	Middle of Ig-like	3'-Ig-like	CRD	Kringle
1-4A5		✓			
G11		✓			
H11		✓			
2G3		✓			
3-D10			✓		

FIG. 12

Anti-R0R1 Mab Binding Site Determination

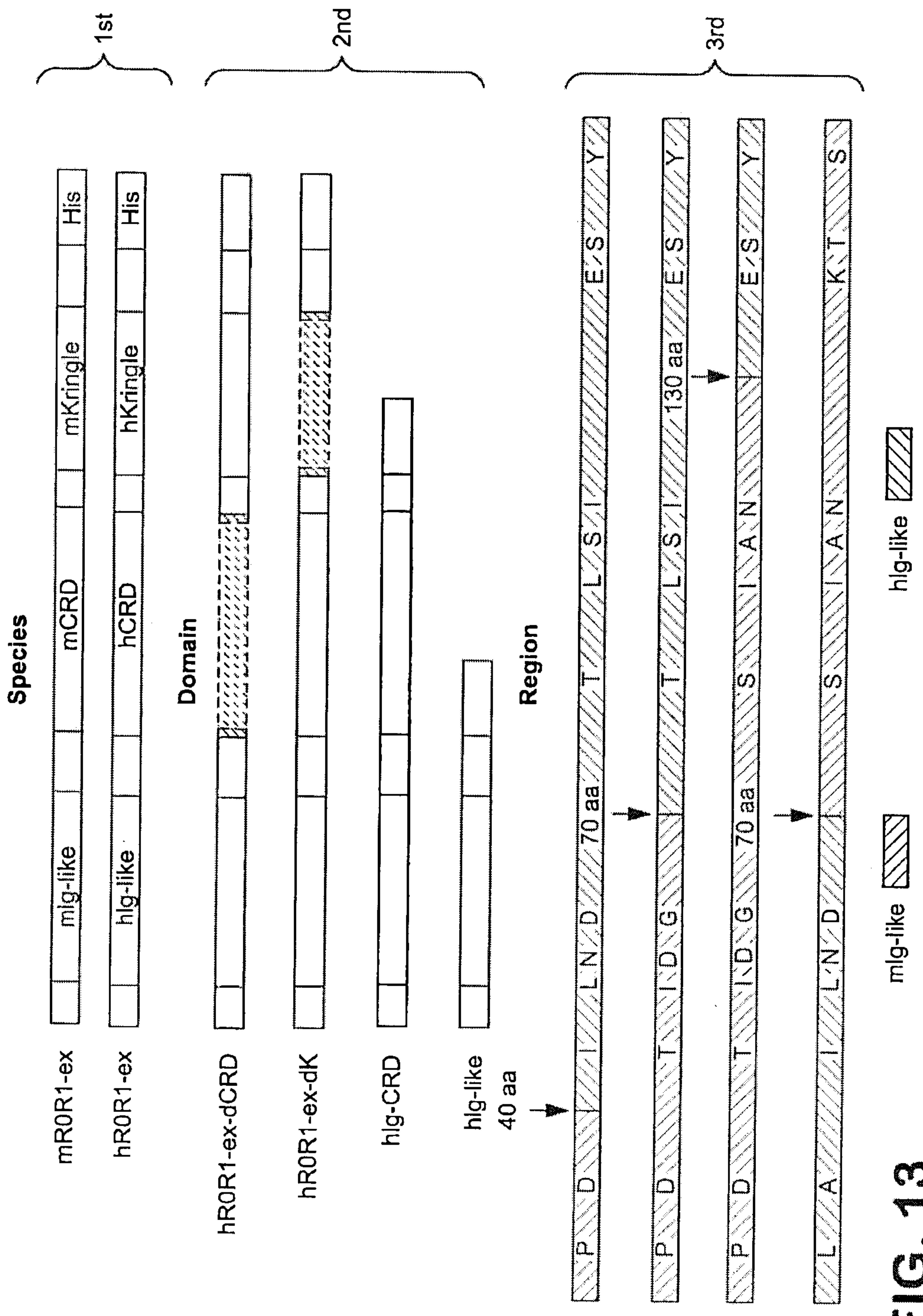


FIG. 13

Anti-ROR1 Mab Binding Region Determination

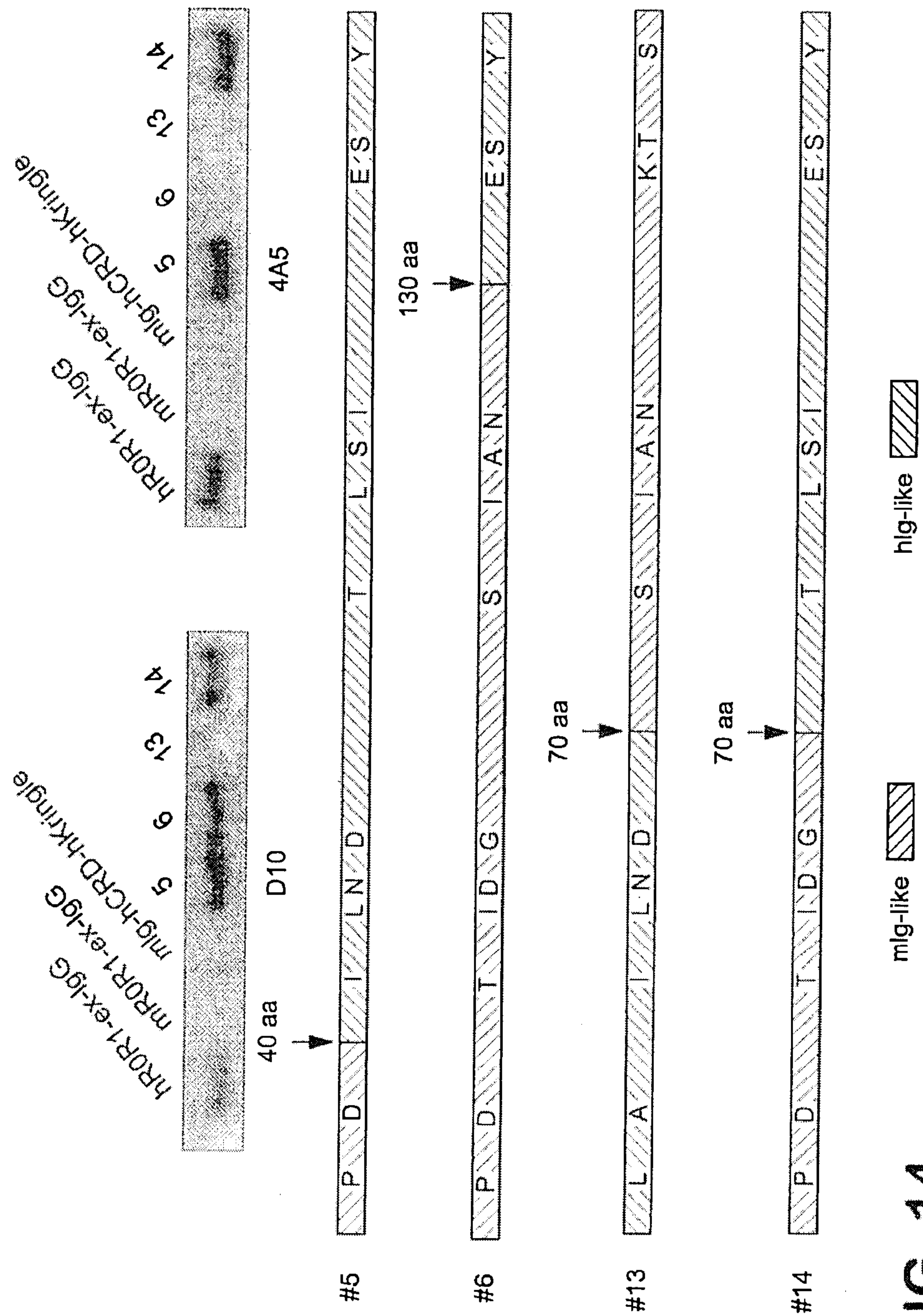
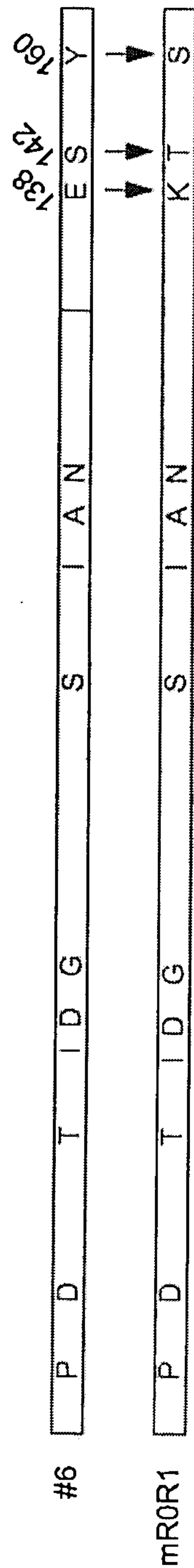


