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(54) **GENETICALLY ENCODED BIOSENSORS**

Publication Classification

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C07K 14/195 (2006.01)
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C07K 14/435 (2006.01)
- (52) **U.S. Cl.**
CPC *G01N 33/557* (2013.01); *G01N 33/68* (2013.01); *G01N 33/582* (2013.01); *C07K 14/195* (2013.01); *C07K 14/245* (2013.01); *G01N 2400/00* (2013.01); *C07K 14/43595* (2013.01); *G01N 2458/00* (2013.01); *C07K 2319/20* (2013.01); *C07K 2319/24* (2013.01); *C07K 2319/60* (2013.01); *G01N 33/6812* (2013.01)

Related U.S. Application Data

- (60) Division of application No. 16/902,160, filed on Jun. 15, 2020, now Pat. No. 11,162,942, which is a continuation of application No. 16/002,697, filed on Jun. 7, 2018, now Pat. No. 10,684,282, which is a continuation-in-part of application No. 15/904,574, filed on Feb. 26, 2018, now Pat. No. 10,060,920, which is a division of application No. 15/664,326, filed on Jul. 31, 2017, now Pat. No. 9,939,437, which is a division of application No. 14/350,199, filed on Nov. 18, 2014, now Pat. No. 9,719,992, filed as application No. PCT/US2012/059219 on Oct. 8, 2012.
- (60) Provisional application No. 61/544,867, filed on Oct. 7, 2011.

(57) **ABSTRACT**

The present disclosure provides, inter alia, genetically encoded recombinant peptide biosensors comprising analyte-binding framework portions and signaling portions, wherein the signaling portions are present within the framework portions at sites or amino acid positions that undergo a conformational change upon interaction of the framework portion with an analyte.

Specification includes a Sequence Listing.

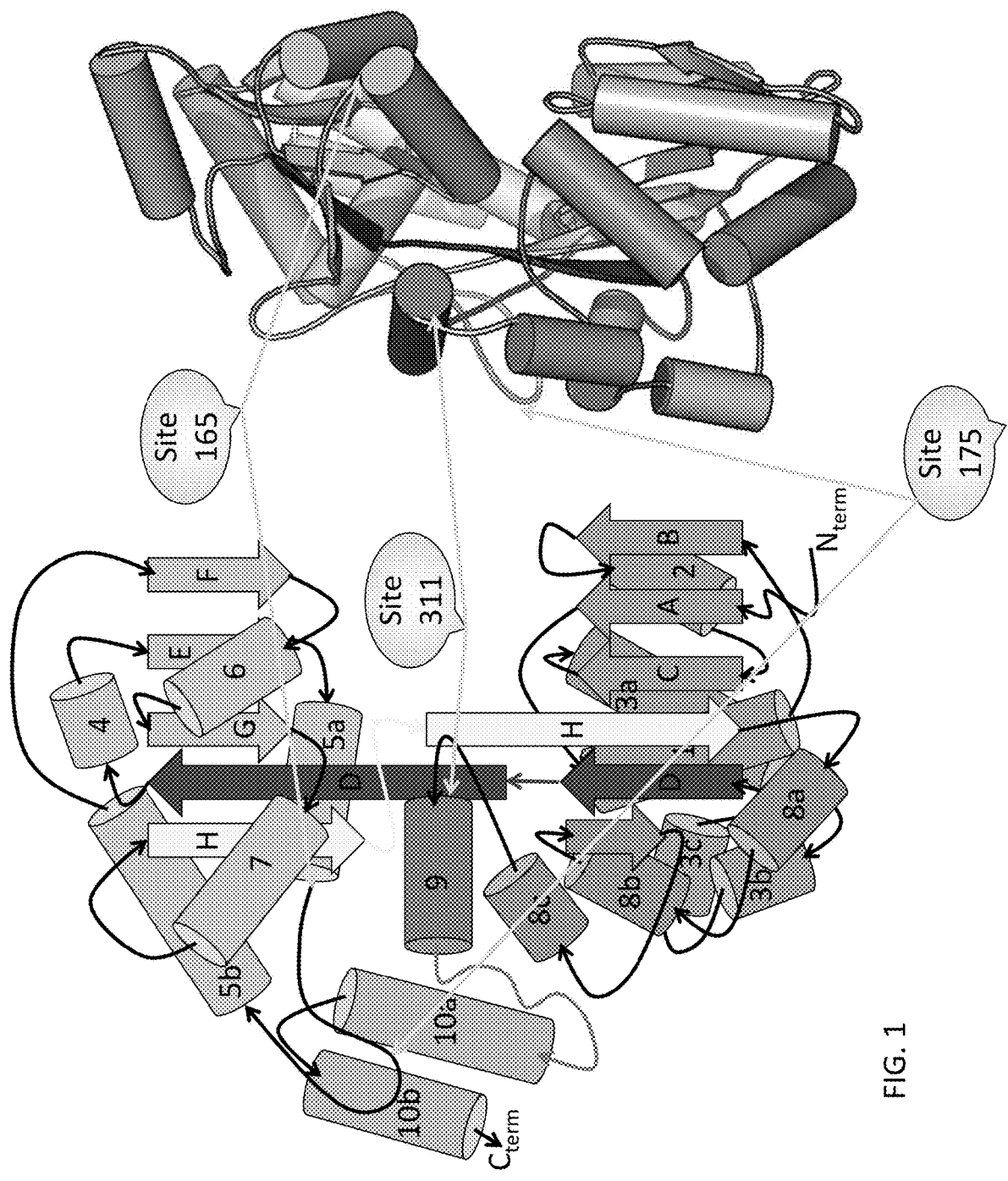


FIG. 1

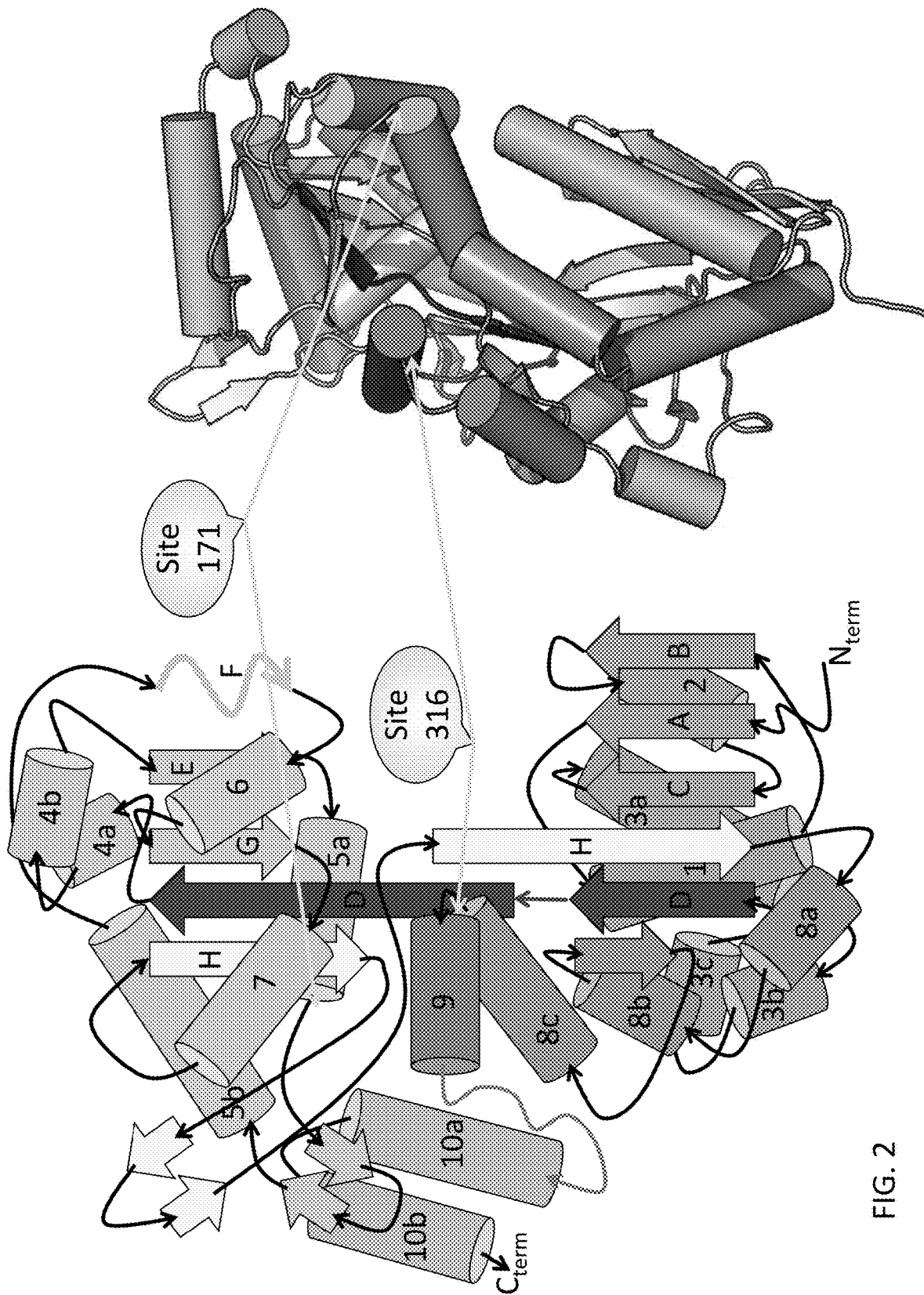


FIG. 2

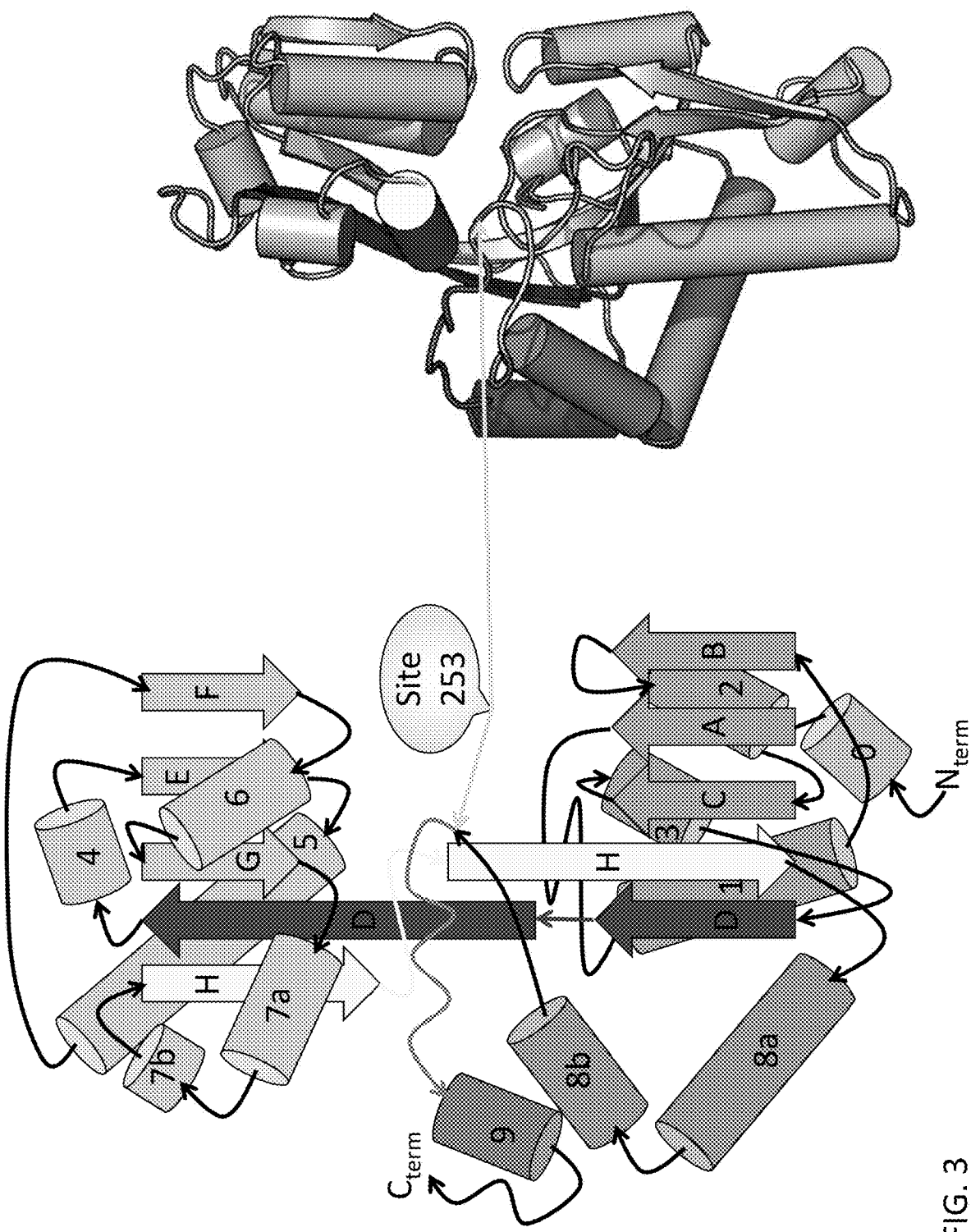


FIG. 3

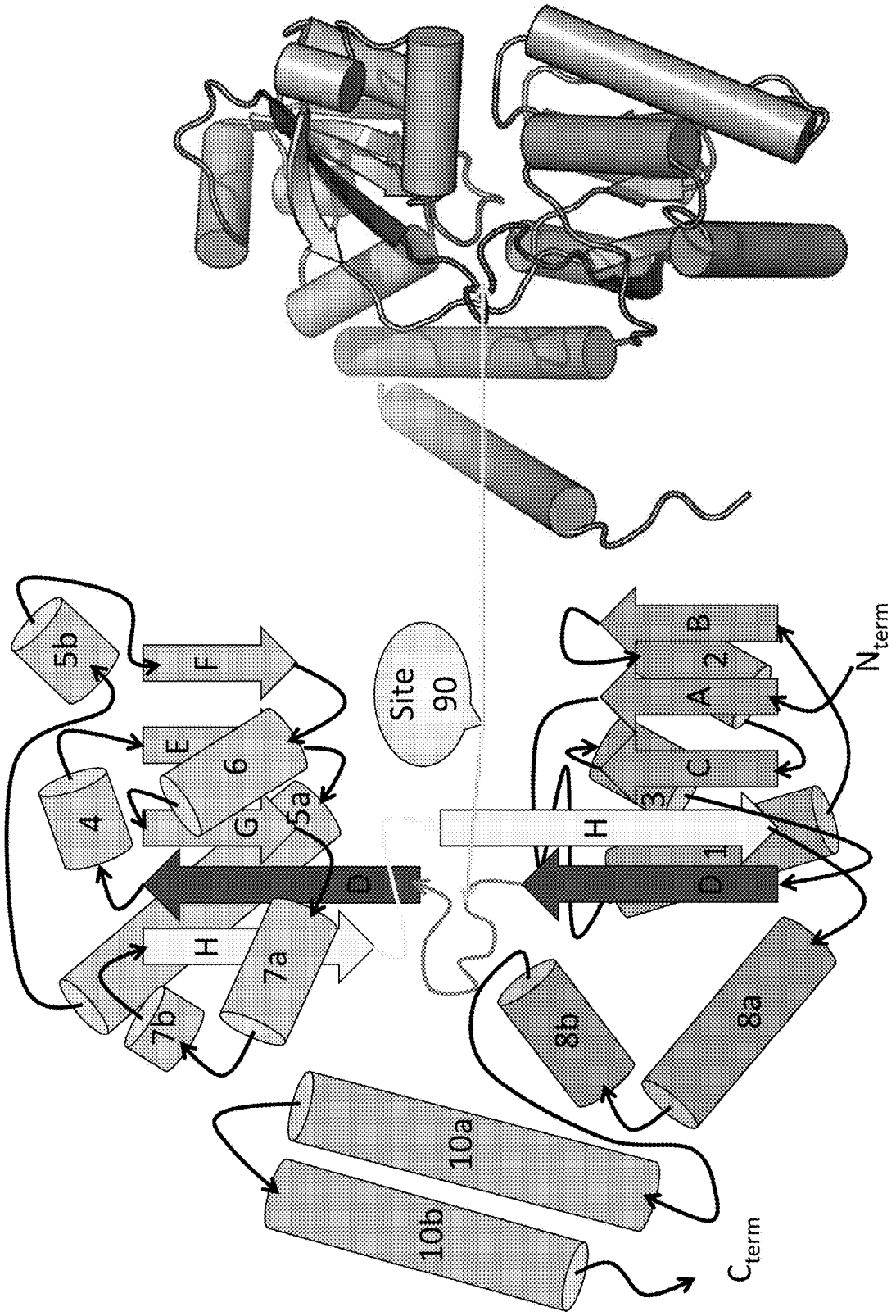


FIG. 4

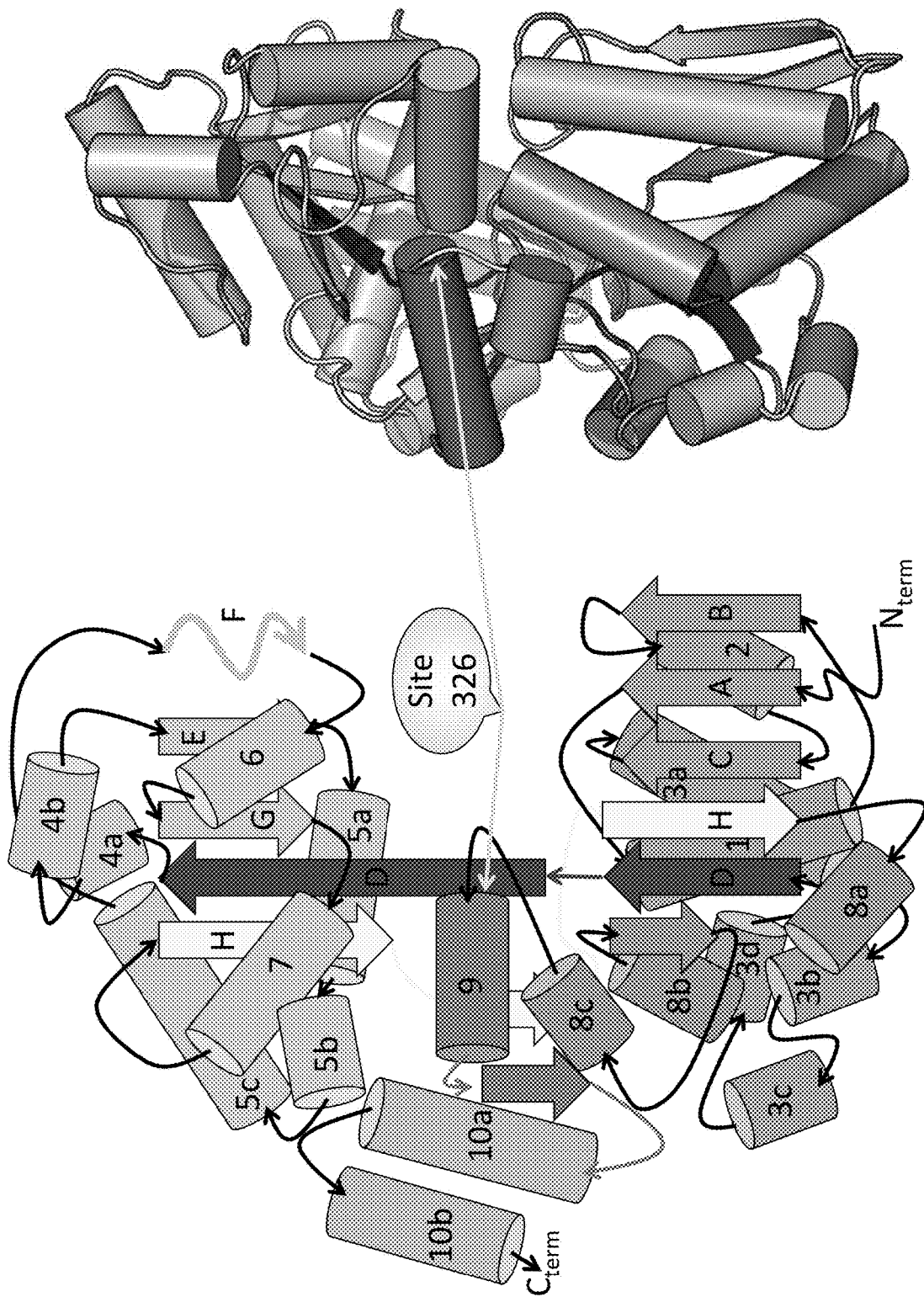


FIG. 5

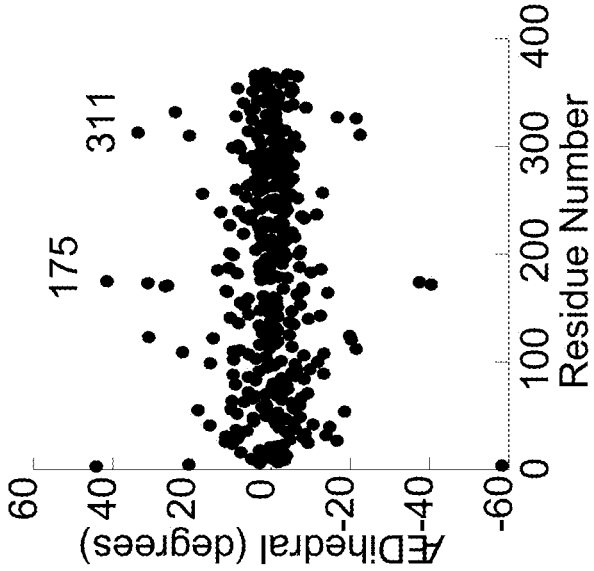


FIG. 6B

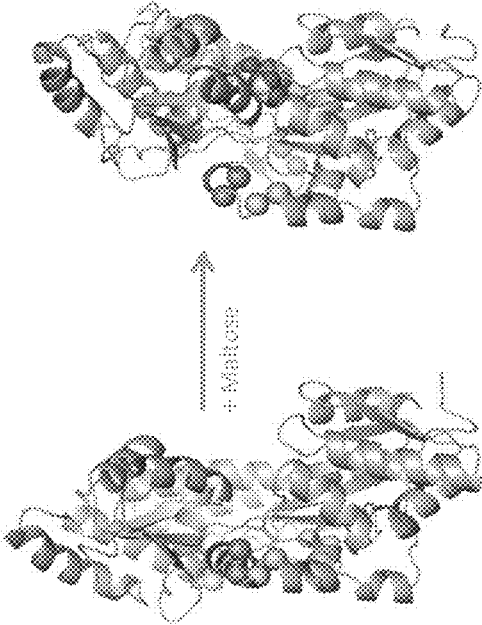


FIG. 6A

c
 N MFGSHHHHGMASNTGCGQWERLYDDEDEKRWGSKIEEGKIWIWINGDKYVWELAEVCKKFEKDTGLKVTVEHPDKLE 80
 c
 c
 c MBP 146
 N ERFPQAAAGDGEDIIIFWAHDRFGYAQSGLLAEITPDAFCDEKLYPFTWDAVYNGKLAAPYIAVEALSIIINRDLFPN 160
 c
 c MBP 145 cpGFP 147-238
 N FEKTWEELFALDKELKAKGKSALMENLQEPYFTWELLAADPFSYVFIIMADKQNGIKANFYIRHNIEDGCVQLAYHYQQ 240
 c
 c spGFP 147-238 Linker spGFP 146
 N NTFIIGCGPVLLPDMBYLSTQSKLSKOPNEKEDHWLLEVPVTAAGITLQMDELYKGTGSSKVKGEELFTGVVPLVELD 320
 c
 c spGFP 146
 N GEVNGHKFVSGEGEDATYGNLILKFKICTGKLFVPPFLVTLVTLTYGVOCGRYEDRHKQDFKSAPEGYIERTIF 400
 c
 c spGFP 146 MBP 166-379
 N FKDDGNYKTRAEVYEGDTLVNRLELKGIDFEDDGNILGHKLEINFGGYAYKENGKYLKDVQVDNAGAKAGLEFLVD 480
 c
 c MBP 166-379
 N LKMKRHMADHYSLAEAEVNGETAMTINGMANSNIDHSKVNYGVTVLPTKGGQPSKPFVGLSAGINASPNKELAK 560
 c
 c MBP 166-379
 N EPLENYLITDEGLEAVNEKPLGAVALKSYEESLVKDKDLAAUMENACKREINPNI PQNSAFVYAVFVAVINAASGEQIV 640
 c
 c MBP 166-379
 N DEELKDAQTNLTKGSRHHHHHG. 720
 c

