

US 20190124896A1

(19) United States (12) Patent Application Publication (10) Pub. No.: US 2019/0124896 A1

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(54) HUMANIZED DIPEPTIDYL-PEPTIDASE IV (DPP4) ANIMALS

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- (21) Appl. No.: 16/239,028
- (22) Filed: Jan. 3, 2019

Related U.S. Application Data

- (63) Continuation of application No. 15/295,955, filed on Oct. 17, 2016, now Pat. No. 10,212,923, which is a continuation of application No. 14/723,855, filed on May 28, 2015, now Pat. No. 9,497,945.
- (60) Provisional application No. 62/072,692, filed on Oct. 30, 2014, provisional application No. 62/051,626, filed on Sep. 17, 2014, provisional application No. 62/005,476, filed on May 30, 2014.

Publication Classification

(51) Int. Cl.

A01K 67/027	(2006.01)
C12Q 1/6883	(2006.01)

May 2, 2019 (43) **Pub. Date:**

C12Q 1/68	(2006.01)
C12Q 1/70	(2006.01)
G01N 33/569	(2006.01)
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(52) U.S. Cl. CPC A01K 67/0278 (2013.01); A01K 67/0276 (2013.01); C12Q 1/6883 (2013.01); C12Q 1/68 (2013.01); C12Q 1/70 (2013.01); G01N 33/56983 (2013.01); A01K 2267/0337 (2013.01); A01K 2267/01 (2013.01); A01K 2207/15 (2013.01); A01K 2267/03 (2013.01); A01K 2217/072 (2013.01); A01K 2227/105 (2013.01); C12Y 304/14005 (2013.01)

(57)ABSTRACT

Non-human animals comprising a human or humanized DPP4 nucleic acid sequence are provided. Non-human animals that comprise a replacement of the endogenous Dpp4 gene with a human or humanized DPP4 gene, or non-human animals comprising a human or humanized DPP4 gene in addition to the endogenous Dpp4 gene are described. Nonhuman animals comprising a human or humanized DPP4 gene under control of human or non-human DPP4 regulatory elements is also provided, including non-human animals that have a replacement of non-human Dpp4-encoding sequence with human DPP4-encoding sequence at an endogenous non-human Dpp4 locus. Non-human animals comprising human or humanized DPP4 gene sequences, wherein the non-human animals are rodents, e.g., mice or rats, are provided. Methods for making and using the non-human animals are described.

Specification includes a Sequence Listing.

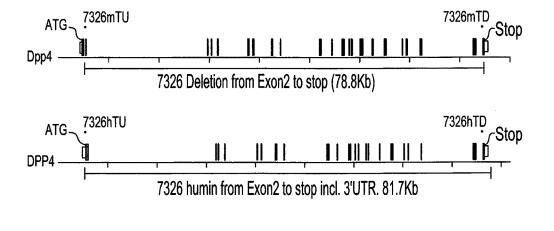


FIG. 1A

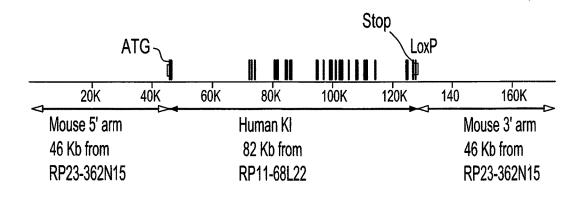


FIG. 1B

** MK <u>TPWKVLLG</u>	LLGAAALVTI	ITVPVVLLNK	GTDDATADSR	KTYTLTDYLK	NTYRLKLYSL
RWISDHEYLY	KQENNILVFN	AEYGNSSVFL	ENSTFDEFGH	SINDYSISPD	GQFILLEYNY
VKQWRHSYTA	SYDIYDLNKR	QLITEERIPN	NTQWVTWSPV	GHKLAYVWNN	DIYVKIEPNL
PSYRITWTGK	EDIIYNGITD	WVYEEEVFSA	YSALWWSPNG	TFLAYAQFND	TEVPLIEYSF
YSDESLQYPK	TVRVPYPKAG	AVNPTVKFFV	VNTDSLSSVT	NATSIQITAP	ASMLIGDHYL
CDVTWATQER	ISLQWLRRIQ	NYSVMDICDY	DESSGRWNCL	VARQHIEMST	TGWVGRFRPS
EPHFTLDGNS	FYKIISNEEG	YRHICYFQID	KKDCTFITKG	TWEVIGIEAL	TSDYLYYISN
EYKGMPGGRN	LYKIQLSDYT	KVTCLSCELN	PERCQYYSVS	FSKEAKYYQL	RCSGPGLPLY
TLHSSVNDKG	LRVLEONSAL	DKMLQNVQMP	SKKLDFIILN	ETKFWYQMIL	PPHFDKSKKY
PLLLDVYAGP	CSQKADTVFR	INWATYLAST	ENIIVASFDG	RGSGYQGDKI	MHAINRRLGT
FEVEDQIEAA	RQFSKMGFVD	NKRIAIWGWS	YGGYVTSMVL	GSGSGVFKCG	IAVAPVSRWE
YYDSVYTERY	MGLPTOEDNL	DHYRNSTVMS	RAENFKQVEY	LLIHGTADDN	VHFQQSAQIS
KALVDVGVDF	QAMWYTDEDH	GIASSTAHQH	IYTHMSHFIK	QCFSLP	

* RESIDUES CODED BY MOUSE EXON 1, ARE THE SAME IN HUMAN. UNDERSCORED RESIDUES CODED BY INTRODUCED HUMAN EXONS. HUMANIZED PROTEIN SEQUENCE IS 100% HUMAN.

FIG. 2

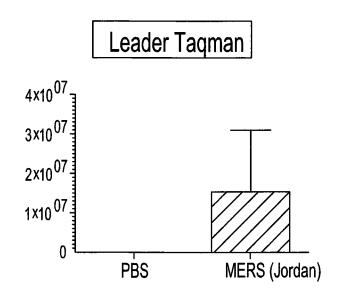
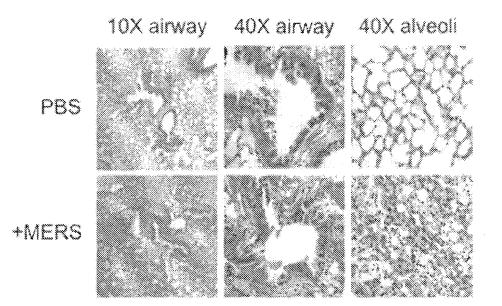


FIG. 3

FIG. 4



Day 4

mDpp4 LLIHGTADDNVHFQQSAQISKALVDAGVDFQAMWYTDEDHGIASSTAHQHIY**S**HMSHFLQQCFSLH hDPP4 LLIHGTADDNVHFQQSAQISKALVDVGVDFQAMWYTDEDHGIASSTAHQHIY**T**HMSHFLQQCFSLP

mDpp4 levedqieaarqfvkmgfvdskrvaiwgwsyggyvtsmvlgsgsgvfxcgiavapvsrweyydsvyterymglp1pednldhyrnstvmsRae**H**fkqvey hdpp4 fevedqieaarqfskmgfvdnkriaiwgwsyggyvtsmvlgsgsgvfkcgiavapvsrweyydsvyterymglp1pednldhyrnstvmsRae**n**fkqvey

mDpp4 DRMLQDVQMPSKKLDFIVLNETRFWYQMILPPHFDKSKKYPLLLDVYAGPCSQKADASFRLNWATYLASTENIIVASFDGRGSGYQGDKIMHAINRRLGT hDpP4 DRMLQNVQMPSKKLDFIILNETRFWYQMILPPHFDKSKKYPLLLDVYAGPCSQKADTVFRLNWATYLASTENIIVASFDGRGSGYQGDKIMHAINRRLGT

mDpp4 AWEVISIEALTSDYLYYISNQYKEMPGGRNLYKIQLTDHTNYKCLSCDLNPERCQYYAVSFSKEAKYYQLGCWGPGLPLYTLHRSTDHKELRVLEDNSAL hDPP4 TWEVIGIEALTSDYLYYISNBYKEMPGGRNLYKIQLSDYTKYTCLSCELNPERCQYYSVSFSKEAKYYQLRCSGPGLPLYTLHSSYNDKGLPVLEDNSAL

mDpp4 CDVVWATEERISLQWLRRIQNYSVMAICDYDKINLTWNCPSEQQHVEMSTTGWVGRFR?AEPHFTSDGSSFYKIISDKDGYKHICHFPKDKKDCTFITKG hDpp4 CDVTWATQERISLQWLRRIQNYSVMDICDYDESSGRWNCLVARQHIEMSTTGWVGRFRPSEPHFTLDGNSFYKIISNEEGYRHICYFQIDKKDCTFITKG

UDEL4 MAIRFEALZUIZHMAEWGUUTZAAFUALZAAFUEL2:J2DF2PÖLKKIAKAAAALAKEAAMID2P22AIUU121ÖLUH2UTGPUT

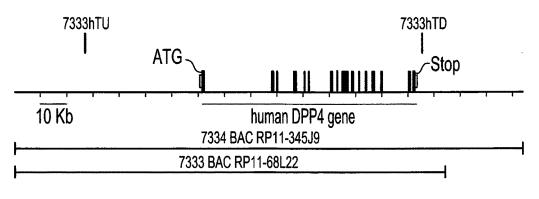
mDpp4 WVYEEEVFGAYSALWWSPN**N**TFLAYAQFNDTGVPJIEYSFYSDESLQYPKTVWIPYPKAGAVNPTVKFFIVNIDSLSSS**S**SA**A**PIQIPAPASVARGDHYL hDpp4 wvyEeEvFSAYSALWWSPNGTFLAYAQFNDTEVPIIEYSFYSDESLQYPKTVRVPYPKAGAVNPTVKFFVVNTDSLSSVTNATSIQITAPASMLIGDHYL

mDpp4 ----HSVSPDRLFVLLEYNYVKQWRHSYTASYLIYDVNKRQLITEEKIPNNTQWITWSPEGHKLAYVWKNDIYVKVEPHLPSHRITSTGEENVIYNGITD hDpp4 sindysispdgofilleynyvkqwrhsytasydiydinkrqliteeripnntqwvtwspyghklayvwnndiyvkiepnlpsyritwtgkediiyngitd

mDpp4 MKTPWKVLLGLLGVAALVTIITVPIVLLSK--DEAAADSRRTYSLADYLKSTFRVKSYSLWWVSDFEYLYKQENNILLINAEHGNSSIFLENSTFESFGY hDPP4 MKTPWKVLLGLLGAAALVTIITVPVVLLNKGTDDATADSRRTYTLTDYLKNTYRIKLYSLRWISDHEYLYKQENNILVENAEYGNSSVFLENSTFDEFGH

	Official Symbol	NCBI GenelD	Primary source	RefSeq mRNA ID		Genomic Assembly	
Human	DPP4	1803	HGNC:3009	NM_001935.3	P27487	GRCh38	chr2:162,848,755-162,931,052 - strand.

FIG. 6





ਤ ਸ	nan Taq	Human Taqman Gain of allele assays	Human Taqman Gain of allele assays	all galil of allere assays
	Fwd	TGGCTTATTCTCTATTCCTCACCTA (SEQ ID NO: 18)	Fwd	TGCAGACTTGTCTTGACATTCATA (SEQ ID NO: 21)
7333 hTII	Probe (BH	Probe (BHQ) TGCTTTCCCTCCTCCCTTCTGA (SEQ ID NO: 19)	7333 Probe (BHQ) hTD	7333 Probe (BHQ) AGCCTCTGCAGACACAGGAATGGC (SEQ ID NO: 22)
	Rev	GGCCTTAGCCCAGAAACTG (SEQ ID NO: 20)	Rev	TCTGGGCACTGGTGTACTC (SEQ ID NO: 23)

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FIG. 9

MKTPWKVLLG	LLGAAALVTI	ITVPVVLLNX	GTDDATADSR	KTYTLTDYLK	NTYRLKLYSL
RWISCHEYLY	KQENNILVFN	AEYGNSSVFL	ENSTFDEFGH	SINDYSISPD	GQFILLEYNY
VKQWRHSYTA	SYDIYOLNKR	QLITEERIPN	NTQWVTWSPV	GHKLAYVWNN	DIYVKIEPNL
PSYRITWTGK	EDIIYNGITD	WVYEEEVFSA	YSALWWSPNG	TFLAYAQFND	TEVPLIEYSF
YSDESLQYPK	TVRVPYPKAG	AVNPTVKFFV	VNTDSLSSVT	NATSIQITAP	ASMLIGDHYL
CDVTWATQER	ISLQWLRRIQ	NYSVMDICDY	DESSGRWNCL	VARQHIEMST	TGWVGRFRPS
EPHFTLDGNS	FYKIISNEEG	YRHICYFQID	KKDCTFITKG	TWEVIGIEAL	TSDYLYYISN
EYKGMPGGRN	LYKIQLSDYT	KVTCLSCELN	PERCQYYSVS	FSKEAKYYQL	RCSGPGLPLY
TLHSSVNDKG	LRVLEDNSAL	DKMLQNVQMP	SKKLDFIILN	ETKFWYQMIL	PPHFDKSXKY
PLLLDVYAGP	CSQKADTVFR	LNWATYLAST	ENIIVASFDG	RGSGYQGDKI	MHAINRRLGT
FEVEDQIEAA	RQFSKMGFVD	NKRIAIWGWS	YGGYVTSMVL	GSGSGVFKCG	IAVAPVSRWE
YYDSVYTERY	MGLPTPEDNL	DHYRNSTVMS	RAENFKQVEY	LLIHGTADDN	VHFQQSAQIS
KALVDVGVDF	QAMWYTDEDH	GIASSTAHQH	IYTHMSHFIK	QCFSLP (SEC	2 ID NO: 24)

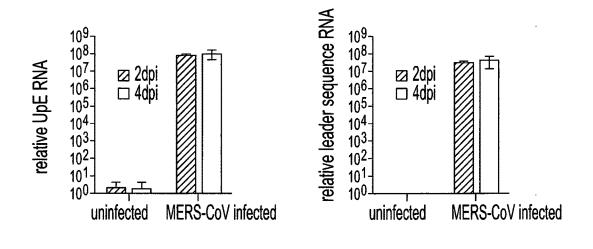
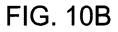


FIG. 10A



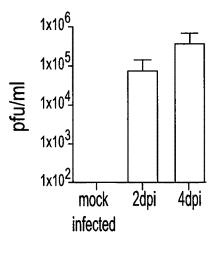


FIG. 10C

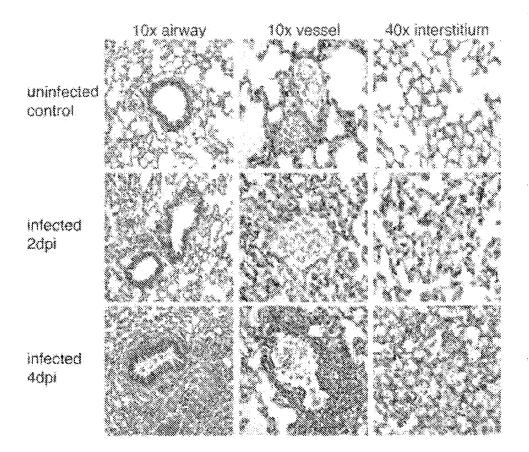


FIG. 10D

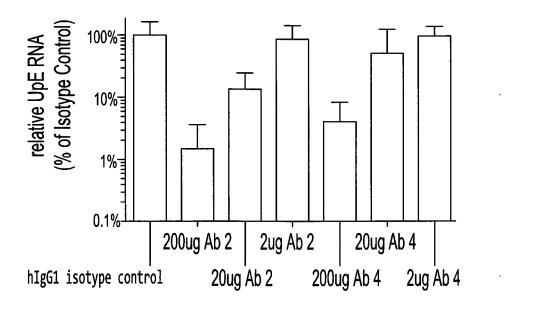


FIG. 11A

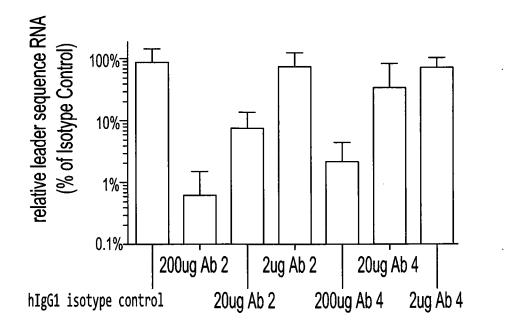


FIG. 11B

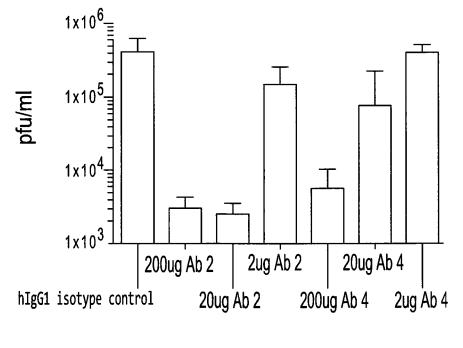
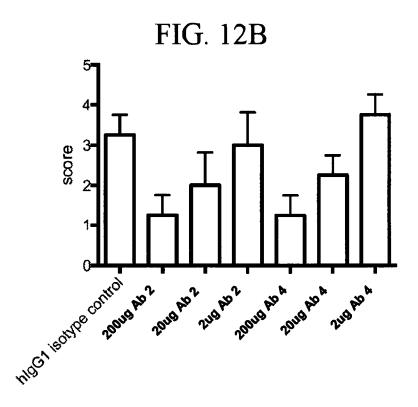