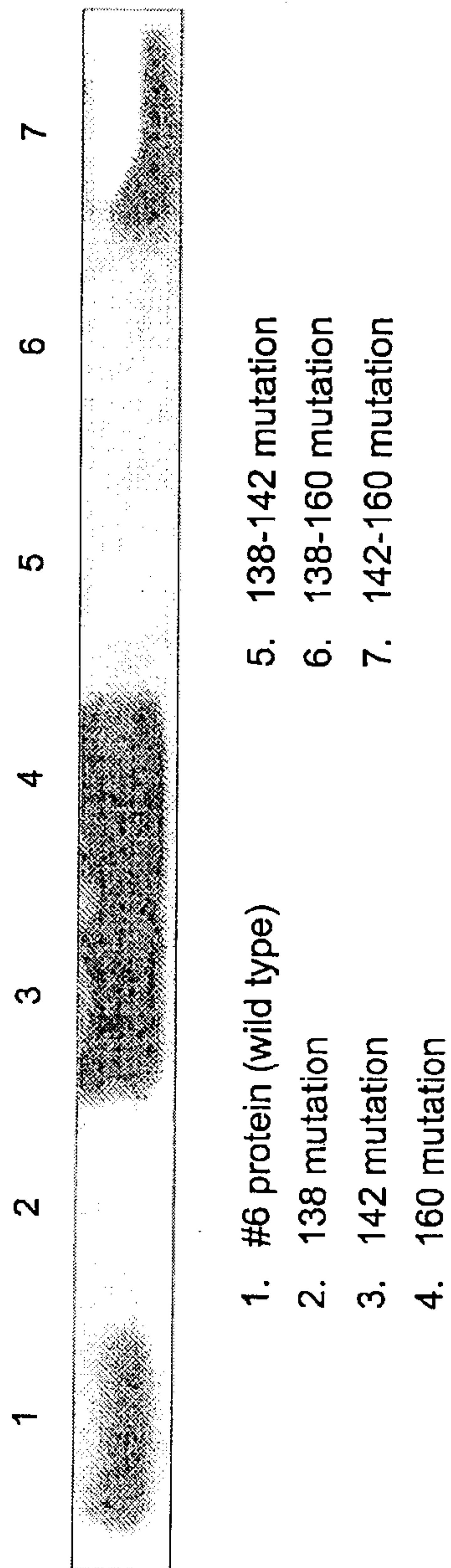
FIG. 14

3-D-10 Binds to the Human Glutamic Acid Residue



138, 142 and 160 amino acids were mutated individually and doubly.

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- 1. #6 protein (wild type)
- 2. 138 mutation
- 3. 142 mutation
- 4. 160 mutation
- 5. 138-142 mutation
- 6. 138-160 mutation
- 7. 142-160 mutation

FIG. 15

3-D10 Kd Determination

Analysis(x)

Baseline / Endpoints:
 5 to 10 [sec] from beginning
 10 to 5 [sec] from end

Ignore	Binding Signal (V)	Concentration
	0.0499	500nM
	0.0805	250nM
	0.1525	125nM
	0.2084	62.5nM
	0.2405	31.25nM
	0.4173	15.63nM
	0.4760	7.81nM
	0.5175	3.90nM
	0.5015	1.95nM
	0.5390	976.56pM
	0.5496	488.28pM
✓	0.5581	0
	0.0012	0
	0.0082	500nM
	0.0777	250nM
✓	0.1245	125nM
✓	0.1684	62.5nM
	0.2908	31.25nM
	0.3642	15.63nM
	0.4170	7.81nM
	0.4595	3.90nM
	0.4782	1.95nM
	0.4804	976.56pM
	0.4870	488.28pM
	0.4968	0
✓	0.0049	0

Kd = 40.47nM
 ABC = 40.47pM
 Ratio = 0.0010
 Sig 100% = 0.54
 Drift = 0.7514
 (%/run)
 NSB = 0.00
 Drift = -0.7086
 (mV/run)
 %Error = 0.92

Kd = 40.47nM
 95% confidence interval
 Kd High = 43.50nM
 Kd Low = 26.25nM

ABC = 40.47pM
 95% confidence interval
 ABC High = 0.70nM
 ABC Low = Less than 146.19fM