FIG. 7B

c
 N MKGSSHHHHGMAKMTGGQQMRDLYDDEDKRWSKIEEGALVWINGDKYNGLAEYCKKSFENDGSIKVTVEREDKLE 80
 c
 c
 c
 N EKEPQVAATGDDGEDIIFWAHDEGGYAQSGLLAEITPKAQQDKLYPFTWDAVYNGRLIAYPIAVEALSILYNNDLLEN 160
 c
 c
 N EFKTWEELPALKDELAAGKKSALNENLQEYFTWPLIADFCSHNVFIMADKQNGIKANFKIRHNIEDEGGVQLATHYQQ 240
 c
 c
 N NTFICGGVLLPEMNYLSTQSKLSQDPAREKEDHWLLEFVTAAGTLDMDLXKGGTGGSWVSKRELFTGVVPIVELD 320
 c
 c
 N GAVNGKFSVSGEGEDATYGLIKLFICTGKLFVPELIVPLTYGVQCSFVPEDRNKQDFPKSANPEGTQERTIF 400
 c
 c
 N FRDDGNYKTRAEVKEGDTLVNKLKGLDFKEDGNLUGHKLEZVNGGYAFKYENSKYDKDYVVDNAGAKAGLITFVD 480
 c
 c
 N LLYKNRERADTDYSINAEAKNGETAMTINGPMAWSNLDISKVNYGVTVLPEKQPSKFFVGLSACINAAENKELAK 560
 c
 c
 N EFLERYLMTDEGLEAVNKDKPLGAVALESEYEEELVKDFKLAIDMENAQKGEINPRIFQNSAFWYAVRTAVINAASGRQIV 640
 c
 c
 N DEFLDAQFKLTKGSHHHHHHG.
 c
 c

FIG. 7C

○ **prSET Leader** **MBP 1175**
 N MRGSHRHHRHGKASMTGGQQMGRILYDDDRKDRGSKLEKGLVWINGDAGYGLAEVTKKFKEDTGIKVTVEPRKLE
 ○ ++++++
 ○ ++++++ 80
 ○ ++++++
 ○ **MBP 1175**
 N ERFQVARTGCPDIIETWADRFGYIAQSGLLAEIETDKAQOKLYFFTWDVYNGKLIAYPIAVEALSIIYKDKLEN
 ○ ++++++ 150
 ○ ++++++
 ○ **MBP 1175** **spGFP 147-238**
 N PPKTWEIIPALDSELRKAKGKSAIENFLOEYFPIWPLIADGGVAFKYEYNGSSHVYIADKRGNGIKANFKIRPNIEDGG
 ○ **MBP 1175** **spGFP 147-238**
 N PPKWEEIPALDKELKARGSAIENLOEYFTWELIADGGVAFKYEYENHLSHVYIADKQKNGIKANFKIRNIEDGG
 ○ ++++++ 240
 ○ ++++++
 ○ ++++++ 320
 ○ ++++++
 ○ **spGFP 1-146**
 N VPELVELDGVNKHKFSYSGEGGDATYGLHLKFLICHTGKLFVWPVTAVTLTYGVCFSRFDNMKQHDFFKSAPE
 ○ ++++++ 400
 ○ ++++++
 ○ **spGFP 1-146** **MBP 176-376**
 N GVIGENITIFPKDDGNKTRAEVKFEGDTLVNRRIELKGIHFKEDGNILGHKLEYNFNGSKYDIEDVGVDNAGAKGLTIV
 ○ ++++++ 480
 ○ ++++++
 ○ **MBP 176-376**
 N DLINKHBMAUTDYSIABAFNGETAMTINGPWASNLDTSKVXGYIVLFFTKQCFKPFVSVLSAGIIRASPNKELA
 ○ ++++++ 560
 ○ ++++++
 ○ **MBP 176-376**
 N KEFLENYLLDEGLEAVNKDKPLGAVALKSYEEELVQDPRIAAATNENAGKGEIMENIFQMSAFVYAVRTAVINAASGQET
 ○ ++++++ 640
 ○ ++++++
 ○ **MBP 176-376**
 N VDELDKDAZTRITKGSHHHHHG.
 ○ ++++++
 ○ ++++++

FIG. 8B

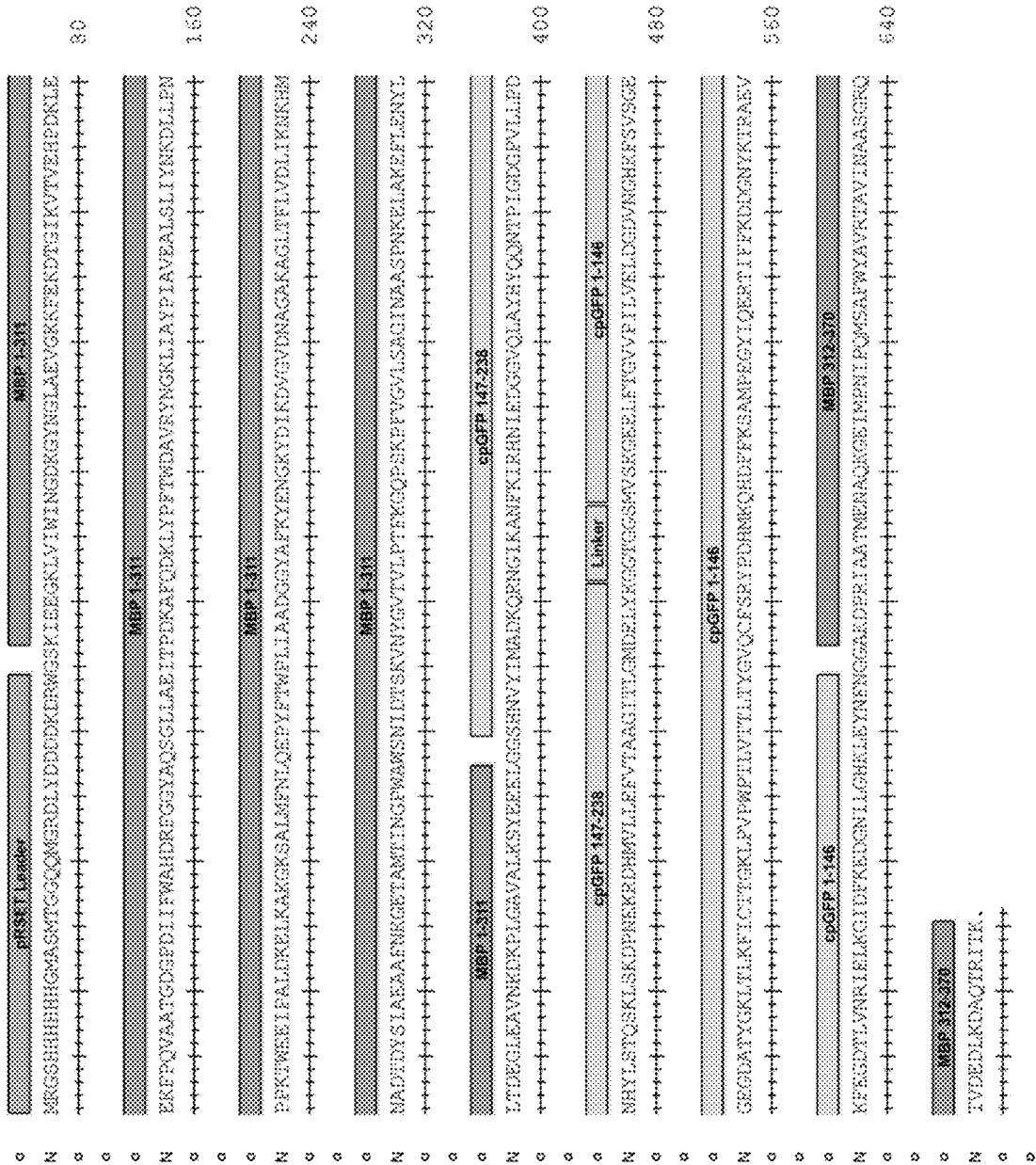


FIG. 9A

○ pRE1 Leader
 N MRGSHHHHGMASNIIGGQMGFDLYDDDKDRWGSRIIEGKIVIWINGKGYNGLAEVGKFKEDTGLNVTVEHPKLE 60
 ○ ++++++
 ○ MBP 1311
 N EKFPVAALGNGPRILFWAHDPRFGYAQSGLLAEIIFDKAFQDKLYFFTDVAVYMERKLIAYFIAVEALSIAVTKDLEEN 160
 ○ ++++++
 ○ MBP 1311
 N PPTWELIHALDKELAKGSALMENLQEPYFTWPLAAGGVAKYENGKDYIKUVGVNAGAKAGLTFVLVLLINKHM 240
 ○ ++++++
 ○ MBP 1311
 N NADTDYSIARAAFPKGETAMTINGPWMSNIDTSKVVYGVTVLTYKQPSKFFVGVLSAGLRAASPNEKELAEKLENYL 320
 ○ ++++++
 ○ MBP 1311
 N LTDEGLAVNKKPLGAVALKSYEEELGGSHNVVIMADKQRNGIKANFKIRHNIEDGGVQLAVHYQQNPIIGDGFVLLPD 400
 ○ ++++++
 ○ epGFP 147-238
 N MBYLSIQKLSKDFNEKREHNVLEFVTAAGITLGMDELYKGGTGGSMVSKGELFTGVVPIILVELDGVNHHKFSVSGE 480
 ○ ++++++
 ○ epGFP 146
 N GEGDATYGNLTLAEICTKLEVPWFVTLTYGQCFSRYPDRMKQHDDEKFSAMPEGYICERTIFFKDDGNVTRAEV 560
 ○ ++++++
 ○ epGFP 146
 N KFEGDTLVNRIELKGIIDFKEDGNILGHKLEYNFNPAKDFRIATMENAQNGEIMENI FQMSAFWYAVKFAVINAAGRQ 640
 ○ ++++++
 ○ MBP 312-378
 N TVDEDLKDAQTRIEK.
 ○ ++++++

FIG. 9B

80
 MKGSHHHHGMASNTGGCCQWEDLYDDDKURWGSKAEEGKLVWINGDKVNSLAEVCKGFEKDFGIKVTVEHPDKLE
 MBP 1317
 160
 EKPEQVAATGDGEDILEFWAHDREGYAQSGLLAEITPEKAFQDKLYPFTWDAVRVYNGKLIAYPIAVEALSIIYNSDLLEN
 MBP 1317
 240
 PEKTWEEIFALDKELKAKGKSAIMFNLOEYFTWFLLAAGGSAFKYENGYDINDVYDNAGAKAGLTFVLDIKNNHM
 MBP 1317
 320
 NADTDYSAAEAEMGETAMTINGPNAWSNIDTSKYNVGTVLPEKGGPSPFVGLSAGINAAAPNKELAKEELENYL
 MBP 1317
 400
 LTDEGLEAVNKDKPLGAVALKGYEELVQPKSHNVYIMADKQKNGIKANFKLRNIEDGGVQLAYHQONTPIGGDFVL
 spGFP 147-238
 480
 LPIAHYLSFGSKLSADPNEKRDHEWVLEFVTAAGLITCNDLXYGCGTGGSMVSKGEEELFTGVVPIIVELDGDVNGHHSV
 spGFP 1-146
 560
 SGGEGDAPYQKLLKFLCTYQKLVFPWPLVTLTYGVQCFSPFDHMKQHFKSNPEGYIQERTIFFKLDGNYKTR
 spGFP 1-146
 640
 AELVKFSGDTLVWKILKGLDFKSDGNLLGHELEYNENAATMENACKGELMPEWIFQNSAFWAVRTAVINAASGRTVDED
 MBP 318-379
 LKDRQTRIFKGSHHHHHG.
 800