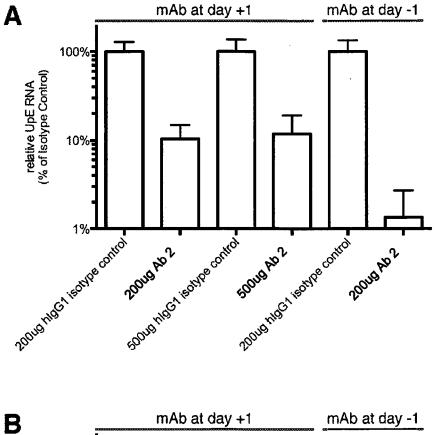
FIG. 11C

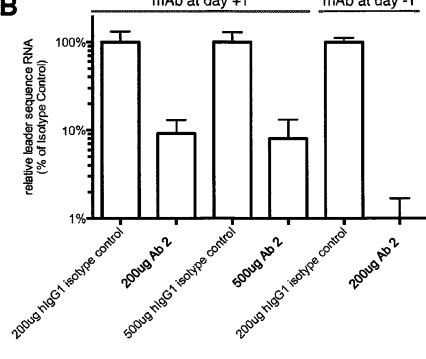
FIG. 12A

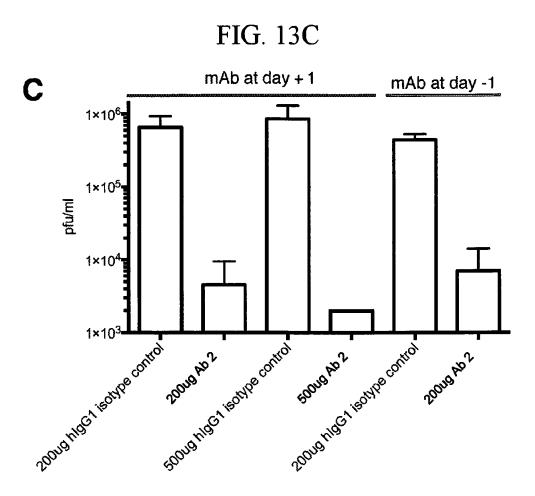
	10x airway 오페이지 아파 아파 아파	t0x vessel 《 · · · · · · · · · · · · · · · · · · ·	40x interstitium
uninfected control	8-		
higG1 isotype contro			
200ug			
Ab 2		14S	
20ug		1. 1. 1. 1. 1. 1. 1. 1. 1. 1. 1. 1. 1. 1	XXX.
Ab 2			
2ug		X:A.K.	S.M.
Ab 2	KS (
200ug			
Ab 4			
20ug			
Ab 4			
Zug		N-S.	
Ab 4			



FIGS. 13A-13B







A

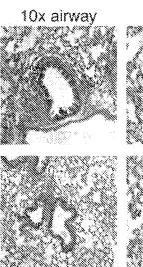
FIGS. 14A-14B

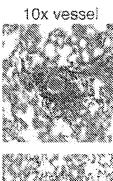
hlgG1 isotype control

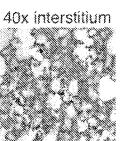
200ug

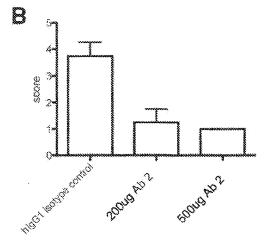
Ab 2

500ug Ab 2









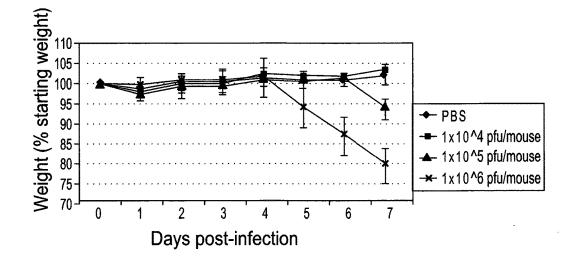


FIG. 15

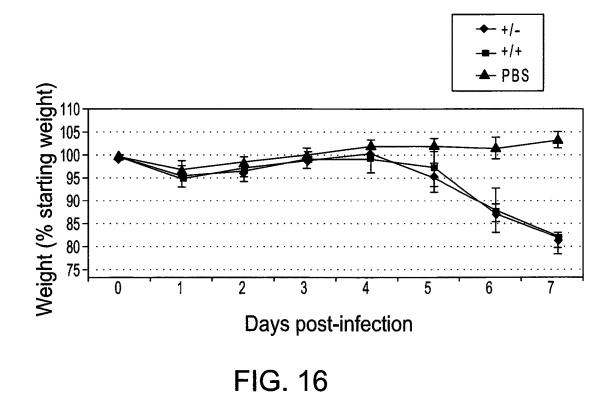
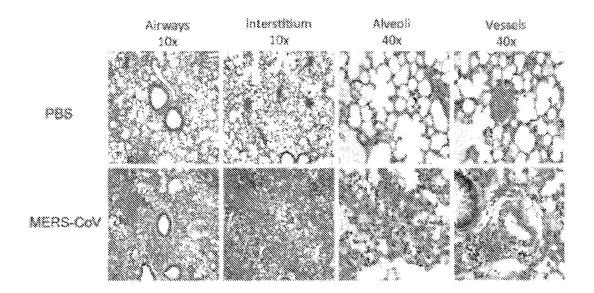


FIG. 17



41560167v.1

HUMANIZED DIPEPTIDYL-PEPTIDASE IV (DPP4) ANIMALS

CROSS-REFERENCE TO RELATED APPLICATIONS

[0001] This application is a continuation of U.S. patent application Ser. No. 15/295,955, filed Oct. 17, 2016, which is a continuation of U.S. patent application Ser. No. 14/723, 855, filed May 28, 2015, now U.S. Pat. No. 9,497,945, which claims priority to U.S. Provisional Patent Application No. 62/005,476, filed May 30, 2014, U.S. Provisional Patent Application No. 62/051,626, filed Sep. 17, 2014, and U.S. Provisional Patent Application No. 62/072,692, filed Oct. 30, 2014, the disclosures of each of which are incorporated by reference herein in their entireties.

INCORPORATION BY REFERENCE

[0002] The content of the following submission on ASCII text file is incorporated herein by reference in its entirety: a computer readable form (CFR) of the Sequence Listing (file name: 47206_501C02US_Sequence_Listing_122016, date created: Dec. 30, 2016, size: 31, 792 bytes).

FIELD OF INVENTION

[0003] Non-human animals comprising nucleic acid sequences encoding a dipeptidyl peptidase IV (DPP4) protein that comprise a human sequence. Transgenic non-human animals comprising a DPP4 gene that is human in whole or in part. Non-human animals that express human or humanized DPP4 proteins. Methods for making and using non-human animals comprising human or humanized DPP4 nucleic acid sequences.

BACKGROUND

[0004] Dipeptidyl peptidase IV (DPP4) is a therapeutic target for the treatment of a variety of human diseases, disorders and conditions, including, for example, hyperglycemia (see, e.g., Gerich (2013) Pathogenesis and Management of Postpandrial Hyperglycemia: Role of Incretin-Based Therapies, Intl. J. Gen. Med. 6:877-895) and Middle East respiratory syndrome coronavirus (MERS-CoV) infection (see, e.g., Raj et al. (2013) Dipeptidyl Peptidase 4 is a Functional Receptor for the Emerging Human Coronovirus-EMC, Nature 495(7440):251-254).

[0005] The evaluation of the pharmacokinetics (PK) and pharmacodynamics (PD) of therapeutic molecules that specifically target human DPP4 protein are routinely performed in non-human animals, e.g., rodents, e.g., mice or rats. However, the PD of such molecules cannot properly be determined in certain non-human animals if these therapeutic molecules also do not target the endogenous Dpp4 protein.

[0006] Moreover, the evaluation of the in vivo therapeutic efficacy of human DPP4-specific small molecule, peptide or protein (i.e., biologic) antagonists in non-human animal models of diseases is problematic in certain non-human animals in which the species-specific antagonist does not interact with the endogenous Dpp4 protein. Furthermore, the evaluation of the in vivo therapeutic efficacy of small molecule, peptide or protein (i.e., biologic) antagonists that target molecules that specifically interact with human DPP4 protein is also problematic in certain non-human animals in

which the therapeutic target molecule itself does not interact with the endogenous Dpp4 protein.

[0007] Accordingly, there is a need for non-human animals, e.g., rodents, e.g., mice or rats that comprise a human or humanized DPP4 gene. For example, there is a need for non-human animals, e.g., rodents, e.g., mice or rats, in which the Dpp4 gene of the non-human animal is humanized in whole or in part or replaced (e.g., at the endogenous nonhuman loci) with a human DPP4 gene comprising sequences encoding human or humanized DPP4 protein.

[0008] There is also a need for non-human animals comprising a DPP4 gene (e.g., human or humanized) in which the DPP4 gene is under control of non-human regulatory elements (e.g., endogenous regulatory elements), for example, in the 5' flanking region, e.g., promoter and enhancer(s), or in the 3' untranslated region, of the DPP4 gene.

[0009] There is also a need for non-human animals comprising a DPP4 gene (e.g., human or humanized) in which the DPP4 gene is under control of human regulatory elements, for example, in the 5' flanking region, e.g., promoter or enhancer(s), or in the 3' untranslated region, of the human DPP4 gene.

[0010] There is also a need for humanized non-human animals that express human or humanized DPP4 protein on the surface of immune cells, e.g., T cells, and/or on the surface of cells in one or more tissues, e.g., placenta, kidney, lung, liver, skeletal muscle, heart, brain and/or pancreas, at a level similar to that of Dpp4 protein on the surface of immune cells, e.g., T cells, and/or on the surface of cells in one or more tissues, e.g., placenta, kidney, lung, liver, skeletal muscle, heart, brain and/or pancreas, of an agematched non-human animal that expresses functional Dpp4 protein, but does not comprise the human or humanized DPP4 gene.

[0011] In addition, there is a need for humanized nonhuman animals that express human or humanized DPP4 protein on the surface of immune cells, e.g., T cells, and/or on the surface of cells in one or more tissues, e.g., placenta, kidney, lung, liver, skeletal muscle, heart, brain and/or pancreas, at a level higher than or lower than that of Dpp4 protein on the surface of immune cells, e.g., T cells, and/or on the surface of cells in one or more tissues, e.g., placenta, kidney, lung, liver, skeletal muscle, heart, brain and/or pancreas, of an age-matched non-human animal that expresses functional Dpp4 protein, but does not comprise the human or humanized DPP4 gene.

[0012] Throughout this specification, various patents, patent applications and other types of publications (e.g., journal articles, electronic database entries, etc.) are referenced. The disclosure of all patents, patent applications, and other publications cited herein are hereby incorporated by reference in their entirety for all purposes.

SUMMARY

[0013] Non-human animals comprising nucleic acid sequences encoding a DPP4 protein that comprises a human sequence are provided.

[0014] Transgenic non-human animals comprising a DPP4 gene that is human in whole or in part are provided.

[0015] Non-human animals that express human or humanized DPP4 protein are provided. **[0016]** Non-human animals having a replacement (in whole or in part) of the endogenous non-human animal Dpp4 gene are provided.

[0017] Non-human animals comprising a DPP4 humanization (in whole or in part) at an endogenous non-human Dpp4 locus are provided.

[0018] Non-human animals are provided that have a human or humanized DPP4 gene, wherein the non-human animals do not express endogenous Dpp4 protein, and wherein the non-human animals express human or humanized DPP4 protein on the surface of immune cells, e.g., T cells, and/or on the surface of cells in one or more tissues, including placenta, kidney, lung, liver, skeletal muscle, heart, brain and/or pancreas, at a level similar to that of Dpp4 protein present on the surface of immune cells, e.g., T cells, and/or on the surface of cells in one or more tissues, including placenta, kidney, lung, liver, skeletal muscle, heart, brain and/or pancreas, at a level similar to that of Dpp4 protein present on the surface of cells in one or more tissues, including placenta, kidney, lung, liver, skeletal muscle, heart, brain and/or pancreas, of an age-matched non-human animal that expresses functional endogenous Dpp4 protein, but does not comprise the replacement.

[0019] In one aspect, non-human animals comprising a human or humanized DPP4 nucleic acid sequence are provided.

[0020] In one aspect, genetically modified non-human animals are provided that comprise a replacement at an endogenous Dpp4 locus of a gene encoding an endogenous Dpp4 gene encoding a human or humanized DPP4 protein. Rodents, e.g., mice or rats, are provided that comprise a replacement of an endogenous Dpp4 gene, at an endogenous Dpp4 locus, with a human Dpp4 gene. In one embodiment, the rodent is heterozygous for a replacement at an endogenous Dpp4 locus of an endogenous Dpp4 gene encoding a human or humanized DPP4 protein. In one embodiment, the rodent is homozygous for a replacement at an endogenous Dpp4 locus of an endogenous Dpp4 gene encoding a human or humanized DPP4 protein. In one embodiment, the rodent is none embodiment, the rodent is a mouse. In one embodiment, the rodent is a rat.

[0021] In one aspect, genetically modified rodents, e.g., mice or rats, are provided comprising a humanization of an endogenous rodent Dpp4 gene, wherein the humanization comprises a replacement at the endogenous rodent Dpp4 locus of a rodent gene encoding an exon of an Dpp4 gene with a nucleic acid sequence encoding at least one human DPP4 gene to form a modified DPP4 gene, wherein expression of the modified DPP4 gene is under control of rodent regulatory elements at the endogenous rodent Dpp4 locus. **[0022]** In one embodiment, the rodent is heterozygous for the nucleic acid sequence encoding at least one exon of a human DPP4 gene to form a modified DPP4 gene. In one embodiment, the rodent is homozygous for the nucleic acid sequence exon of a human DPP4 gene to form a modified DPP4 gene. In one embodiment, the rodent is homozygous for the nucleic acid sequence exon of a human DPP4 gene to form a modified DPP4 gene.

[0023] In one embodiment, the rodent is a mouse or a rat. In one embodiment, the rodent is a mouse. In one embodiment, the rodent is a rat.

[0024] In one embodiment, the human DPP4 gene encoding a human or humanized DPP4 protein comprises exon 2 through exon 26 of the human DPP4 gene.

[0025] In one embodiment, the humanized DPP4 protein comprises the extracellular domain of the human DPP4 protein.

[0026] In one embodiment, the humanized DPP4 protein comprises the transmembrane domain and cytoplasmic domain of the mouse Dpp4 protein.

[0027] In one embodiment, the rodent is a mouse that is incapable of expressing a mouse Dpp4 protein.

[0028] In one embodiment, the rodent is a mouse wherein a contiguous genomic fragment of mouse Dpp4 sequence encoding exon 2 through exon 26 of mouse Dpp4 is replaced with a contiguous genomic fragment of human DPP4 sequence encoding exon 2 through exon 26 of human DPP4. [0029] In one aspect, genetically modified rodents, e.g., a mouse or rat, are provided that express a human or humanized DPP4 protein, wherein the rodent that expresses a human or humanized DPP4 protein comprises a normal immune system, i.e., the number of immune cells, e.g., T cells, in the blood, plasma or serum of the rodent expressing human or humanized DPP4 protein are similar to the number of immune cells, e.g., T cells, in the blood, plasma or serum of a rodent that expresses functional endogenous Dpp4 protein. In one embodiment, the rodent is a mouse. In one embodiment, the rodent is a rat.

[0030] In one embodiment, the blood of the rodent that expresses a human or humanized DPP4 protein has approximately the same number of immune cells, e.g., T cells, as a rodent that expresses a functional, endogenous Dpp4 protein, e.g., a wild-type mouse or rat. In one embodiment, the rodent is a mouse. In one embodiment, the rodent is a rat. [0031] In one embodiment, the mouse expressing human or humanized DPP4 on the surface of T cells has an amount of T cells present in the blood of at least about 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90%, 100%, 110%, 120%, 130%, 140%, 150%, 160%, 170%, 180%, 190% or 200% of the amount of T cells present in the blood of an age-matched mouse that expresses functional endogenous Dpp4 protein, but does not comprise a replacement of an endogenous Dpp4 gene, at an endogenous mouse Dpp4 locus, with a human DPP4 gene.

[0032] In one embodiment, the mouse expressing human or humanized DPP4 protein on the surface of T cells has an amount of T cells in the blood of between about 20% and about 200%, between about 40% and about 160%, or between about 80% and about 120% of the amount of T cells present in the blood of an age-matched mouse that expresses functional endogenous Dpp4 protein, but does not comprise a replacement of an endogenous Dpp4 gene, at an endogenous mouse Dpp4 locus, with a human DPP4 gene.

[0033] In one aspect, genetically modified rodents, e.g., a mouse or rat, are provided that express a human or humanized DPP4 protein, wherein the rodent expresses a human or humanized DPP4 protein on the surface of immune cells, e.g., T cells, and/or on the surface of cells in one or more tissues, e.g., placenta, kidney, lung, liver, skeletal muscle, heart, brain and/or pancreas, of an age-matched rodent that expresses functional endogenous Dpp4 protein. In one embodiment, the rodent is a mouse. In one embodiment, the rodent is a rat.

[0034] In one embodiment, the immune cells, e.g., T cells, of the rodent that expresses a human or humanized DPP4 protein have approximately the same level of DPP4 protein on its surface as the immune cells, e.g., T cells, of a rodent that expresses a functional, endogenous Dpp4 protein, e.g., a wild-type mouse or rat. In one embodiment, the rodent is a rat.

[0035] In one embodiment, the mouse expresses human or humanized DPP4 protein on the surface of immune cells, e.g., T cells, at a level of at least about 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90%, 100%, 110%, 120%, 130%, 140%,

150%, 160%, 170%, 180%, 190% or 200% of the level of Dpp4 protein on the surface of immune cells, e.g., T cells, of an age-matched mouse that expresses functional endogenous Dpp4 protein, but does not comprise a replacement of an endogenous Dpp4 gene, at an endogenous mouse Dpp4 locus, with a human DPP4 gene.