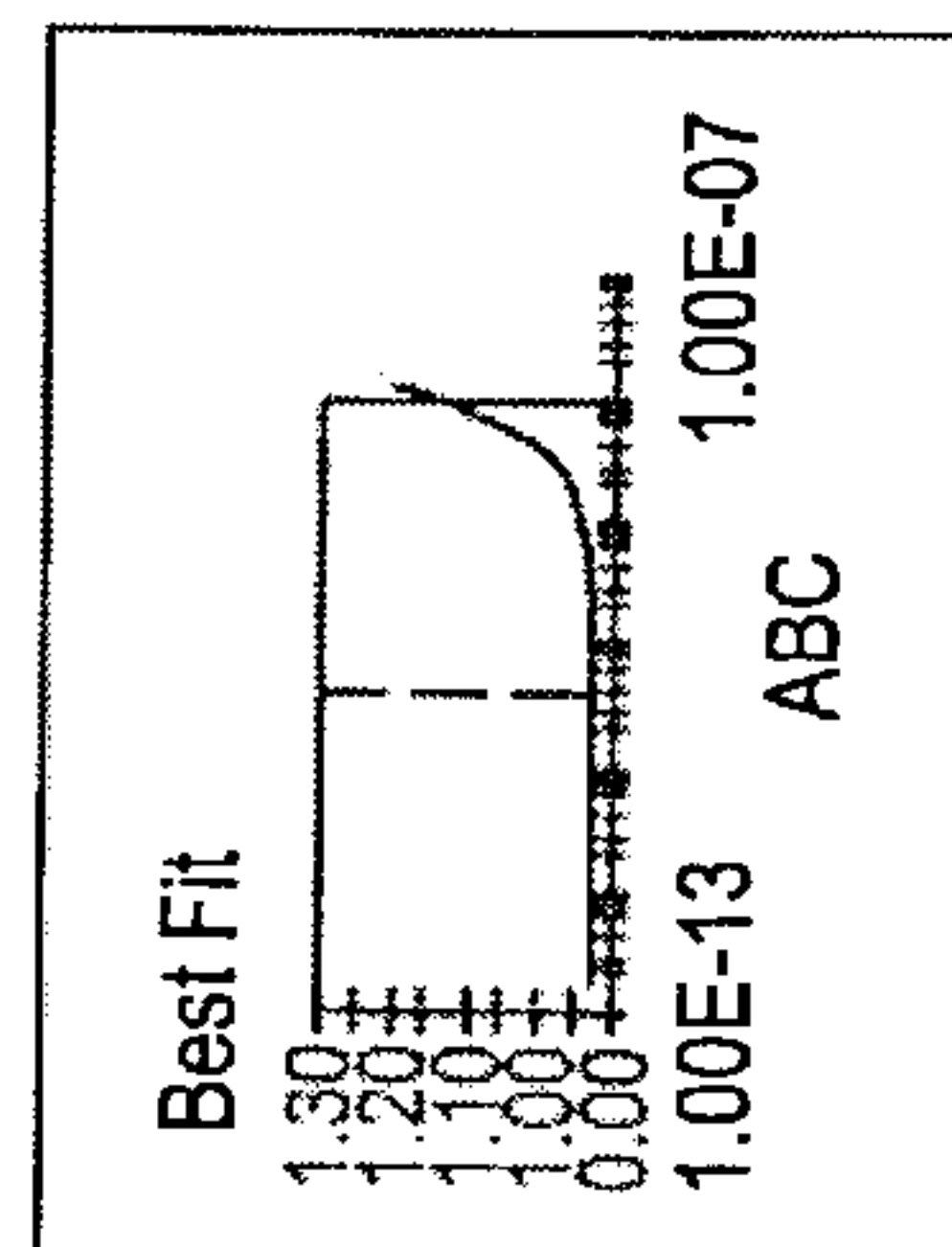
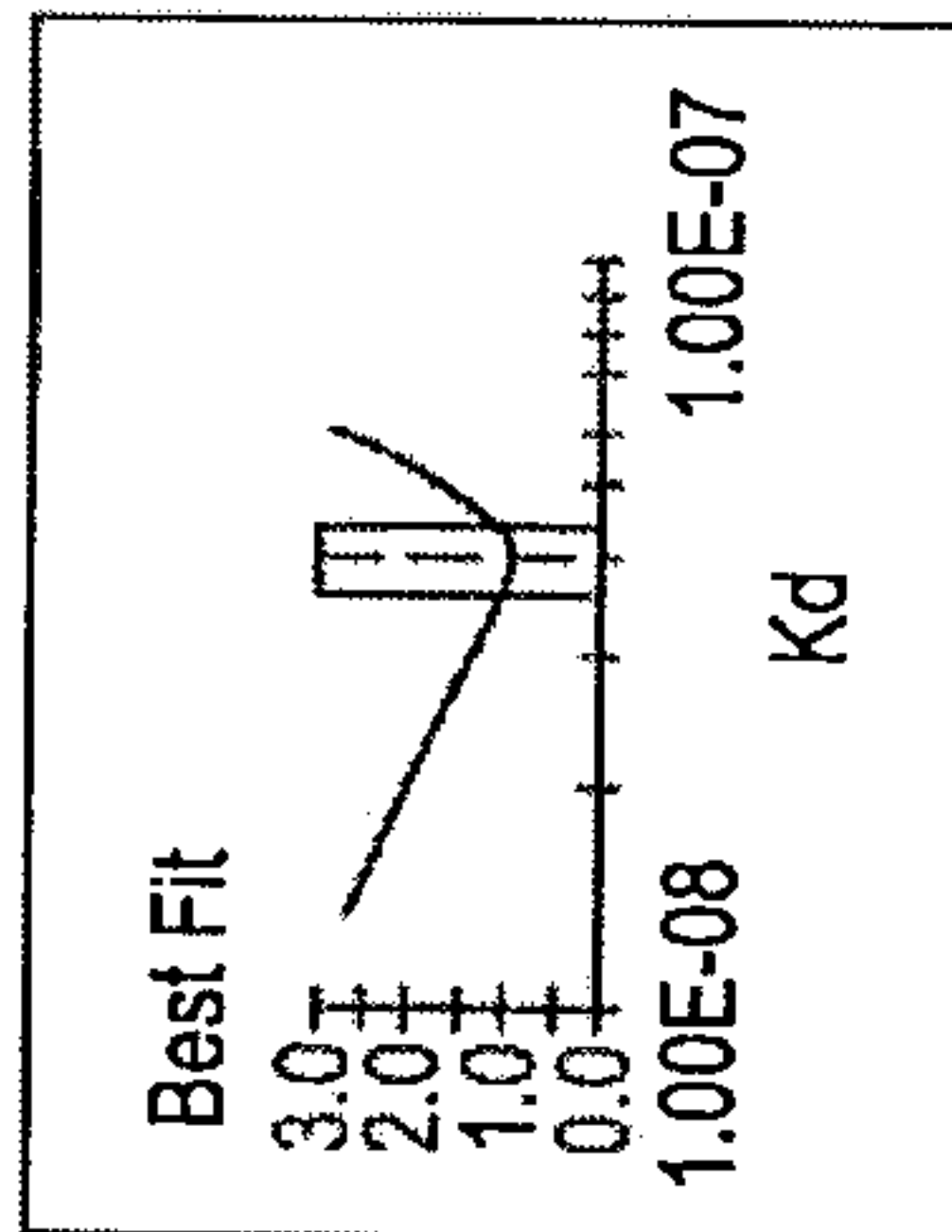
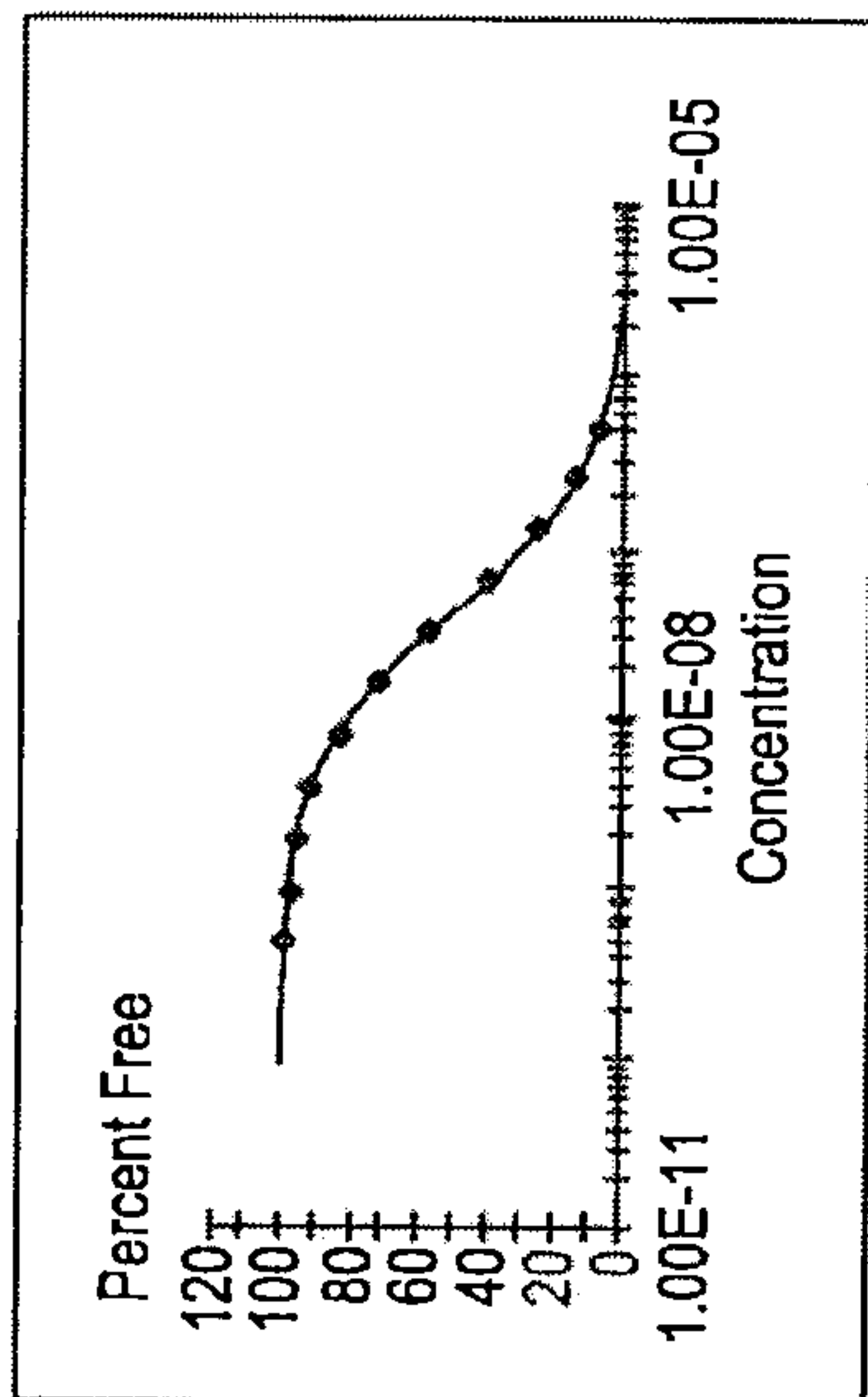


FIG. 16A

1-4A5 Kd Determination

Analysis(x)