FIG. 10

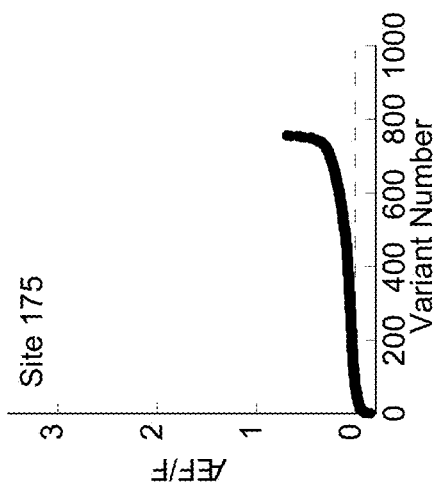


FIG. 11B

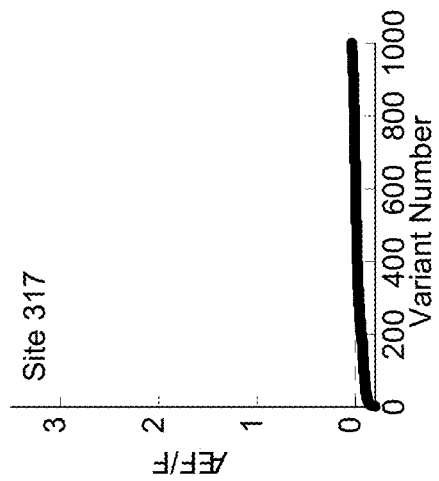


FIG. 11D

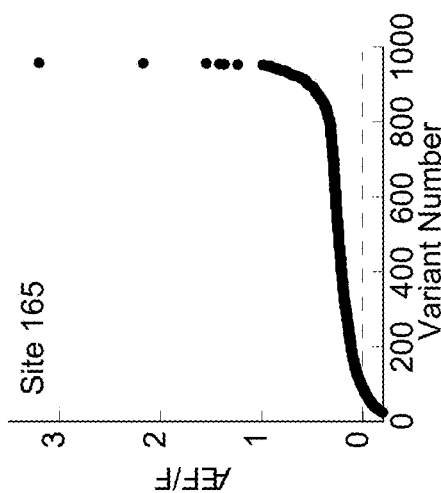


FIG. 11A

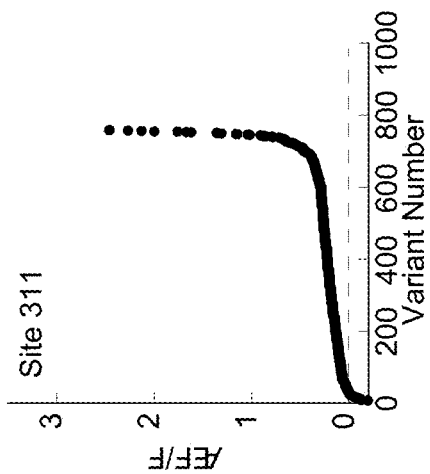


FIG. 11C

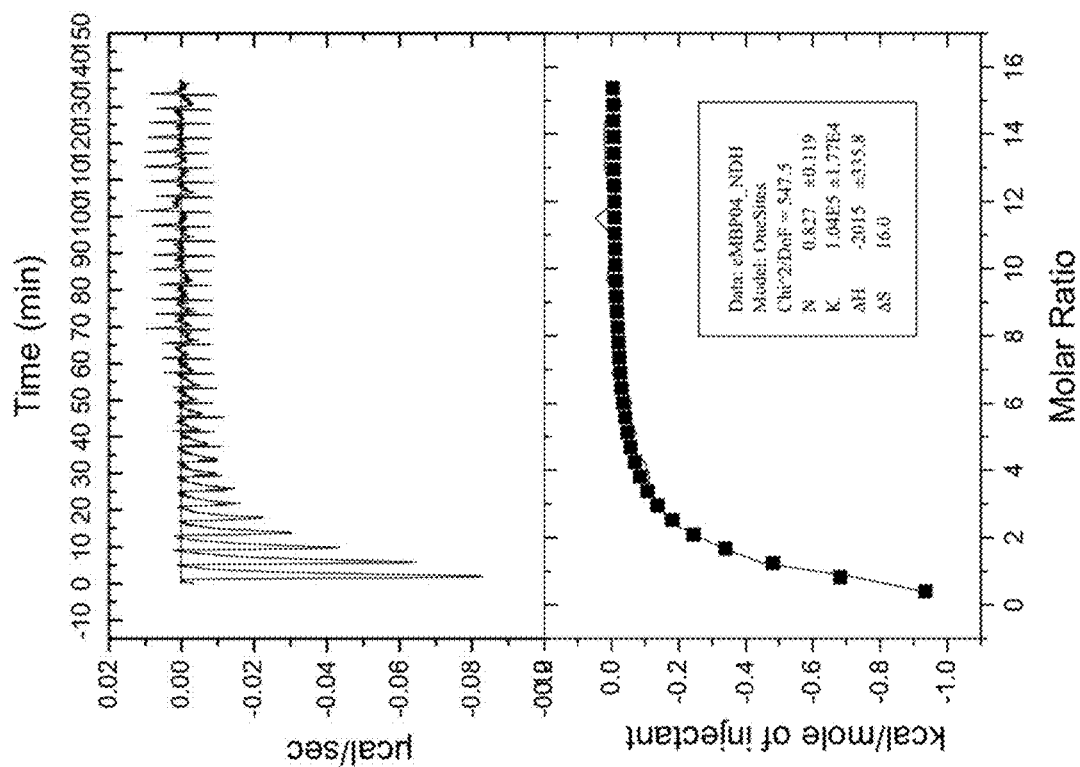


FIG. 12

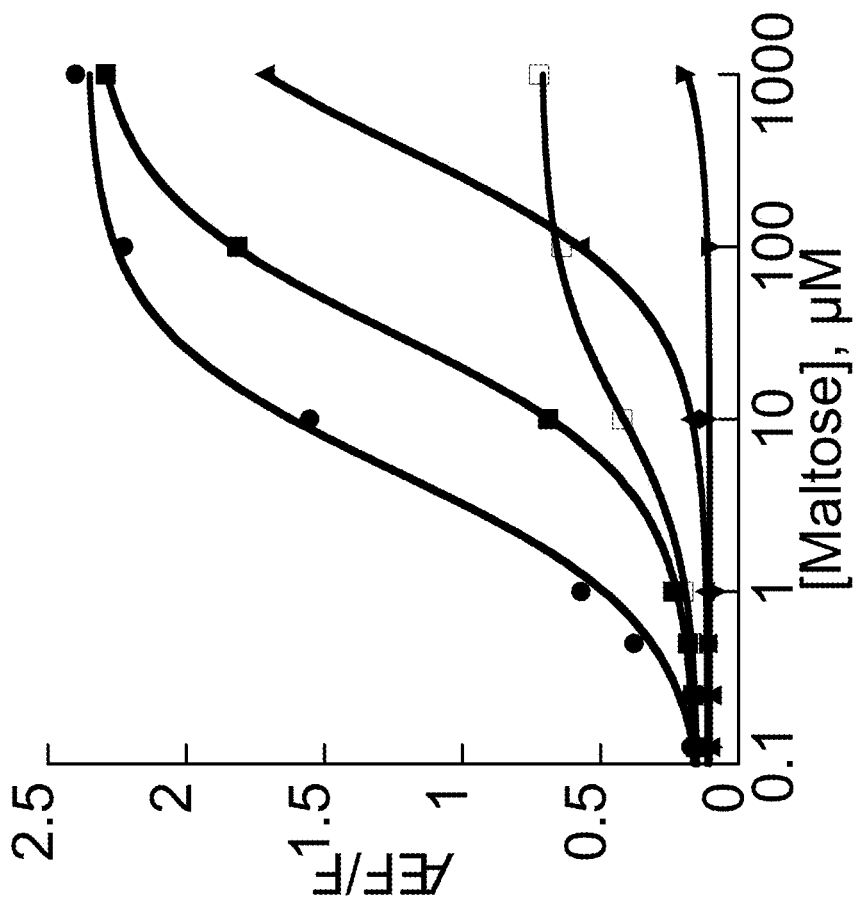


FIG. 13

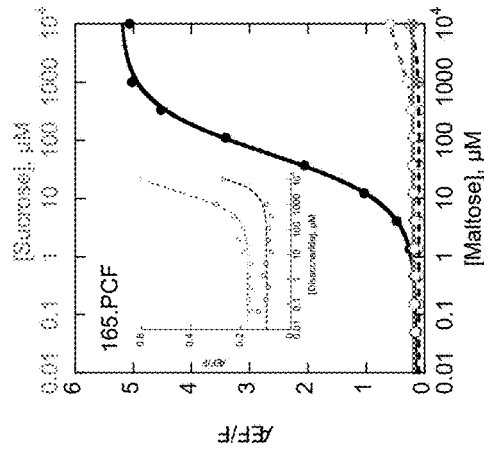


FIG. 14A

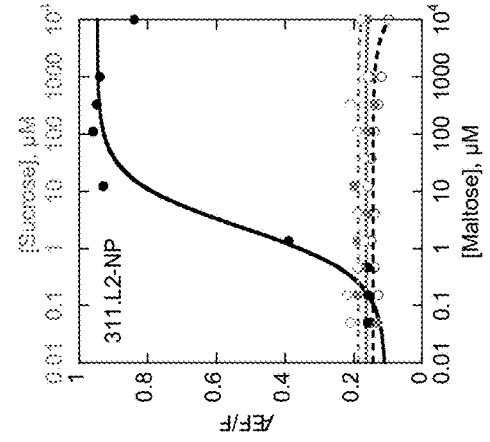


FIG. 14B

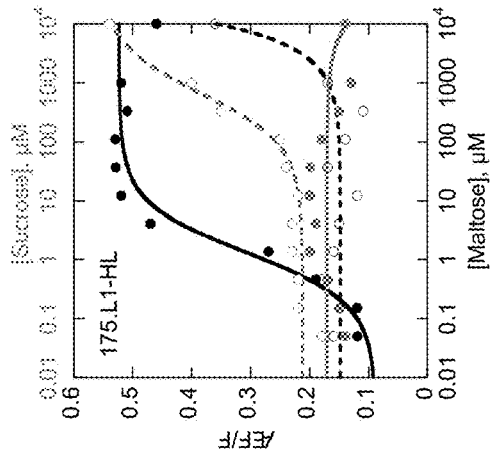


FIG. 14C

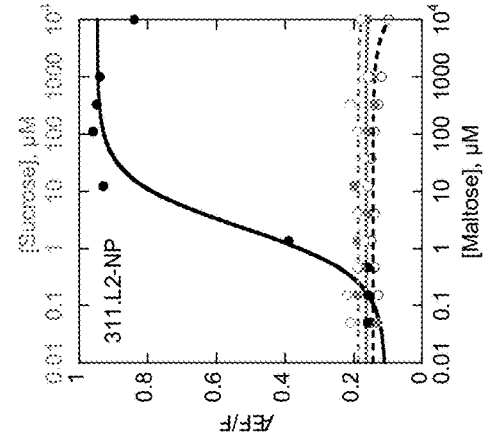


FIG. 14D

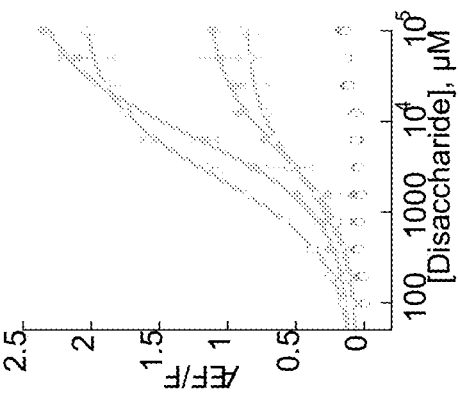


FIG. 15A

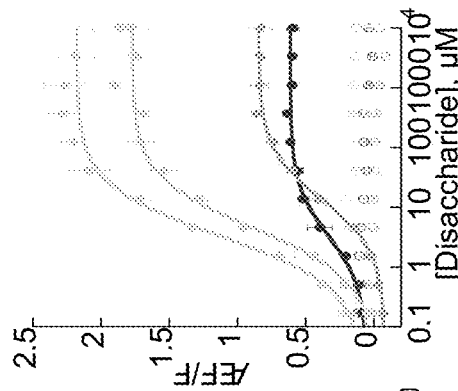


FIG. 15B

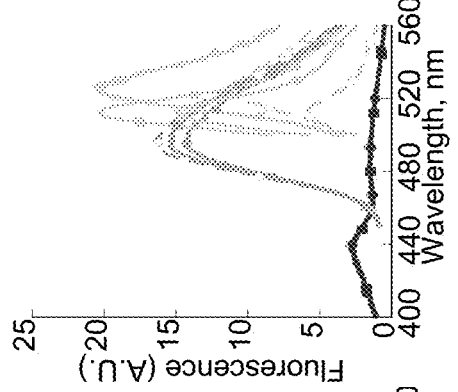


FIG. 15C

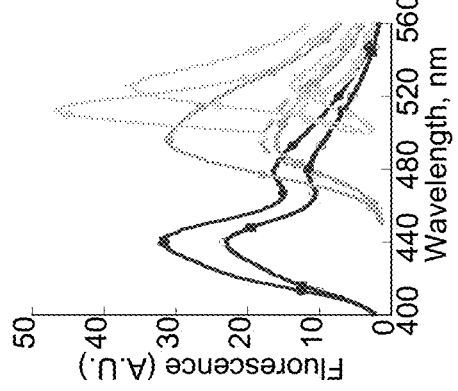


FIG. 15D



FIG. 15E

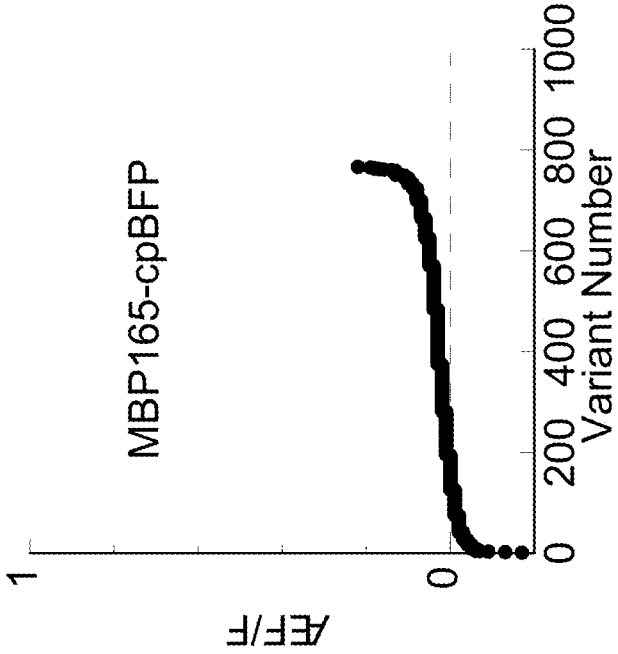


FIG. 16

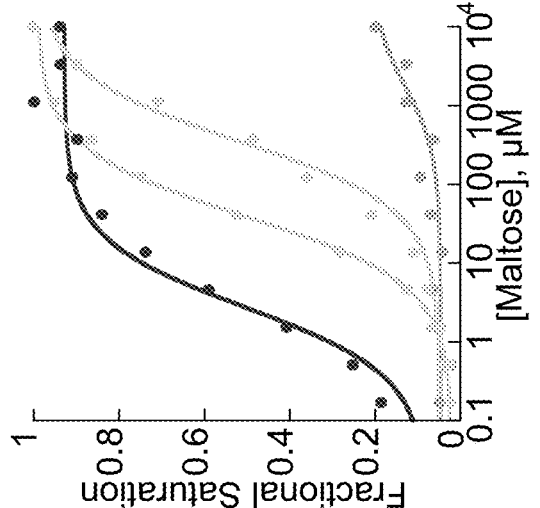


FIG. 17B

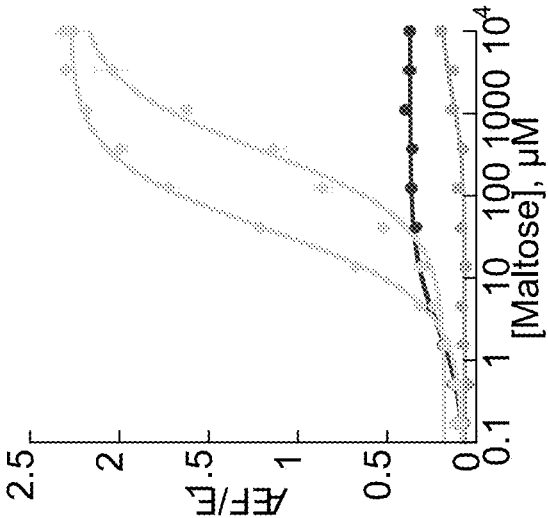


FIG. 17A

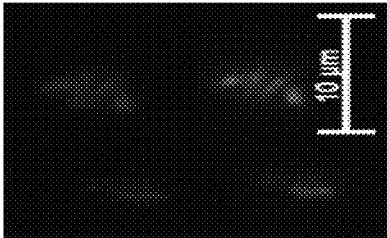


FIG. 18C

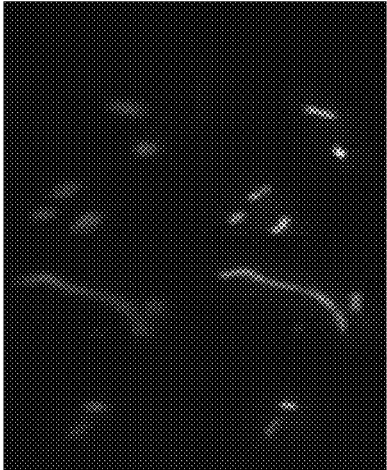


FIG. 18B

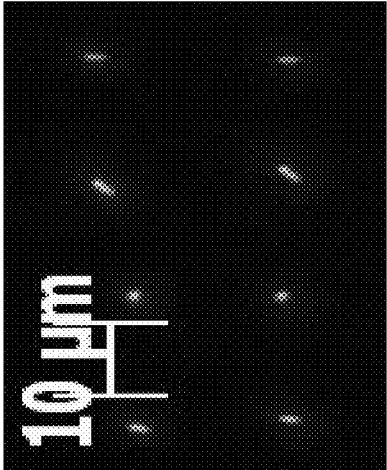


FIG. 18A

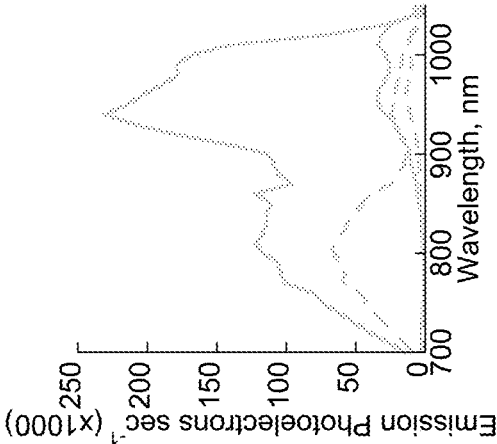


FIG. 19B

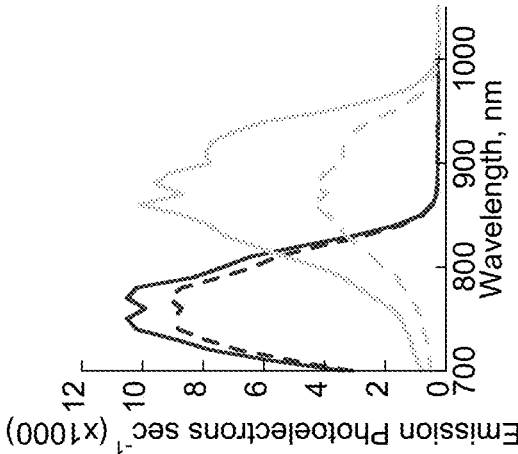


FIG. 19A

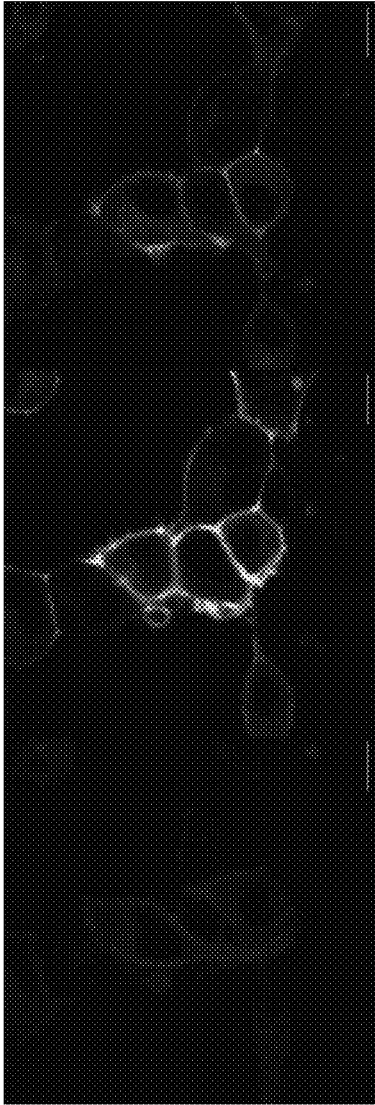


FIG. 20C

FIG. 20B

FIG. 20A

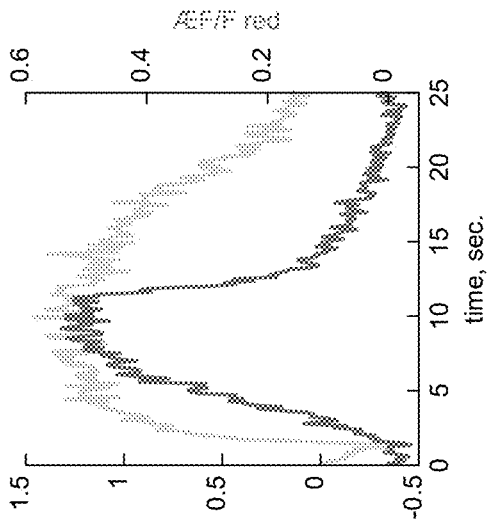


FIG. 21B

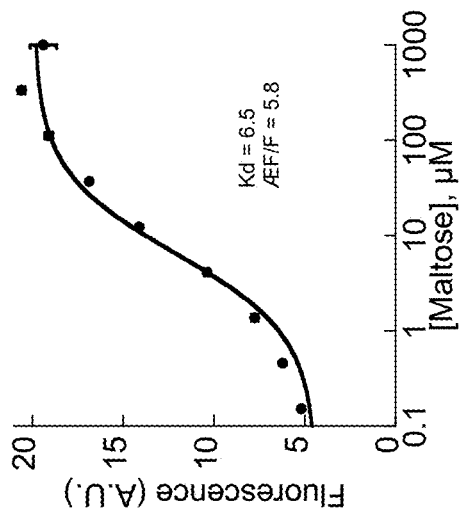


FIG. 21A



FIG. 22A

FIG. 22B

FIG. 22C

FIG. 22D

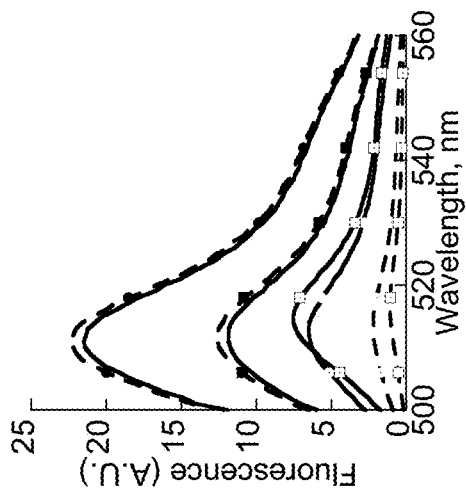


FIG. 23A

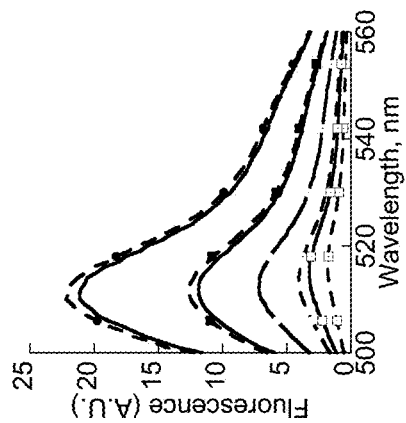


FIG. 23C

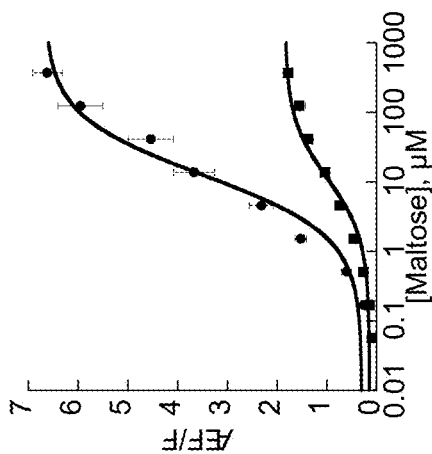


FIG. 23B

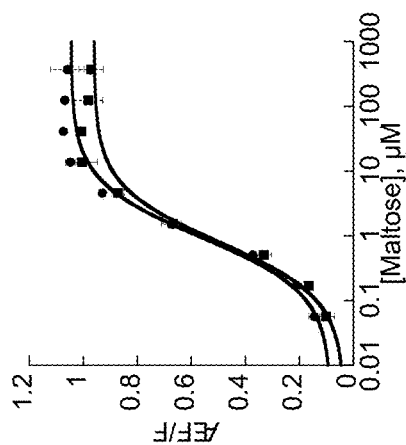


FIG. 23D


```

1  MRCSHHHHHAGMTGGQQRDLDDDKDRKSKIEEGKVYIMIMQNELEVFQSLAEEYMALCPVEVIVFEOKPN 80
2
3  LEDALKAALPTGQGPOLFIRAHIDWIKFAFAGLLEPDEYVTELLINEFAPMAQDMQKHYALPFAETVAILYSKE 160
4
5  IWSEPKTEDEMKALMEKYIDANKEYGLAWPINAYFISALAQAGGSHNYIMAKQKNGKIKANFKLRHNIEDGGVOLA 240
6
7  YHQDNEPLGQGVLLFDNHLVLSQSKLQKDPNEKSDHMLLEVTANGILQMDLNLKCGGSMVSKGELSTGVVPI 320
8
9  LVLEDDVNGHKSVSCEGDATYCKELAFICTYCKLEVPWFVLYTLLVGVQCFSEYEDHMQHDFKSNAPGEGYIQ 400
10
11  ERSLFFKDDGNYKTRAEYKFECDTLVNRIELKQIDFKEDONLGHKLEYNENFEYEDNTEQGLKPKETIECFKFSFT 480
12
13  ELPNPAPTGQYNTQGSIFLEGRAPMNGPWSINRYKACIAGVWPLPFLKDCREYKRPYGVKLIYSAGLKNKO 560
14
15  AAKKFAWLTSESIKTLALSAGYIPVLEKVLDDZFLKNDPVIYGFQQAQYHAYLMPKSPKNSAYWGVGDCALNELIQD 640
16
17  PNADIEGILKKYQOEILANQQGSHHHHHG. 872

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FIG. 24B

◊ pRSET Leader

N MEGSHHHHGMASMTGGCKWGRDLYEDDDKORWGSKLEEGKVIWIMQPSNELEVFSQLAEEYMWALCFEVLVTEQKFN 80

◊

◊

N LEDALKAAIFTCQCPDLPIWADHWLCKSAEACLEPIDEVYTEELNEEPAPWQDAMQYAGHYIALPFAAEVVALIYSKZ 160

◊

◊

N MVSEPKTFDENKALMEKYIDENKEYCIANFVNAFISALIAQAFGGYTFDCKTEQCLDAPETIEGKFTTEIMPYMA 240

◊

◊

N FTGDYNTQOSIFLEGRAPAVNGFWSINDVKKAGINFGWPLPFIADCKEYWERSPYGGVKLIVFAAGCIKSKDAAWKEAK 320

◊

◊

N KLTISEESIKTLALELGYIFVLKVLGDFEISHVVIWADKQKNGIKANPKIRUNIEDGCVQLAVHYQQNTFICGGFVLL 400

◊

◊

N PDNHYLSTQSKLQKDPNKRDHMVLEFVTAGITLQDELKCKGCTGSSMYSKQEELEFQVVPILVELDGDYNGHKFSVS 480

◊

◊

N GECEGDATYCKLILKPICTTYKELPVMWPTLVTLTYGQCFSRYPDMQKHDFFKSAMPEGYIQERTIFFKDDGNKTKRA 560

◊

◊

N EVKFECDYLNRIELKGLDFKEDGNILCHKLEYNFKNDVPIVGFQAVQIHTYMPKSPKMSAVWGCVCAGINEILQDFQ 640

◊

◊

N NADIEGILKVKYQELLANNMQGS . 663

◊

FIG. 25A

*
 N ERGSHHHHMGASMTGGQWGRDLYDDEDDKDRWSKIEGKVIWHAQPNLELVQSLAEIYMALCEVELIVTEQKEN 80
 *
 *
 *
 *
 N LEDALKAAIPTGQGPOLFIMANDWIGKFAEAGLEPIDEIYVTEELINEFAWADQAMQYKGHYYALPFAETVALIYSKE 160
 *
 *
 *
 *
 N MYSEPPKTFDEMKALMEKYDFANEKCYIAWPIINAVFISALQAQFSGYFDKTEQPLKRFETIEGFAFFTEINPIMA 240
 *
 *
 *
 *
 N PFGDYNTOOSIFLECHMFWNVNCFWSINDVRRKACINFCVWVLEPFLIKCKEYWRPFGGKLIYFAACIKKDAWKYAK 320
 *
 *
 *
 *
 N KLTSEESIKTLALELGYIFVLRKYLDDPELPEPESHVIVMADKQKNGIKANEXIRINIEDCGVQLAYHYKONTPIGCGPV 400
 *
 *
 *
 *
 N LLDPNHYLSTQSKLSDPNEKRDHNVILLEPVTAGITLGMDELYKGGTGGSWKGGELFTGVWPIILVELDGVNKHKS 480
 *
 *
 *
 *
 N VSECEGDATYCKLTKSICITTKLVPWPPTLVTLTYGQCFGRFDMWQHDFTKGMPECYIQERTIEFKDDQNYKE 560
 *
 *
 *
 *
 N MAVKPEGDTLNNIELKCIDFRDGNILCHKLEYFNKNDPVIYGFQAVQIAYILMFKSPMSAVWGCYDCAINELQD 640
 *
 *
 *
 *
 N EQNADIEGILMKYQOEILNNQGS . 665
 *
 *

FIG. 25B

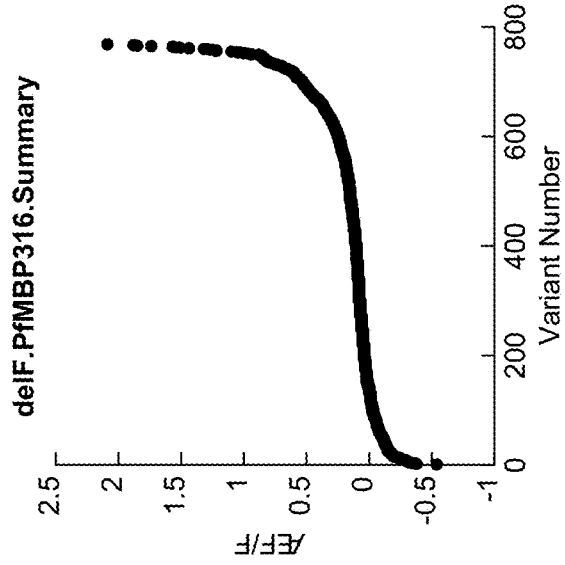


FIG. 26B

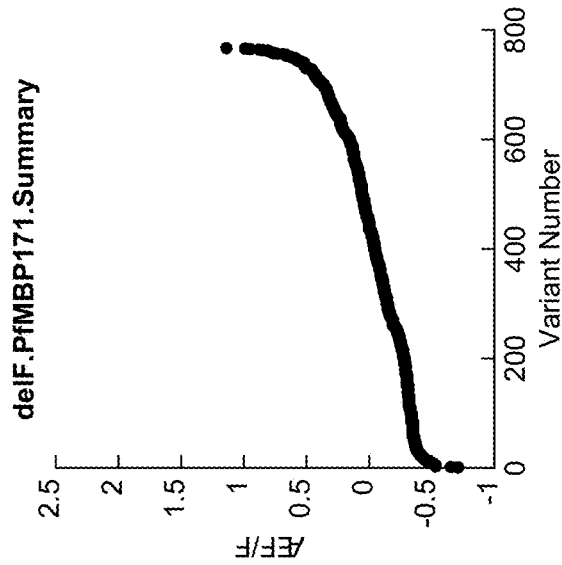


FIG. 26A

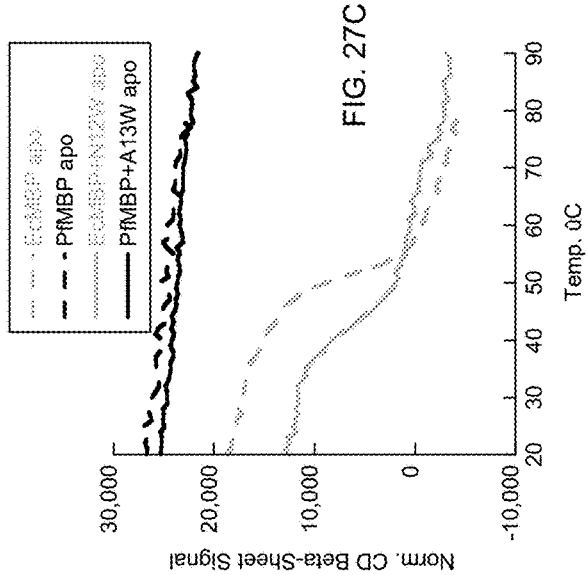
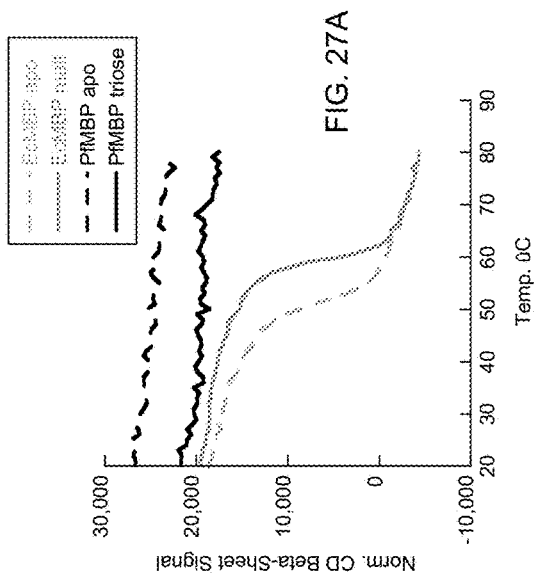
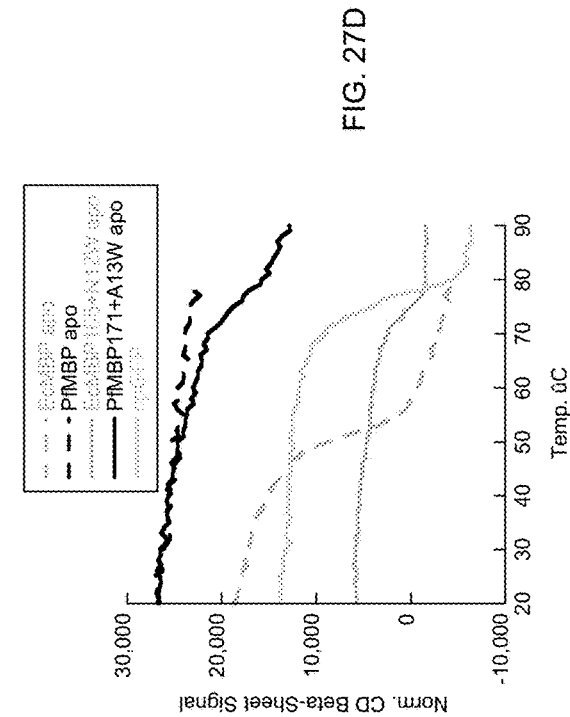
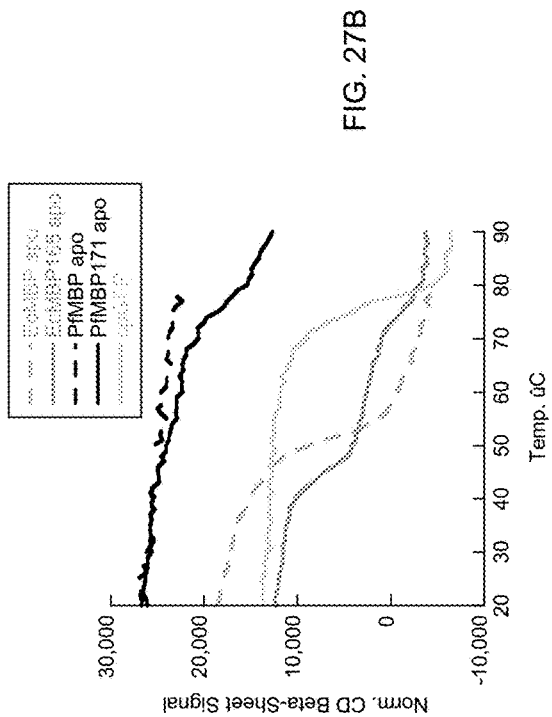


FIG. 27B

FIG. 27D

FIG. 27A

FIG. 27C

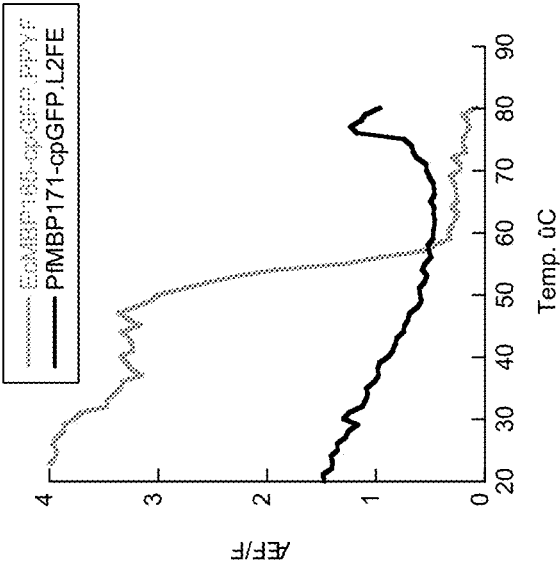


FIG. 28B

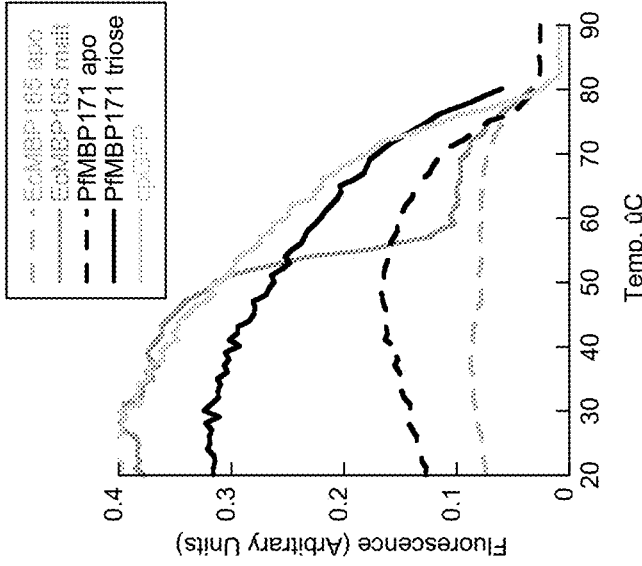


FIG. 28A

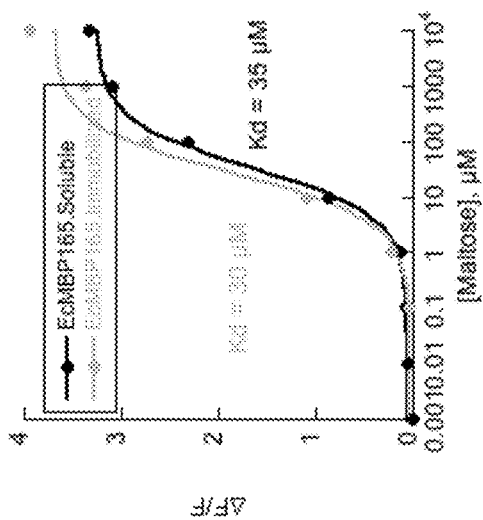


FIG. 28E

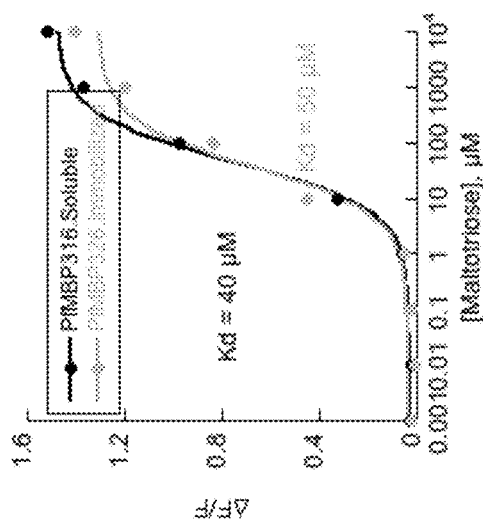


FIG. 28D

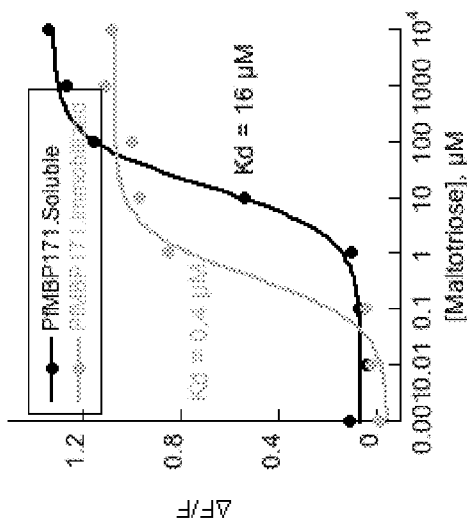


FIG. 28C