[0036] In one embodiment, the mouse expresses human or humanized DPP4 protein on the surface of immune cells, e.g., T cells, at a level of between about 20% and about 200%, between about 40% and about 160%, or between about 80% and about 120% of the level of mouse Dpp4 protein present on the surface of immune cells, e.g., T cells, of an age-matched mouse that expresses functional endogenous Dpp4 protein, but does not comprise a replacement of an endogenous Dpp4 gene, at an endogenous mouse Dpp4 locus, with a human DPP4 gene.

[0037] In one embodiment, the cells in one or more tissues, e.g., placenta, kidney; lung, liver, skeletal muscle, heart, brain and/or pancreas, of the rodent that expresses a human or humanized DPP4 protein have approximately the same level of DPP4 protein on its surface as the cells in one or more tissues, e.g., placenta, kidney, lung, liver, skeletal muscle, heart, brain and/or pancreas, of a rodent that expresses a functional, endogenous Dpp4 protein, e.g., a wild-type mouse or rat. In one embodiment, the rodent is a mouse. In one embodiment, the rodent is a rat.

[0038] In one embodiment, the mouse expresses human or humanized DPP4 protein on the surface of cells in one or more tissues, e.g., placenta, kidney, lung, liver, skeletal muscle, heart, brain and/or pancreas, at a level of at least about 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90%, 100%, 110%, 120%, 130%, 140%, 150%, 160%, 170%, 180%, 190% or 200% of the level of Dpp4 protein on the surface of cells in one or more tissues, e.g., placenta, kidney, lung, liver, skeletal muscle, heart, brain and/or pancreas, of an age-matched mouse that expresses functional endogenous Dpp4 protein, but does not comprise a replacement of an endogenous Dpp4 gene, at an endogenous mouse Dpp4 locus, with a human DPP4 gene.

[0039] In one embodiment, the mouse expresses human or humanized DPP4 protein on the surface of cells in one or more tissues, e.g., placenta, kidney, lung, liver, skeletal muscle, heart, brain and/or pancreas, at a level of between about 20% and about 200%, between about 40% and about 160%, or between about 80% and about 120% of the level of mouse Dpp4 protein present on the surface of cells in one or more tissues, e.g., placenta, kidney, lung, liver, skeletal muscle, heart, brain and/or pancreas, of an age-matched mouse that expresses functional endogenous Dpp4 protein, but does not comprise a replacement of an endogenous Dpp4 gene, at an endogenous mouse Dpp4 locus, with a human DPP4 gene.

[0040] In one aspect, a genetically modified rodent is provided, comprising a humanized DPP4 gene comprising a replacement of rodent Dpp4 extracellular domain-encoding sequence with human DPP4 extracellular domain-coding sequence, wherein the humanized DPP4 gene comprises a rodent Dpp4 transmembrane sequence and a rodent Dpp4 cytoplasmic sequence, wherein the humanized DPP4 gene is under control of endogenous rodent Dpp4 regulatory elements at the endogenous Dpp4 locus.

[0041] In one embodiment, the rodent is heterozygous for the humanized DPP4 gene. In one embodiment, the rodent is homozygous for the humanized DPP4 gene.

[0042] In one embodiment, the rodent is a mouse or a rat. In one embodiment, the rodent is a mouse. In one embodiment, the rodent is a rat.

[0043] In one embodiment, the mouse is incapable of expressing a mouse Dpp4 protein.

[0044] In one embodiment, the rodent regulatory elements or sequences at the endogenous rodent Dpp4 locus are from a mouse or a rat.

[0045] In one embodiment, the rodent regulatory elements or sequences are endogenous rodent regulatory elements or sequences at the rodent Dpp4 locus are from a mouse or a rat.

[0046] In one aspect, a non-human animal, e.g., a rodent, e.g., a mouse or rat, is provided that expresses human or humanized DPP4 protein, wherein the non-human animal expresses human or humanized DPP4 protein from an endogenous non-human Dpp4 locus. In an embodiment, the non-human animal is a rodent. In an embodiment, the rodent is a mouse. In an embodiment, the rodent is heterozygous for the endogenous non-human Dpp4 locus expressing a human or humanized DPP4 protein. In one embodiment, the rodent is homozygous for the endogenous non-human Dpp4 locus expressing a human or humanized DPP4 protein.

[0047] In one aspect, a genetically modified mouse is provided that expresses human or humanized DPP4 protein from an endogenous mouse Dpp4 locus, wherein the endogenous mouse Dpp4 gene has been replaced, in whole or in part, with a human DPP4 gene.

[0048] In one embodiment, about 78.8 kb at the endogenous mouse Dpp4 locus, including exon 2 through the stop codon in exon 26, is deleted and replaced with about 81.8 kb of human DPP4 gene sequence comprising exon 2 through exon 26 and a portion of the 3' untranslated sequence of the human DPP4 gene. In a specific embodiment, the human DPP4 gene comprises exon 2 through exon 26 and a portion of the 3' untranslated sequence of the human DPP4 gene of human BAC RP11-68L22. In a specific embodiment, the DPP4 gene comprises mouse Dpp4 gene 5' regulatory elements, mouse Dpp4 exon 1, including the first two amino acids, Met and Lys, of the mouse Dpp4 protein, and mouse Dpp4 3' regulatory elements (e.g., 3' untranslated region), and human DPP4 gene exon 2 through exon 26, i.e., the human DPP4 protein coding sequences, except for the first two amino acids, which are derived from mouse Dpp4 exon 1.

[0049] In one aspect, a genetically modified mouse is provided that comprises a nucleotide sequence encoding a human or humanized DPP4 protein, wherein the nucleotide sequence encoding the human or humanized DPP4 protein replaces, in whole or in part, an endogenous nucleotide sequence encoding an endogenous mouse Dpp4 protein.

[0050] In one embodiment, the mouse is heterozygous for the nucleotide sequence encoding a human or humanized DPP4 protein. In one embodiment, the mouse is homozygous for the nucleotide sequence encoding a human or humanized DPP4 protein.

[0051] In one aspect, a method is provided for making a humanized DPP4 rodent, comprising replacing a rodent Dpp4 gene sequence encoding rodent Dpp4 protein with a human DPP4 gene sequence comprising one or more exons of the human DPP4 gene sequence encoding human or humanized DPP4 protein, wherein the replacement is at an endogenous rodent Dpp4 locus and the human DPP4 gene

sequence comprising one or more exons of the human DPP4 gene sequence encoding human or humanized DPP4 protein is operably linked to rodent regulatory elements or sequences at the endogenous rodent Dpp4 locus.

[0052] In one embodiment, the rodent is heterozygous for the nucleotide sequence encoding a human or humanized DPP4 protein. In one embodiment, the rodent is homozygous for the nucleotide sequence encoding a human or humanized DPP4 protein.

[0053] In one embodiment, the rodent is a mouse or a rat. In one embodiment, the rodent is a mouse. In one embodiment, the rodent is a rat.

[0054] In one embodiment, the rodent regulatory elements or sequences are derived from a mouse. In one embodiment, the rodent regulatory elements or sequences are derived from a rat.

[0055] In one embodiment, the rodent regulatory elements or sequences are endogenous rodent regulatory elements or sequences at the rodent Dpp4 locus. In one embodiment, the rodent is a mouse. In one embodiment, the rodent is a rat. [0056] In one embodiment, the human DPP4 gene sequence replacing the rodent Dpp4 gene sequence comprises at least one exon of the human DPP4 gene sequence. In other embodiments, the human DPP4 gene sequence replacing the rodent Dpp4 gene sequence comprises at least 2, at least 3, at least 4, at least 5, at least 6, at least 7, at least 8, at least 9, at least 10, at least 11, at least 12, at least 13, at least 14, at least 15, at least 16, at least 17, at least 18, at least 19, at least 20, at least 21, at least 22, at least 23, at least 24, or at least 25 exons of the human DPP4 gene sequence. In one embodiment, the human DPP4 gene sequence replacing the rodent Dpp4 gene sequence comprises all 26 exons of the human DPP4 gene sequence. In one embodiment, the rodent is a mouse. In one embodiment, the rodent is a rat.

[0057] In one embodiment, the human or humanized DPP4 gene sequence replacing the rodent Dpp4 gene sequence encodes a protein that is about 85%, 90%, 95%, 96%, 97%, 98%, or about 99% identical to a human DPP4

[0058] In one embodiment, the human or humanized DPP4 gene sequence replacing the rodent Dpp4 gene sequence comprises at least one exon of the human DPP4 gene sequence encoding the extracellular domain of the human DPP4 protein. In other embodiments, the human DPP4 gene sequence replacing the rodent Dpp4 gene sequence comprises at least 2, at least 3, at least 4, at least 5, at least 6, at least 7, at least 8, at least 9, at least 10, at least 11, at least 12, at least 13, at least 14, at least 15, at least 16, at least 17, at least 18, at least 19, at least 20, at least 21, at least 22, or at least 23 exons of the human DPP4 gene sequence encoding the extracellular domain of the human DPP4 protein. In one embodiment, the human DPP4 gene sequence replacing the rodent Dpp4 gene sequence comprises all 24 exons of the human DPP4 gene sequence encoding the extracellular domain of the human DPP4 protein. In one embodiment, the rodent is a mouse. In one embodiment, the rodent is a rat.

[0059] In one embodiment, the human or humanized DPP4 gene sequence replacing the rodent Dpp4 gene sequence encodes an extracellular domain of the DPP4 protein that is about 85%, 90%, 95%, 96%, 97%, 98%, or about 99% identical to the extracellular domain of a human DPP4 protein.

[0060] In one embodiment, the replacement is at an endogenous rodent Dpp4 locus and the human DPP4 gene sequence comprising one or more exons of the human DPP4 gene sequence encoding human or humanized DPP4 protein is operably linked to endogenous rodent regulatory elements or sequences at the endogenous rodent Dpp4 locus.

[0061] In one aspect, a method is provided for making a humanized DPP4 mouse, comprising replacing a mouse Dpp4 gene sequence encoding mouse Dpp4 protein with a human DPP4 gene sequence encoding human or humanized DPP4 protein.

[0062] In one embodiment, the replacement is at an endogenous mouse Dpp4 locus, and the human DPP4 gene encoding human or humanized DPP4 protein is operably linked to mouse regulatory elements or sequences at the endogenous mouse Dpp4 locus.

[0063] In one embodiment, the replacement is at an endogenous mouse Dpp4 locus, and the human DPP4 gene encoding human or humanized DPP4 protein is operably linked to endogenous mouse regulatory elements or sequences at the endogenous mouse Dpp4 locus.

[0064] In various aspects, the genetically modified nonhuman animals, e.g., rodents, e.g., mice or rats, described herein comprise the genetic modifications in their germ-line. [0065] In one aspect, a non-human animal, e.g., rodent, e.g., a mouse or rat, embryo comprising a genetic modifi-

e.g., a mouse or rat, embryo comprising a genetic modification as described herein is provided.

[0066] In one aspect, a non-human animal, e.g., rodent, e.g. a mouse or rat, host embryo is provided that comprises a donor cell that comprises a genetic modification as described herein.

[0067] In one aspect, a pluripotent or totipotent nonhuman animal, e.g., rodent, e.g., mouse or rat, cell comprising a genetic modification as described herein is provided. In one embodiment, the cell is a rodent cell. In one embodiment, the cell is a mouse cell. In one embodiment, the cell is a rodent embryonic stem (ES) cell. In one embodiment, the cell is a mouse ES cell.

[0068] In one aspect, a non-human animal, e.g., rodent, e.g., mouse or rat, egg is provided, wherein the non-human animal egg comprises an ectopic non-human animal chromosome, wherein the ectopic non-human animal chromosome comprises a genetic modification as described herein. In one embodiment, the non-human animal is a rodent. In one embodiment, the rodent is a mouse. In one embodiment, the rodent is a rat.

[0069] In one aspect, the mouse embryo, egg, or cell that is genetically modified to comprise a human DPP4 gene is of a mouse that is of a C57BL strain selected from C57BL/ A, C57BL/An, C57BL/GrFa, C57BL/KaLwN, C57BL/6, C57BL/61, C57BL/6ByJ, C57BL/6NJ, C57BL/10, C57BL/ 10ScSn, C57BL/10Cr, and C57BL/Ola. In another embodiment, the mouse is a 129 strain selected from the group consisting of a strain that is 129P1, 129P2, 129P3, 129X1, 129S1 (e.g., 129S1/SV, 129S1/Svlm), 129S2, 129S4, 129S5, 129S9/SvEvH, 129S6 (129/SvEvTac), 129S7, 129S8, 129T1, 129T2 (see, e.g., Festing et al. (1999) Revised nomenclature for strain 129 mice, Mammalian Genome 10:836, see also, Auerbach et al (2000) Establishment and Chimera Analysis of 129/SvEv- and C57BL/6-Derived Mouse Embryonic Stem Cell Lines). In a specific embodiment, the genetically modified mouse is a mix of an aforementioned 129 strain and an aforementioned C57BL/6 strain. In another specific embodiment, the mouse is a mix

of aforementioned 129 strains, or a mix of aforementioned BL/6 strains. In a specific embodiment, the 129 strain of the mix is a 129S6 (129/SvEvTac) strain. In another embodiment, the mouse is a BALB strain, e.g., BALB/c strain. In yet another embodiment, the mouse is a mix of a BALB strain and another aforementioned strain. In one embodiment, the mouse is Swiss or Swiss Webster mouse.

[0070] In various aspects, the non-human animals comprising a human or humanized DPP4 nucleic acid sequence are selected from mammals and birds. In one embodiment, the non-human animals are mammals. In one embodiment, the mammals are rodents. In one embodiment, the rodents are mice or rats.

[0071] In one aspect, a rodent is provided that comprises a nucleic acid sequence comprising a human DPP4 gene or fragment thereof, where the human DPP4 gene or fragment thereof comprises at least one exon of the human DPP4 gene, and where the human DPP4 gene or fragment thereof encodes a human or humanized DPP4 protein.

[0072] In one embodiment, the human DPP4 gene or fragment thereof comprises at least 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 18, 20, 21, 22, 23, 24, or 25 exons of the human DPP4 gene.

[0073] In one embodiment, the human DPP4 gene or fragment thereof comprises all 26 exons of the human DPP4 gene.

[0074] In one embodiment, the nucleic acid sequence further comprises a 5' flanking region of the human DPP4 gene. In one embodiment, the human DPP4 gene or fragment thereof is operably linked to the 5' flanking region of the human DPP4 gene. In one embodiment, the 5' flanking region of the human DPP4 gene comprises at least 1 kb in length (e.g., at least 1, 2, 3, 4, 5, 10, 20, 30, 40, 50 kb, or greater, in length). In one embodiment, the 5' flanking region of the human DPP4 gene comprises at least 10 kb in length. In one embodiment, the 5' flanking region of the human DPP4 gene comprises at least 10 kb in length.

[0075] In one embodiment, expression of the human DPP4 gene or fragment thereof is under control of the 5' flanking region of the human DPP4 gene.

[0076] In one embodiment, the human or humanized DPP4 protein comprises the amino acid sequence of SEQ ID NO: 24 or a fragment thereof.

[0077] In one embodiment, the rodent expresses the human or humanized DPP4 protein on the surface of T cells in a level that is at least about 20% (e.g., at least about 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90%, 95%, 98%, 99%, or greater) of the level of rodent Dpp4 protein present on the surface of T cells of an age-matched rodent that expresses functional endogenous rodent Dpp4 protein but that does not comprise the human DPP4 gene or fragment thereof.

[0078] In one embodiment, the rodent expresses the human or humanized DPP4 protein on the surface of cells in one or more tissues selected from the group consisting of placenta, kidney, lung, liver, skeletal muscle, heart, brain, and pancreas, in a level that is at least about 20% (e.g., at least about 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90%, 95%, 98%, 99%, or greater) of the level of rodent Dpp4 protein present on the surface of one or more tissues of an age-matched rodent that expresses functional endogenous rodent Dpp4 protein but that does not comprise the human DPP4 gene or fragment thereof.

[0079] In one embodiment, the rodent expresses functional endogenous rodent Dpp4 protein.

[0080] In one embodiment, the rodent is a mouse or a rat. **[0081]** In one aspect, provided herein is a method for making a humanized transgenic rodent, comprising integrating a nucleic acid sequence comprising one or more exons of a human DPP4 gene sequence into a chromosome of a rodent, where the one or more exons of the human DPP4 gene sequence encodes a human or humanized DPP4 protein.

[0082] In one embodiment, the human DPP4 gene or fragment thereof comprises at least 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 18, 20, 21, 22, 23, 24, or 25 exons of the human DPP4 gene.

[0083] In one embodiment, the human DPP4 gene or fragment thereof comprises all 26 exons of the human DPP4 gene.

[0084] In one embodiment, the nucleic acid sequence further comprises a 5' flanking region of the human DPP4 gene.

[0085] In one embodiment, the human DPP4 gene sequence is operably linked to the 5' flanking region of the human DPP4 gene.

[0086] In one embodiment, the human or humanized DPP4 protein comprises the amino acid sequence of SEQ ID NO: 24 or a fragment thereof.

[0087] In one embodiment, the rodent is a mouse or a rat. In further aspects, methods for determining the in vivo therapeutic efficacy of a human-specific DPP4 antagonist in any of the humanized DPP4 rodents described herein are provided, the method comprising administering to the rodent a DPP4 antagonist, wherein the rodent is infected with Middle East respiratory syndrome coronavirus (MERS-CoV); and determining if whether the DPP4 antagonist treats or prevents one or more symptoms of MERS-CoV infection compared to control rodents infected with MERS-CoV who have not been administered the DPP4 antagonist. [0088] In one embodiment, the DPP4 antagonist is selected from the group consisting of small molecules, peptides and antibodies.