Baseline / Endpoints:
 0 to 10 [sec] from beginning
 10 to 5 [sec] from end

Ignore	Binding Signal (V)	Concentration
	0.0781	100nM
	0.0990	50nM
	0.1213	25nM
	0.1554	12.5nM
	0.2060	6.25nM
	0.2768	3.13nM
	0.3167	1.56nM
	0.3613	781.25pM
	0.3702	390.63pM
	0.3790	195.31pM
	0.3895	97.66pM
	0.4102	48.63pM
	0.3928	0
	0.0616	100nM
	0.0646	50nM
	0.0995	25nM
	0.1257	12.5nM
	0.1807	6.25nM
	0.2415	3.13nM
	0.2903	1.56nM
	0.3300	781.25pM
	0.3533	390.63pM
	0.3650	195.31pM
	0.3636	97.66pM
	0.3682	48.63pM
	0.3758	0

Kd = 4.53nM
 ABC = 62.92pM
 Ratio = 0.0139
 Sig 100% = 0.40
 Drift = -0.0110
 (%/run) = 0.04
 NSB Drift = -2.0021
 (mV/run)
 %Error = 1.53

Kd = 4.53nM
 95% confidence interval
 Kd High = 5.09nM
 Kd Low = 3.33nM

ABC = 62.92pM
 95% confidence interval
 ABC High = 2.59nM
 ABC Low = Less than 227.30fM