```

*
* N MRGSHHHHHGASMTGCGQMRGLYDDDDKRWGSAAGSTLKIANGVIVGHRESSVPSYVDNQQKVVGYSQDYSN 80
*
*
* N AIIVEAVKKLNKPDLOVKLEPLTESQNRIPLLQNGTFDEECGTTNNVERQKQAAESDTLIVVGFALLLKKGGELKDFANL 160
*
*
* N KDKAVVTSSTSEYLLNKLNEQKQANPLISAKDHGDSFRTLFESGRAVAFNEDVLLAGEBAKAKKPNWELVCKKFSQ 240
*
*
* N EAYECMERKDDPQKKLNDDTLAQVQTSCEAKKWFQKFNPILVSHNVYIMADKQKNGIKANFKIHNIEDCGVQLAYH 320
*
*
* N YQONTPIGDFVLLFDNNILSTQSKLSKQFNKRDIMVLEFVTAGITLQMDLYKKEITGGSMYSKGEELFTGVVPIIV 400
*
*
* N ELDGIVNGHKFSVSGEGEDATYKGLTLKFCITTKLWVWPVTLVTTLYGVQCFSRYPDHMKQHDFTKSAHFECYIQER 480
*
*
* N TIFKDDGNKTRAEVREEDTLVNRILKLGIDFKEDGNILGHKLEYNENPLNNEELSDENKALEEENDKALK. 557
*

```

FIG. 29A

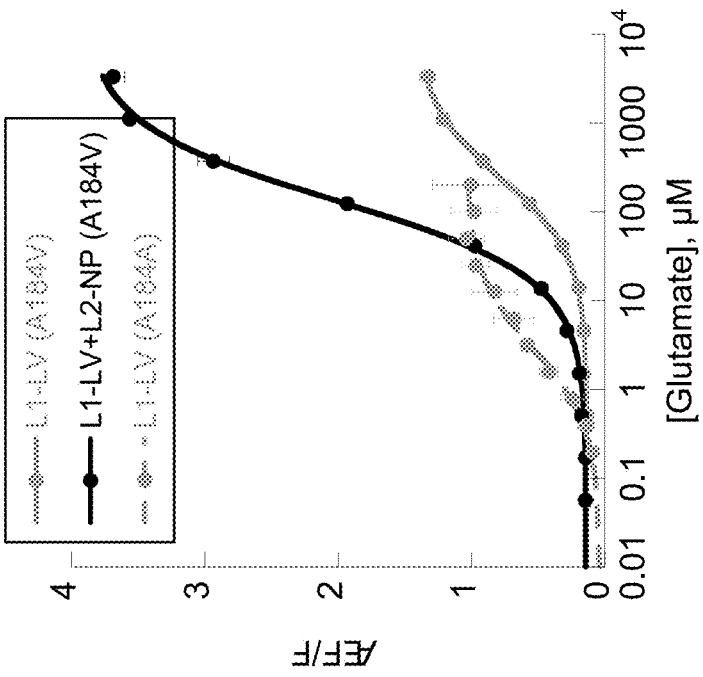


FIG. 30

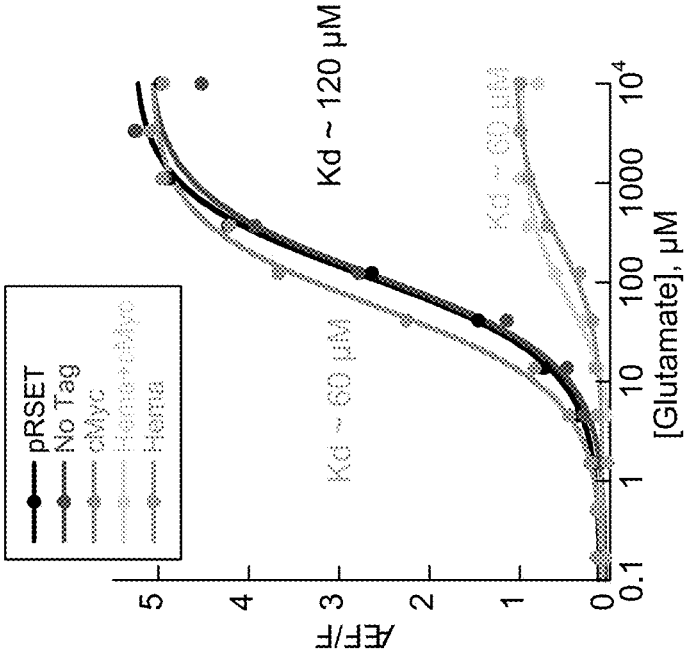


FIG. 31

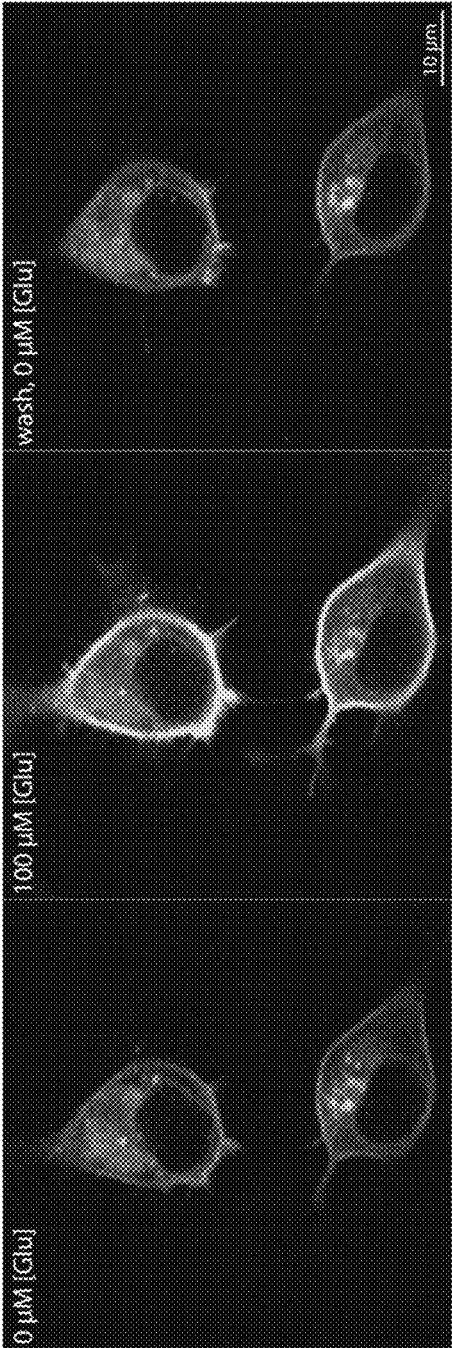


FIG. 32A

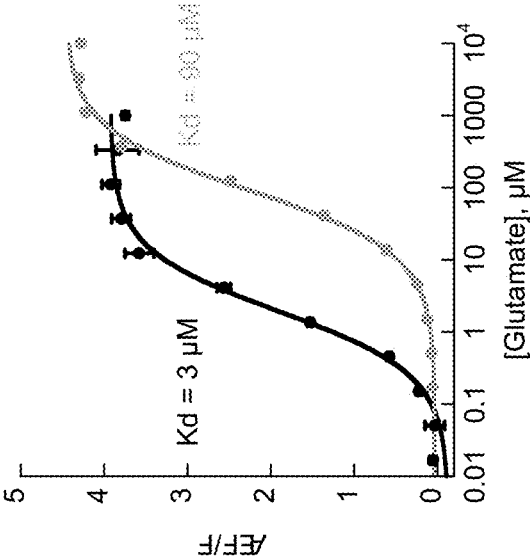


FIG. 32B

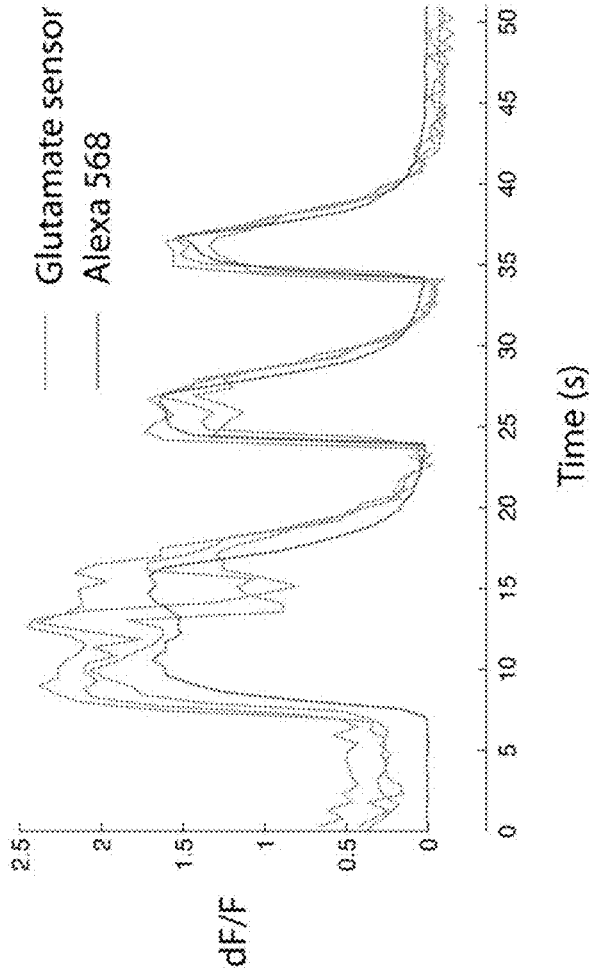


FIG. 33

* **Emb 146** **Emb 0 139**
 N MHHHHHGSZEDEKALNFCIISTESQQNLKQWTFPLQDMKKLVQVNAFFAPDYACIIQQHFNKVDLAWYCNLSAME 80
 *
 *
 * **Emb 0 139** **spGFP 147-239**
 N AVDRANGOVENQTVAADSSHNVYIMADKQRNGLIKANKIRHNIEDGGVQLAYHYQQNTFIGGVPVLLFDNHVLSQSKL 160
 *
 *
 * **spGFP 147-239** **Linker** **spGFP 1-146**
 N SKOPNEKRDRHMVLLSEVTAAGIPLGMDELYKGGTGGSMVSKGELLFTGVVPIVLELDGVDVNGHKFSVSGEGCDNTYQKL 240
 *
 *
 * **spGFP 1-146**
 N TLKFICTYTKLFFVWFVTLVTLVYGVQCFSRNFDIMKQHDFFKSAMPEGYIQERTIFFDDGNYKRAEVKFEQDTLVNR 320
 *
 *
 * **spGFP 1-146** **Emb 0 31-319**
 N IELKGIQDFWEDCNILGHKLEYNFGYWSVLIVNKDPSPINVLDLAKRRDLTFGNDPNSSTGFLVFCYVFRKNNISA 400
 *
 *
 * **Emb 0 31-319**
 N SDFKRTVANACHETNALAVANKQVDVANTNTENLDKLTSAPELKLKVIWKSPLIFGDFIVWRNLSLSEYTKDKLIYDFFM 480
 *
 *
 * **Emb 319** **Emb 0 31-319**
 N NYGKTPPEEKAVLERLGWAPBPASSDILQVPIRQLALEKEMQSVKDKGLNEQDKLAKTTAIQACLDLDRLNALSMS 560
 *
 *
 * **Emb 319**
 N VSKAVQ.
 *
 *
 * 567

FIG. 34A

```

*   *
N   MHHHHHGGSEEQEKALNFCIIISTESQONLKPORTFLQDMKKLVKYNATFAPDYAGIIQQMHNKVDIAWYCNLSAME
*   *
*   *
*   *
N   AVDRANGOVENQIVAAADAEVYIMADKQKNGIKANFKIRHNIEKGVQLAYHYQQNTPIGDIPVLLPENHYLSTQSKLSK
*   *
*   *
*   *
N   SKDENEKRDIHMVLLLEFVTAAGITLQMDLNLKGGTGGSMVSKGEELEFGVPIVLELDGGVNGHKSVSCEGEGDATYQKL
*   *
*   *
*   *
N   TLKFICTTKLQVWPFLVTELYGVQCFSRYPDHNKQNDFFKSNMPEGYIQERTIFFKDDGNKTRAEVKFEQDTLVNR
*   *
*   *
*   *
N   IELKCIDFKEDCNILGHKLEFVENVPGYWSVLIIVKDSPINNLDLAKRKDLFGNGDPNSTSGELVPGYVYFAKNNISA
*   *
*   *
*   *
N   GKTPEEKAVLERLQWAPFRASSDLQVPIRQALFKEMQSVKNGKGLNEQDKLAKTALQALDLDLDRNNARSAMSSVS
*   *
*   *
*   *
N   NYKTPPEEKAVLERLQWAPFRASSDLQVPIRQALFKEMQSVKNGKGLNEQDKLAKTALQALDLDLDRNNALSAMSS
*   *
*   *
N   VSKAVQ.
*   *
*   *

```

FIG. 34B

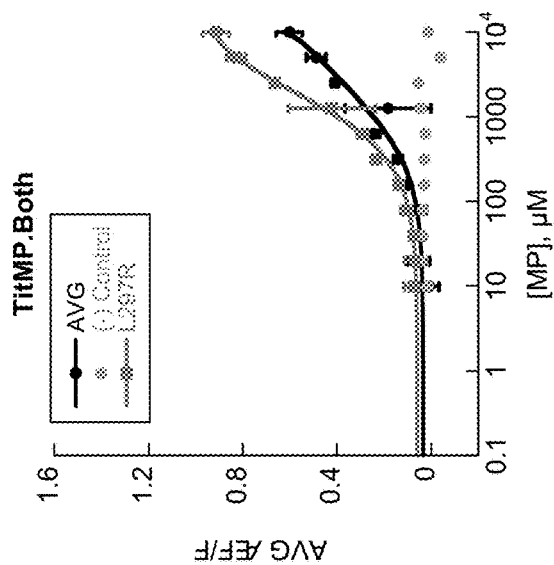


FIG. 35C

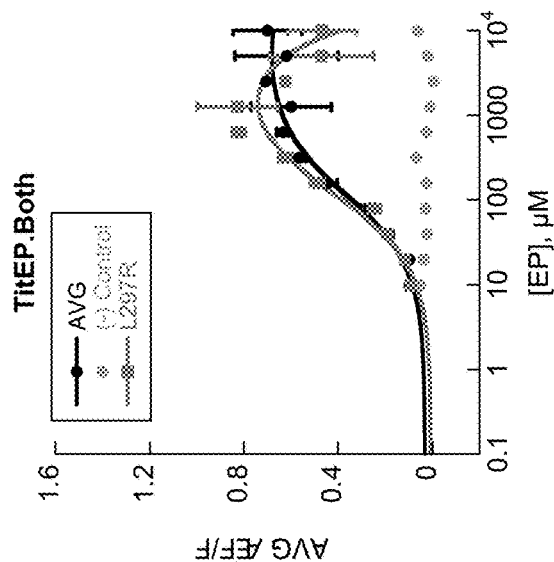


FIG. 35B

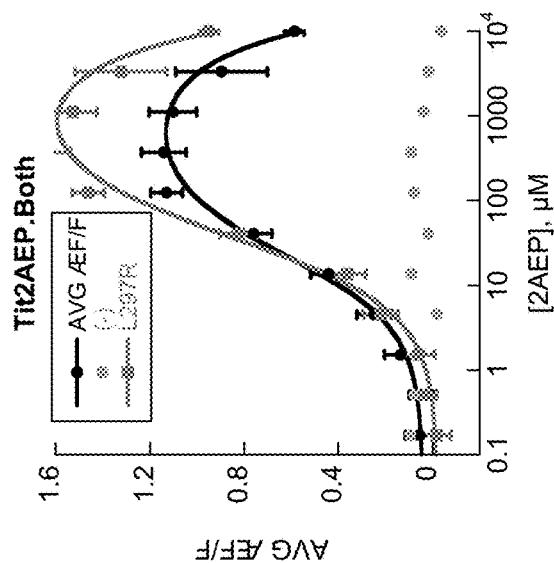


FIG. 35A

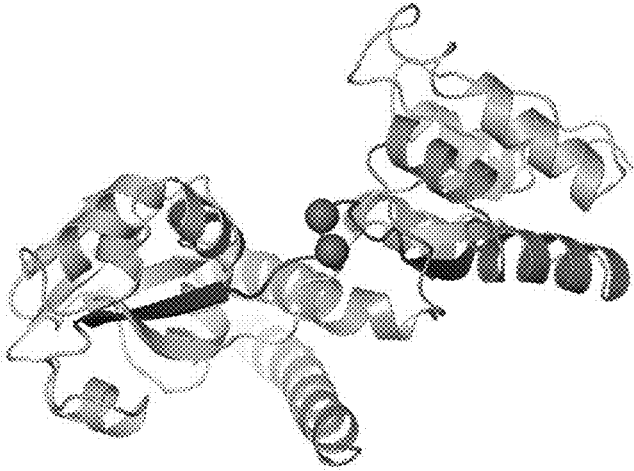


FIG. 36A

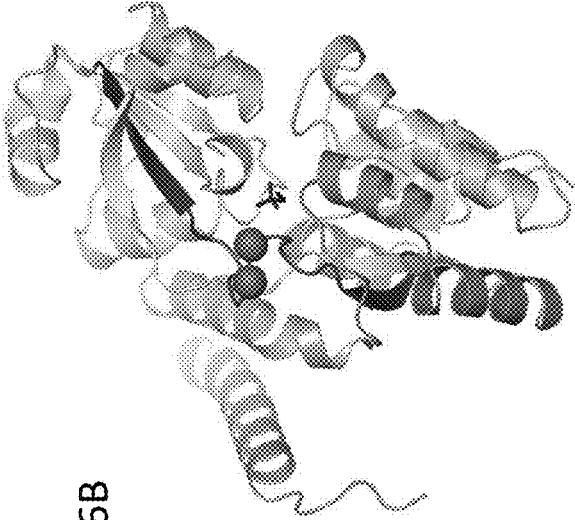


FIG. 36B

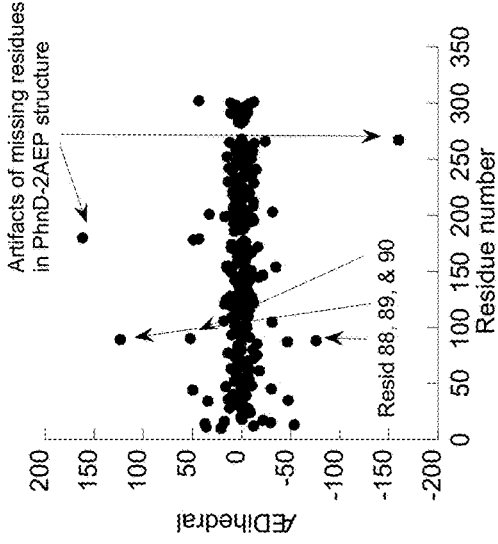


FIG. 36C

80
180
240
320
400
480
560
640
678

spFF 1-146
MPCSHHHHGGASMTGGQMGEDLYDDDDKDRWGSKLEIFSWAGDEGEALALRLKQKYPQVEVINALVTCAGVN
spFF 1-146
ARAVLKRMLAGEPDTFQVHAGMELIGVAVANMELLSALFRQEWLDAEPKGLIDLISYKGIWSVFNIIHRSNVMW
spFF 1-146
YLPKLRKGVNRPFWDEFACOTLAKGKLEFLALGENWVQSHLWESVALAVLCPDRNNLAKKLRFTDFKAVRW
spFF 1-146
EVTQRVLDCAKWDAGLSWQQVDRVQSGKAFVWGDWAGVFTTLKKECTDFWASDFGTCYGVFMUSDSFGLPKG
spFF 1-146
AKRQKALNWLRYGSKGQDFENFLKESLAARLSDPSKYCGSNVYIMAKQKNGIKAKFKIRHNLEDGGVQLAYHQ
spFF 1-146
QNTPIGDPVLLPDRHYLSTQSKLSKQFNEKRDHWLLEFVTAAGITLGMDELYKGGTGGSWSKGELEFGVYFLLVEL
spFF 1-146
DGDVNSHKFSVSEGECDATYKGLTLKFICTTKLQVFWPTLVTLTYGQCSRYPDHMKQDFEFGKAMPEGYIQERUL
spFF 1-146
EFKDDGNYKTRAEVRFEGDTLVNRIELKGIQDFKEDNLELHKLEYNFNNAVQSANRDKWSRIVGLSLVHGAVAPESE
spFF 1-146
NSQFQVREIIFQTQNPQANNAQAIAQVGLCRLLGQ.

FIG. 37A

* **spFF 137**
 N KRGGHHHGHGMASTGGQMRDLYDDDKORWESKLEIFSWAGDEGPALEALIRLKKQKYPQVEVINATVGGACVN 80
 *
 * **IGBP 138**
 N ARVLTINLGGPDETFQVHAGMELIGNVVANMEDLSALFQEGNLDAPFEGCLIDLSYKGGINSVFNIIHRSNWNK 160
 *
 * **IGBP 139**
 N YLPAKLKENVNPRINDEFLATQQLKQNGLEAFALALGENWQCHILMESVALAVLSPDDMNNLANSKLTQPKAVPAW 240
 *
 * **IGBP 138**
 N EYFGRVLDCAKNDANGLSMQAVDRVYQKAAFNVMIDMAGNMTTLKLPQDFANAPSPGTCGVNWLSDSFLKQ 320
 *
 * **IGBP 138** **spFF 147-238**
 N AKNQVALNWLAVGSKGQDTENPLKGSIAARLSDPSKIPNSHNVYIMADKQNGEKANFKIRINIEGGVQLAYHYQ 400
 *
 * **spFF 147-238** **Linker** **spFF 146**
 N QNPIGDKFVLLPNDHYLSYQSKLSKPNERRDHVLEFVHAGITLGGDLKGGTGGSMWKGKELFTGVVPIVEL 480
 *
 * **spFF 146**
 N DGGVNGHKEVSGEGEDRFPYKLEKSICTTKRLEFVFWVLTLYGQCFSRYPDMMKQNDFFKSNPEQYIDERTI 560
 *
 * **spFF 146** **IGBP 277-284**
 N EYFDGQNYKTRAEVKFEGDLVNRILELKGIDFEDENILCHKLEYNFNFNAYGQSANRQWSRIVGSLVHGAVAPESP 640
 *
 * **IGBP 277-284**
 N MSQCTVMEIEIQTRNFOANNAQAIAQVGLRGLGQ. 679
 *

FIG. 37B