[0089] In one embodiment, the DPP4 antagonist is an antibody to a MERS-CoV protein.

[0090] In one embodiment, the MERS-CoV protein is MERS-CoV spike protein.

[0091] In one embodiment, the rodent is infected with one or more strains of MERS-CoV selected from the group consisting of Al-Hasa_1, Al-Hasa_2, Al-Hasa_3, Al-Hasa_4, Al-Hasa_12, Al-Hasa_15, Al-Hasa_16, Al-Hasa_17, Al-Hasa_18, Al-Hasa_19, Al-Hasa_21, Al-Hasa_25, Buraidah_1, EMC/2012, FRA/UAE, Hafr-Al-Batin_1, Hafr-Al-Batin_2, Hafr-Al-Batin_6, Jeddah_1, Jordan-N3/2012, Munich, Riyadh_3, Riyadh_4, Riyadh_5, Riyadh_14, Taif_ 1, Wadi-Ad-Dawasir_1, Riyadh_9, KFU-HKU 1, KFU-HKU 13, Qatar3, Qatar4, England 1, England-Qatar/2012, Bisha_1, Riyadh_1, and Riyadh_2.

[0092] In one embodiment, the antagonist is administered before MERS-CoV infection. In one embodiment, the antagonist is administered after MERS-CoV infection.

[0093] In one embodiment, the antagonist is administered simultaneously with MERS-CoV infection.

[0094] In one embodiment, the symptom of MERS-CoV infection is viral titer or RNA level.

[0095] In one embodiment, the viral titer or RNA level is assessed by one or more methods selected from the group consisting of qPCR, Northern Blot, plaque assay, and in situ hybridization,

[0096] In one embodiment, the symptom of MERS-CoV infection is lung inflammation.

[0097] In one embodiment, the lung inflammation is assessed histochemically

[0098] In one embodiment, the symptom of MERS-CoV infection is weight loss.

[0099] In one embodiment, the rodent is a mouse or a rat. In one embodiment, the rodent is a mouse. In one embodiment, the rodent is a rat.

[0100] Each of the aspects and embodiments described herein are capable of being used together, unless excluded either explicitly or clearly from the context of the embodiment or aspect.

BRIEF DESCRIPTION OF THE DRAWINGS

[0101] FIG. 1A and FIG. 1B provide illustrations, not to scale, of the strategy for humanization of the Dpp4 locus. FIG. 1A is a schematic showing that 78.8 kb of the mouse Dpp4 gene (top) spanning exon 2 through the stop codon in exon 26 are deleted and replaced with 81.7 kb of the human DPP4 gene (bottom) spanning exon 2 through exon 26 and a portion of the 3' untranslated region, as indicated. FIG. 1B is a schematic showing that the humanized DPP4 mouse comprises (i) the mouse Dpp4 gene 5' flanking region, including the regulatory sequences, e.g., promoter and transcription start site, and exon 1, including the initiation ATG codon, (ii) the human DPP4 gene spanning exon 2 through exon 26, including the Stop codon, and a portion of the 3'untranslated region, including the loxP site, and (iii) the mouse Dpp4 gene 3' untranslated region starting from just 3' to the Stop codon, as indicated.

[0102] FIG. **2** shows the amino acid sequence (SEQ ID NO:17) of the humanized DPP4 protein expressed in the humanized DPP4 mice.

[0103] FIG. **3** shows the results of real-time PCR performed on RNA obtained from lung tissue from either mock (PBS)-infected, or MERS-CoV (Jordan strain)-infected, F0 humanized DPP4 mice 4 days after infection.

[0104] FIG. **4** shows the HU staining of airway (10x and 40x magnification) and alveoli (40x magnification) from the lungs of either mock (PBS)-infected, or MERS-CoV (Jordan strain)-infected, F0 humanized DPP4 mice 4 days after infection.

[0105] FIG. **5** is a sequence alignment of the mouse Dpp4 (mDpp4) amino acid sequence (SEQ ID NO: 25) with the human DPP4 (hDPP4) protein encoded by the transgenic MAID 7326/7327 mice (SEQ ID NO: 26). Non-homologous residues that differ between the sequences are underlined, homologous residues that differ between the sequences are bolded and italicized, and gaps are indicated by hyphens. Residues that are identical between the two sequences are shown in unformatted text.

[0106] FIG. **6** is a table displaying gene, sequence, and chromosomal information for human DPP4.

[0107] FIG. 7 is a schematic showing the coverage of the human DPP4 gene and flanking genomic sequences for each of the BACs, BAC RP11-345J9 and BAC RP11-68L22. The locations within the human DPP4 gene and promoter regions at which the human TaqManTM primer-probe sets anneal are also shown (7333hTU for the upstream set and 7333hTD for the downstream set).

[0108] FIG. **8** is a table displaying the primer-probe sets used for the human TaqManTM gain of allele assays, where 7333 hTU refers to the upstream set and 7333 hTD refers to the downstream set.

[0109] FIG. **9** is the amino acid sequence of the humanized DPP4 protein encoded by the transgenic MAID 7333 and 7334 mice (SEQ ID NO: 24).

[0110] FIG. **10**A is a bar graph showing quantitative PCR measurements of MERS-CoV genome (UpE RNA) in infected mice 2 and 4 days post-infection (dpi). FIG. **10**B is a bar graph showing quantitative PCR measurements of MERS-CoV mRNA transcript (leader RNA) in infected mice on 2 dpi and 4 dpi. FIG. **10**C is a bar graph quantifying MERS-CoV viral titer of infected mouse lung at 2 dpi and 4 dpi. MERS-CoV levels in homogenized mouse lung were quantified by 50% Tissue Culture Infective Dose (TCID50) assay and expressed as plaque forming units (pfu) per mL. FIG. **10**D is a panel of histological images, stained with Hematoxylin and Eosin, of lungs from MERS-CoV infected mice. Airway (10×), vasculature (10×) and interstitium (40×) are shown for PBS, 2 dpi, and 4 dpi mice.

[0111] FIG. **11**A is a bar graph showing quantitative PCR measurements of MERS-CoV genome (UpE RNA) from lungs of mice pre-treated with anti-MERS-CoV spike protein antibodies (Ab 2 or Ab 4) before viral infection. FIG. **11**B is a bar graph showing quantitative PCR measurements of MERS-CoV mRNA transcript (leader RNA) from lungs of mice pre-treated with anti-MERS-CoV spike protein antibodies before viral infection. RNA was quantified using primers directed against the MERS-CoV genome and compared to hlgG 1 isotype control treated mice. All samples were compared to hlgG 1 control set at 100%. FIG. **11**C is a bar graph showing the viral titer in lungs of mice pre-treated with anti-MERS-CoV spike protein antibodies before viral infection. Viral titer was quantitated by plaque assay and reported as pfu/mL.

[0112] FIG. **12**A is a panel of histological images of lungs of MERS-CoV infected B6/hDPP4 mice with anti-MERS-CoV spike protein antibody (Ab 2 or Ab 4) pre-treatment. Hematoxylin and Eosin stained sections of mouse lung showing airway, vasculature and interstitium of a representative mouse from each group. FIG. **12**B is a bar graph showing the histological scoring of the mouse lungs shown in FIG. **12**A. The scores were the average scores of all mice in each experimental group and time point.

[0113] FIG. 13A is a bar graph showing quantitative PCR measurements of MERS-CoV genome (UpE RNA) from infected lungs. Effects of anti-MERS-CoV spike protein antibodies (Ab 2 or Ab 4) injected one day before or one day after viral infection were compared. FIG. 13B is a bar graph showing quantitative PCR measurements of MERS-CoV mRNA transcript (leader RNA) from infected lungs. Effects of anti-MERS-CoV spike protein antibodies (Ab 2 or Ab 4) injected one day before or one day after viral infection were compared. For FIG. 13A and FIG. 13B, RNA was quantified using primers directed against the MERS-CoV genome and compared to hIgG 1 isotype control treated mice. All samples were compared to hIgG 1 control set at 100%. FIG. 13C is a bar graph showing the viral titer in lungs of mice treated with anti-MERS-Co V spike protein antibodies (Ab 2 or Ab 4) after viral infection. Viral titer was quantitated by plaque assay and reported as PFU/mL lung. Effects of antibodies injected one day before or one day after viral infection were compared.

[0114] FIG. **14**A is a panel of histological images of lungs from MERS-CoV infected B6/hDPP4 mice with anti-MERS-CoV spike protein Ab 2 antibody treatment at 1 day post-infection. Hematoxylin and Eosin stained sections of mouse lung show airway, vasculature, and interstitium of a representative mouse from each group. FIG. **14**B is a bar graph showing the histological scoring of the mouse lungs of FIG. **14**A.

[0115] FIG. **15** depicts a dose-response study of weight as a function of time post-infection with MERS-CoV in humanized DPP4 mice. 4-5 mice are used per group.

[0116] FIG. **16** depicts a dose-response study of weight as a function of time post-infection with MERS-CoV $(1 \times 10^6 \text{ pfu/mouse}; 4-5 \text{ mice per group})$ in heterozygotic and homozygotic humanized DPP4 mice.

[0117] FIG. **17** depicts pathology (inflammation) as seen by histological examination at day 7 in a humanized DPP4 mouse exposed to a high dose of virus $(1 \times 10^6 \text{ pfu/mouse})$ versus PBS-treated controls.

DETAILED DESCRIPTION

DPP4 Gene and Protein

[0118] The DPP4 gene encodes the type II transmembrane protein, tripeptidyl peptidase IV (DPP4) (also known as CD26, adenosine deaminase complexing protein-2 (ADCP2), adenosine deaminase binding protein (ADABP), and TP103), which has serine exopeptidase activity, and which plays an important role in the activation of T cells and in intracellular signal transduction cascades in several other cell types.

[0119] Human DPP4.

[0120] NCBI Gene ID: 1803; Primary source: HGNC: 3009; RefSeq transcript: NM_001935.3; UniProt ID: P27487; Genomic assembly: GRCh38; Location: chr2:162, 848,755-162,931,052-strand. (See FIG. **6**).

[0121] The human DPP4 gene is located on chromosome 2, at 2q24.3. The human DPP4 gene has 26 exons and encodes a type II transmembrane polypeptide of 766 amino acids in length, including an N-terminal 6 amino acid cytoplasmic domain, a 22 amino acid transmembrane domain, and a C-terminal 738 amino acid extracellular domain. The extracellular domain (i.e., ectodomain) of the human DPP4 protein is encoded by coding exons 3 through 26 of the human DPP4 gene.

[0122] Mouse Dpp44.

[0123] NCBI Gene ID: 13482; Primary source: MGI: 94919; RefSeq transcript: NM_010074.3; UniProt ID: P28843; Genomic assembly: GRCm38; Location: chr2:62, 330,073-62,412,591-strand.

[0124] The mouse Dpp4 gene is located on chromosome 2, at 2 35.85 cM. The mouse Dpp4 gene has 26 exons and encodes a type II transmembrane polypeptide of 760 amino acids in length, including an N-terminal 6 amino acid cytoplasmic domain, a 22 amino acid transmembrane domain, and a C-terminal 732 amino acid extracellular domain. The extracellular domain (i.e., ectodomain) of the mouse Dpp4 protein is encoded by coding exons 3 through 26 of the mouse Dpp4 gene.

Species Specificity of DPP4 Protein

[0125] As discussed below in Example 2, the human, but not the mouse, DPP4 protein is a functional receptor for the Middle East respiratory syndrome coronavirus (MERS-CoV) infection.

[0126] Candidate therapeutic molecules that target the human DPP4 protein in a species-specific manner, or target molecules, such as MERS-CoV, which interact with the human DPP4 protein in a species-specific manner, are typically evaluated for pharmacokinetics (PK) and pharmacodynamics (PK) in non-human animals, e.g., rodents, e.g., mice or rats. Such therapeutic molecules are also tested for in vivo therapeutic efficacy in non-human animal, e.g., rodent, e.g., mouse or rat, models of human diseases, disorders and conditions in which DPP4 plays a role.

[0127] However, therapeutic molecules that are specific for the human DPP4 protein, e.g., human-specific DPP4 inhibitors, cannot be adequately evaluated for PD and/or in vivo therapeutic efficacy in rodents, in particular mice, because the targets of these therapeutic molecules are missing.

[0128] Moreover, therapeutic molecules that are specific for targets that specifically interact with human DPP4 protein, e.g., human DPP4-specific MERS-CoV, cannot be adequately evaluated for in vivo therapeutic efficacy in rodents, in particular mice, because the targets (e.g., receptor, interaction partner) of these therapeutic target molecules are missing.

[0129] Accordingly, in various embodiments, to assess the PD and/or in vivo therapeutic efficacy of a human-specific DPP4 protein antagonist or inhibitor in non-human animals, e.g., rodents, e.g., mice or rats, it is desirable to replace the endogenous Dpp4 protein with human or humanized DPP4 protein. In various embodiments, to assess the in vivo therapeutic efficacy of small molecules, peptides or biologic antagonists or inhibitors of a target molecule that specifically interacts with a human DPP4 protein in non-human animals, e.g., rodents, e.g., mice or rats, it is desirable to replace the endogenous Dpp4 protein with human or humanized DPP4 protein.

[0130] Further, in various embodiments, in order to avoid potential problems of the over- or under-expression of the human or humanized DPP4 protein, and/or the inappropriate expression of the human or humanized DPP4 protein in cells or tissues in which the endogenous Dpp4 protein is not normally expressed, it is desirable to insert the human DPP4 gene, in whole or in part, into the genome of the non-human animals, e.g., rodents, e.g., mice or rats, at the endogenous Dpp4 gene loci, and to express the human or humanized DPP4 protein in non-human animals, e.g., rodents, e.g., mice or rats, under the control, at least in part, of the endogenous Dpp4 regulatory elements.

[0131] In some embodiments, targeted replacement of the endogenous, e.g., mouse or rat, Dpp4 gene by the human DPP4 gene or fragment thereof is desirable.

[0132] In other embodiments, the human DPP4 gene or fragment thereof is randomly inserted into the rodent, e.g., mouse or rat, genome instead of replacing the endogenous Dpp4 gene with a human DPP4 gene or fragment thereof. In some embodiments, in rodents, e.g., mice or rats, in which the human DPP4 gene or fragment thereof has been randomly inserted into the genome, expression of endogenous rodent Dpp4 is retained.

[0133] Provided herein are non-human animals, e.g., rodents, e.g., mice or rats, that comprise a human DPP4 gene or fragment thereof either at (i.e., replacing) the endogenous Dpp4 locus, or at one or more other loci. Also provided herein are non-human animals, e.g., rodents, e.g., mice or rats, that comprise a human DPP4 gene or fragment thereof both at (i.e., replacing) the endogenous Dpp4 locus, and at an additional locus/loci.

[0134] In some embodiments, a fragment of a human DPP4 gene contains 200 kilobases (kb) or fewer nucleotides, e.g., 180, 160, 140, 120, 100, 80, 70, 60, 50, 40, 30, 20, 10, 5, 2.5, 1 kb or fewer nucleotides, e.g., 1000, 800, 600, 400, 200, or fewer nucleotides.

Generation of Cells and Non-Human Animals with Human DPP4

[0135] For targeted replacement of an endogenous nonhuman Dpp4 gene or fragment with a human DPP4 gene or fragment, a targeting construct is generated. See, e.g., Valenzuela et al. Nature Biotech, 21.6(2003):652-659; U.S. Pat. Nos. 6,586,251; and 8,759,105. For example, a targeting construct comprises homology arms flanking a replacement human DPP4 gene or fragment thereof.

[0136] In some embodiments, the replacement human DPP4 gene or fragment thereof comprises the entire human DPP4 gene. In other embodiments, the replacement human DPP4 gene or fragment thereof comprises a portion of the human DPP4 gene. For example, the replacement human DPP4 gene or fragment thereof comprises one or more exons of human DPP4 gene, e.g., 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, or 26 exons of the human DPP4 gene. For example, the replacement human DPP4 gene or fragment thereof comprises the exons 1-26 of the human DPP4 gene. In other embodiments, the replacement human DPP4 gene or fragment thereof comprises exons 2-26 of the human DPP4 gene. For example, the replacement human DPP4 gene or fragment thereof comprises intron 1 upstream of exon 2 through exon 26 of the human DPP4 gene. In some embodiments, the replacement human DPP4 gene or fragment thereof further comprises a human regulatory element(s), e.g., a portion of the human 3' untranslated region (UTR) downstream of the human DPP4 gene, for example at least 1 kb of downstream region (e.g., at least 1, 2, 3, 4, 5, 10, 20, 30, 40 Kb, or greater), and/or a human promoter or enhancer region upstream of the human DPP4 gene, for example, at least 10 kb of upstream region (e.g., at least 10, 20, 30, 40, 50 Kb, or greater).

[0137] Homology arms are sequences that are homologous to endogenous chromosomal nucleic acid sequences flanking the desired genetic modification/replacement, e.g., flanking the endogenous Dpp4 gene or fragment that is to be replaced. Homologous nucleic acid sequences can be two or more nucleic acid sequences that are either identical or similar enough that they are able to hybridize to each other or undergo intermolecular exchange. Due to the homology between the homology arms and the corresponding endogenous sequence, the homology arms direct the targeting construct to a specific chromosomal location within the genome, e.g., the endogenous Dpp4 gene locus. See, e.g., Valenzuela et al. Nature Biotech, 21.6(2003):652-659; U.S. Pat. Nos. 6,586,251; and 8,759,105.