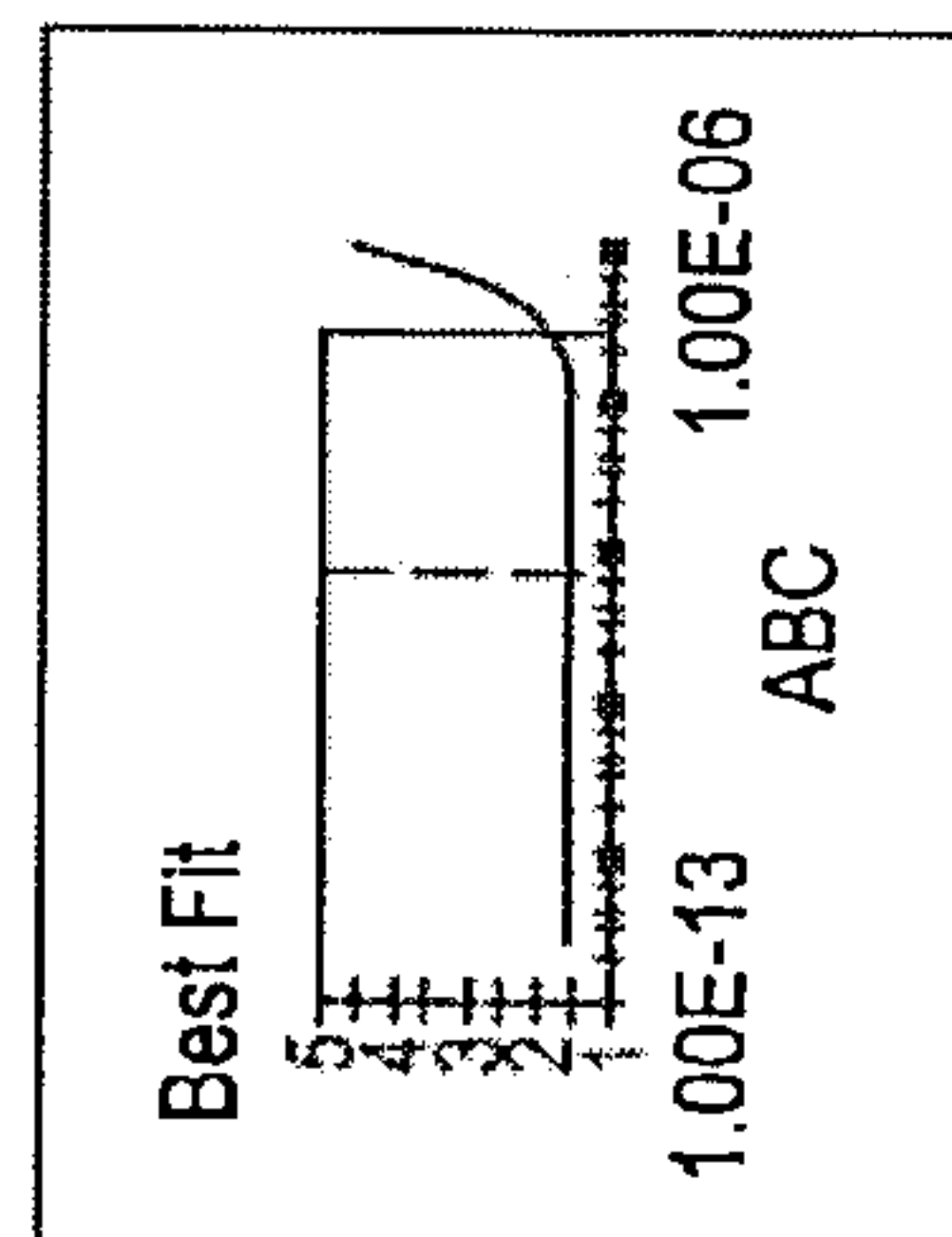
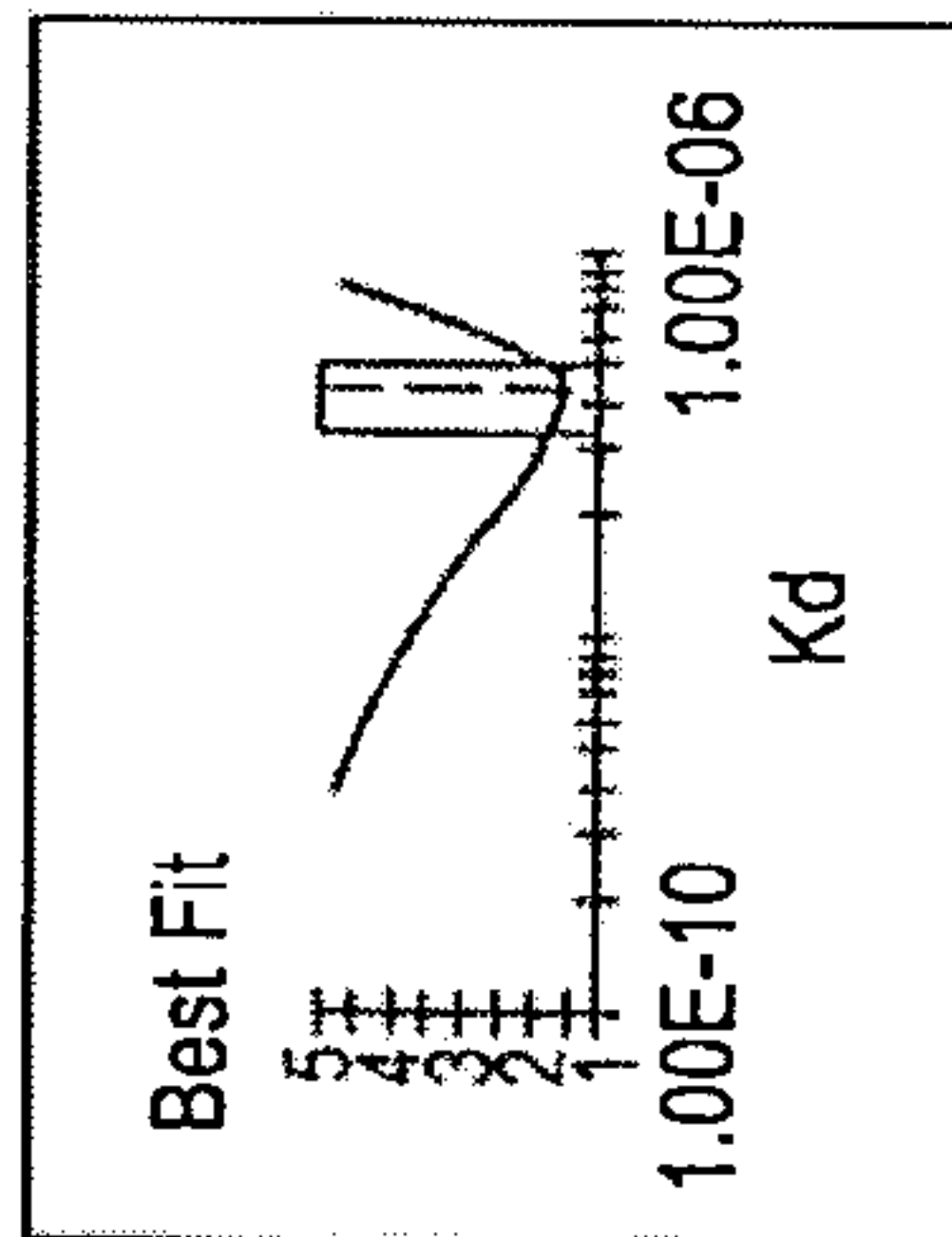
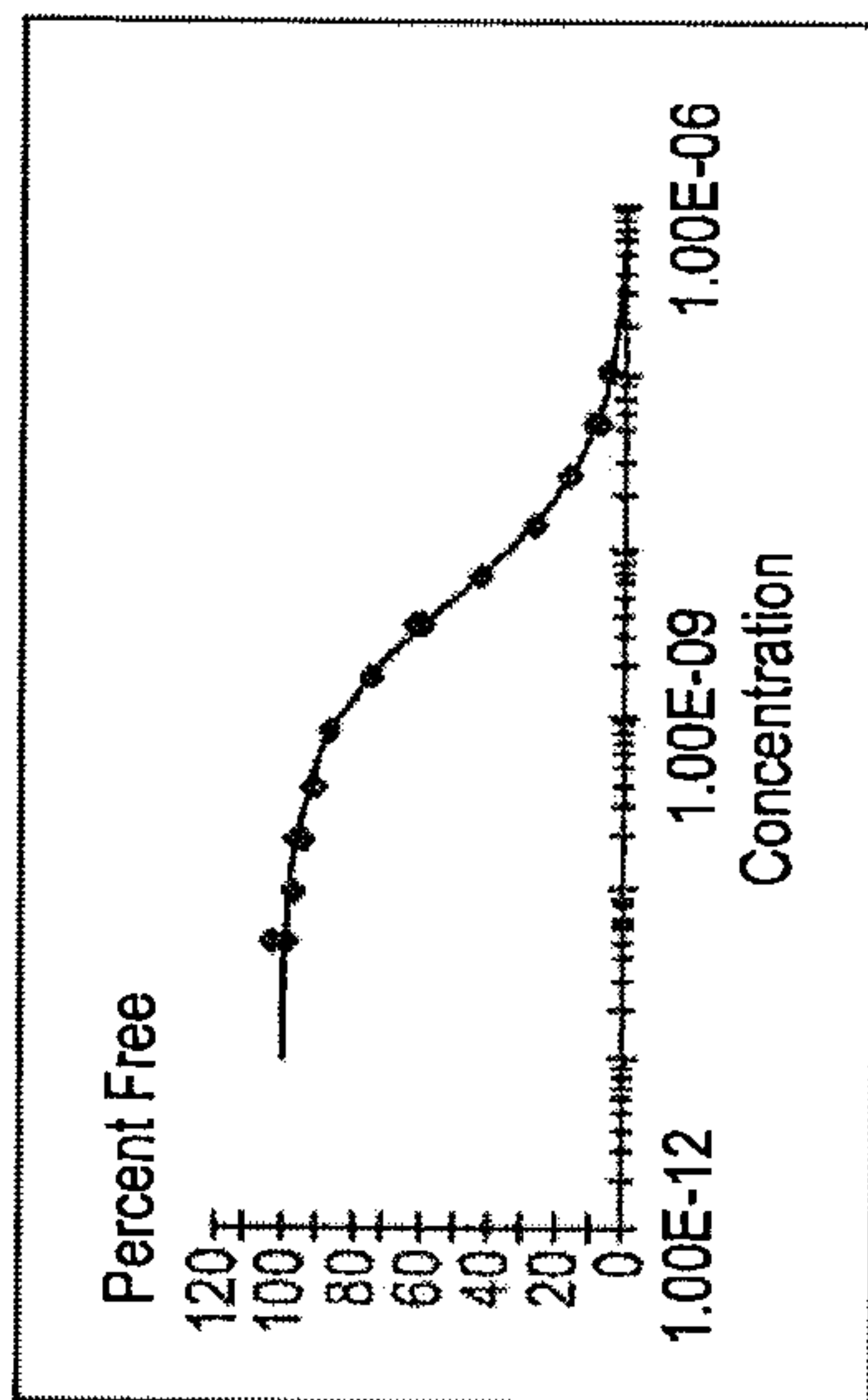
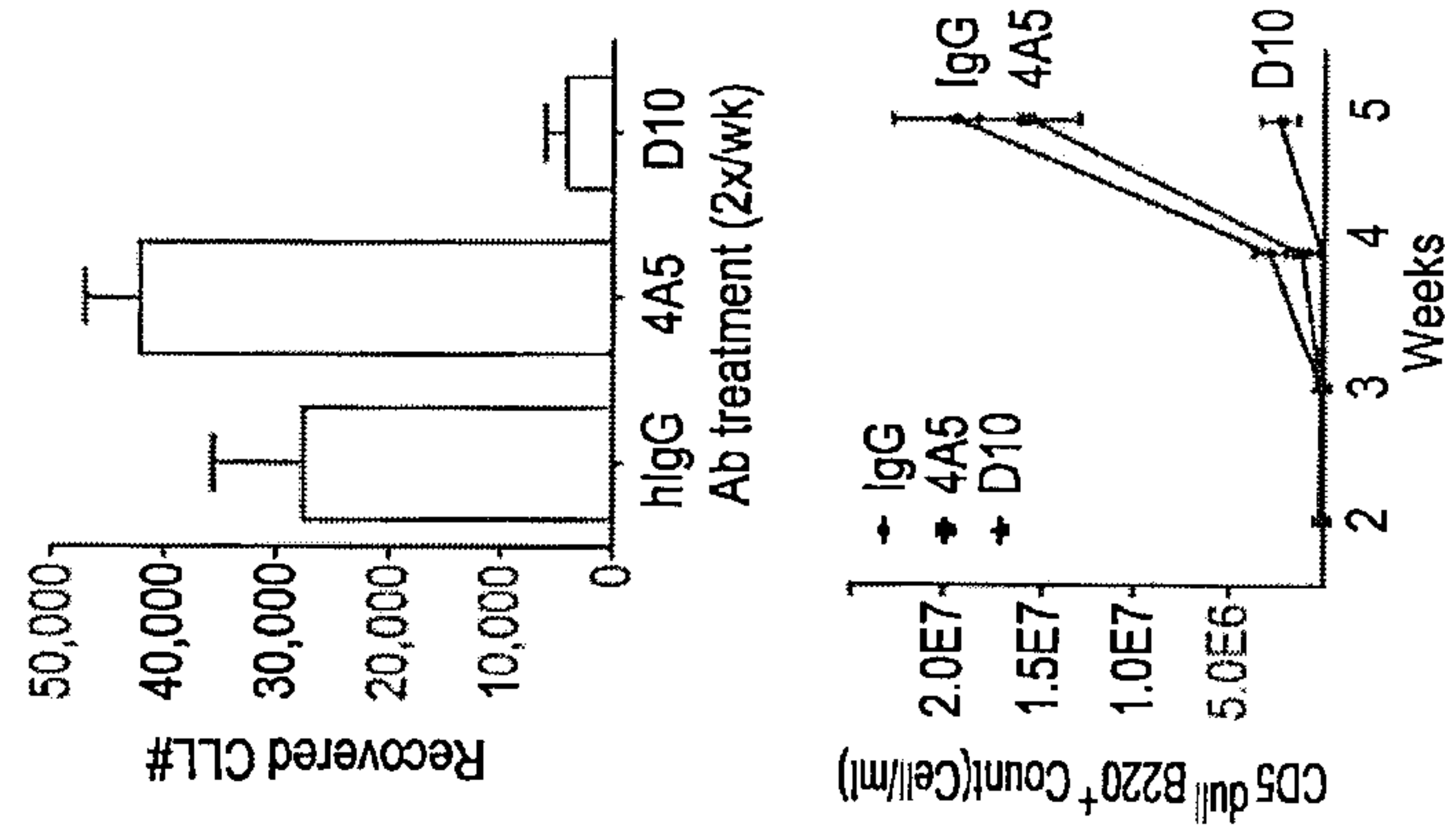