```

*
N      pHEP Leader
*      MKGSHHHHGMASMTGCGKGRDLYDDDDKDKAGSKLEIFGWNQDEGPALEALLRLYKQKYPGVEVINALTGCGAVN
*      +-----+-----+-----+-----+-----+-----+-----+-----+-----+-----+
*      80
*
*      TGGP 1378
*      ARAVLAKTRMLGGDFPDTFQVHNGMELIGTWVANNEDLSALEQEQWLNQAPFKGLIDLSYKGGINSVFNHRSNVNM
*      +-----+-----+-----+-----+-----+-----+-----+-----+-----+-----+
*      160
*
*      TGGP 1378
*      YLPKLEKGVNPPKINDEFLAYCQLKQGLKPLALGENTVQOHNESVALVLEPDDNNLWNGKLFYDFKAVNM
*      +-----+-----+-----+-----+-----+-----+-----+-----+-----+-----+
*      240
*
*      TGGP 1378
*      ARAVLAKTRMLGGDFPDTQVAANGMELIGTWVANNEDLSALFRQEGWLNQAPFKGLIDLSYKGGINSVFNHRSNVNM
*      +-----+-----+-----+-----+-----+-----+-----+-----+-----+-----+
*      320
*
*      TGGP 147-238
*      AKNNONAINKLRLVGSKEGDTENPLKGSIAALDSDFSKGCSINVTYIMDKQKNGIKANFKYIRHNIEDGGVQLAYHYQ
*      +-----+-----+-----+-----+-----+-----+-----+-----+-----+-----+
*      400
*
*      spGFP 147-238
*      QMTPIGDQVLLPDNHYLSTQSKLQKQPNKRDHMKVLEFFVYKAGITLNGELYKGGTGGSMVSKLELFTGVYPILEVEL
*      +-----+-----+-----+-----+-----+-----+-----+-----+-----+-----+
*      480
*
*      spGFP 1-148
*      EGDWNGHKESVSGEGDATYCKLKLKPICTIKLVPWFLVTLTYCYCCFSRYPDKMKOHDFFKSAMFESIQERTI
*      +-----+-----+-----+-----+-----+-----+-----+-----+-----+-----+
*      560
*
*      spGFP 1-148
*      FEYDDGNFKERAEVKEGDTLWNRLELKGIDFKEDGNILGHKLEYNFNNAAYGQSMEDWRNRYVGSLSVHGAVPESF
*      +-----+-----+-----+-----+-----+-----+-----+-----+-----+-----+
*      640
*
*      TGGP 37-204
*      MSQGTWELFELQTSQQAANNAQALADQVSLGCGQ.
*      +-----+-----+-----+-----+-----+-----+-----+-----+-----+-----+
*      678
*

```

FIG. 37C

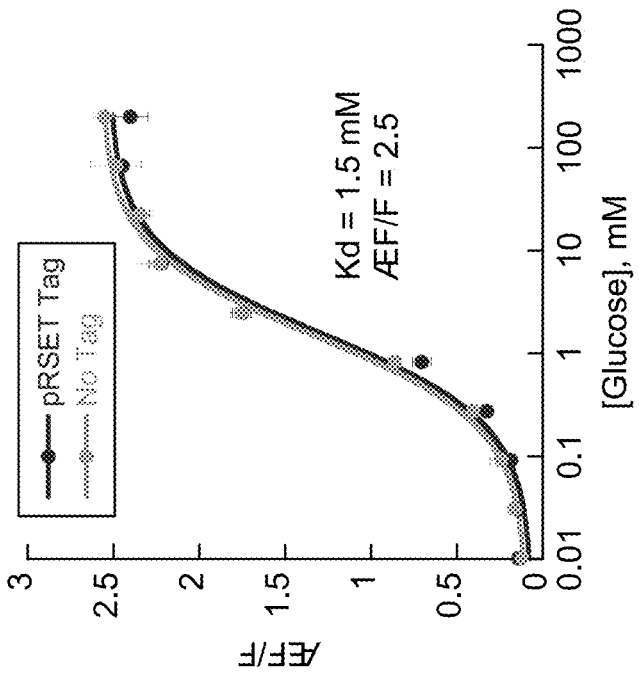


FIG. 38

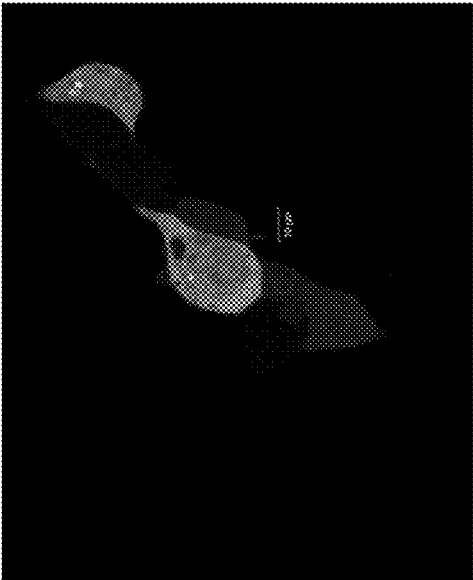


FIG. 39

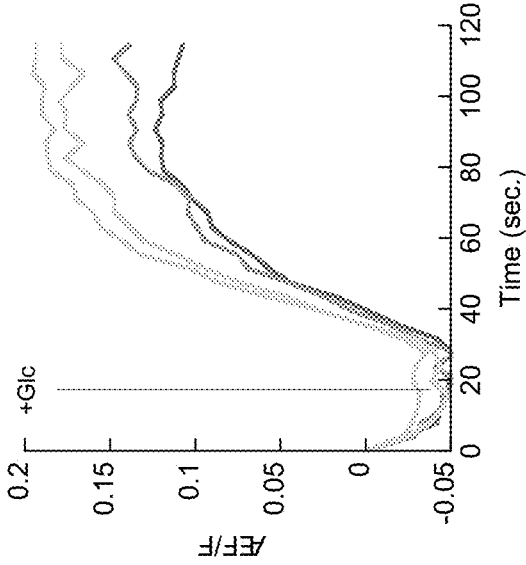


FIG. 40B

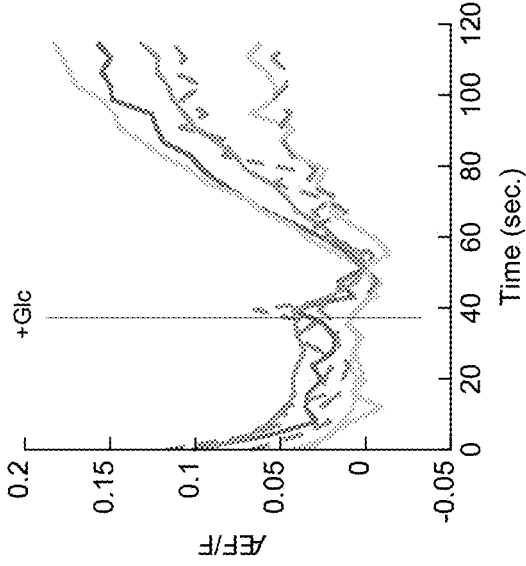


FIG. 40A

MKIKTGARIL ALSALTTMMF SASALAKIEE GKLVIIWINGD
KGYNGLAEVG KKFEEKDTGK VTHEHPDKLE EKFPQVAATG
DGPDIIFWAH DRFGGYAQSG LLAETTPDKA FQDKLYPFTW
DAVRYNGKLI AYP IAVEALS LIYNKDLLPN PPKTWEEIPA
LDKELKAKGK SALMFNLQEP YFTWPLIAAD GYAFKYENG
KYDIKDVGVD NAGAKAGLTF LVDLIKNKHM NADTDYSIAE
AAFNKGETAM TINGPWAWSN IDTSKVNYGV TVLPTFKGQP
SKPFVGVLSA GINAASPKE LAKEFLENYL LTDEGLEAVN
KDKPLGAVAL KSYEEELAKD PRIAATMENA QKGEIMPNI P
QMSAFWYAVR TAVINAASGR QTVDEALKDA QTRITK

FIG. 41

MRRATYAFAL LAILLVLGVVA SGCIGGGTTT
PTQTSPTQP TTTQTPTQTE TQAVECGSGK
VVIWHAMQPN ELEVFQSLAE EYMALCPEVE
IVFEQKPNLE DALKAAIPTG QGPDLEFIWAH
DWIGKFAEAG LLEPIDEYVT EDLLNEFAPM
AQDAMQYKGH YYALPFAAET VAI IYNKEMV
SEPPKTFDEM KAIMEKYYPD ANEKYGIAMP
INAYFISAIA QAFGGYFDD KTEQPGLDKP
ETIEGFKFFF TEIWPYMAPT GDYNTQQSIF
LEGRAPMMVN GPWSINDVKK AGINFGVVPL
PPIIKDGKEY WPRPYGGVKL IYFAAGIKNK
DAAWKFAKWL TTSEESIKTL ALELGYIPVL
TKVLLDDPEIK NDPVIYGFQ AVQHAYLMPK
SPKMSAVWGG VDGAINIILQ DPQNADIEGI
LKKYQQEILN NMQG

FIG. 42

MNAKIIASLA FTSMFSLSTL LNPAYAEEQE
KALNFGIIST ESQQNLKPQW TPFLQDMEKK
LGVKVNFAFA PDYAGIIQGM RFNKVDIAWY
GNLSAMEAVD RANGQVFAQT VAADGSPGYW
SVLIVNKDSP INNLDLLAK RKDLTFGNGD
PNSTSGFLVP GYYVFAKNNI SASDFKRTVN
AGHETNALAV ANKQVDVATN NTENLDKTKT
SAPEKLELKV IWKSPILIPG DPIVWRKNLS
ETTKDKIYDF FMNYGKTPEE KAVLERLGWA
PFRASSDLQL VPIRQLALFK EMQSVKDNKG
LNEQDKLAKT TAIQAQLDDL DRLNALSAM
SSVSKAVQ

FIG. 43

MQLRKPATAI LALALSAGLA QADDAAPAAG
STLDKIAKNG VIVVGHRESS VPFSYDNQO
KVVGYSQDYS NAIVEAVKKK LNKPDLOVKL
IPITSQNRIP LLQNGTFDFE CGSTNNVER
QKQAAFSDTI FVVGTRLLTK KGGDIKDFAN
LKDKAVVVTG GTTSEVLLNK LNEEQKMNMR
IISAKDHGDS FRTLESRAV AFMDDALLA
GERAKAKKPD NWEIVGKPKS QEAYGCMLRK
DDPQFKKLLMD DTIAQVQTSQ EAEKWFDKWF
KNPIPPKNLN MNFELSDEMKA ALFKEPNDKA
LN

FIG. 44

MRKWLLAIGM VLGLSALAQQ GKLEIFSWWA
GDEGPALEAL IRLYKQKYPG VEVINATVTG
GAGVNARAVL KTRMLGGDPP DTFQVHAGME
LIGTWVVANR MEDLSALFRQ EGWLQAFPKG
LIDLISYKGG IWSVPVNIHR SNVMWYLPK
LKEWGVNPPR TWDEFLATCQ TLKQKGLEAP
LALGENWTQQ HLWESVALAV LGPDDWNNLW
NGKLFKTFDPK AVRAREVEFGR VLDCANKDAA
GLSWQQAVDR VVQKKAAFNV MGDWAAGYMT
TTLKLPKPGTD FAWAPSPGTQ GVFMMLSDSF
GLPKGAKNRQ NAINWLRLVG SKEGQDTFNP
LKGSIAARLD SDPSKYNAYG QSAMRDWRSN
RIVGSLVHGA VAPESFMSQF GTVMEIFLQT
RNPQAAANAA QAIADQVGLG RLGQ

FIG. 45

MIRTLCLKFM LAGAVCMATL TAGSAFAAEP