[0138] Optionally, the targeting construct further comprises a selection marker, e.g., in between the two homology arms. Exemplary selection markers include antibiotic resis-

tance markers (e.g., neomycin or kanamycin) and fluorescent proteins. In some embodiments, the selection marker is foxed, i.e., flanked by two loxP sites. The floxed selection marker can be removed by the addition of Cre recombinase, which catalyzes the excision of the floxed segment, e.g., including the selection marker.

[0139] Vector/Constructs

[0140] The transgenic non-human animals (e.g., rodents, e.g., mice or rats) of the invention can be made by using various vectors and/or constructs. In some embodiments, the targeting construct is in the form of a circular piece of double-stranded DNA, e.g., a bacterial artificial chromosome (BAC), plasmid, or P1-derived artificial chromosome (PAC).

[0141] To generate a non-human cell comprising a targeted replacement of the endogenous Dpp4 locus, a targeting construct containing a human DPP4 gene or fragment described herein is introduced into a non-human (e.g., rodent, e.g., mouse or rat) cell, e.g., embryonic stem (ES) cell.

[0142] To generate a non-human cell comprising a human DPP4 gene or fragment randomly inserted into the genome, a circular DNA construct, e.g., BAC, containing a human DPP4 gene or fragment thereof, is introduced into a nonhuman (e.g., rodent, e.g., mouse or rat) cell, e.g., ES cell. In some cases, the circular DNA construct, e.g., BAC, further contains a human DPP4 regulatory element, e.g., a human promoter or enhancer region upstream and/or downstream of human DPP4 gene. For example, the circular DNA construct contains at least 10 kb (e.g., at least 10, 20, 30, 40, 50 kb or greater) of promoter/enhancer region upstream of the ATG start codon of the human DPP4 gene. In addition or alternatively, the circular DNA construct contains at least 1 kb (e.g., at least 1, 2, 3, 4, 5, 10, 20, 30, 40 kb or greater) of untranslated region downstream of the human DPP4 gene. For example, the human DPP4 gene or fragment is operably linked to the human DPP4 regulatory element.

[0143] In some embodiments, the human DPP4 gene or fragment thereof in the circular DNA construct (e.g., BAC) comprises the entire human DPP4 gene. In other embodiments, the human DPP4 gene or fragment thereof comprises a portion of the human DPP4 gene. For example, the human DPP4 gene or fragment thereof comprises one or more exons of human DPP4 gene, e.g., 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, or 26 exons of the human DPP4 gene. For example, the human DPP4 gene or fragment thereof comprises the exons 1-26 of the human DPP4 gene. In other embodiments, the human DPP4 gene or fragment thereof comprises exons 2-26 of the human DPP4 gene. For example, the human DPP4 gene or fragment thereof comprises exons 2-26 of the human DPP4 gene. For example, the human DPP4 gene or fragment thereof comprises exons 2-26 of the human DPP4 gene. For example, the human DPP4 gene or fragment thereof comprises exons 2-26 of the human DPP4 gene. For example, the human DPP4 gene or fragment thereof comprises exons 2-26 of the human DPP4 gene. For example, the human DPP4 gene or fragment thereof comprises exons 2-26 of the human DPP4 gene. For example, the human DPP4 gene or fragment thereof comprises exons 2-26 of the human DPP4 gene. For example, the human DPP4 gene or fragment thereof comprises intron 1 upstream of exon 2 through exon 26 of the human DPP4 gene.

[0144] For example, the introduction step into the cell is done by electroporation or lipid-mediated transfection.

[0145] Optionally, the circular DNA construct, e.g., BAC, is linearized before introduction into the cell. For example, linearization is performed with rare-cutting restriction enzymes, e.g., SgrDI, SfiI, NotI, PacI, or SwaI.

[0146] In cases in which the targeting construct comprises an antibiotic selection marker (e.g., neomycin), cells that have taken up the targeting construct are optionally selected in neomycin/G418-containing media. Cells that survive and/ or proliferate in neomycin/G418-containing media are selected and positive for the targeting construct. **[0147]** In some embodiments, the cell population is screened for those cells that have incorporated into their genome a human DPP4 gene or fragment thereof, e.g., randomly inserted into the genome or targeted (e.g., by the targeting construct) to the endogenous Dpp4 locus.

[0148] Methods for screening include quantitative PCR and fluorescence in situ hybridization. See, e.g., U.S. Pat. No. 6,586,251 B2 and U.S. Pat. No. 8,759,105 B2. For example, methods of screening include detecting for the presence of a human DPP4 gene or fragment. In some embodiments, methods of screening include detecting for a loss of copy number of endogenous Dpp4 gene or fragment and/or gain of copy number of human DPP4 gene or fragment. Exemplary methods of screening are described in Examples 1 and 3.

[0149] In some embodiments in which the targeting construct comprises a floxed selection marker, correctly targeted cells are optionally further electroporated with a Cre-expressing vector, e.g., transiently expressing Cre recombinase, to remove the floxed selection marker.

[0150] To generate transgenic animals, positive ES cell clones, e.g., without floxed selection marker, containing a human DPP4 gene or fragment thereof, are introduced into a rodent embryo, e.g., a mouse or rat embryo, such as an 8-cell stage mouse embryo. For example, the introduction step is done by blastocyst injection technology, aggregation techniques, nuclear transfer and cloning, and/or the Veloci-Mouse® method. See, e.g., U.S. Pat. No. 8,759,105 B2, U.S. Pat. Nos. 7,294,754, 7,576,259, and 7,659,442. For example, an ES cell clone is a subpopulation of cells derived from a single cell of the ES cell population following introduction of DNA and subsequent selection.

[0151] In some cases, DNA from transgenic non-human animals are screened in similar ways as described above to confirm transmittance of the human DPP4 gene/fragment through the germline.

[0152] In some embodiments, the humanized DPP4 rodents described herein are heterozygous for the human DPP4 allele. As such, these rodents have one human DPP4 allele and one wild-type rodent DPP4 allele. In other embodiments, the humanized DPP4 rodents are homozygous for the human DPP4 allele.

Uses for Humanized DPP4 Rodents

[0153] Humanized DPP4 rodents, e.g., mice or rats, are useful to evaluate the pharmacodynamics (PD) of human-specific DPP4 antagonists, e.g., small molecule, peptide or biologic inhibitors, useful for the treatment of hyperglycemia.

[0154] Pharmacokinetics (PK) and PD assays in humanized DPP4 rodents, e.g, mice or rats, are performed according to standard procedures known in the art.

[0155] Humanized DPP4 rodents, e.g., mice or rats, are useful to evaluate the in vivo therapeutic efficacy of human-specific DPP4 antagonists, e.g., small molecule, peptide or biologic inhibitors, in the treatment of hyperglycemia.

[0156] Humanized DPP4 rodents, e.g., mice or rats, are useful to test the in vivo therapeutic efficacy of antagonists, e.g., small molecule, peptide or biologic inhibitors, e.g., neutralizing antibodies, that are specific for target molecules, e.g., MERS-CoV (e.g., spike protein (S) of MERS-CoV, e.g., receptor binding domain of the spike protein of MERS-CoV), which specifically interact with human DPP4, in the treatment or prevention (or prophylaxis) of MERS-CoV infection. In some embodiments, rodents that are heterozygous for the human DPP4 allele are used to test the in vivo therapeutic efficacy of one or more antagonists in the treatment or prevention (or prophylaxis) of ME RS-Co V infection. In other embodiments, DPP4 rodents that are homozygous for the human DPP4 allele are used to test the in vivo therapeutic efficacy of one or more antagonists in the treatment or prevention (or prophylaxis) of MERS-CoV infection.

[0157] Exemplary MERS-CoV strains include MERS-CoV Jordan strain (GenBank accession no. KC776174.1, MERS-CoV-Hu/Jordan-N3/2012) and MERS-CoV EMC/ 2012 strain (GenBank accession no. JX869059.2). In some embodiments, a MERS-CoV virus described herein comprises a MERS-CoV clinical isolate. In other embodiments, a MERS-CoV virus described herein comprises a strain comprising the same spike protein receptor binding domain (RBD) sequence as a clinical isolate described herein. Exemplary clinical isolates are shown in the table below. The table shows the amino acid sequence variation within the receptor binding domain (RBD) of the spike protein of several MERS-CoV clinical isolates. National Center for Biotechnology Information (NCBI)-deposited sequences of MERS-CoV clinical isolates were aligned at amino acids 367-606 and compared to that of the EMC/2012 strain. Clinical isolates harboring the A431P, S457G, S460F, A482V, L506F, D509G, and V534A substitutions (where the amino acid (single letter designation) preceding the number is that of the EMC/2012 strain, and the amino acid (single letter designation) following the number is that of the clinical isolate) are shown in the table.

TABLE I

No variation from EMC/2012 sequence	A431P	S457G	S460F	A482V	L506F	D509G	V534A
Al-Hasa_1 Al-Hasa_2 Al-Hasa_3 Al-Hasa_4 Al-Hasa_12 Al-Hasa_15 Al-Hasa_16 Al-Hasa_17 Al-Hasa_18 Al-Hasa_19	Riyadh_9	KFU-HKU 1 KFU-HKU 13	Qatar3 Qatar4	Riyadh_9	England 1 England- Qatar/2012	Bisha_1 Riyadh_1	Riyadh_2

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No variation from EMC/2012 sequence	A431P	S457G	S460F	A482V	L506F	D509G	V534A
Al-Hasa_21							
Al-Hasa_25 Buraidah_1							
EMC/2012							
FRA/UAE							
Hafr-Al-Batin_1							
Hafr-Al-Batin_2							
Hafr-Al-Batin_6							
Jeddah_1							
Jordan-N3/2012							
Munich							
Riyadh_3 Riyadh_4							
Riyadh_5							
Riyadh_14							
Taif_1							
Wadi-Ad-Dawasir_1							

[0158] In some embodiments, an antagonist is administered before (e.g., at least 1, 2, 4, 6, 12, 24, 48 hours, 2, 3, 4, 5, 6, or 7 days, or more before) a MERS-CoV infection in the rodent. In other embodiments, the antagonist is administered after (e.g., at least 1, 2, 4, 6, 12, 24, 48 hours, 2, 3, 4, 5, 6, or 7 days, or more after) a MERS-CoV infection in the rodent.

[0159] In some embodiments, where an antagonist is administered to a rodent after MERS-CoV infection, a lower viral titer or RNA level (e.g., viral UpE or leader sequence RNA level) in the rodent after administration of the antagonist, e.g., lower by at least 5-fold (e.g., at least 5, 10, 50, 100, 500, 1000, 10^4 , 10^5 , 10^6 , 10^7 -fold or more) compared to a control level indicates that the antagonist is effective in treating a MERS-CoV infection. For example, a control level is the viral titer or RNA level in the rodent prior to administration of the antagonist. In other examples, a control level is the viral titer or RNA level in a virus-infected rodent that is untreated with the antagonist.

[0160] In some embodiments, where an antagonist is administered to a rodent prior to MERS-CoV infection, a lower viral titer or RNA level (e.g., viral UpE or leader sequence RNA level) in the rodent after administration of the antagonist, e.g., lower by at least 5-fold (e.g., at least 5, 10, 50, 100, 500, 1000, 10^4 , 10^5 , 10^6 , 10^7 -fold or more) compared to a control level indicates that the antagonist is effective in preventing a MERS-CoV infection. For example, a control level is the viral titer or RNA level of a rodent infected with MERS-CoV that was not treated with the antagonist.

[0161] In some embodiments, viral RNA levels in a rodent lung can be determined by extracting RNA from the rodent lung by homogenization in a solution containing phenol, e.g., a solution containing phenol and guanidinium isothiocyanate (e.g., Trizol® (Life Technologies, Inc)). For example, the lung can be homogenized using a Magnalyzer (Roche) according to the manufacturers' instructions. In some embodiments, levels of MERS-CoV RNA can be assessed using quantitative PCR (qPCR) using primers that target the MERS-CoV genome and/or primers that target the MERS-CoV mRNA transcript. For example, the Taqman® Fast virus one-step master mix (Applied Biosystems) can be used in qPCR according to the manufacturers' instructions using a duplex of primers obtained from Life Technologies targeting a region of the genome upstream of the envelope gene (UpE) or the leader sequence of the nucleocapsid messenger RNA (leader primer), and compared to an endogenous control, such as rodent (e.g., mouse) 18S rRNA. For example, qPCR reactions in Microamp® fast optical reaction plates (Applied Biosystems) can be read on a 7500 fast DX real-time PCR instrument (Applied Biosystems). In some examples, qPCR data can be analyzed using the delta Ct method, with an uninfected control set to 1. For example, percent MERS-CoV RNA detected in infected mice treated with control antagonist, e.g., isotype-matched control antibodies, in cases where the antagonist is an antibody against MERS-CoV.

[0162] In some embodiments, the viral titer in a rodent lung can be determined by homogenizing the rodent lung in a buffer (e.g., phosphate buffered saline (PBS)), centrifuged (e.g., at 10,000 rpm). The supernatant can be analyzed by plaque assay on mammalian cells, such as Vero cells (e.g., Vero E6 cells) to quantitate levels of virus remaining after treatment with an antagonist. A standard plaque assay can be used, e.g., a plaque assay described in Page et al. (2012) Induction of alternatively activated macrophages enhances pathogenesis during severe acute respiratory syndrome coronavirus infection, J Virol 86:13334-13349. In some examples, the Page et al. plaque assay can be modified by leaving plates for 3 days for plaques to appear.

[0163] In some embodiments, less inflammation (e.g., interstitial or peri-vascular inflammation), fewer inflammatory cells, and/or less bronchiolar cuffing, e.g., determined by histological analysis, in a lung sample of a rodent treated with an antagonist prior to or after MERS-CoV infection compared to a control lung sample can indicate that the antagonist is effective in treating or preventing a MERS-CoV infection. For example, a control lung sample can be from a rodent infected with MERS-CoV that is untreated with an antagonist, e.g., untreated before and/or after infection.

[0164] The rodents described herein are useful for the efficient testing of drugs and vaccines for MERS-CoV, e.g., to demonstrate safety and efficacy prior to clinical testing in

humans. They permit rapid identification and/or validation of therapeutics/prophylactics, e.g., within several weeks of testing.

Definitions

[0165] Homologous amino acid residues are residues that share similar characteristics or properties. Characteristics or properties of an amino acid residue are based on, e.g., the structure of the polypeptide backbone, for example, a sheet or helical conformation, the charge or hydrophobicity of the residue, and/or the bulk of the side chain(s). For example, homologous residues are similar in side chain properties, e.g., polarity, charge, size, aromaticity, and/or hydrophobicity.

[0166] The term, "about" refers to a stated value plus or minus another amount; thereby establishing a range of values. In certain embodiments, "about" indicates a range relative to a base (or core or reference) value or amount plus or minus up to 15%, 14%, 13%, 12%, 11%, 10%, 9%, 8%, 7%, 6%, 5%, 4%, 3%, 2%, 1%, 0.75%, 0.5%, 0.25% or 0.1%. For example, about refers to a range of $\pm -5\%$ below and above the recited numbers, e.g., numbers of nucleotide bases.

[0167] The term "operably linked" as used herein refers to positions of components so described, e.g., nucleotide sequences, are in a relationship permitting them to function in their intended manner.

[0168] As used herein, the term "protein" includes polypeptides, peptides, fragments of polypeptides, and fusion polypeptides.

[0169] As used herein, a "nucleic acid" refers to two or more deoxyribonucleotides and/or ribonucleotides covalently joined together in either single or double-stranded form.

[0170] The term "replacement" in reference to gene replacement refers to placing exogenous genetic material at an endogenous genetic locus, thereby replacing all or a portion of the endogenous gene with an orthologous or homologous nucleic acid sequence. In one instance, an endogenous non-human gene or fragment thereof is replaced with a corresponding human gene or fragment thereof is a human gene or fragment that is an ortholog of, a homolog of, or is substantially identical or the same in structure and/or function, as the endogenous non-human gene or fragment thereof that is replaced.

[0171] As used herein, the term "rodent" refers to any member of the order Rodentia. Examples of rodents include, without limitation, mice, rats, squirrels, prairie dogs, porcupines, beavers, guinea pigs, and hamsters. In one embodiment, a rodent is a rat. In another embodiment, a rodent is a mouse.

[0172] Unless defined otherwise herein, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention pertains.

[0173] As used herein, the singular terms "a," "an," and "the" include the plural reference unless the context clearly indicates otherwise.

[0174] It is intended that every maximum numerical limitation given throughout this specification includes every lower numerical limitation, as if such lower numerical limitations were expressly written herein. Every minimum numerical limitation given throughout this specification will include every higher numerical limitation, as if such higher numerical limitations were expressly written herein. Every numerical range given throughout this specification will include every narrower numerical range that falls within such broader numerical range, as if such narrower numerical ranges were all expressly written herein.

[0175] The following examples are provided for illustrative purposes only and not intended to limit the invention in any manner.

EXAMPLES

Example 1

Replacement of the Endogenous Mouse Dpp4 Gene with a Human DPP4 Gene

[0176] The 81.8 kb human DPP4 gene containing exon 2 through exon 26 and a portion of the 3' untranslated region of the human DPP4 gene replaced 78.8 kb of the murine Dpp4 gene locus spanning exon 2 through the stop codon in exon 26. See FIGS. **1**A and **1**B.