FIG. 16B

3-D10 Anti-ROR1 mAb is Highly Active in *in vivo* Assays

- 3-D10 Mab highly active in *in vivo* niche dependent activity model
 - Substantial reduction in leukemic burden using 4 primary CLL patient products tested in 76 mice
 - Activity much greater than other anti-ROR1 Mabs (4A5)
- 3-D10 Mab active in *in vivo* immune competent mouse model
 - Substantial reduction in spontaneous human ROR1 expressing leukemia model
 - Activity much greater than other anti-ROR1 Mabs (4A5)



3-D10 Mab has greatest anti-ROR1 activity in *in vivo* assay systems

FIG. 17

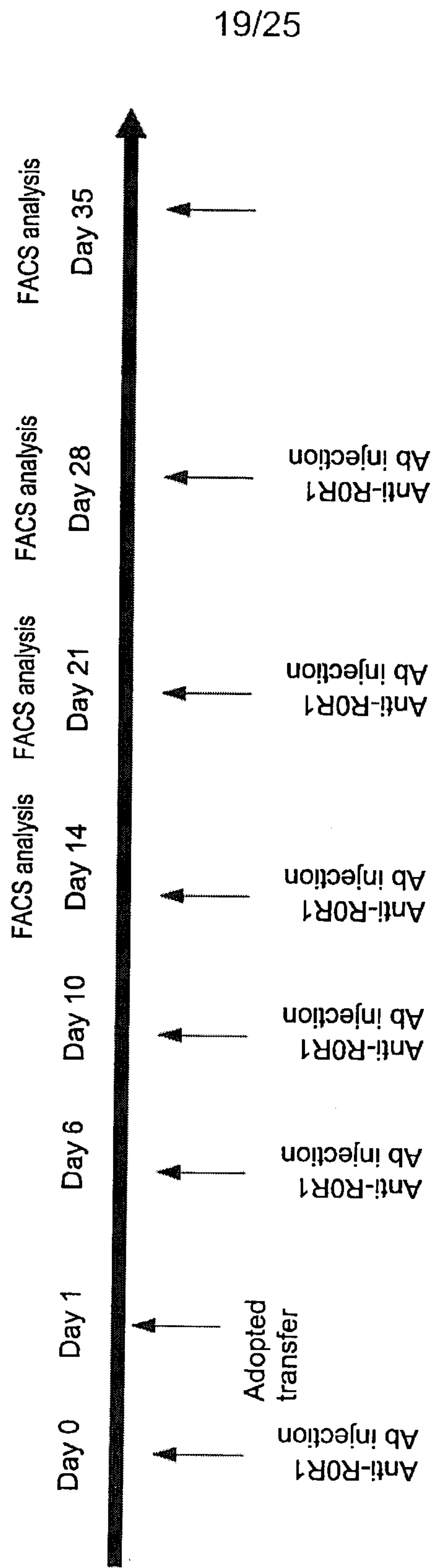


FIG. 18

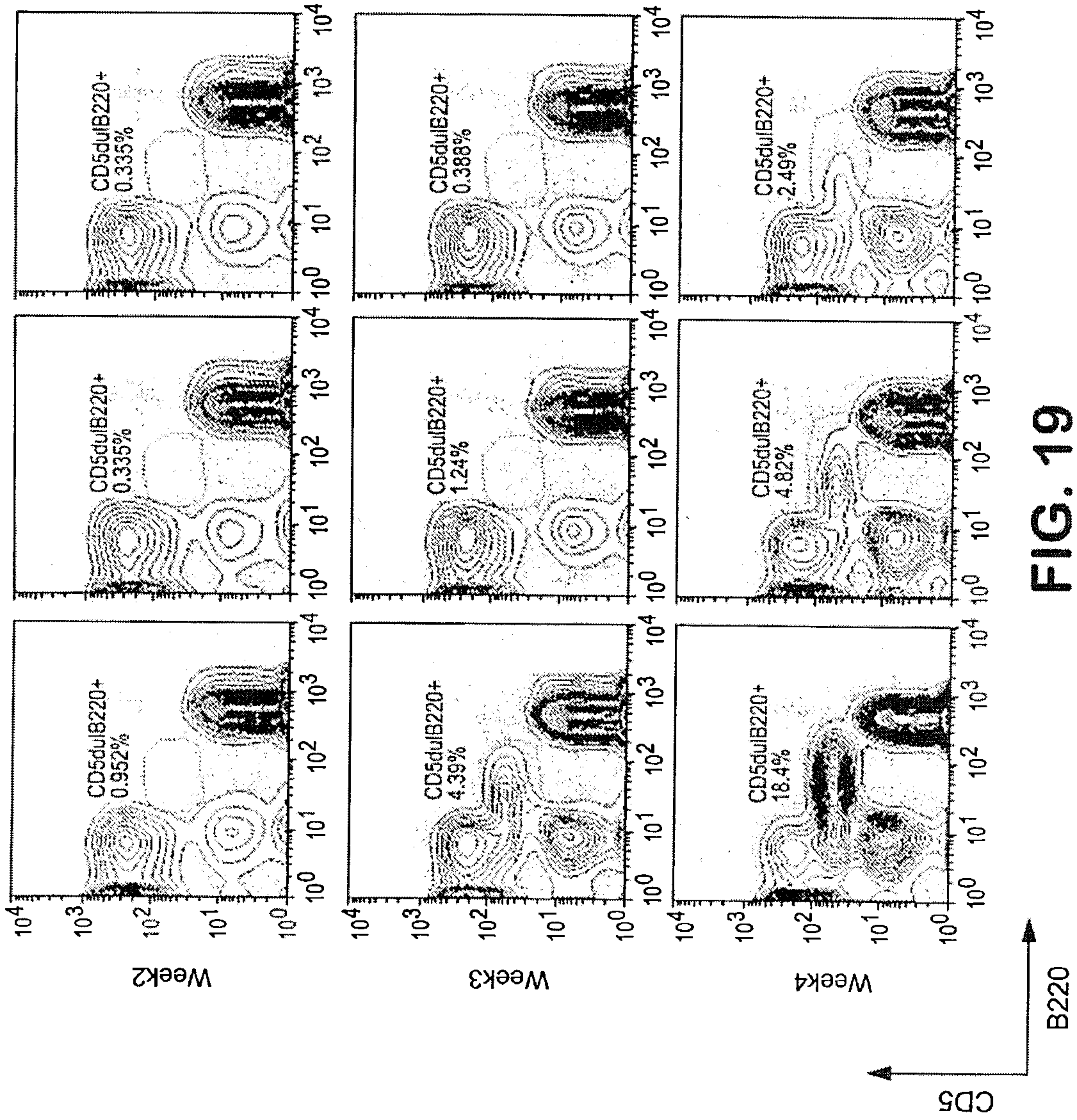


FIG. 19

CD5^{dim} B220⁺ Leukemia development
(absolute leukemia cell count)