ESCGTVRFSD VGWTDITATT ATATTILEAL
GYETDVKVLV VPVYTSLKN KDIDVFLGNW
MPTMEADIAP YREDKSVETV RENLAGAKYT
LATNAKGAEL GIKDFKIDIAA HKDELKGKIY
GIEPGNDGNR LIIDMVEKGT FDLKGFVEVE
SSEQGMLAQV ARAEKSGDPI VFLGWEPHPM
NANFKLTYLS GGDDVFGPNY GGATVHTNVR
AGYTTECPNV GKLLQNLISFS LQMEINEIMGK
ILLNDGEDPEK AAAAWLKDNP QSIEPWLSGV
ATKDGGDGLA AVKKAALGL

FIG. 46

MGGGRSTETS SSSGGDGGAT KKKVVVGTDA
AFAPFEYMQK GKIVGFDVDL LDAVMKAAGL
DYELKNIGWD PLFASLQSKV VDMGISGITI
TDERKQSYDF SDPYFEATQV IIVKQGSQVK
NALDLKGTI GVQNAITGQE AAELKFGKGP
HIKKFETTIV AIMELLNGGV DAVITDNAVVA
NEYVKNNPNK KLQVIEDPKN FASEYYGMIF
PKNSELKAKV DEALKNVINS GKYTEIYKKW
FGKEPKLDRL

FIG. 47

MKKSLLSAVA LTAMVAFGGS AWADVIAVG
APLTGPNAAF GAQIQKGAEQ AAKDINAAGG
INGEQIKIVL GDDVSDPKQG ISVANKFVAD
GVKFFVGHFN SGVSI PASEV YAENGILEIT
PAATNPVFTE RGLWNTFRTC GRDDQQGGIA
GKYLADHFKD AKVAIIHDKT PYGQGLADET
KKAANAAGVT EVMYEGVNVG DKDFSALISK
MKEAGVSI IY WGGLHTEAGL IIRQAADQGL
KAKLVSGDGI VSNELASIAG DAVEGTLNTF
GPDPTLRPEN KELVEKFKAA GENPEAYTLY
SYAAMQAIAG AAKAAGSVEP EKVAEALKKG
SFPTALGEIS FDEKGDPKLP GYVMYEWKKG
PDGKFTYIQQ

FIG. 48

MNIK GKALLA GCIALAFSNM ALAEDIKVAV
VGAMSGPVAQ YGDQEF TGAE QAVADINAKG
GIKGNKLQIV KYDDACDPKQ AVAVANKVVN
DGIKYVIGHL CSSSTQPPASD IYEDEGILMI
TPAATAPELT ARGYQLILRT TGLDSDQGPT
AAKYILEKVK PQRIAVHDK QYGEGLARA
VQDGLKKGNA NVVFFDGITA GEKDFSTLVA
RLKKENIDFV YYGGYHPFMG QILRQARAAG
LKTQFMGPEG VANVSLSNIA GESAEGLLVT
KPKNYDQVPA NKPIVDAIKA KKQDPSGAFV
WTTYAALQSL QAGLNQSDDP AEIAKYLKAN
SVDITVMGPLT WDEKGDILKGF EFGVFDWHAN
GTATDAK

FIG. 49

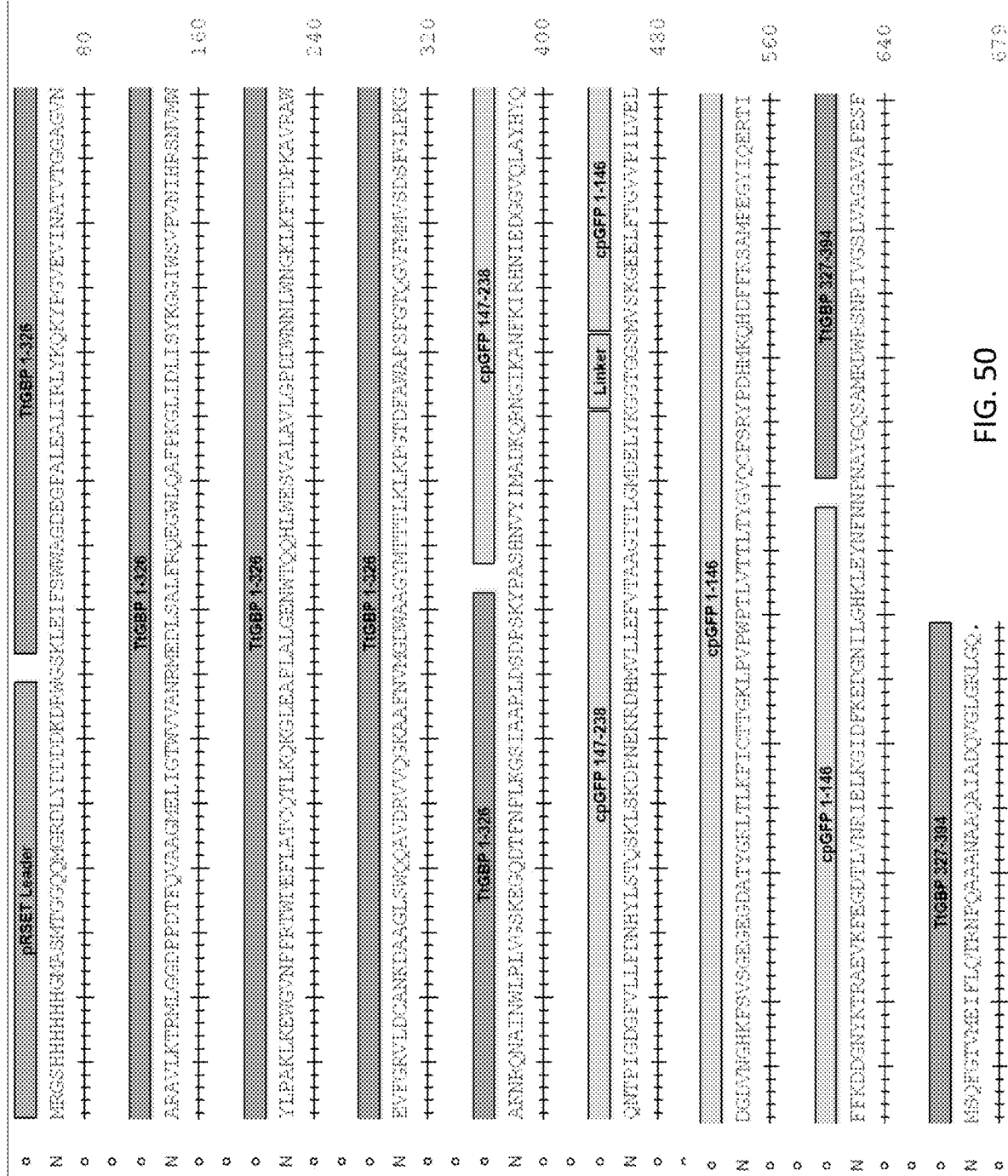


FIG. 50

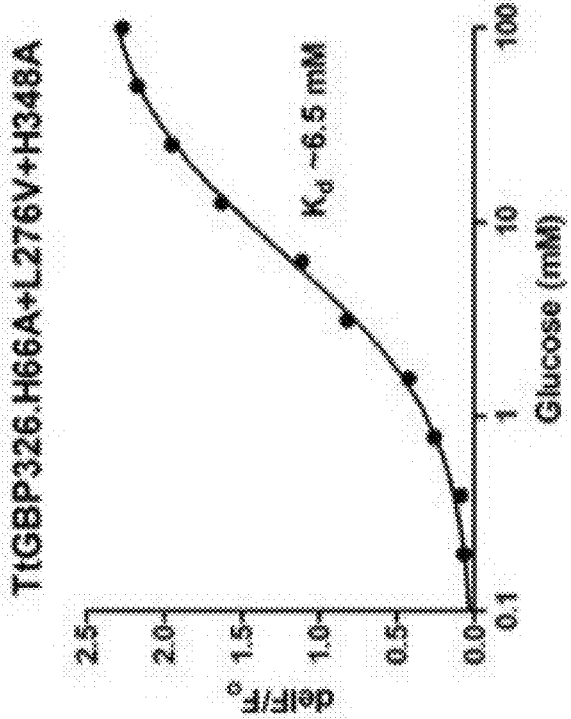


FIG. 51

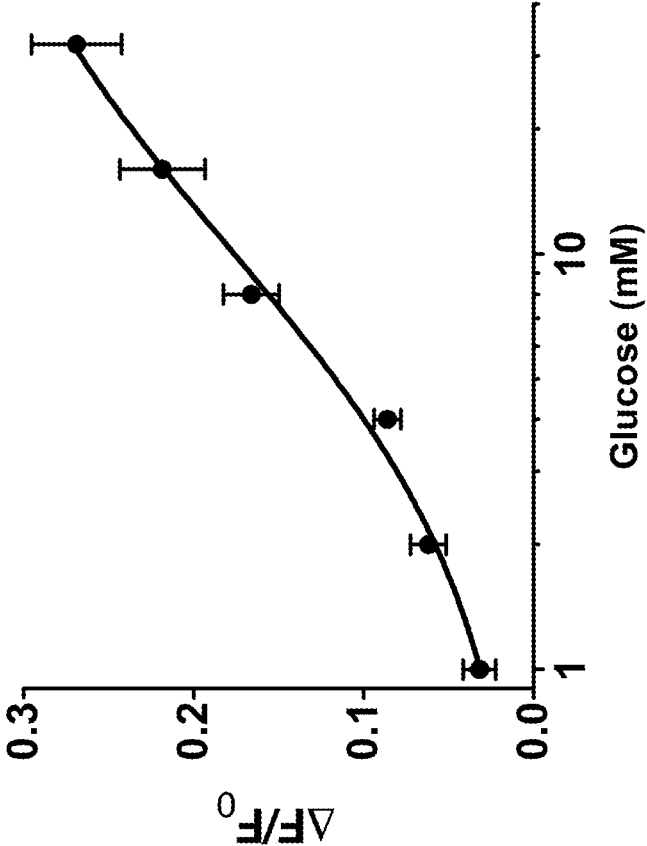


FIG. 52

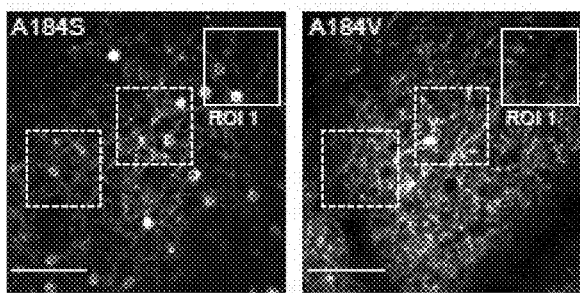


FIG. 53A

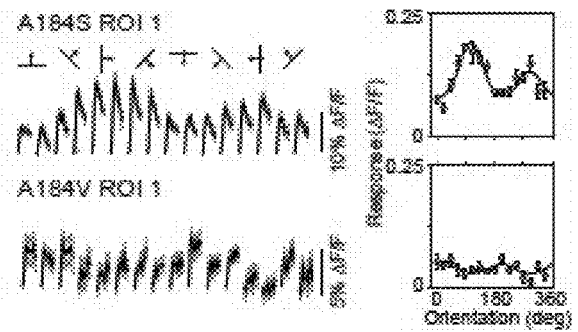


FIG. 53B

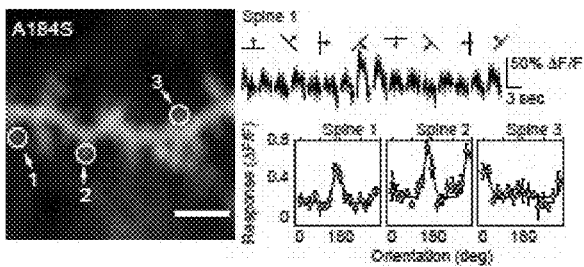


FIG. 53C

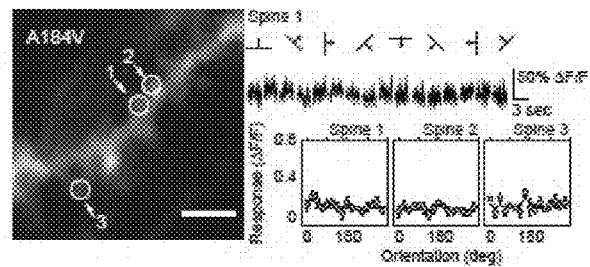


FIG. 53D

FIG. 54A

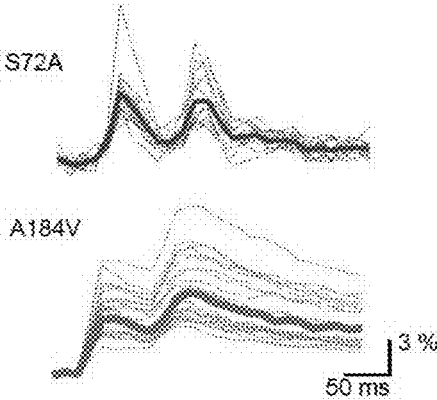


FIG. 54B

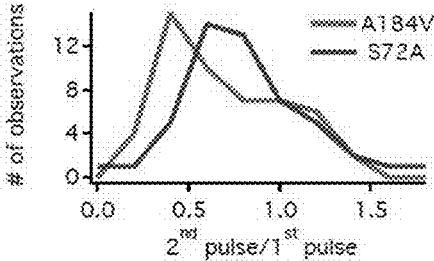


FIG. 54C

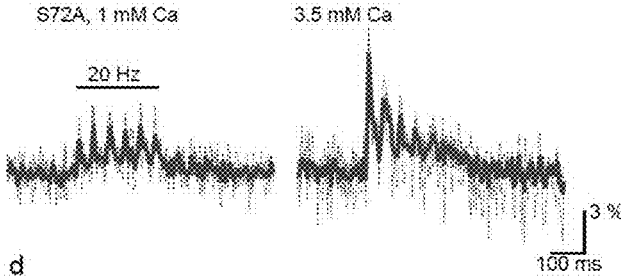
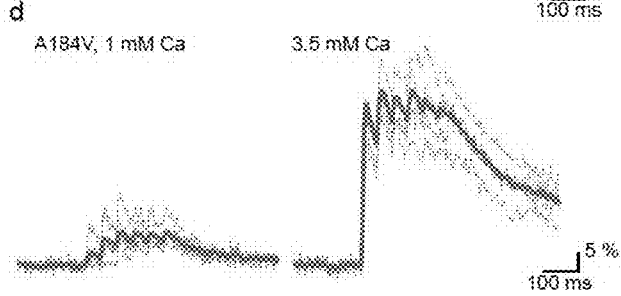


FIG. 54D



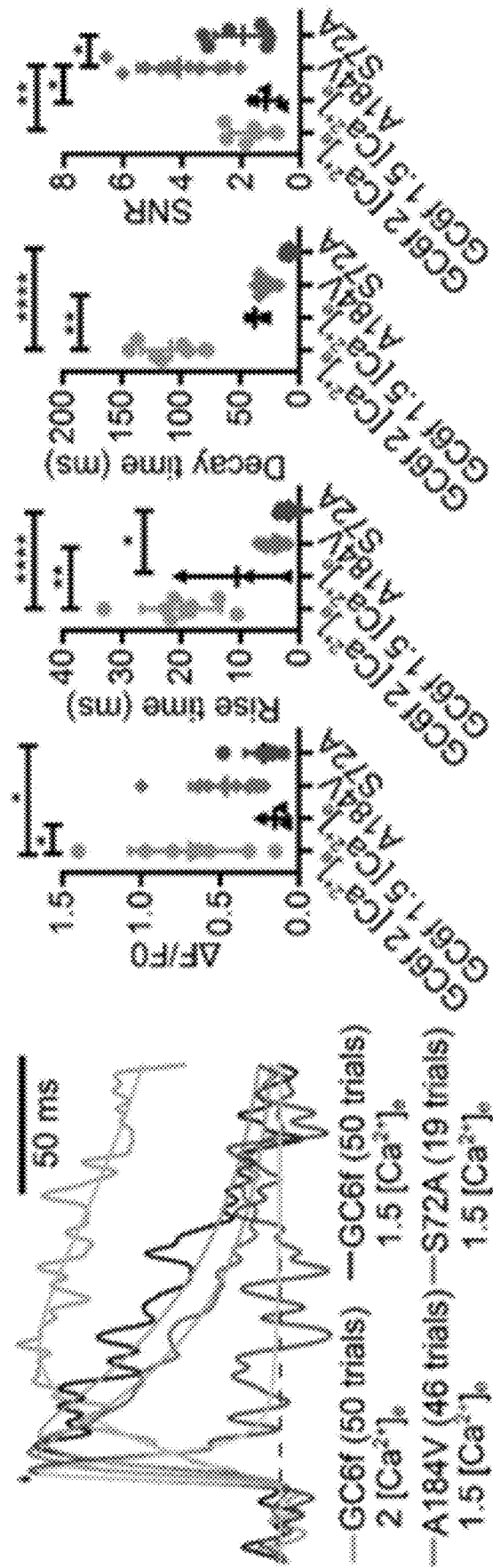


FIG. 55A

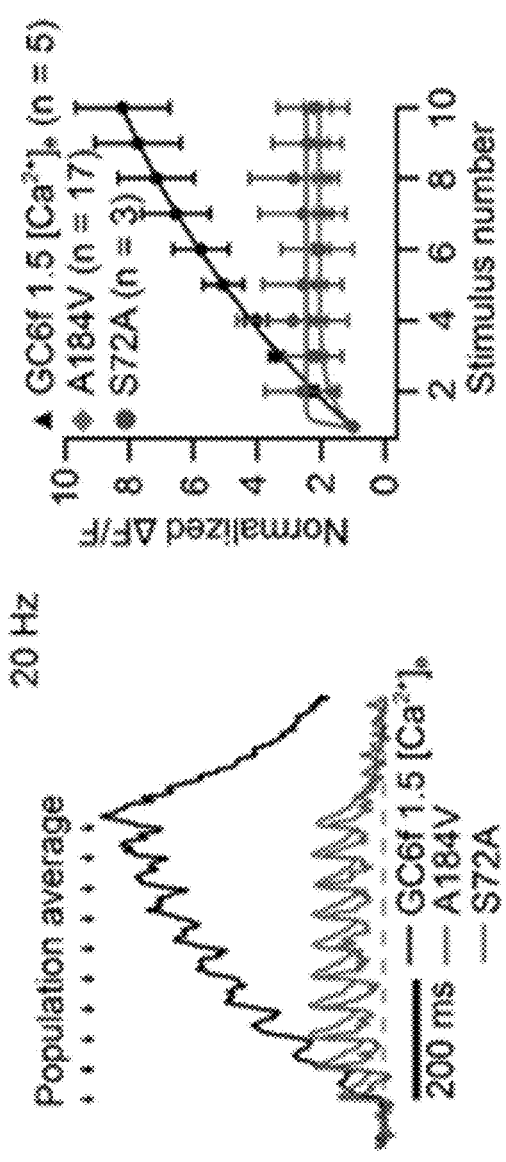


FIG. 55C

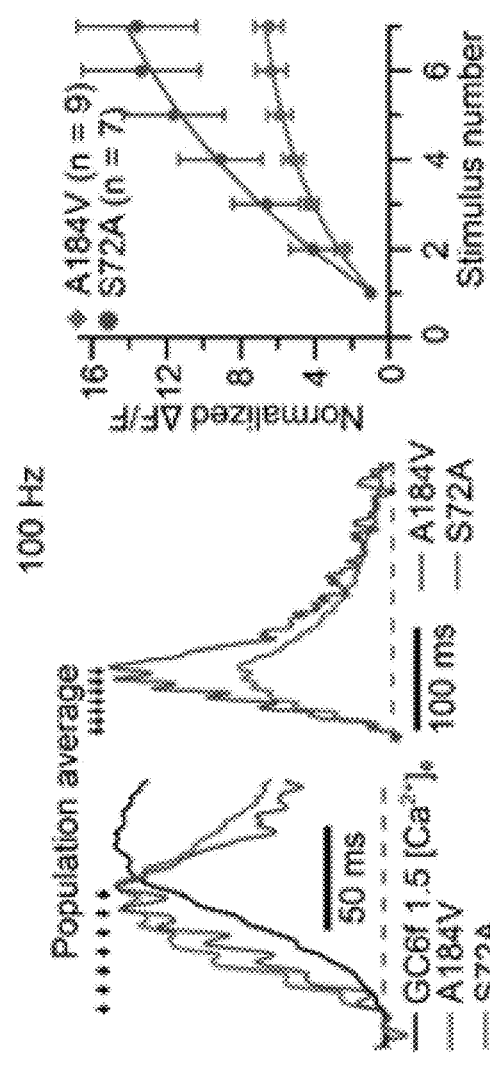


FIG. 55D

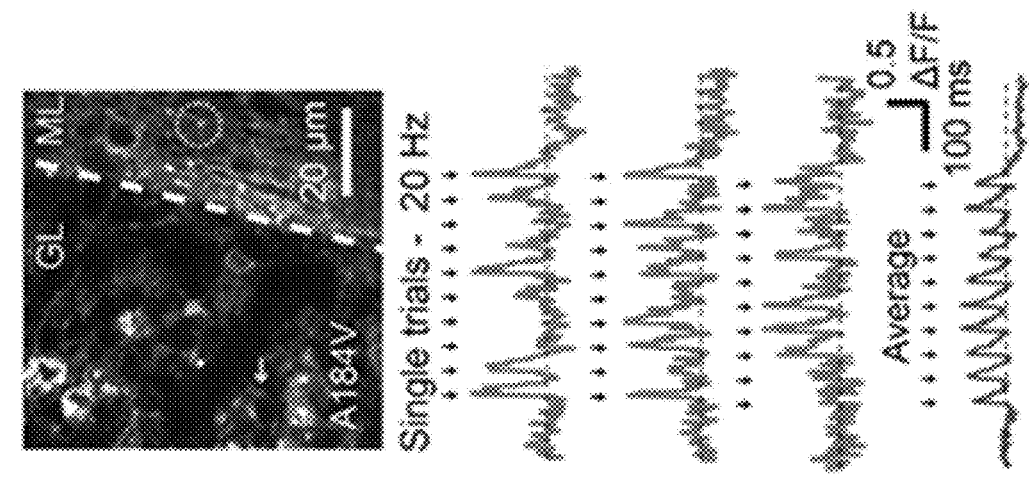


FIG. 55B

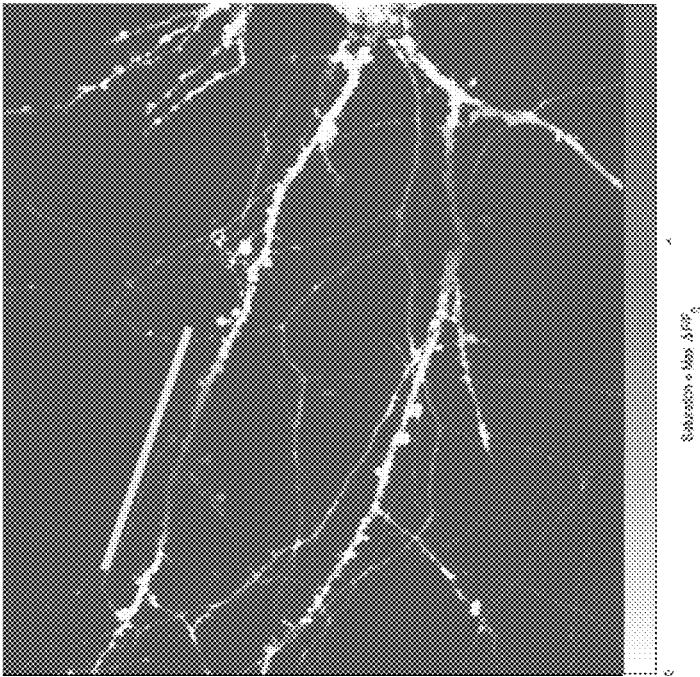


FIG. 56A

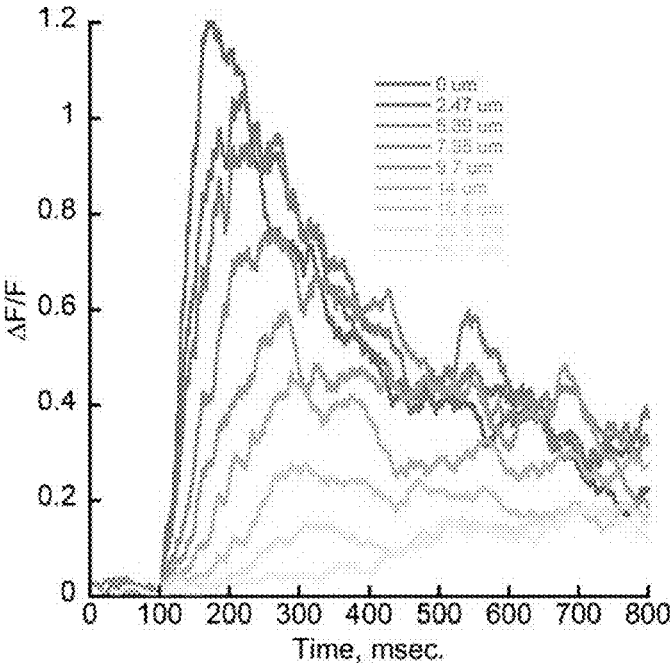


FIG. 56B

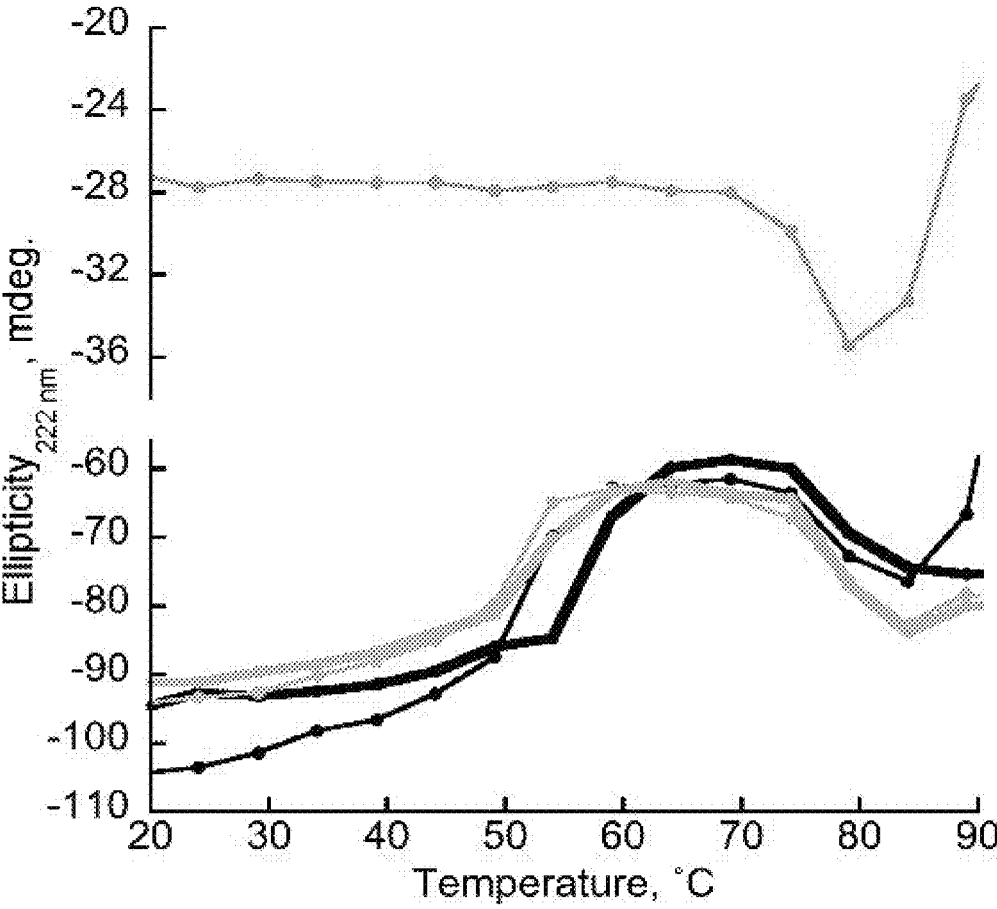


FIG. 57

FIG. 58A

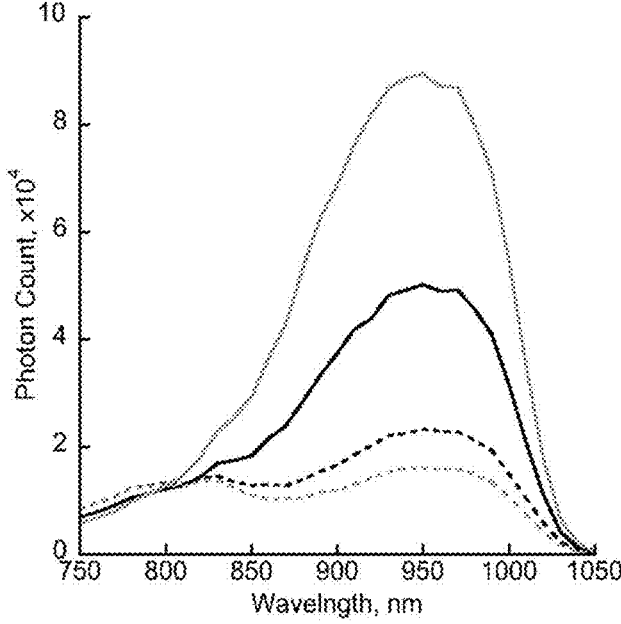


FIG. 58B

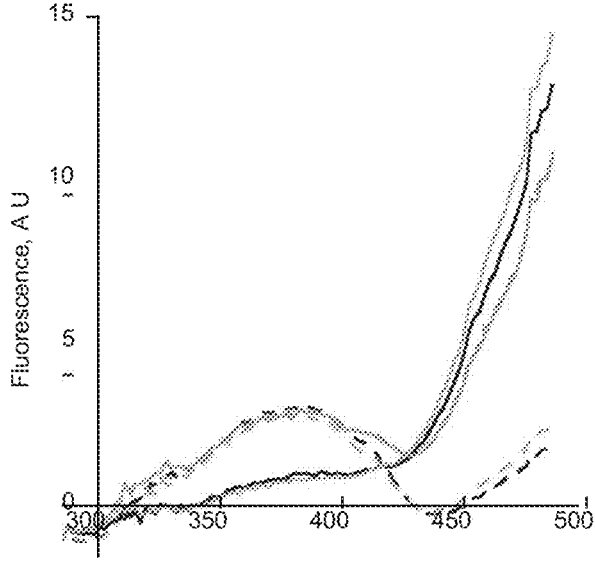


FIG. 58C

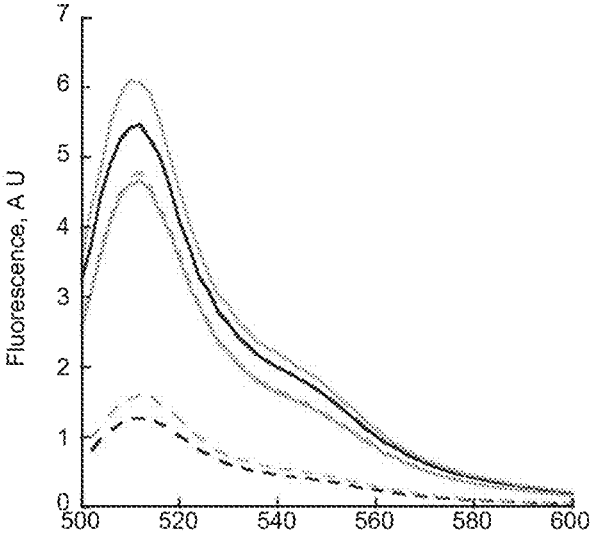
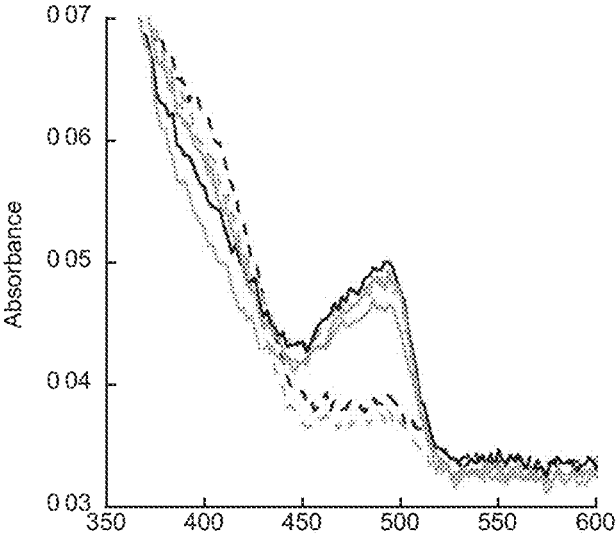


FIG. 58D



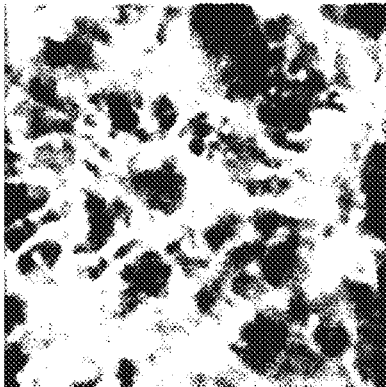


FIG. 59A

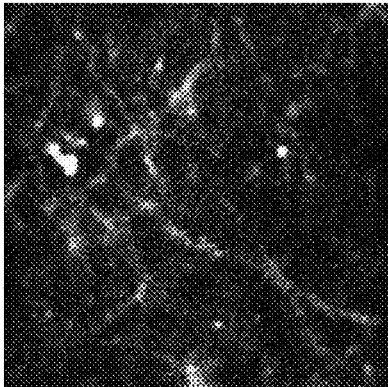


FIG. 59B

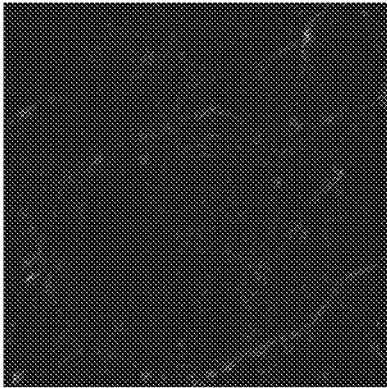


FIG. 59C

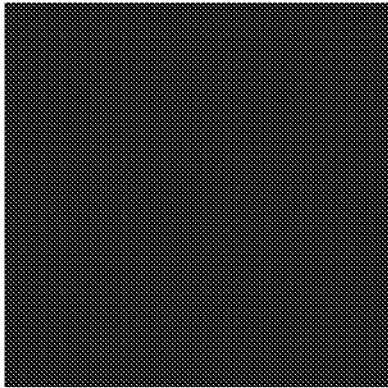


FIG. 59D

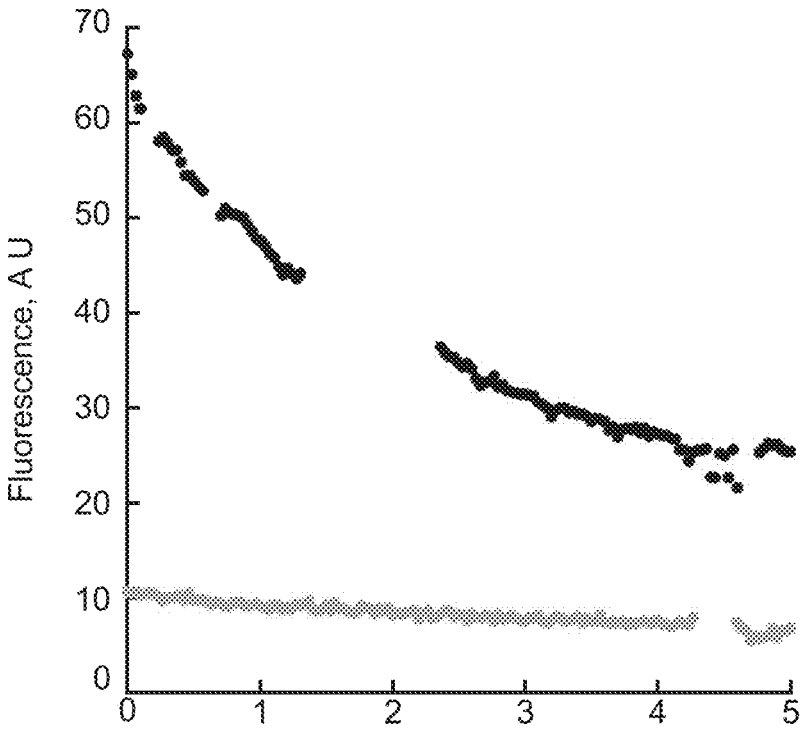


FIG. 59E

FIG. 60A

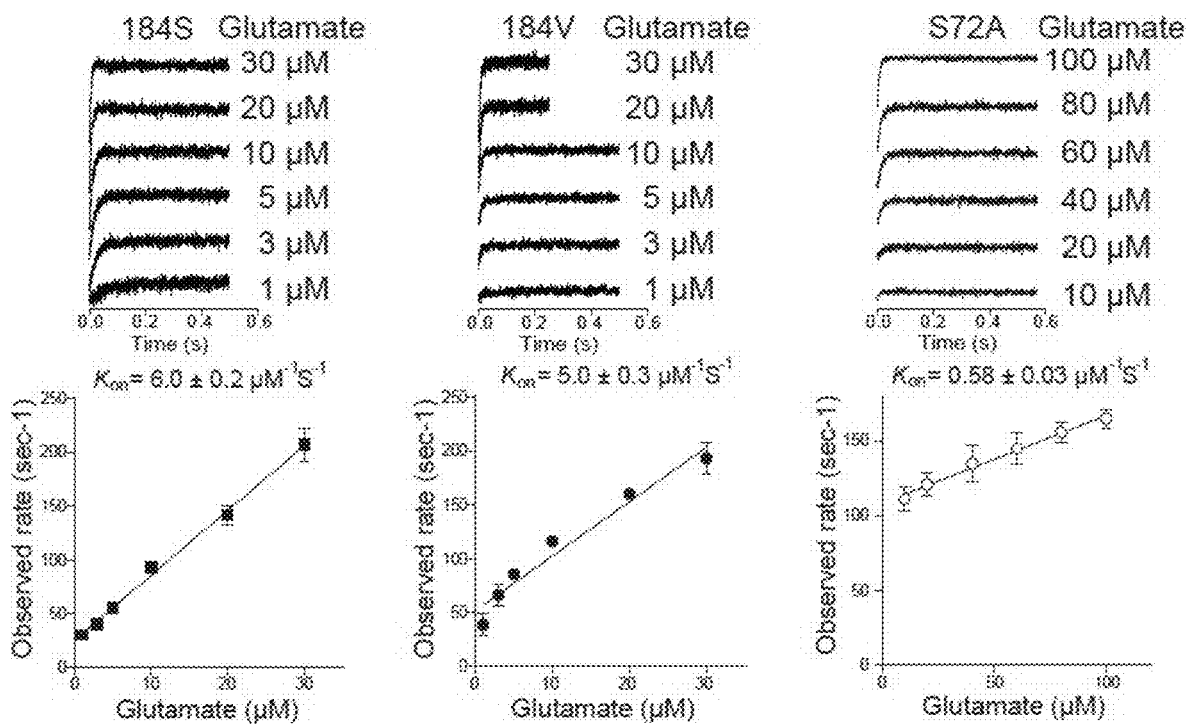
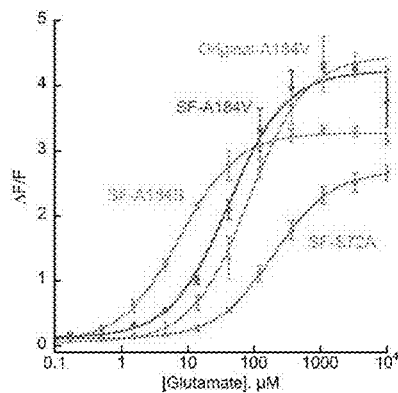


FIG. 60B

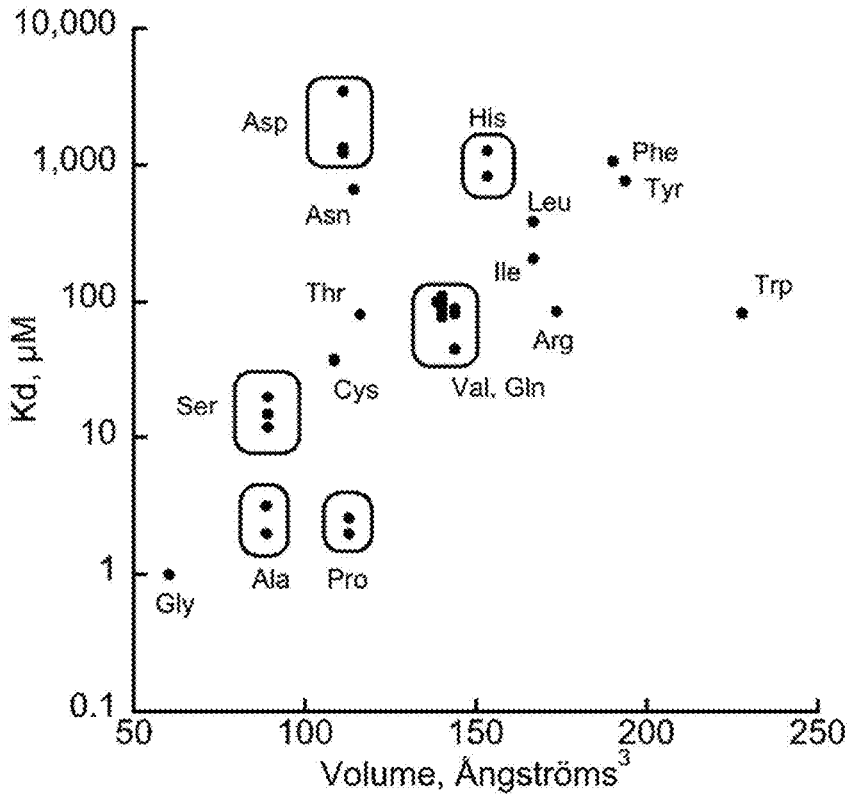


FIG. 61

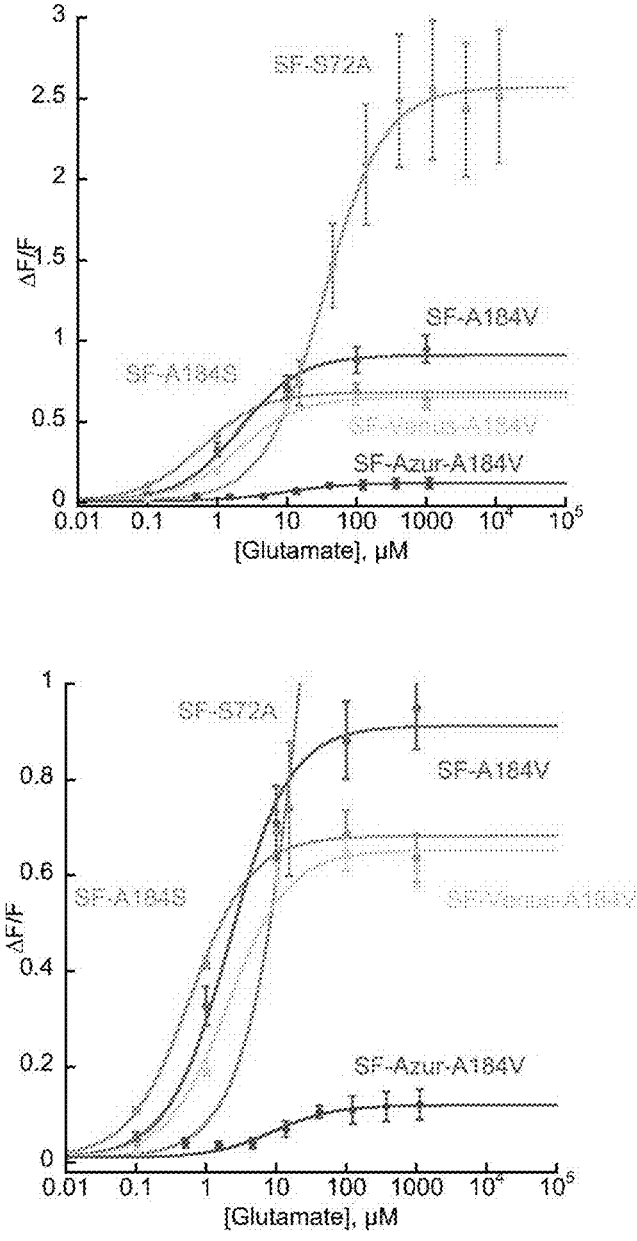


FIG. 62

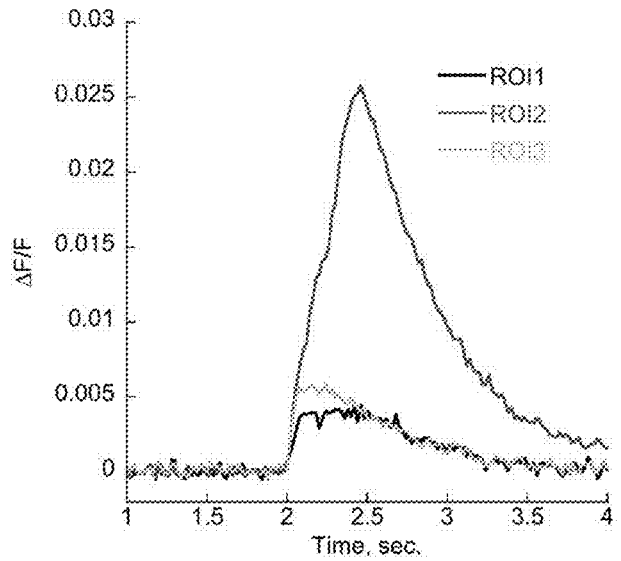
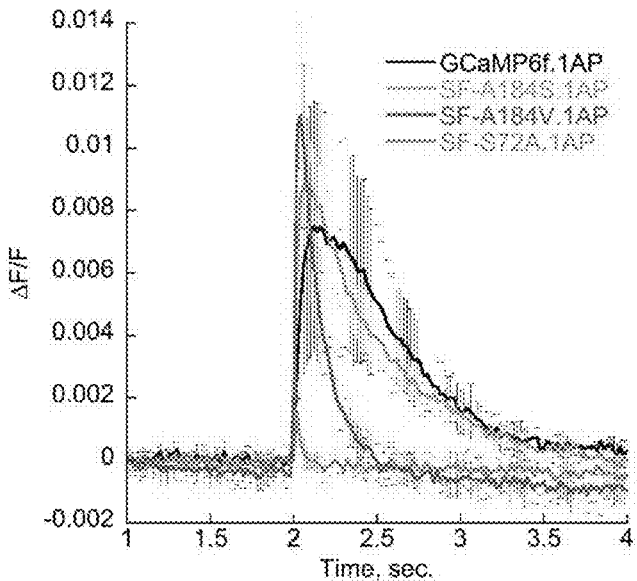
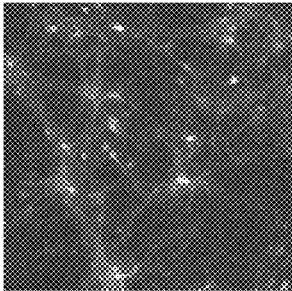
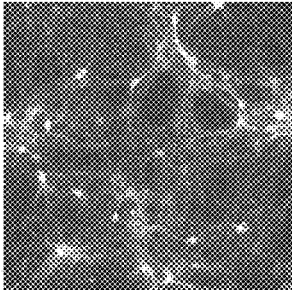


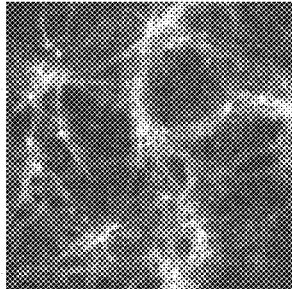
FIG. 63A



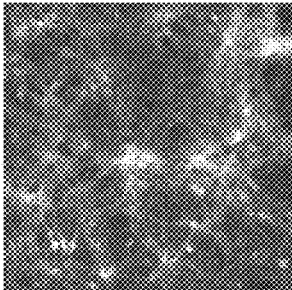
GCaMP6f



A184Sf



A184V



S72A

FIG. 63B

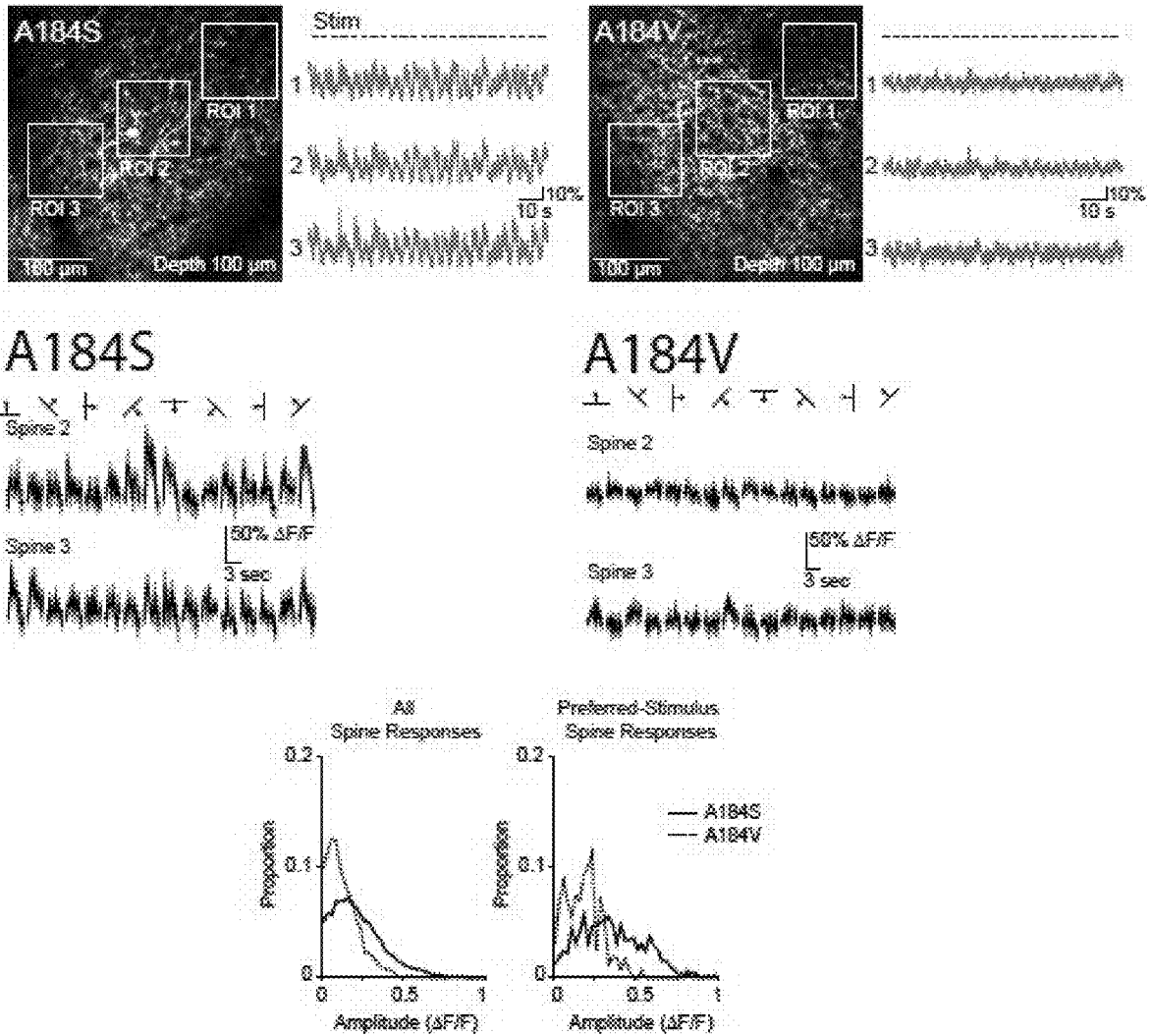


FIG. 64

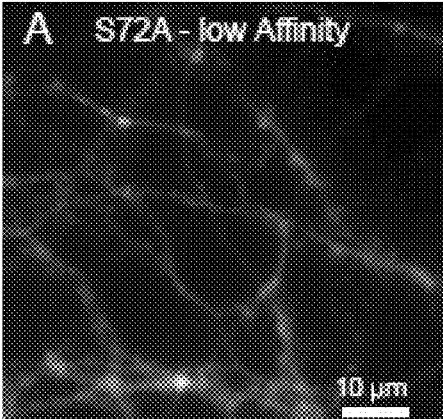


FIG. 65A

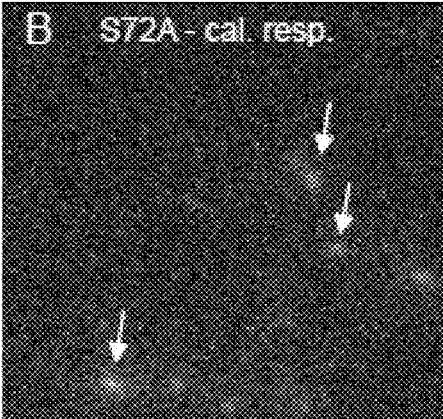


FIG. 65B

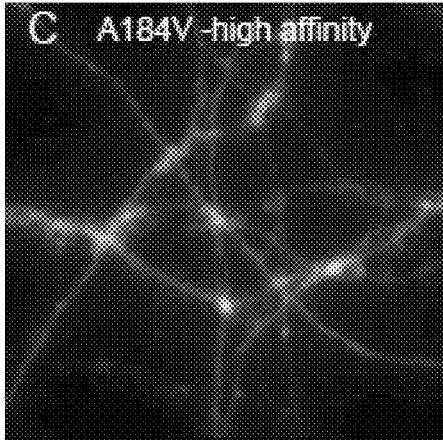


FIG. 65C

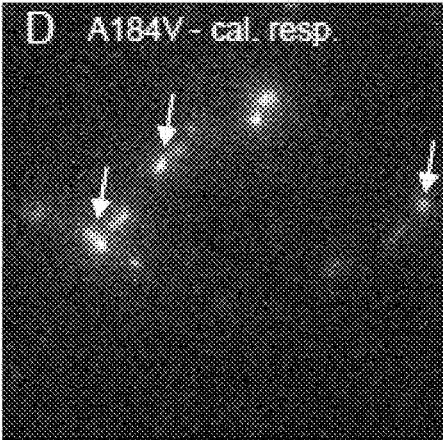


FIG. 65D

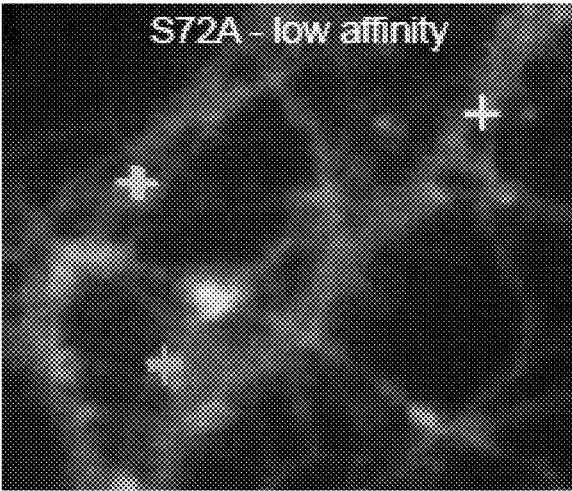


FIG. 66A

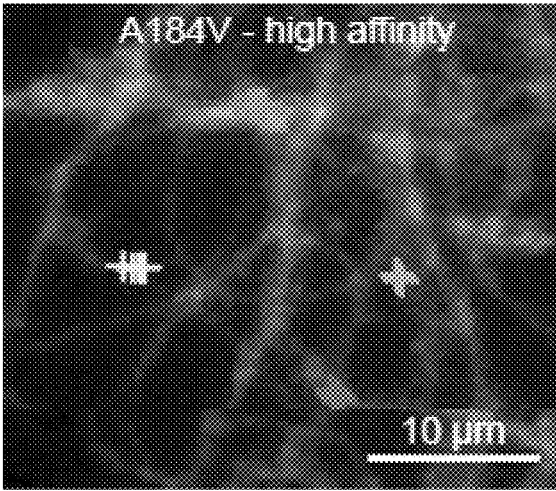


FIG. 66B

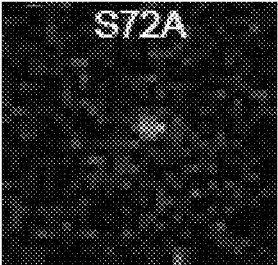


FIG. 66C

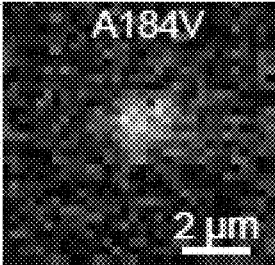


FIG. 66D

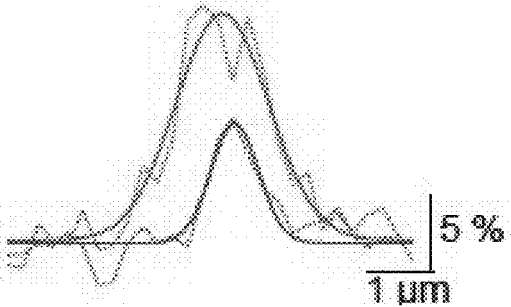


FIG. 66E

FIG. 66I

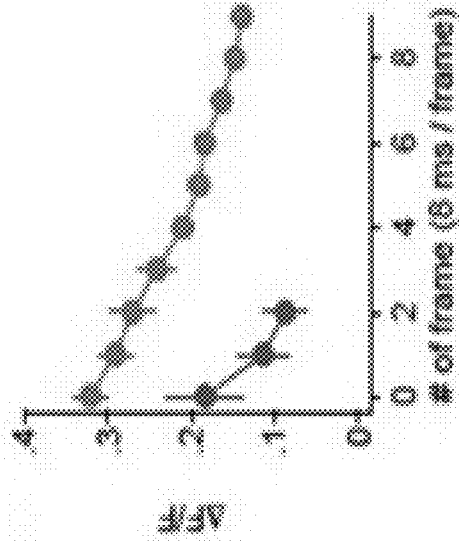


FIG. 66H

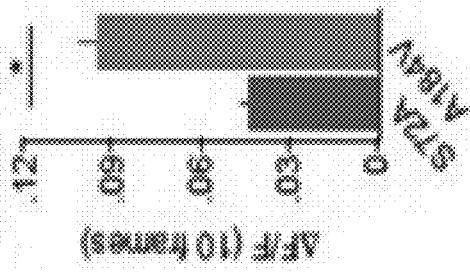