[0177] A targeting construct for replacing the mouse with the human DPP4 gene in a single targeting step was constructed using VelociGene® genetic engineering technology (see Valenzuela et al. (2003) High-throughput engineering of the mouse genome coupled with high-resolution expression analysis, Nature Biotech, 21(6):652-659). Mouse and human DPP4 DNA were obtained from bacterial artificial chromosome (BAC) clones RP23-362N15 and RP11-68L22, respectively. Briefly, an SgrDI linearized targeting construct generated by gap repair cloning containing mouse Dpp4 upstream and downstream homology arms flanking 82 kb of human DPP4 sequence extending from intron 1 upstream of exon 2 through exon 26, including the stop codon and a portion of the 3' untranslated region (genomic coordinates of the entire human DPP4 gene: GRCh38: chr2:162,848,755-162,931,052 (- strand)), and a floxed neo selection cassette, was electroporated into VGB6 mouse embryonic stem (ES) cells (derived from C57BL/6N mice). Correctly targeted ES cells (MAID 7326) were further electroporated with a transient Cre-expressing vector to remove the drug selection cassette. Targeted ES cell clones without drug cassette (MAID 7327) were introduced into an 8-cell stage SW mouse embryo by the VelociMouse® method (see, U.S. Pat. Nos. 7,294,754, 7,576,259, 7,659, 442, and Poueymirou et al. (2007) F0 generation mice that are essentially fully derived from the donor gene-targeted ES cells allowing immediate phenotypic analyses Nature Biotech. 25(1):91-99). VelociMice® (F0 mice fully derived from the donor ES cell) bearing the humanized DPP4 gene were identified by genotyping for loss of mouse allele and gain of human allele using a modification of allele assay (see, Valenzuela et al. (2003)).

[0178] Correctly targeted ES cell clones were identified by a loss-of-native-allele (LONA) assay (Valenzuela et al. 2003) in which the number of copies of the native, unmodified Dpp4 gene were determined by two TaqManTM quantitative polymerase chain reactions (qPCRs) specific for sequences in the mouse Dpp4 gene that were targeted for deletion. The qPCR assays comprised the following primerprobe sets (written 5' to 3'): upstream forward primer, TCGCCACTGT GCCTAACATA G (SEQ ID NO:1); upstream reverse primer, CCGGGACTAA ACTGGAACAT TC (SEQ ID NO:2); upstream probe, FAM-TCAGTCAACT TCTTCTGGGT TGTTTCC-BHQ (SEQ ID NO:3); downstream forward primer, CAGCTCTGGT GGAGAACTAG AC (SEQ ID NO:4); downstream reverse primer, GGAG-GTCCTC GGTCTTTAGA AG (SEQ ID NO:5); downstream probe, FAM-TCACACTTAG GCTTATAAAC CAT-TCCCGT-BHQ (SEQ ID NO:6); in which FAM refers to the 5-carboxyfluorescein fluorescent probe and BHQ refers to the fluorescence quencher of the black hole quencher type (Biosearch Technologies). DNA purified from ES cell clones that have taken up the targeting vector and incorporated in their genomes was combined with TaqMan[™] Gene Expression Master Mix (Life Technologies) according to the manufacturer's suggestions in a 384-well PCR plate (Micro-Amp[™] Optical 384-Well Reaction Plate, Life Technologies) and cycled in an Applied Biosystems Prism 7900HT, which collects fluorescence data during the course of the PCRs and determines a threshold cycle (Ct), the fractional PCR cycle at which the accumulated fluorescence reaches a pre-set threshold. The upstream and downstream DPP4-specific qPCRs and two qPCRs for non-targeted reference genes were run for each DNA sample. The differences in the Ct values (Mt) between each DPP4-specific qPCR and each reference gene qPCR were calculated, and then the difference between each \mathbb{P} Ct and the median Δ Ct for all samples assayed was calculated to obtain $\Delta\Delta$ Ct values for each sample. The copy number of the DPP4 gene in each sample was calculated from the following formula: copy number= $2 \times 2^{-\Delta \Delta Ct}$. A correctly targeted clone, having lost one of its native copies, will have a Dpp4 gene copy number equal to one. Confirmation that the human DPP4 gene sequence replaced the deleted mouse Dpp4 gene sequence in the humanized allele was confirmed by a TaqMan[™] qPCR assay that comprises the following primer-probe sets (written 5' to 3'): human upstream forward primer, GCG-GTCTCCC TCTTCTAACG (SEQ ID NO:7); human upstream reverse primer, GCAAGCCGAG CAGATCAAG (SEQ ID NO:8); human upstream probe, FAM-ACTC-CCACCT GCAAATCCTG CTGC-BHQ (SEQ ID NO:9); human downstream forward primer, AACCGCACTG GCATATGGA (SEQ ID NO:10); human downstream reverse primer, TACAAGGTAG TCTGGGATTA CTAACAAAA (SEQ ID NO:11); human downstream probe, FAM-ACATTTATCT AGAAAGGCTC-BHQ (SEQ ID NO:12).

[0179] The same LONA assay is used to assay DNA purified from tail biopsies for mice derived from the targeted ES cells to determine their DPP4 genotypes and confirm that the humanized DPP4 allele is transmitted through the germline. Two pups heterozygous for the replacement are bred to generate a mouse that is homozygous for the replacement of the endogenous mouse Dpp4 gene by the human DPP4 gene. Pups that are homozygous for the replacement are used for phenotyping.

[0180] The upstream junction of the murine Dpp4 locus and the sequence containing the human DPP4 gene is designed to be within 5'-AGGAGAGAAG CCAACAAGAT CATAAGATCA TGCTCAGGGC CAAAAATTCAA GGGCTTCTGC (CGTCGACG) GCCTTAGAGA ACTC-CAACTG GCGCACTCCA GACGCCACCC CCAC-CCCCAG CCCGCGGTCT CCCTCTTCTA ACG-CACTCCC ACCTGCAAAT (SEQ ID NO:13), wherein the human DPP4 sequences are italicized, and the SgrDI restriction site is bracketed. The downstream junction of the sequence containing the human DPP4 gene and the floxed neo selection cassette is designed to be within 5'-TTATTC-CAGG GAACTATGAT GAGGCTTATA TAAGAACGAA TAAGATCAGA AATATCATTC TGGCAGTTCT TATG-GCTCAG ctcgag(ataa cttcgtataa tgtatgctat acgaagttat) atgcatggcc tccgcgccgg gttttggcgc ctcccgcggg (SEQ ID NO:14), wherein the human DPP4 sequences are italicized, the neo cassette sequences are in lower case, and the loxP site is bracketed. The downstream junction of the sequence of the floxed neo selection cassette and the murine Dpp4 locus is designed to be within 5'-atgtctgga(a taacttcgta taatgtatgc tatacgaagt tat)gctagta actataacgg tcctaaggta gcgagctagc CAGCATAGCT CTCCATAGCT TATTTAAGAC CACATTTGTT CTCATTATCT CAAAAGTGCA CTGT-TAAGAT GAAGATCTTA (SEQ ID NO:15), wherein the neo cassette sequences are in lower case, and the loxP site is bracketed. After removal of the neo selection cassette, the junction of the sequence containing the human DPP4 gene, the loxP site remaining after removal of the neo selection cassette, and the murine Dpp4 locus is designed to be within 5'-TATTCCAGGG AACTATGATG AGGCTTATAT AAGAACGAAT AAGATCAGAA ATATCATTCT GGCA-GTTCTT ATGGCTCAG ctcgag(ataa cttcgtataa tgtatgctat acgaagttat) gctagtaact ataacggtcc taaggtagcg agctagcCA GCATAGCTCT CCATAGCTTA TTTAAGACCA CATTT-GTTCT CATTATCTCA AAAGTGCACT GTTAAGATGA AGATCTTAAT AATGTTGCAT TGAGACATTT CAG-GCTGCTT TCTCCAGTTT TACACCTGCA ATC-CTAACTA AGGATGCCTG TCCCCAGAAC (SEQ ID NO:16), wherein the human DPP4 sequences are italicized, the neo cassette sequences are in lower case, and the loxP site is bracketed.

[0181] FIG. **2** shows the amino acid sequence of DPP4 encoded by the humanized DPP4 nucleic sequence in MAID 7326/7327 (SEQ ID NO:17) is the same as human DPP4 because mouse Dpp4 codon 1, encodes only the first two amino acids of DPP4, Met and Lys, which are the same as those encoded by human DPP4 codon 1.

Example 2

Infection of Humanized DPP4 Mice by MERS-CoV

[0182] Middle East Respiratory Syndrome-Coronavirus (MERS-CoV) is a newly emergent virus that causes severe acute respiratory disease. The receptor for MERS-CoV is dipeptidyl peptidase IV (DPP4) (see Raj et al. (2013) Dipeptidyl Peptidase 4 is a Functional Receptor for the Emerging Human Coronavirus-EMC, Nature 495(7440): 251-254). In vivo testing of anti-viral molecules requires an animal model, e.g., a small animal model, such as a rodent (e.g., mouse or rat), that is susceptible to MERS-CoV infection. However, recent studies have shown that mouse Dpp4 cannot support MERS-CoV infection (see, e.g., Cockrell et al. (2014) Mouse Dipeptidyl Peptidase is Not a Functional Receptor for Middle East Respiratory Syndrome Coronavirus (MERS-CoV) Infection, J. Virol. 88(9):5195-5199; and Coleman et al. (2014) Wild-Type and Innate Immune-Deficient Mice are Not Susceptible to the Middle East Respiratory Syndrome Coronavirus, J. Gen. Virol. 95(2):408-412), at least in part because the MERS-CoV Spike protein interacts with human, but not mouse, DPP4 (see, e.g., Coleman et al. (2013); and Raj et al. (2013)). Sequence comparison of the sequences of mouse and human DPP4 revealed that the amino acids that have previously

been identified as contact sites between MERS-CoV spike (S) protein and its receptor differ between the two species. In addition, expression of human DPP4 in mouse cells allows for MERS-CoV virus entry and propagation, indicating that entry of the virus is the limiting step in infection of mouse cells, and that the lack of interaction between mouse DPP4 and the MERS-CoV glycoprotein defines the species tropism in vitro. See, e.g., Lu et al. (2013) Molecular basis of binding between novel human coronavirus MERS-CoV and its receptor CD26, Nature. 500(7461):227-31; and Cockrell et al. (2014) Mouse Dipeptidyl Peptidase is Not a Functional Receptor for Middle East Respiratory Syndrome Coronavirus (MERS-CoV) Infection, J. Virol. 88(9):5195-5199.

[0183] As a consequence, normal mouse strains cannot be used to measure the efficacy of therapeutics targeting MERS-CoV. Zhao et al. (2014) Rapid Generation of a Mouse Model for Middle East Respiratory Syndrome, Proc. Natl. Acad. Sci. USA 111(13):4970-4975 have expressed human DPP4 in mice by adenovirus transduction, thereby allowing for MERS-CoV infection. However, this adenovirus model has several limitations, including: (a) the virus is cleared rapidly from infected mice; (b) there is a loss of human DPP4 expression over time; (c) the tissue distribution of virally-transduced DPP4 does not reflect expression seen in mice or humans; and (d) adenovirus infection induces an interferon response.

[0184] To generate a mouse model for MERS-CoV infection, humanized DPP4 mice were generated as described above in Example 1. As shown in FIG. 1B, exons 2 through 26 of mouse Dpp4 were replaced by the corresponding sequences of human DPP4. Because the remaining mouse Dpp4 coding exon 1 encodes only the first two amino acids of Dpp4, Met and Lys, which are the same as those in corresponding human DPP4 exon 1, the DPP4 protein expressed in humanized DPP4 mice is completely human (see FIG. 2, SEQ ID NO:17). Thus, humanized DPP4 mice express a fully human DPP4 under the control of the endogenous mouse regulatory sequences, i.e., 5' flanking region (promoter and enhancer(s)), and 3' untranslated region sequences. It is expected that the humanized DPP4 is expressed in the same cells and tissues, and at the same or similar levels, as mouse Dpp4 is expressed in wild-type mice lacking human DPP4 nucleic acid sequences.

[0185] Two F0 humanized DPP4 mice were infected intranasally with MERS-CoV (Jordan strain) or mock treated with PBS. Four days post-infection, MERS-CoV RNA was quantified in the lungs by real-time PCR using primers specific for the replicative form of the MERS-CoV genome. Data was normalized to the amount of PCR product obtained from the lungs of the mock-infected mice (arbitrarily set at 1). FIG. **3** shows that MERS-CoV RNA could be amplified from lungs of the MERS-CoV-infected humanized DPP4 mice. H&E staining was also performed using lung tissue from mock- and MERS-CoV-infected mice.

[0186] FIG. **4** shows that MERS-CoV infection of humanized DPP4 mice did not affect the airway, but resulted in thickening of the walls of the alveoli and less space between alveolar cells, indicating inflammation in the lungs associated with MERS-CoV infection.

[0187] In addition, 6 to 8 week old mice were inoculated intranasally with MERS-CoV, e.g., 2×10^5 pfu of MERS-CoV (Jordan). No mortality or clinical signs of disease were observed up to day 4 after inoculation. On days 2 and 4

post-inoculation, mice were euthanized and their lungs were dissected. To obtain virus RNA levels, lungs were homogenized in Trizol®, RNA extracted, and analyzed by real-time PCR using primers specific to MERS-CoV (FIGS. **10**A and **10**B). A set of primers was specific to a region of the viral genome upstream of the envelope gene (UpE), and another set of primers was specific to the leader sequence of the nucleocapsid mRNA (leader primer). Mouse 18S rRNA was used as endogenous control.

[0188] To obtain virus titers, lungs were homogenized in phosphate buffered saline (PBS), clarified by centrifugation, and titered on Vero E6 cells (FIG. **10**C). For example, the supernatant was analyzed by a plaque assay on VeroE6 cells to quantitate the levels of virus present in the lungs. For example, plaque assays were performed as described in Page et al. (2012) Induction of alternatively activated macrophages enhances pathogenesis during severe acute respiratory syndrome coronavirus infection, J Virol 86:13334-13349, with plates left for 3 days for plaques to appear.

[0189] Robust MERS-CoV replication in the lungs was evident at 2 and 4 days post-infection. RNA quantification, using a primer set specific for MERS-CoV leader, which was designed to only amplify replicating MERS-CoV, demonstrated high levels of MERS-CoV replicating RNA in lungs collected at day 2, and these levels were maintained through day 4 post-infection (FIGS. **10**A-B). Plaque assay of lung homogenate on Vero E6 cells quantified MERS-CoV (Jordan) levels of ~ 7.27×10^4 pfu/mL lung at day 2 and ~ 3.75×10^5 pfu/mL lung at 4 days post-infection (FIG. **10**C), demonstrating active replication of MERS-CoV in the lungs of the infected humanized DPP4 mice.

[0190] Also, lungs from humanized DPP4 mice intranasally inoculated with either MERS-CoV (Jordan strain) or PBS (mock infected) were analyzed for pathological changes (FIG. **10**D). At day 2 post-infection, peri-bronchiolar inflammation was evident with alterations in bronchiolar cell structure found throughout the lungs. Minimal perivascular inflammation or effects on alveolar structures were observed at this time point. At 4 days post-infection, interstitial infiltration was observed with peri-vascular cuffing and extensive alveolar thickening. Bronchiolar alterations were present as well. See FIG. **10**D. This pathology is consistent with the radiographic findings of development of interstitial pneumonia and significant lung disease seen in humans with MERS-CoV.

[0191] The above data shows that humanized DPP4 mice, such as those described herein, are susceptible to MERS-CoV infection. The data also demonstrate that the humanized DPP4 mice described herein are an in vivo model of MERS-CoV infection that recapitulates the pathology, e.g., pathological sequelae, that is seen in MERS-CoV infection of humans.

[0192] Thus, humanized DPP4 mice are a robust model of MERS-CoV that is useful to assess MERS-CoV treatment in vivo. For example, the humanized DPP4 mice are appropriate host animals to measure the pharmacokinetics, pharmacodynamics and therapeutic efficacy of therapeutic molecules that target MERS-CoV.

[0193] FIG. **5** shows a protein sequence alignment of mouse Dpp4 (SEQ ID NO: 25) and human DPP4 (encoded by the 7326/7327 transgenic mice) (SEQ ID NO: 26).

[0194] Next, a dose-response study of weight as a function of time post-infection of MERS-CoV was conducted in humanized DPP4 mice. Mice were infected with either

MERS-CoV (Jordan strain) or PBS (mock infected) as described above and were analyzed for weight loss, which is a sign of productive infection, over a period of seven days. As shown in FIG. 14, humanized DPP4 mice exhibited productive infection (i.e., manifested disease pathology), with weight loss beginning 4 days post-infection. Four to five mice were used per group. FIG. 15 shows that mice that were heterozygotic for the humanized DPP4 allele were equally susceptible to infection by MERS-CoV, as they exhibited a similar degree of weight loss when compared to homozygotes. This finding is significant because it indicates that studies can be conducted using heterozygous mice. Additionally, use of heterozygous mice avoids any potential issue related to functional mouse Dpp4 knockouts that could potentially be present in homozygous humanized DPP4 mice.

[0195] The lungs of these mice were also examined histologically for inflammation according to the methods described above. As shown in FIG. **16**, hDPP4 mice exposed to a high dose of virus (1×10exp6 pfu/mouse) exhibited increased pathology relative to PBS controls.

Example 3

Generation of Transgenic Mice Containing the Human DPP4 Gene Using Random Insertion of BACs

[0196] Transgenic mice were generated that contain the human DPP4 gene, for which the sequence and genomic information is shown in FIG. **6**. Two different overlapping BACs containing the human DPP4 gene were used: BAC RP11-68L22 and BAC RP11-345J9 (FIG. 7). Both BACs contained the coding region of the human DPP4 gene, as well as over 40 kb of promoter region upstream of the ATG start codon of the DPP4 gene and several kilo bases downstream of the stop codon of the DPP4 gene (FIG. 7).