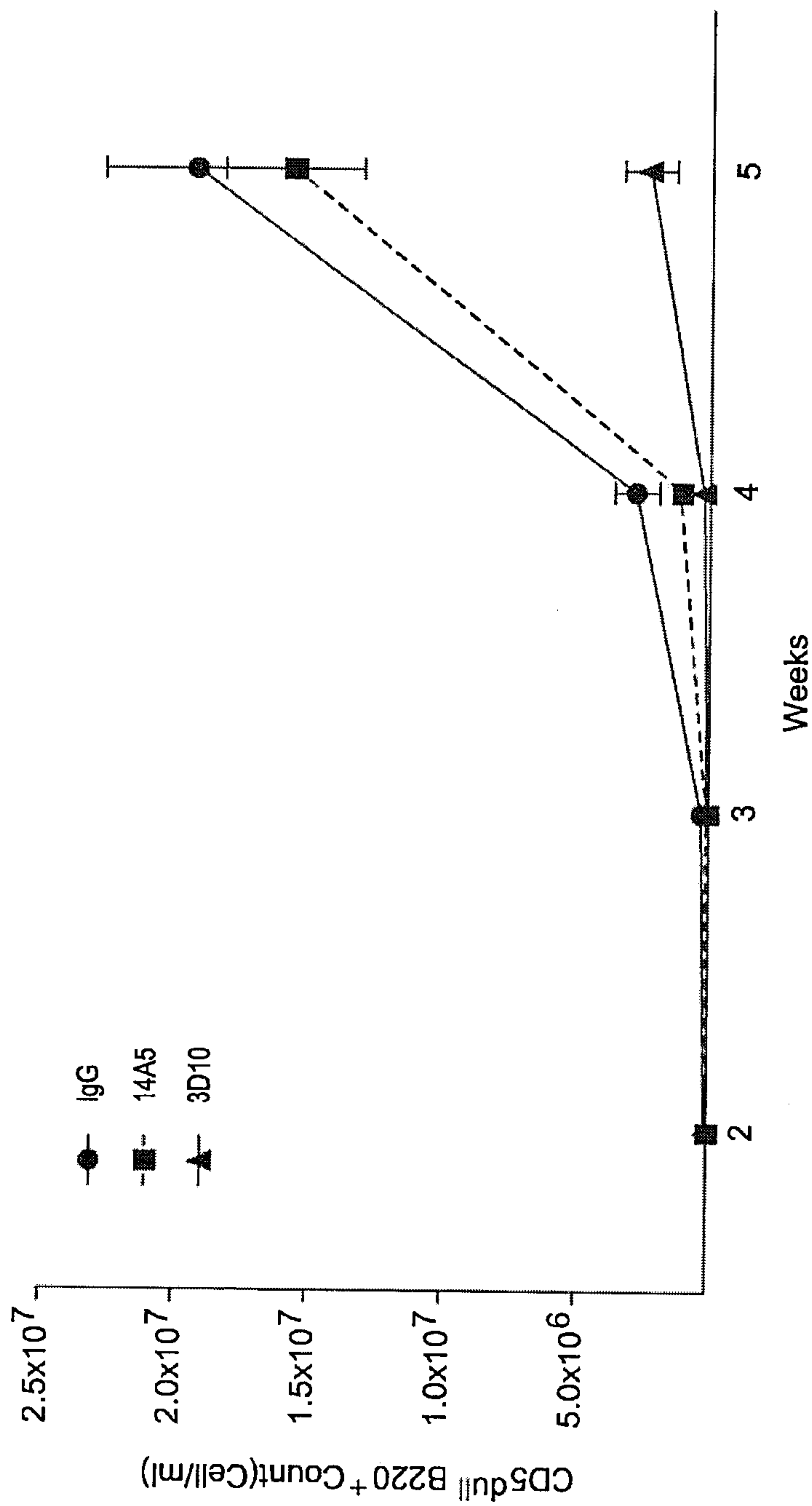


FIG. 20

Rapid Anti-ROR1 3-D10 Ab Internalization into CLL Cells

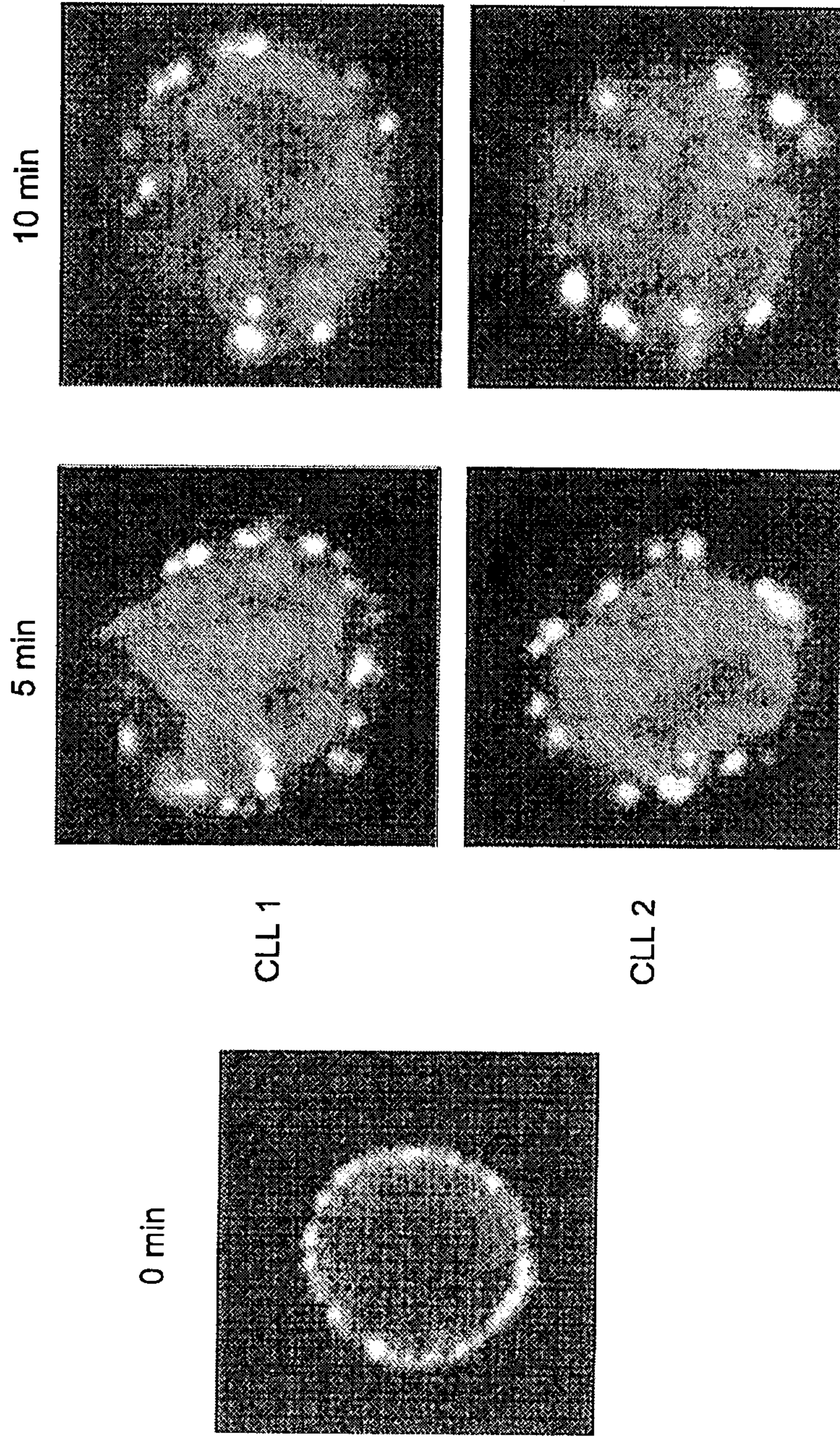


FIG. 21

Anti-hROR1 antibody internalization study

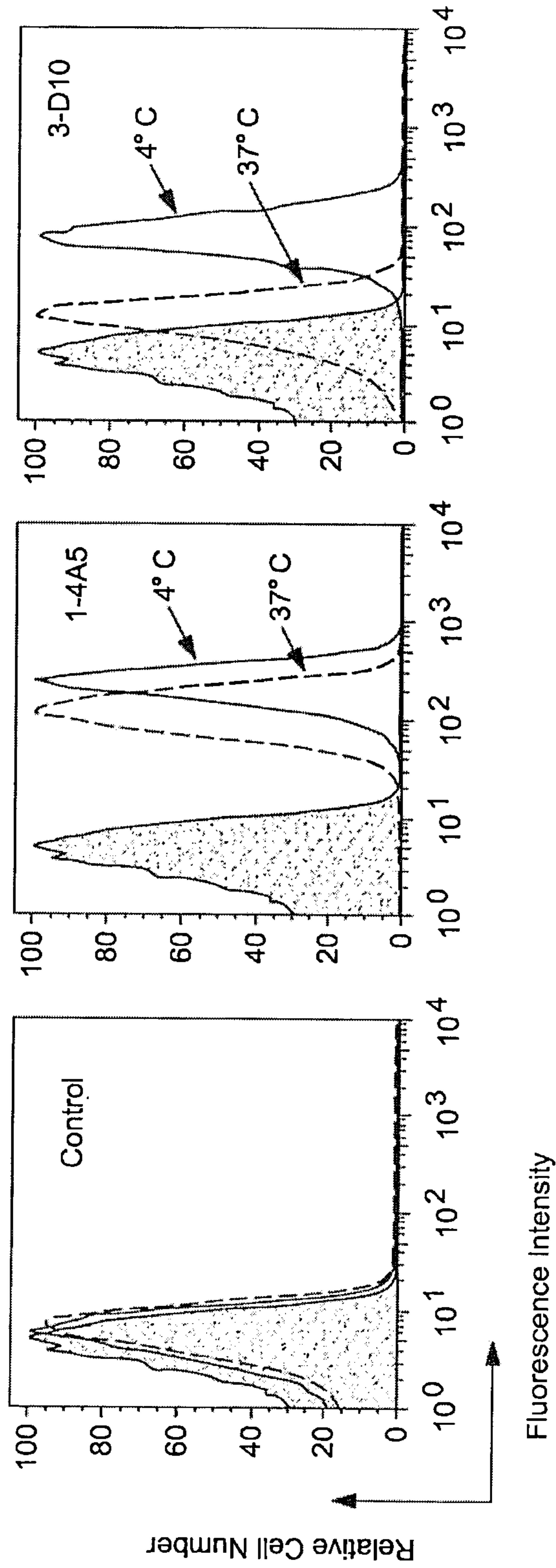


FIG. 22

1-4A5 and 3-D10 internalization studies and kinetics of ROR1 antibody internalization.