FIG. 66G

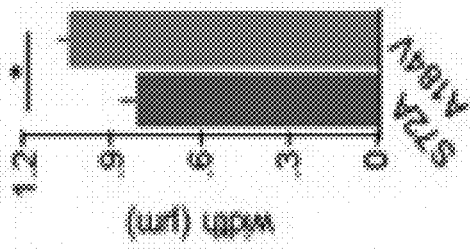


FIG. 66F

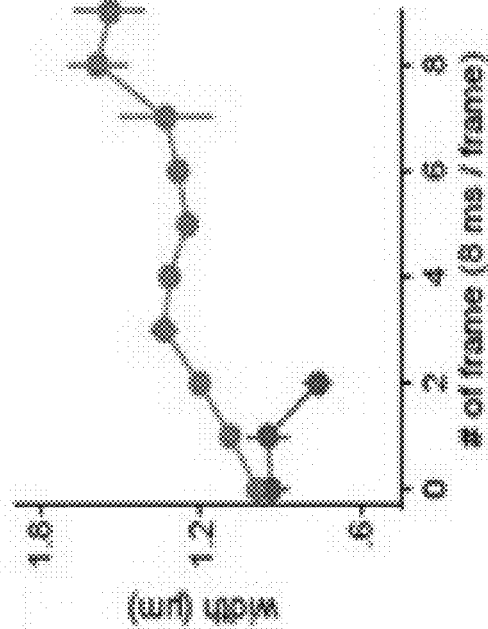
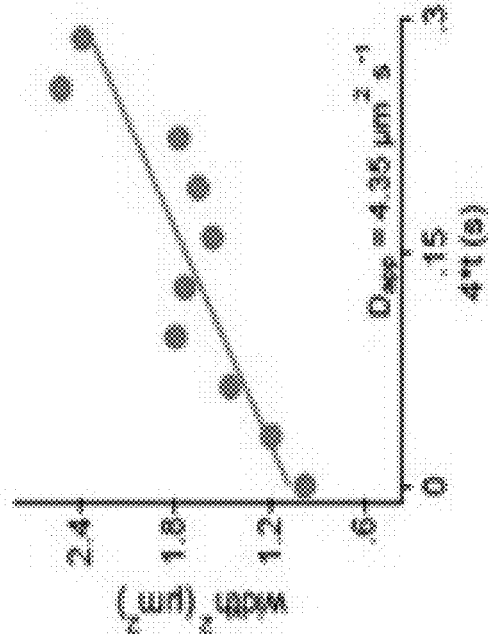
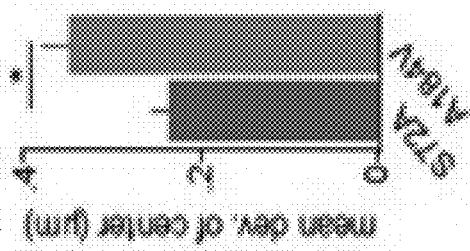


FIG. 66J



FIG. 67A

B SF-Venus-iGluSnFR
Mutations in GFP: T203Y and Y65G to shift the color. Mutations F46L and S72A to increase chromophore maturation.



FIG. 67B

SF-Azurite-iGluSnFR

Mutations in GFP: T65S and Y66H to shift the color. V150I and V224R to improve maturation and brightness.

Linker1 mutations: GltI-cpSFGFP connection from P₁LV₁NV to P₁LC₁NV

Linker2 mutations: cpSFGFP-GltI connection from YNFNN₂N to YNFNE₂N

MTFDLLEWVLEWVEGSGERS
[REDACTED]
[REDACTED]
[REDACTED]
[REDACTED]
[REDACTED]
[REDACTED]
[REDACTED]GGTEGS[REDACTED]
[REDACTED]
[REDACTED]
[REDACTED]LQVDEOK
LISEEDLNAVG
QDTQEVIVPHSLPFKVVVISAILALVVLTIIISLIIILMLWQKKPK

FIG. 67C

FIG. 68A

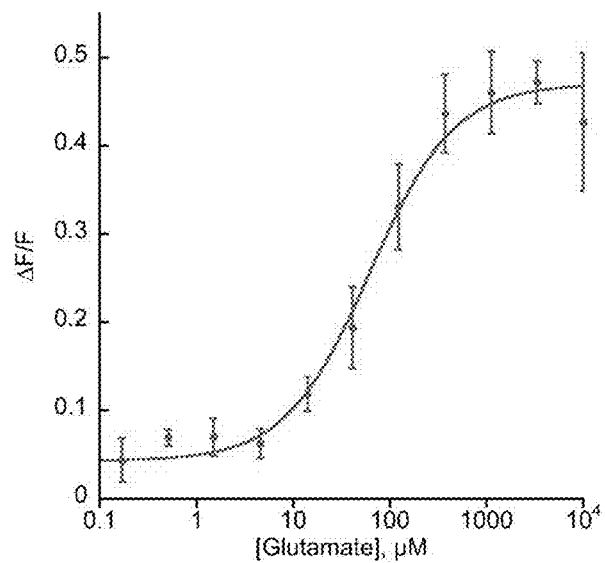


FIG. 68B

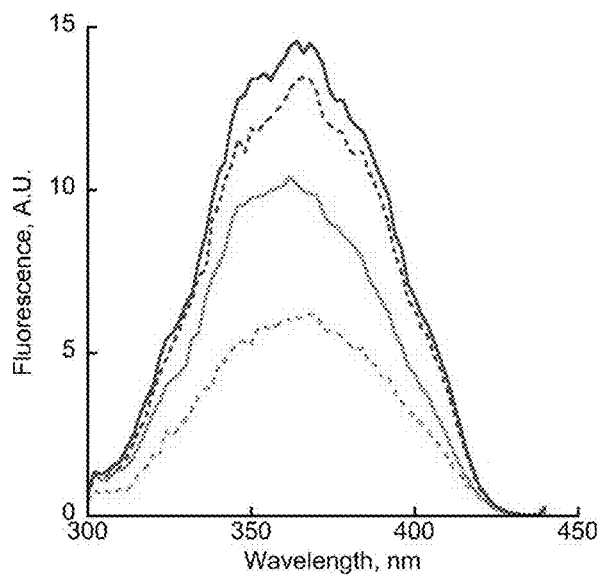


FIG. 68C

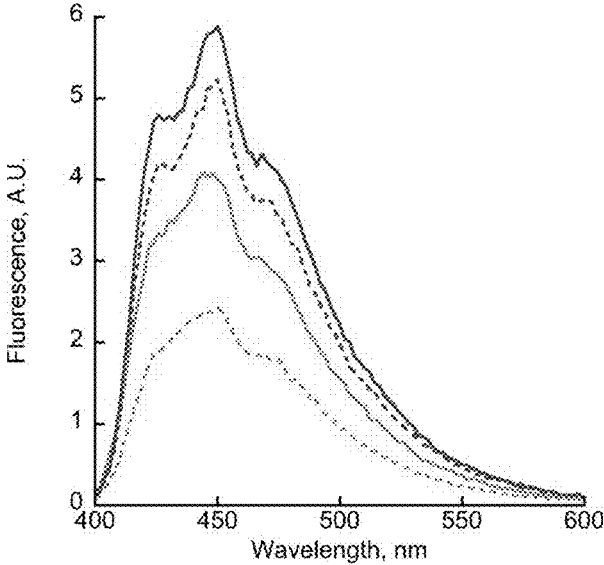


FIG. 68D

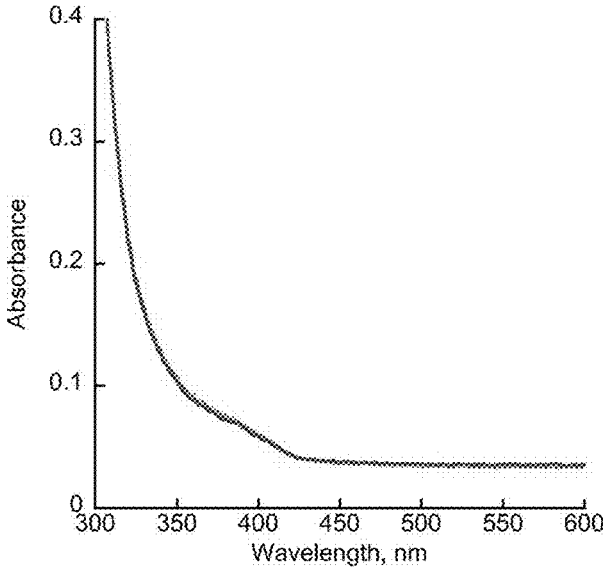


FIG. 69A

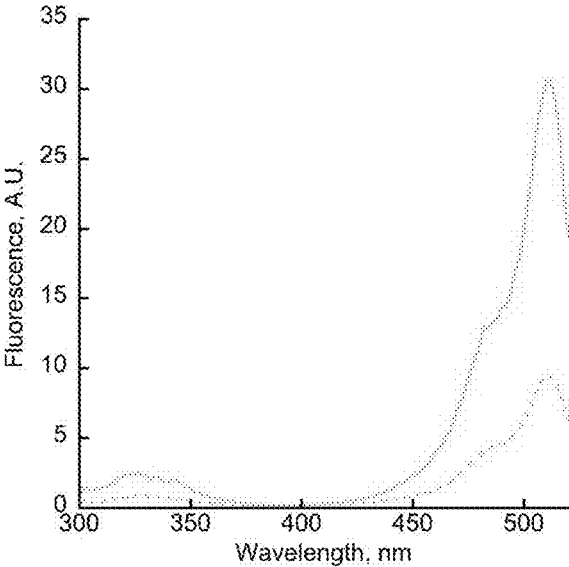


FIG. 69B

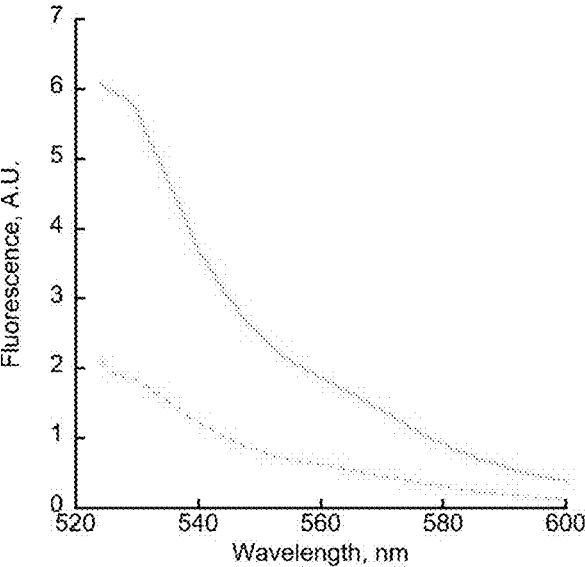


FIG 69C

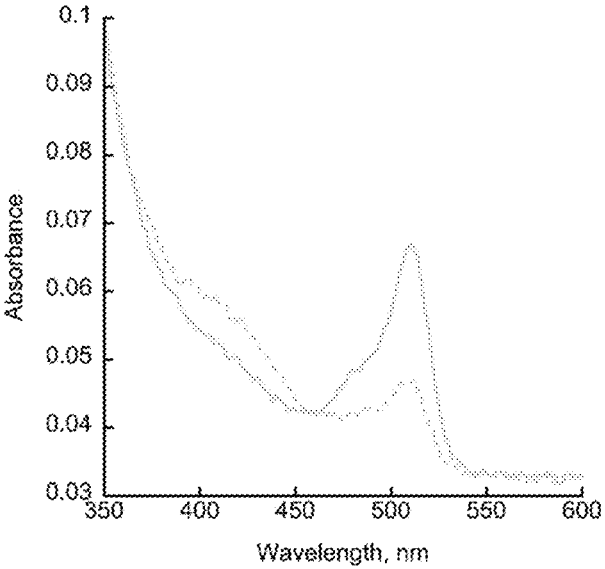
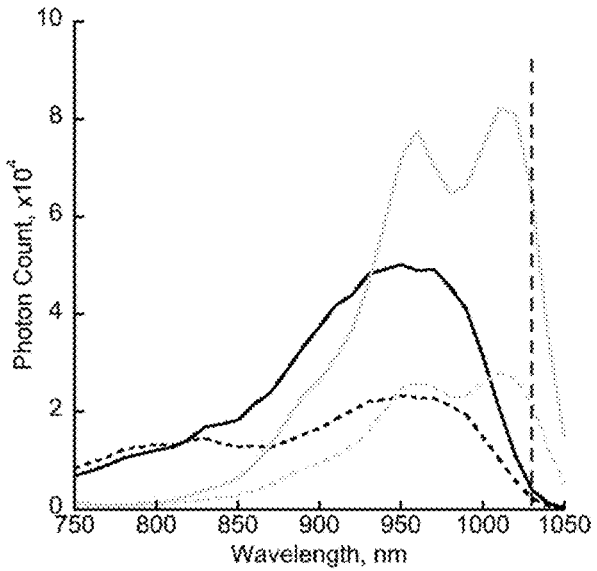


FIG. 69D



SF-iGluSnFR.A184V (SEQ ID NO: 176)

METDTELLLVVLLLVVPGSTGDRSAAAGSTLDKIAKNGVIVVGHRESSVVPFSYYDNQOKVVGYSQDYSNATV
EAVKKKLNKFDLQVKLIPITSONRIPILLQNGTFEEFCGSTTNNVERQKQAAFSDTIEVVGTRLLTKKGGD
IKDFANLKDKAVVVTSGTTSEVILNKINEEQKMMRILISAKDHGDSFRTLESGRAVAFMMDVLLAGERA
KAKKPDNWEIVGKPGSQBAYGCMLRKDDPQFKKLMDDTIAQVQTSGEAEKWEDKWEKNEPLVSHNVYITA
KOKNGIKANEKIRHNVEDSVOADHYOQNTFEGDGEVLLPDNHYLSTOSVLSKDPNEKRDHMVILEF
AAAGTTCMDELVGGTGGSSKQPELFTGVEELVEEDGAVNGHKFSVRGECEGATNCKILMPEICT
SKLVEVPAFTLVVTLIVGVCESFYEDHNEQHDPEKSAMPEAVQERTISKEDDGYKTRAEVKEEGDTLY
NRTELKGDIDKEDGNILCHKLEYWNNPFLNMNEELSDEMKALEKPEPNDKALKLQVDEQKLISEEDLNAV
QDTQEVIVVPHSLPEFKVVVISAILALVVVLTIIISLIILIMLWQKKPR

SF-iGluSnFR.A184S (SEQ ID NO: 177)

METDTELLLVVLLLVVPGSTGDRSAAAGSTLDKIAKNGVIVVGHRESSVVPFSYYDNQOKVVGYSQDYSNATV
EAVKKKLNKFDLQVKLIPITSONRIPILLQNGTFEEFCGSTTNNVERQKQAAFSDTIEVVGTRLLTKKGGD
IKDFANLKDKAVVVTSGTTSEVILNKINEEQKMMRILISAKDHGDSFRTLESGRAVAFMMDPSLLAGERA
KAKKPDNWEIVGKPGSQBAYGCMLRKDDPQFKKLMDDTIAQVQTSGEAEKWEDKWEKNEPLVSHNVYITA
KOKNGIKANEKIRHNVEDSVOADHYOQNTFEGDGEVLLPDNHYLSTOSVLSKDPNEKRDHMVILEF
AAAGTTCMDELVGGTGGSSKQPELFTGVEELVEEDGAVNGHKFSVRGECEGATNCKILMPEICT
SKLVEVPAFTLVVTLIVGVCESFYEDHNEQHDPEKSAMPEAVQERTISKEDDGYKTRAEVKEEGDTLY
NRTELKGDIDKEDGNILCHKLEYWNNPFLNMNEELSDEMKALEKPEPNDKALKLQVDEQKLISEEDLNAV
QDTQEVIVVPHSLPEFKVVVISAILALVVVLTIIISLIILIMLWQKKPR

FIG. 70A

SF-iGluSnFR.S72A (SEQ ID NO: 178)

METDTLLLWVLLLWVPGSTGDRS AAGSTLDKIAKNGVIVVGHRESSVPPFSYYDNOCKVVGYSQDYSNATV
EAVKKKLNKPDLOVKLIPIITSONRIPLLONCTFDPECGSTTNNVEROKQAAFSDTIEVVGTRLLTKKGGD
IKDFANLKDKAVVVTSGTTSEVILNKLNFEQKMMRIISAKDHGDSFRTLESCRAVAFMMDVILAGERA
KAKKPNWEIVGKFGSQRAYGCMERKDDPOFKKLMDDTIAQVOTSGEAEKWEDKWEKNIIVSNHWYITK
IKQKNGIKANFKIRHNVEDGAVQLADHYQONTPEGDPVLLIPDNHYLSYQSVLSKDPENKRDHMVILEEV
IAAGITLGMDELKGGTGGSSRSGEELFTGVVPELVEEDGCVNGHKFSVRGEGEGDAPNGKLTLLKICT
KLRVWFETLVTTLYGVQCFASYPDMKQDDEKSAHPGEGVQERTISPKDDGTAKTRAEVKEEGDTLV
NRLEKGIIDFKEDGNLGHKLEYNFNPLMNMNFELSDEMKALEKPEPKALKLQVDEQKLISEEDLNAV
GQDTQEVIVVPHSLPFKVVVISAILALVVLTIISLIILIMLWQKKPR

SF-Venus-iGluSnFR.A184V (SEQ ID NO: 179)

METDTLLLWVLLLWVPGSTGDRS AAGSTLDKIAKNGVIVVGHRESSVPPFSYYDNOCKVVGYSQDYSNATV
EAVKKKLNKPDLOVKLIPIITSONRIPLLONCTFDPECGSTTNNVEROKQAAFSDTIEVVGTRLLTKKGGD
IKDFANLKDKAVVVTSGTTSEVILNKLNFEQKMMRIISAKDHGDSFRTLESCRAVAFMMDVILAGERA
KAKKPNWEIVGKFGSQRAYGCMERKDDPOFKKLMDDTIAQVOTSGEAEKWEDKWEKNIIVSNHWYITK
IKQKNGIKANFKIRHNVEDGAVQLADHYQONTPEGDPVLLIPDNHYLSYQSVLSKDPENKRDHMVILEEV
IAAGITLGMDELKGGTGGSSRSGEELFTGVVPELVEEDGCVNGHKFSVRGEGEGDAPNGKLTLLKICT
KLRVWFETLVTTLYGVQCFASYPDMKQDDEKSAHPGEGVQERTISPKDDGTAKTRAEVKEEGDTLV
NRLEKGIIDFKEDGNLGHKLEYNFNPLMNMNFELSDEMKALEKPEPKALKLQVDEQKLISEEDLNAV
GQDTQEVIVVPHSLPFKVVVISAILALVVLTIISLIILIMLWQKKPR

FIG. 70B

SF-Venus-iGluSnFR.A184S (SEQ ID NO: 180)

METDTLLLWVLLLWVPGSTGDRS AAGSTLDKIAKNGVIVVGHRESSVPPFSYYDNQOKVVGYSQDYSNATV