[0197] To generate the BAC transgenic mice, each BAC DNA was electroporated into VGB6 mouse embryonic stem (ES) cells (derived from C57BL/6N mice). ES cells containing the coding region of the human DPP4 gene, as well as promoter regions of the gene, were introduced into an 8-cell stage SW mouse embryo by the SW mouse embryo by the VelociMouse® method (see, e.g., U.S. Pat. Nos. 7,294, 754, 7,576,259, 7,659,442, and Poueymirou et al. (2007) F0 generation mice that are essentially fully derived from the donor gene-targeted ES cells allowing immediate phenotypic analyses Nature Biotech. 25(1):91-99).

[0198] Human gain of allele assays were used to screen ES cell clones for ones that contained copies of the human DPP4 gene along with promoter regions of the gene. Human gain of allele assays were also used to identify VelociMice® (F0 mice fully derived from the donor ES cell) bearing the humanized DPP4 gene along with promoter regions of the gene.

[0199] Briefly, genomic DNA was extracted from ES cell clones using standard methods and tested in a Taq ManTM quantitative PCR (q PCR) assay using two sets of primerprobes to detect a human DNA sequence upstream (7333 hTU) and downstream hTD) of the human DPP4 coding sequence (FIG. **8**). The locations within the human DPP4 gene and flanking regions (e.g., promoter regions) at which each primer-probe set annealed is shown in FIG. **7**. A fluorescent read-out above background in the TaqManTM qPCR assay indicated the presence of the human DPP4 gene and at least 40 kb of the 5' flanking region of the human DPP4 gene that had been integrated into the transgenic mouse genome.

[0200] The 7333 hTU primer-probe set (written 5' to 3') was: human upstream forward primer, TGGCTTATTCTC-TATTCCTCACCTA (SEQ ID NO: 18); human upstream FAM-TGCTTTCCCTCCTCCCTTCTGA-BHQ probe, (SEQ ID NO: 19); human upstream reverse primer, GGC-CTTAGCCCAGAAACTG (SEQ ID NO: 20). The 7333 hTD primer-probe set (written 5' to 3') was: human downstream forward primer, TGCAGACTTGTCTTGACAT-TCATA (SEQ ID NO: 21); human downstream probe, CAL-AGCCTCTGCAGACACAGGAATGGC-BHQ (SEQ ID NO: 22); and human downstream reverse primer, TCTGGGCACTGGTGTACTC (SEQ ID NO: 23); in which FAM and CAL refer to the 5-carboxyfluorescein and CAL Orange fluorescent probes, respectively, and BHQ refers to the fluorescence quencher of the black hole quencher type (Biosearch Technologies).

[0201] For example, genomic DNA from ES cell clones was combined with TaqManTM Gene Expression Master Mix (Life Technologies) according to the manufacturer's suggestions in a 384-well PCR plate (MicroAmp[™] Optical 384-Well Reaction Plate, Life Technologies) and cycled in an Applied Biosystems Prism 7900HT, which collects fluorescence data during the course of the PC Rs and determines a threshold cycle (Ct), the fractional PCR cycle at which the accumulated fluorescence reaches a pre-set threshold. The upstream and downstream DPP4-specific qPCRs and two qPCRs for non-DPP4 reference genes were run for each DNA sample. The differences in the Ct values (\square Ct) between each DPP4-specific qPCR and each reference gene qPCR were calculated, and then the difference between each at and the median 2 Ct for all samples assayed was calculated to obtain 2 Ct values for each sample. The copy number of the DPP4 gene in each sample was calculated from the following formula: copy number= $2 \times 2^{-1} \square \square^{Ct}$. A clone containing at least one copy of the human DPP4 plus promoter regions integrated into the chromosome had a DPP4 gene copy number equal to or greater than one.

[0202] The same human gain of allele assay was used to assay DNA purified from tail biopsies for mice derived from the ES cells to confirm that the humanized DPP4 allele along with the human 5' flanking regions were transmitted through the germline.

[0203] Using the BAC insertion and screening methods described herein, two transgenic mice with DNA encoding human DPP4 were confirmed. BAC RP11-68L22 was used to generate ES cell clones and transgenic mice referred to as MAID 7333, and BAC RP11-345J9 was used to generate ES cell clones and transgenic mice referred to as MAID 7334. **[0204]** The protein encoded by the humanized DPP4 nucleic acid sequence in the MAID 7333 and 7334 mice had the amino acid sequence shown in FIG. **9** (SEQ ID NO: 24), which is the same as human DPP4 (as encoded by the transcript, NM 001935.3).

Example 4

Treatment of Humanized DPP4 Mice that were Infected with the MERS-CoV Virus

[0205] Transgenic mice with the humanized DPP4 gene and flanking promoter regions were tested for their ability to

be infected by MERS-CoV and to serve as a model for assessing therapeutic molecules for treating or preventing MERS-CoV.

[0206] Transgenic MAID 7333 mice (e.g., generated by the methods described in Example 3) were treated with 200 µg of antibodies directed against MERS-CoV spike protein or isotype controls by intraperitoneal injection (ip). One day after antibody injection, the mice were infected intranasally with MERS-CoV. Four days after infection, lungs of the mice were harvested, and viral RNA levels were measured using real-time PCR (RT-PCR). In particular, levels of the genomic RNA (UpE) or replicating RNA (leader) (specific for the replicative form of the MERS-CoV genome) of MERS-CoV were measured.

[0207] The RT-PCR data is shown in the table below.

Antibody	$\mathrm{Up}\mathrm{E}^{1}$	Leader ¹
Anti-MERS-CoV spike protein 1 (Ab 1)	0.356839562	0.273565089
Anti-MERS-CoV spike protein 2 (Ab 2)	0.254493202	0.206006238
Anti-MERS-CoV spike protein 3 (Ab 3)	1.989548316	1.112094283
(IgG1) isotype control	104.0889287	101.2578723
(IgG4) isotype control	100	100

¹Averages (% of isotype control)

[0208] Treatment of transgenic mice with the antibodies decreased viral RNA levels (both UpE and Leader) by about 50-fold to about 500-fold.

[0209] The data described herein show that transgenic mice generated by targeted integration methods (Example 1) and random BAC insertion methods (Example 3) with human DPP4 were susceptible to infection by MERS-CoV. In addition, anti-MERS-CoV antibodies blocked infection in transgenic mice in vivo. Thus, transgenic mice with human DPP4 (e.g., generated by the methods described herein) are useful for evaluating the efficacy of therapeutics (e.g., antibodies) that target the MERS-CoV virus.

Example 5

Prophylactic Effects of Anti-MERS-CoV Antibodies on MERS-CoV Infection in Humanized DPP4 Mice

[0210] The humanized DPP4 mice described herein were used to evaluate the prophylactic capability of the two monoclonal antibodies in vivo. Mice were i.p. injected with a dose range of anti-MERS-CoV antibodies 200 µg, 20 µg or 2 µg of anti-MERS-CoV spike protein antibody 2 (Ab 2), anti-MERS-CoV spike protein antibody 4 (Ab 4), or 200 µg of human IgG 1 (hIgG 1) isotype control antibody-at 24 hours before intranasal infection with 1×10⁵ pfu of MERS-CoV (Jordan strain). Ab 2 and Ab 4 were fully human anti-MERS-CoV spike protein antibodies. RNA was extracted from the mouse lungs and analyzed by quantitative PCR as described above. For example, qPCR data was analyzed using the delta Ct method, with an uninfected control set to 1. Percent MERS-CoV RNA detected was expressed relative to levels of RNA detected in infected mice treated with isotype-matched control antibodies (FIGS. 11A-B). Also, viral titers from mouse lungs were determined as described above.

[0211] Both antibodies significantly decreased MERS-CoV specific RNA levels in the lungs by over 2 logs at the 200 µg per mouse dose, compared to the isotype-matched control antibody (FIGS. **11**A-B). Ab 2 was more effective at

reducing MERS-CoV RNA levels at the 20 μ g dose compared to Ab 4 at the same dose. The 2 μ g dosing of either antibody was ineffective at reducing viral RNA levels compared to isotype control treated mice. When MERS-CoV titer was analyzed in the lungs (FIG. 11C), both the 200 μ g and 20 μ g dose of Ab 2 reduced virus levels to near the level of detection in the assay (2×10³ pfu/ml). Ab 4 was equally efficient at the 200 μ g dose as Ab 2, while the 20 μ g and 2 μ g doses displayed a dose dependent inhibition of viral inhibition. These data show that anti-MERS-CoV antibodies, e.g., Ab 2 and Ab 4, effectively blocked MERS-CoV infection in vivo.

[0212] Histological analysis was also performed at 4 days post-infection on lungs from mice treated at 24 hours pre-infection with Ab 2, Ab 4, or hIgG 1 isotype control antibody (FIG. **12**A). For example, the degrees of interstitial, peribronchiolar, and perivascular inflammation were scored from 0 to 5. Other histologic features, such as the presence of bronchiolar epithelial and alveolar damage, pleural changes and the extent of peribronchovascular inflammation, were also analyzed. An overall inflammatory score for each mouse was averaged for each experimental group, and the scores were presented as average scores of all mice in each group and time point (FIG. **12**B).

[0213] Lungs from mice pre-treated with hIgG 1 isotype control mice displayed significant lung pathology with increased interstitial inflammation, perivascular cuffing, and thickening of alveolar septa. Mice treated with 200 µg of either Ab 2 or Ab 4 had reduced inflammation with minimal foci of inflammatory cells in the interstitium, minor bronchiolar cuffing, and less alveolar wall thickening. In mice pre-treated with 20 µg of Ab 2 and Ab 4, there were moderate levels of perivascular cuffing and interstitial inflammation compared to the higher dose antibody group. The 2 µg antibody pre-treated group had similar pathology to the hIgG1 isotype control, displaying significant interstitial inflammation and predominant peri-vascular inflammation. Blinded histological scoring demonstrated reduced inflammation scores for treated mice (FIG. 12B). These findings demonstrate that anti-MERS-CoV antibodies, such as Ab 2 and Ab 4, confer a dose-dependent reduction in lung pathology following MERS-CoV infection, corroborating viral RNA levels and virus titers in the mice.

[0214] Thus, anti-MERS-CoV antibodies, such as Ab 2 and Ab 4, were effective in an in vivo model of MERS-CoV infection—the antibodies blocked MERS-CoV infection and disease in vivo when injected before infection, e.g., 1 day before infection.

Example 6

Antibody Treatment of Humanized DPP4 Mice that have been Infected with MERS-CoV

[0215] To determine the therapeutic effect (e.g., ability to inhibit MERS-CoV replication and lung pathology after infection) of anti-MERS-CoV antibodies (e.g., Ab 2 or Ab 4), humanized DPP4 mice were infected with MERS-CoV. At 24 hours post-infection, the mice were injected i.p. with either 500 µg of hIgG1 isotype control or Ab 2 at 500 µg or 200 µg. At 4 days post-infection, mice were euthanized and mouse lungs analyzed for viral RNA, virus titer, and lung pathology. Both the 500 µg and 200 µg doses of Ab 2 reduced viral RNA levels by about 10 fold in the lungs of mice compared to control antibody treated mice (FIGS.

13A-B). Lung titers of the same mice demonstrated significant reduction in viral levels in the lungs, with a greater than 2 log reduction at day 4 post-infection (FIG. **13**C). These data demonstrate that after infection, e.g., 24 hours post-infection, an anti-MERS-CoV antibody (e.g., Ab 2) significantly inhibited viral replication.

[0216] Histological analysis was also performed on mice treated 24 hours post-infection with hIgG1 control antibody, 500 µg Ab 2, or 200 µg Ab 2 (FIGS. **14**A-B). Mice treated with control antibody displayed similar pathology to the

controls in Examples 2 and 5, with significant interstitial inflammation, peri-vascular cuffing, and thickening of alveolar septa. Mice treated with either 200 μ g or 500 μ g of Ab 2 had minimal interstitial inflammation with reduced and only focal peri-vascular inflammation throughout the lungs. Blinded histological scoring demonstrated reduced inflammation scores for treated mice (FIG. **14**B). The data demonstrate that therapeutic doses of anti-MERS-CoV antibodies (e.g., Ab 2) reduced MERS-CoV induced lung pathology even when given after infection, e.g., 24 hours post-infection.

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ctagtaacta taacggtcct aaggtagcga gctagccagc atagctctcc atagcttatt	180
taagaccaca tttgttctca ttatctcaaa agtgcactgt taagatgaag atcttaataa	240
tgttgcattg agacatttca ggctgctttc tccagtttta cacctgcaat cctaactaag	300

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19

318

Jacjoorjoo oorajaat															
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Leu	Val	Thr	Ile 20	Ile	Thr	Val	Pro	Val 25	Val	Leu	Leu	Asn	Lys 30	Gly	Thr
Asp	Asp	Ala 35	Thr	Ala	Asp	Ser	Arg 40	Lys	Thr	Tyr	Thr	Leu 45	Thr	Asp	Tyr
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Asp 65	His	Glu	Tyr	Leu	Tyr 70	Lys	Gln	Glu	Asn	Asn 75	Ile	Leu	Val	Phe	Asn 80
Ala	Glu	Tyr	Gly	Asn 85	Ser	Ser	Val	Phe	Leu 90	Glu	Asn	Ser	Thr	Phe 95	Asp
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Thr	Ala 130	Ser	Tyr	Asp	Ile	Tyr 135	Asp	Leu	Asn	Lys	Arg 140	Gln	Leu	Ile	Thr
Glu 145	Glu	Arg	Ile	Pro	Asn 150	Asn	Thr	Gln	Trp	Val 155	Thr	Trp	Ser	Pro	Val 160
Gly	His	Lys	Leu	Ala 165	Tyr	Val	Trp	Asn	Asn 170	Asp	Ile	Tyr	Val	Lys 175	Ile
Glu	Pro	Asn	Leu 180	Pro	Ser	Tyr	Arg	Ile 185	Thr	Trp	Thr	Gly	Lys 190	Glu	Asp
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Asn	Tyr	Ser	Val	Met 325	Asp	Ile	Cys	Asp	Tyr 330	Asp	Glu	Ser	Ser	Gly 335	Arg
Trp	Asn	Сув	Leu 340	Val	Ala	Arg	Gln	His 345	Ile	Glu	Met	Ser	Thr 350	Thr	Gly

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Trp	Val	Gly 355	Arg	Phe	Arg	Pro	Ser 360	Glu	Pro	His	Phe	Thr 365	Leu	Asp	Gly
Asn	Ser 370	Phe	Tyr	Lys	Ile	Ile 375	Ser	Asn	Glu	Glu	Gly 380	Tyr	Arg	His	Ile
Cys 385	Tyr	Phe	Gln	Ile	Asp 390	Гла	Lys	Asp	Суз	Thr 395	Phe	Ile	Thr	Lys	Gly 400
Thr	Trp	Glu	Val	Ile 405	Gly	Ile	Glu	Ala	Leu 410	Thr	Ser	Asp	Tyr	Leu 415	Tyr
Tyr	Ile	Ser	Asn 420	Glu	Tyr	Гла	Gly	Met 425	Pro	Gly	Gly	Arg	Asn 430	Leu	Tyr
ГЛЗ	Ile	Gln 435	Leu	Ser	Asp	Tyr	Thr 440	Lys	Val	Thr	Суз	Leu 445	Ser	Cys	Glu
Leu	Asn 450	Pro	Glu	Arg	Сүз	Gln 455	Tyr	Tyr	Ser	Val	Ser 460	Phe	Ser	Lys	Glu
Ala 465	Lys	Tyr	Tyr	Gln	Leu 470	Arg	Суз	Ser	Gly	Pro 475	Gly	Leu	Pro	Leu	Tyr 480
Thr	Leu	His	Ser	Ser 485	Val	Asn	Asp	Lys	Gly 490	Leu	Arg	Val	Leu	Glu 495	Asp
Asn	Ser	Ala	Leu 500	Asp	ГЛа	Met	Leu	Gln 505	Asn	Val	Gln	Met	Pro 510	Ser	Lya
Lys	Leu	Asp 515	Phe	Ile	Ile	Leu	Asn 520	Glu	Thr	Lys	Phe	Trp 525	Tyr	Gln	Met
Ile	Leu 530	Pro	Pro	His	Phe	Asp 535	Lys	Ser	Lys	Lys	Tyr 540	Pro	Leu	Leu	Leu
Asp 545	Val	Tyr	Ala	Gly	Pro 550	Суз	Ser	Gln	Lys	Ala 555	Asp	Thr	Val	Phe	Arg 560
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Ser	Phe	Asp	Gly 580	Arg	Gly	Ser	Gly	Tyr 585	Gln	Gly	Asp	Lys	Ile 590	Met	His
Ala	Ile	Asn 595	Arg	Arg	Leu	Gly	Thr 600	Phe	Glu	Val	Glu	Asp 605	Gln	Ile	Glu
Ala	Ala 610	Arg	Gln	Phe	Ser	Lys 615	Met	Gly	Phe	Val	Asp 620	Asn	Lys	Arg	Ile
Ala 625	Ile	Trp	Gly	Trp	Ser 630	Tyr	Gly	Gly	Tyr	Val 635	Thr	Ser	Met	Val	Leu 640
Gly	Ser	Gly	Ser	Gly 645	Val	Phe	Lys	Суз	Gly 650	Ile	Ala	Val	Ala	Pro 655	Val
Ser	Arg	Trp	Glu 660	Tyr	Tyr	Asp	Ser	Val 665	Tyr	Thr	Glu	Arg	Tyr 670	Met	Gly
Leu	Pro	Thr 675	Pro	Glu	Asp	Asn	Leu 680	Asp	His	Tyr	Arg	Asn 685	Ser	Thr	Val
Met	Ser 690	Arg	Ala	Glu	Asn	Phe 695	Lys	Gln	Val	Glu	Tyr 700	Leu	Leu	Ile	His
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	Ala	Leu	Val	Asp 725		Gly	Val	Asp	Phe 730		Ala	Met	Trp	Tyr 735	
Asp	Glu	Asp			Ile	Ala	Ser			Ala	His	Gln	His		Tyr
Thr	His	Met	740 Ser	His	Phe	Ile	Lys	745 Gln	Суз	Phe	Ser	Leu	750 Pro		