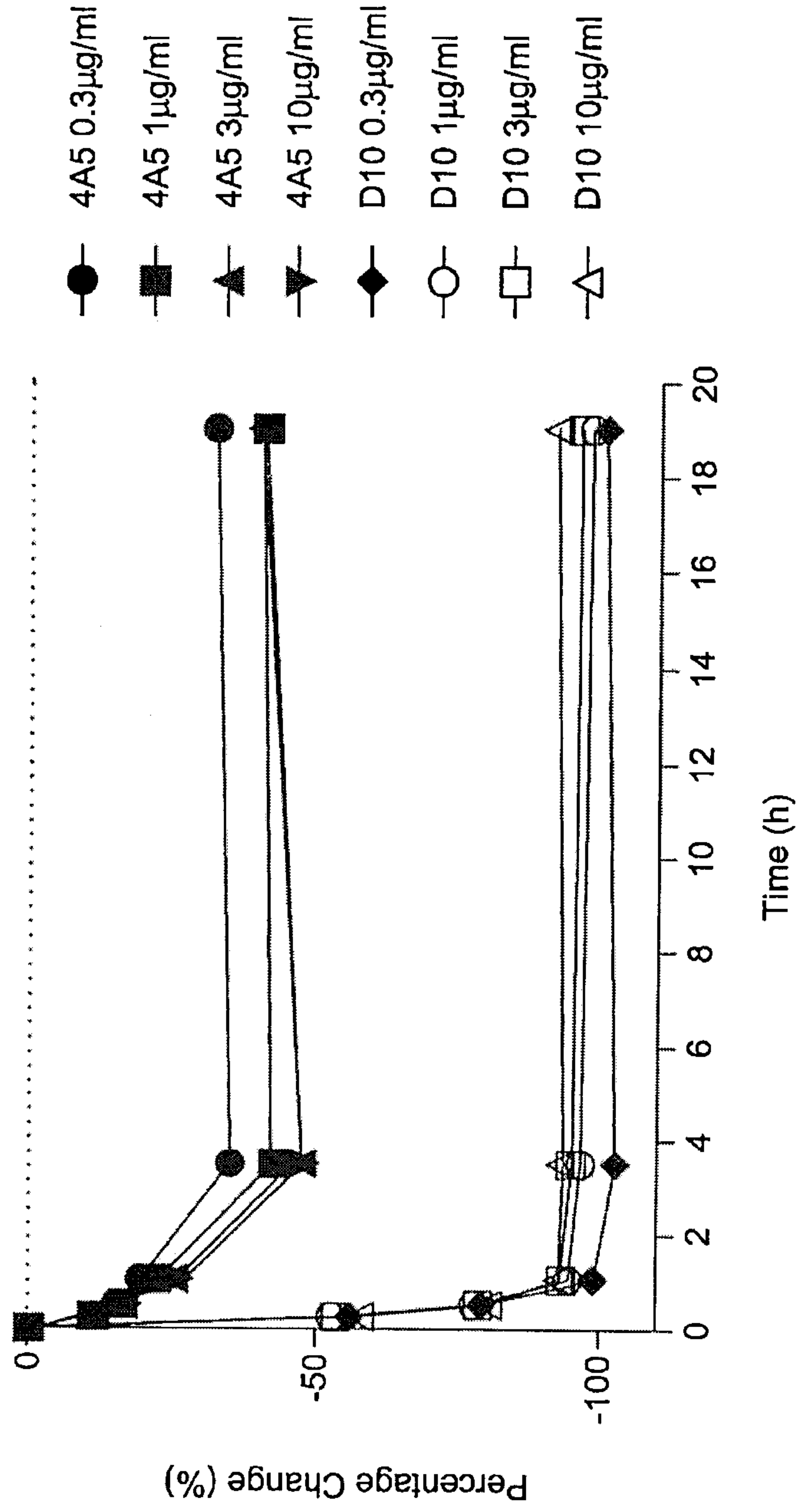
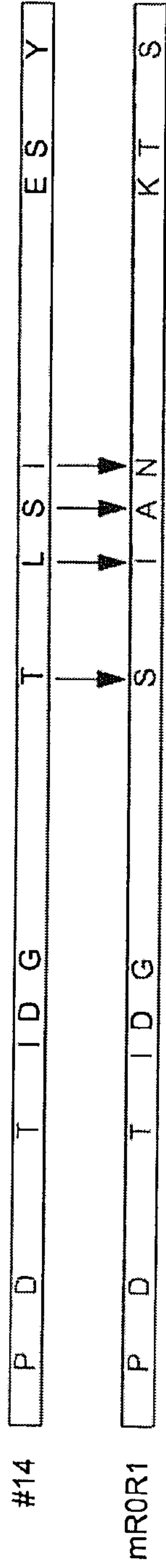
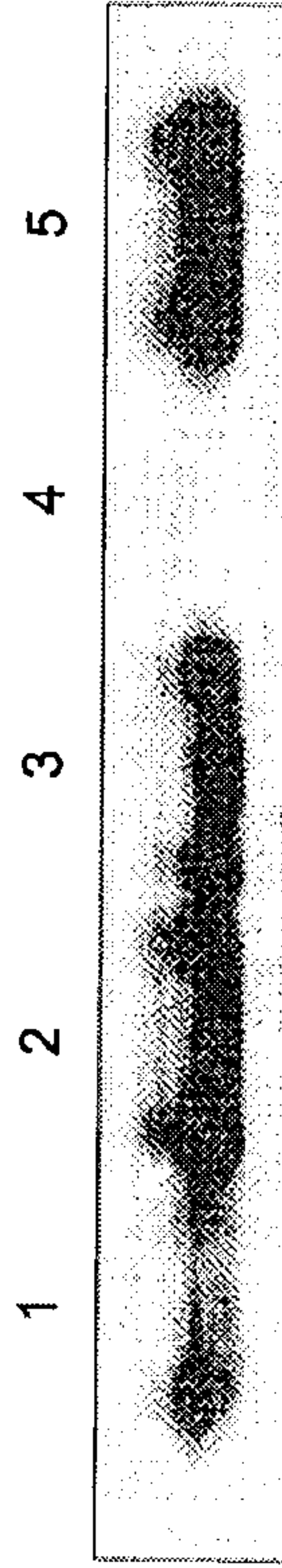


FIG. 23

4A5 binding site



88, 105, 109 and 111 amino acids were mutated individually.



- 1. 88aa mutation
- 2. 105aa mutation
- 3. 109aa mutation
- 4. 111aa mutation
- 5. #14 protein (wild type)

4A5 binds to the 111 amino acid of human R0R1

FIG. 24