21

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Met 1	Lys	Thr	Pro	5	гда	vai	Leu	Leu	GIY 10	Leu	Leu	GIY	AIA	AIA 15	Ala
Leu	Val	Thr	Ile 20	Ile	Thr	Val	Pro	Val 25	Val	Leu	Leu	Asn	Lys 30	Gly	Thr
Asp	Asp	Ala 35	Thr	Ala	Asp	Ser	Arg 40	Гла	Thr	Tyr	Thr	Leu 45	Thr	Asp	Tyr
Leu	Lys 50	Asn	Thr	Tyr	Arg	Leu 55	Lys	Leu	Tyr	Ser	Leu 60	Arg	Trp	Ile	Ser
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Ala	Glu	Tyr	Gly	Asn 85	Ser	Ser	Val	Phe	Leu 90	Glu	Asn	Ser	Thr	Phe 95	Asp
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Phe	Ile	Leu 115	Leu	Glu	Tyr	Asn	Tyr 120	Val	Lys	Gln	Trp	Arg 125	His	Ser	Tyr
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Gly 705	Thr	Ala	Asp	Asp	Asn 710	Val	His	Phe	Gln	Gln 715	Ser	Ala	Gln	Ile	Ser 720
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Thr	His	Met 755	Ser	His	Phe	Ile	Lys 760	Gln	Cys	Phe	Ser	Leu 765	Pro		

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Ala A	Ala	Ala 35	Asp	Ser	Arg	Arg	Thr 40	Tyr	Ser	Leu	Ala	Asp 45	Tyr	Leu	Lys
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His (Gly	Asn	Ser	Ser 85	Ile	Phe	Leu	Glu	Asn 90	Ser	Thr	Phe	Glu	Ser 95	Phe
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Tyr A	Asp 130	Val	Asn	Lys	Arg	Gln 135	Leu	Ile	Thr	Glu	Glu 140	Lys	Ile	Pro	Asn
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Val :	Trp	Lys	Asn	Asp 165	Ile	Tyr	Val	Lys	Val 170	Glu	Pro	His	Leu	Pro 175	Ser
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Thr (225	-				230		-			235		-			240
Gln 7				245					250					255	
Asn I			260	-				265			-		270		
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Lys 385	Lys	Asp	Суз	Thr	Phe 390	Ile	Thr	Гла	Gly	Ala 395	Trp	Glu	Val	Ile	Ser 400
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His	Thr	Asn 435	Val	ГЛа	Суа	Leu	Ser 440	Суз	Asp	Leu	Asn	Pro 445	Glu	Arg	Суз
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Leu	Gln	Gln 755	Суз	Phe	Ser	Leu	His 760								

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	2 > T 3 > OH			Homo	o saj	bien	5								
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Leu	Lys	Acn	Thr	Tur	۸ra	Leu	Lare	Leu	Tur	Cor	Lau	۸ra	Trn	TIA	Cor
Leu	50	mon		1 9 1	mg	55	Цур	ncu	1 y 1	DCI	60	тg	115	110	bei
Asp	His	Glu	Tvr	Leu	Tvr	Lvs	Gln	Glu	Asn	Asn	Ile	Leu	Val	Phe	Asn
65			-1-		70	-1				75					80
Ala	Glu	Tyr	Gly	Asn	Ser	Ser	Val	Phe	Leu	Glu	Asn	Ser	Thr	Phe	Asp
		-	-	85					90					95	-
Glu	Phe	Gly	His	Ser	Ile	Asn	Asp	Tyr	Ser	Ile	Ser	Pro	Asp	Gly	Gln
			100					105					110		
Phe	Ile	Leu	Leu	Glu	Tyr	Asn		Val	Lys	Gln	Trp		His	Ser	Tyr
		115					120					125			
Thr	Ala	Ser	Tyr	Asp	Ile		Asp	Leu	Asn	Lys		Gln	Leu	Ile	Thr
	130					135					140				
Glu 145	Glu	Arg	Ile	Pro	Asn 150	Asn	Thr	Gln	Trp	Val 155	Thr	Trp	Ser	Pro	Val 160
140					100					100					100
Gly	His	Lys	Leu	Ala 165	Tyr	Val	Trp	Asn	Asn 170	Asp	Ile	Tyr	Val	Lys 175	Ile
~ 1	_				~	_				_	-	<i>a</i> 1		~ 1	_
GIu	Pro	Asn	Leu 180	Pro	Ser	Tyr	Arg	Ile 185	Thr	Trp	Thr	GIY	Lys 190	GIu	Asp
T10	T10	T 1 1 1 1	Aan	Clw	T10	Thr	^a an	Trn	Vol	Ture	C1.,	C1.1	C1.1	Vol	Dho
тте	Ile	191 195	ASII	GIY	тте	1111	ASP 200	пр	Val	тут	GIU	205	Gru	Val	Pile
Ser	Ala	Tvr	Ser	Ala	Leu	Tro	Trn	Ser	Pro	Asn	Glv	Thr	Phe	Leu	Ala
001	210	- / -	001		Lou	215		501	110		220		1110	Lou	1124
Tvr	Ala	Gln	Phe	Asn	Asp	Thr	Glu	Val	Pro	Leu	Ile	Glu	Tvr	Ser	Phe
225					230					235			-1-		240
Tyr	Ser	Asp	Glu	Ser	Leu	Gln	Tyr	Pro	Lys	Thr	Val	Arg	Val	Pro	Tyr
-		-		245			•		250					255	-
Pro	Lys	Ala	Gly	Ala	Val	Asn	Pro	Thr	Val	Lys	Phe	Phe	Val	Val	Asn
			260					265					270		
Thr	Asp	Ser	Leu	Ser	Ser	Val	Thr	Asn	Ala	Thr	Ser	Ile	Gln	Ile	Thr
		275					280					285			
Ala	Pro	Ala	Ser	Met	Leu		Gly	Asp	His	Tyr		Cys	Asp	Val	Thr
	290					295					300				
	Ala	Thr	Gln	Glu		Ile	Ser	Leu	Gln		Leu	Arg	Arg	Ile	
305					310					315					320
Asn	Tyr	Ser	Val		Asp	Ile	Cys	Asp	-	Asp	Glu	Ser	Ser	-	Arg
				325					330					335	
Trp	Asn	Суз		Val	Ala	Arg	Gln		Ile	Glu	Met	Ser		Thr	Gly
			340					345					350		
Trp	Val	-	Arg	Phe	Arg	Pro		Glu	Pro	His	Phe		Leu	Asp	Gly
		355					360					365			
Asn	Ser	Phe	Tyr	Lys	Ile	Ile	Ser	Asn	Glu	Glu	Gly	Tyr	Arg	His	Ile

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	370					375					380				
Cys 385	Tyr	Phe	Gln	Ile	Asp 390	Lys	Lys	Asp	Cys	Thr 395	Phe	Ile	Thr	Lys	Gly 400
Thr	Trp	Glu	Val	Ile 405	Gly	Ile	Glu	Ala	Leu 410	Thr	Ser	Asp	Tyr	Leu 415	Tyr
Tyr	Ile	Ser	Asn 420	Glu	Tyr	Lys	Gly	Met 425	Pro	Gly	Gly	Arg	Asn 430	Leu	Tyr
Lys	Ile	Gln 435	Leu	Ser	Asp	Tyr	Thr 440	Lys	Val	Thr	СЛа	Leu 445	Ser	Суз	Glu
Leu	Asn 450	Pro	Glu	Arg	Сүз	Gln 455	Tyr	Tyr	Ser	Val	Ser 460	Phe	Ser	Lys	Glu
Ala 465	Lys	Tyr	Tyr	Gln	Leu 470	Arg	Суз	Ser	Gly	Pro 475	Gly	Leu	Pro	Leu	Tyr 480
Thr	Leu	His	Ser	Ser 485	Val	Asn	Asp	Lys	Gly 490	Leu	Arg	Val	Leu	Glu 495	Asp
			500	-	-			Gln 505					510		-
Lys	Leu	Asp 515	Phe	Ile	Ile	Leu	Asn 520	Glu	Thr	ГЛа	Phe	Trp 525	Tyr	Gln	Met
Ile	Leu 530	Pro	Pro	His	Phe	Asp 535	Lys	Ser	Lys	Lys	Tyr 540	Pro	Leu	Leu	Leu
Asp 545	Val	Tyr	Ala	Gly	Pro 550	Сүз	Ser	Gln	Lys	Ala 555	Asp	Thr	Val	Phe	Arg 560
Leu	Asn	Trp	Ala	Thr 565	Tyr	Leu	Ala	Ser	Thr 570	Glu	Asn	Ile	Ile	Val 575	Ala
Ser	Phe	Asp	Gly 580	Arg	Gly	Ser	Gly	Tyr 585	Gln	Gly	Asp	Lys	Ile 590	Met	His
Ala	Ile	Asn 595	Arg	Arg	Leu	Gly	Thr 600	Phe	Glu	Val	Glu	Asp 605	Gln	Ile	Glu
Ala	Ala 610	Arg	Gln	Phe	Ser	Lys 615	Met	Gly	Phe	Val	Asp 620	Asn	Lys	Arg	Ile
Ala 625	Ile	Trp	Gly	Trp	Ser 630	Tyr	Gly	Gly	Tyr	Val 635	Thr	Ser	Met	Val	Leu 640
Gly	Ser	Gly	Ser	Gly 645	Val	Phe	Lys	Суз	Gly 650	Ile	Ala	Val	Ala	Pro 655	Val
Ser	Arg	Trp	Glu 660	Tyr	Tyr	Asp	Ser	Val 665	Tyr	Thr	Glu	Arg	Tyr 670	Met	Gly
Leu	Pro	Thr 675	Pro	Glu	Asp	Asn	Leu 680	Asp	His	Tyr	Arg	Asn 685	Ser	Thr	Val
Met	Ser 690	Arg	Ala	Glu	Asn	Phe 695	ГÀа	Gln	Val	Glu	Tyr 700	Leu	Leu	Ile	His
Gly 705	Thr	Ala	Asp	Asp	Asn 710	Val	His	Phe	Gln	Gln 715	Ser	Ala	Gln	Ile	Ser 720
Lys	Ala	Leu	Val	Asp 725	Val	Gly	Val	Asp	Phe 730	Gln	Ala	Met	Trp	Tyr 735	Thr
Asp	Glu	Asp	His 740	Gly	Ile	Ala	Ser	Ser 745	Thr	Ala	His	Gln	His 750	Ile	Tyr
Thr	His	Met 755	Ser	His	Phe	Ile	Lys 760	Gln	Суз	Phe	Ser	Leu 765	Pro		

1. A humanized DPP4 rodent embryo comprising a humanization of an endogenous rodent Dpp4 gene, wherein the humanization comprises a replacement at the endogenous rodent Dpp4 locus of at least one exon of the endogenous rodent Dpp4 gene with a nucleic acid sequence encoding at least one exon of a human DPP4 gene to form a modified DPP4 gene, wherein expression of the modified DPP4 gene is under control of rodent regulatory elements at the endogenous rodent Dpp4 locus, and wherein the rodent is a mouse or a rat.

2. The embryo of claim **1**, wherein the human DPP4 gene encoding a human or humanized DPP4 protein comprises exon 2 through exon 26 of the human DPP4 gene.

3. The embryo of claim **2**, wherein the humanized DPP4 protein comprises the extracellular domain of the human DPP4 protein.

4. The embryo of claim **3**, wherein the humanized DPP4 protein comprises the transmembrane domain and cytoplasmic domain of the rodent Dpp4 protein.

5. The embryo of claim **1**, wherein the rodent is a mouse that is incapable of expressing a mouse Dpp4 protein.

6. The embryo of claim 1, wherein the humanization of the endogenous rodent Dpp4 gene is heterozygous.

7. The embryo of claim 1, wherein the humanization of the endogenous rodent Dpp4 gene is homozygous.

8. The embryo of claim **1**, wherein the rodent is a mouse and wherein the mouse is: (a) a C57BL strain selected from the group consisting of C57BL/A, C57BL/An, C57BL/ GrFa, C57BL/KaLwN, C57BL/6, C57BL/6J, C57BL/6ByJ, C57BL/6NJ, C57BL/10, C57BL/10ScSn, C57BL/10Cr, and C57BL/01a; (b) a 129 strain selected from the group consisting of 129P1, 129P2, 129P3, 129X1, 129S1 (e.g., 129S1/ SV, 129S1/SvIm), 129S2, 129S4, 129S5, 129S9/SvEvH, 129S6 (129/SvEvTac), 129S7, 129S8, 129T1, and 129T2; (c) a mixture of a 129 strain and a C57BL/6 strain; (d) a mix of 129 strains; or (e) a mix of BL/6 strains.

9. A rodent host embryo comprising a donor cell that comprises a humanization of an endogenous rodent Dpp4 gene, wherein the humanization comprises a replacement at the endogenous rodent Dpp4 locus of at least one exon of the endogenous rodent Dpp4 gene with a nucleic acid sequence encoding at least one exon of a human DPP4 gene to form a modified DPP4 gene, wherein expression of the modified DPP4 gene is under control of rodent regulatory elements at the endogenous rodent Dpp4 locus, and wherein the rodent is a mouse or a rat.

10. The host embryo of claim **9**, wherein the human DPP4 gene encoding a human or humanized DPP4 protein comprises exon 2 through exon 26 of the human DPP4 gene.

11. The host embryo of claim **10**, wherein the humanized DPP4 protein comprises the extracellular domain of the human DPP4 protein.

12. The host embryo of claim **11**, wherein the humanized DPP4 protein comprises the transmembrane domain and cytoplasmic domain of the rodent Dpp4 protein.

13. The host embryo of claim **9**, wherein the rodent is a mouse that is incapable of expressing a mouse Dpp4 protein.

14. The host embryo of claim 9, wherein the humanization of the endogenous rodent Dpp4 gene is heterozygous.

15. The host embryo of claim **9**, wherein the humanization of the endogenous rodent Dpp4 gene is homozygous.

16. A rodent egg comprising an ectopic non-human animal chromosome, wherein the ectopic non-human animal chromosome comprises a humanization of a endogenous rodent Dpp4 gene, wherein the humanization comprises a replacement at the endogenous rodent Dpp4 locus of at least one exon of the endogenous rodent Dpp4 gene with a nucleic acid sequence encoding at least one exon of a human DPP4 gene to form a modified DPP4 gene, wherein expression of the modified DPP4 gene is under control of rodent regulatory elements at the endogenous rodent Dpp4 locus, and wherein the rodent is a mouse or a rat.

17. The rodent egg of claim **16**, wherein the human DPP4 gene encoding a human or humanized DPP4 protein comprises exon 2 through exon 26 of the human DPP4 gene.

18. The rodent egg of claim **17**, wherein the humanized DPP4 protein comprises the extracellular domain of the human DPP4 protein.

19. The rodent egg of claim **18**, wherein the humanized DPP4 protein comprises the transmembrane domain and cytoplasmic domain of the rodent Dpp4 protein.

20. The rodent egg of claim **16**, wherein the rodent is a mouse that is incapable of expressing a mouse Dpp4 protein.

21. The rodent egg of claim **16**, wherein the humanization of the endogenous rodent Dpp4 gene is heterozygous.

22. The rodent egg of claim **16**, wherein the humanization of the endogenous rodent Dpp4 gene is homozygous.

23. A rodent comprising a germ-line modification, wherein said germ line modification comprises a humanization of a endogenous rodent Dpp4 gene, wherein the humanization comprises a replacement at the endogenous rodent Dpp4 locus of at least one exon of the endogenous rodent Dpp4 gene with a nucleic acid sequence encoding at least one exon of a human DPP4 gene to form a modified DPP4 gene, wherein expression of the modified DPP4 gene is under control of rodent regulatory elements at the endogenous rodent Dpp4 locus, wherein the rodent is a mouse or a rat.

24. The rodent of claim **23**, wherein the human DPP4 gene encoding a human or humanized DPP4 protein comprises exon 2 through exon 26 of the human DPP4 gene.

25. The rodent of claim **24**, wherein the humanized DPP4 protein comprises the extracellular domain of the human DPP4 protein.

26. The rodent of claim **25**, wherein the humanized DPP4 protein comprises the transmembrane domain and cytoplasmic domain of the rodent Dpp4 protein.

27. The rodent of claim **23**, wherein the rodent is a mouse that is incapable of expressing a mouse Dpp4 protein.

28. The rodent of claim **23**, wherein the humanization of the endogenous rodent Dpp4 gene is heterozygous.

29. The rodent of claim **23**, wherein the humanization of the endogenous rodent Dpp4 gene is homozygous.

30. The rodent of claim **23**, wherein the rodent is a mouse and wherein the mouse is: (a) a C57BL strain selected from the group consisting of C57BL/A, C57BL/An, C57BL/ GrFa, C57BL/KaLwN, C57BL/6, C57BL/6J, C57BL/6ByJ, C57BL/6NJ, C57BL/10, C57BL/10ScSn, C57BL/10Cr, and C57BL/01a; (b) a 129 strain selected from the group consisting of a strain that is 129P1, 129P2, 129P3, 129X1, 129S1 (e.g., 129S1/SV, 129S1/SvIm), 129S2, 129S4, 129S5, 129S9/SvEvH, 129S6 (129/SvEvTac), 129S7, 129S8, 129T1, and 129T2; (c) a mixture of a 129 strain and a C57BL/6 strain; (d) a mix of 129 strains; or (e) a mix of BL/6 strains.

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