EAVKKKLNKPFLOVKLIPTITSONRIPLLONGTFDFECGSTTNNVEROKQAAPSDTIEVVGTRILLTKKGGD
IKDFANLKDKAVVVTSGTTSEVILNKLNNEEQKMMMRIISAKDHGDSFRTLES CRAVAFAFMDDSLLAGERA
KAKKPNWEIVGKPGSQRAYGCMIRKDDPQFKKLMDDTIAQVQTSGEAEKWFEDKWEKNEIEVSHNYITD
IKQKNGIKANFKIHHNVEEESVQLADHYQONTPEGGGEVLLPNNHYLSYQSNLSKDPNEKRDHMVLEFV
LAAGITLGMDELY GGTGGS MSYGEELFTGVVEIIVELDESDANGHKEFVRGECEGADPNCKILKLICT
SKLIVPWFNLTLELVGVCQFARYFDHMKQHDNEKSAHPESYQERTISEDDGTAKIRAEVKEEGDTLV
NRTELKGIPTFKEDGNILGHKLEINFRNFINMNEELSDEMKA LKPEPNDKALKLOVDEQKLISEEDLNAV
QDTQEVIVVPHSLPFEKVVVISAILALVVLTIISLIILIMLWQKKPR

SF-Venus-iGluSnFR.S72A (SEQ ID NO: 181)

METDTLLLWVLLLWVPGSTGDRS AAGSTLDKIAKNGVIVVGHRESSVPPFSYYDNQOKVVGYSQDYSNATV
EAVKKKLNKPFLOVKLIPTITSONRIPLLONGTFDFECGSTTNNVEROKQAAPSDTIEVVGTRILLTKKGGD
IKDFANLKDKAVVVTSGTTSEVILNKLNNEEQKMMMRIISAKDHGDSFRTLES CRAVAFAFMDDVLLAGERA
KAKKPNWEIVGKPGSQRAYGCMIRKDDPQFKKLMDDTIAQVQTSGEAEKWFEDKWEKNEIEVSHNYITD
IKQKNGIKANFKIHHNVEEESVQLADHYQONTPEGGGEVLLPNNHYLSYQSNLSKDPNEKRDHMVLEFV
LAAGITLGMDELY GGTGGS MSYGEELFTGVVEIIVELDESDVNGHKEFVRGECEGADPNCKILKLICT
SKLIVPWFNLTLELVGVCQFARYFDHMKQHDNEKSAHPESYQERTISEDDGTAKIRAEVKEEGDTLV
NRTELKGIPTFKEDGNILGHKLEINFRNFINMNEELSDEMKA LKPEPNDKALKLOVDEQKLISEEDLNAV
QDTQEVIVVPHSLPFEKVVVISAILALVVLTIISLIILIMLWQKKPR

FIG. 70C

SF-Azurite-iGluSnFR (SEQ ID NO: 182)

METDTLLLWVLLLWVPGSTGDRS LAGSTLDKIARNGVIVVGHRESSVPPFSYYDMOOKVVGYSODYSNATV
EAVKXKLNKPDLOVKLIPIITSONRIFLLONGTDFEFCGGSTTNNVEROKOAAFSDTIFVVGTRLLTKKGGD
IKDFANLKDKAVVVISCTTSEVLLNKLNEEOKMMNRILISAKDHGDSFRTLESGRPAVAFMMDDVILAGERA
KAKKPNWEIVGKFOEQEAYGCFLRKDDPQFKLMDDTIAQVQTSGEAEKWFDKWFKNPILGYNLYTIA
IKOKNGIKANFKIRHNVEDSSVQLADHYQONTPTGDPVLEPQNHYLSTQSVLSKDPNEKRDHNVLEFF
LAAGTTEGMDELY GGTGGS SNGSELTGVPELWELGSDVNGHKTSVRGECEGDATNGKILKPIIT
KLIPVFWETLVTTLGHSVQCFSRYPDHMKQDDEKSAHPGYSVQERTISTKDDGTYKTRAEVKFESDTE
NRILEKSIDFKEDGNILGHKLEYNFNGLNMMNFELSDMKALFKPEPNDKALKLQVDEQKLISEEDLNAV
QDTQEVIVVPHSLPFKVVVISAILALVVLTIISLIIILIMLWQKKPR

iDexSnFR (SF-GlucoseSensor) (SEQ ID NO: 183)

METDTLLLWVLLLWVPGSTGDRS KLEIFSWWAGDEGPALEALIRLYKQKYPGVEVINATVTGGAGVNARA
VLKTRMLGGDPPDTFQVAAGMELIGTWVVANRMEDLSALFRQEGWLQAFPKGLIDLISYKGGIWSVPVNI
HRSNVMWYLPKCLKEWGVNPPRTWDEFLATCOTLKQKGLEAPLALGENWTQOHLWESVALAVLGPDDWNN
LWNGKLFKFTDPKAVRAWEVFGRVLDCANKDAAGLSWQQAVDRVVQKAAFNVMGDWAAGYMTTTLKLPK
TDFAWAPSPGTQGVFMMLSDSFGLPKGAKNRQAINWLRVGSKEGQDTFNPLKGSTAARLSDSPSKYPA
SENVYITADKQKNGILANPKIRHNVEFGSVQLADHYQONTPTGDPVLEPQNHYLSTQSVLSKDPNEKRL
HMVLEFFVTAAGTTEGMDELY GGTGGS MSKGLLELTGVPELWELGSDVNGHKTSVRGECEGDATNGK
ILKPIITGKLEYEWEPELVTLETYGVQCFSRYPDHMKQDDEKSAHPGYSVQERTISTKDDGTYKTRAEV
TEGDTLVNFTLKGIDFADGRILGHKLEYNFNPNAYGQSAMRDWRSNRIVGSLVAGAVAPESFMSQF
GTVMEIFLQTRNFOAAAANAQAATADQVGLGRLGQLQVDEQKLISEEDLNAVQDTQEVIVVPHSLPFKVV
VISAILALVVLTIISLIIILIMLWQKKPR

FIG. 70D

iGABASnFR (SEQ ID NO:184)

METDILLWVLLWVPGSTGDRSEINPVSNCGSTODAKQAWADPFSKASGITVVQDGPDPYGLKAMV
ESENVCQNDVVDVEADFAALRAAAGLEPLEDFSVIQRDKIDPRFVSDHCVGSFLFSFVLGYNEGKLEASKP
QDWTALFDTKTYPKKALYKWPSFCVLELALLAEGVPADKLYPLELDRAFKKLDTIKKDIVNWGGGCAQSQ
QLLASGEVSMGQFVNGRIHALOEDGAPVGVSWKQMLVMADILVVPKGTKNKAAAMKFLASASSAKGQDDE
SALTAYAPVNIQSVQRLDLSQVFTADKQKNGMANFKIRHNVEDSSVOLADHYQONFAGSPVLELFDN
VILSTQSVLSKDPNEKRDHMLLEFVTAAGITLGMDELYGGTGGSSKGGEEFTGVVPIVVELLGDVNV
KFSVVRGEGEGDATNGKLTLLKFICTTGKLPVPWPTLVTTLTLYGVQCFSRYPDHMKQHDFKAMPEGYV
RFLSFKDDGTYKTRAEVKFEGDTLVNRIELKGIQKEDGNIILGHKLEYNFPPATT
DFAYWAKNGPAIATRWNEWLVKLOVDLQVDEOKLISEEDLNAVQDQTOEVIVVPHSLPFFKVVVISAILAL
VVLTIISLIILIMLWOKKPR

iAChSnFR

SEQ ID NO: 185

MHHHHHHGYPYDVPDYAGAQPARSANDTVVVGSIIIFTEGIIVANMVAEMIEAHTDLKVVRKLNLGGVNVN
FEAIKRGGANNGIDIYVEYTGHLVDILGFEPEPNVYITADKQKNGIKANFKIRHNVEDGSSVOLADHYQON
TPIGDGPVLLPDNHVILSTQSVLSKDPNEKRDHMLLEFVTAAGITLGMDELYKGGTGGSSMSKGEELFTGV
VPILVELDGDVNGHKFSVRGEGEGDATNGKLTLLKFICTTGKLPVPWPTLVTTLTLYGVQCFSRYPDHMKQH
DFFKSAMPEGYVQERTISFKDDGTYKTRAEVKFEGDTLVNRIELKGIQKEDGNIILGHKLEYNFPPATT
DPEGAYETVKKEYKRKWNIVWLKPLGFNNTYTLTVKDELAKQYNLKTFSDLAKISDKLILGATMFFLEGP
DGYPLQKLYNFKFKHTKSMDMGI RYTAIDNNEVQVIDAWATDGLLVSHKLIKILEDDKAFPPYYAAPII
RQDVLDKHPKLDVNLKLANQISLEEMQKLNKVDGEGQDPAKVAKVFLKEKGLILQVDEOKLISEEDLN

FIG. 70E

SEQ ID NO: 186

METDTLLLWVLLLWVPGSTGDRSANDTVVVGSIIFTEGIIVANMVAEMIEAHTDLKVVRKLNLGGVNVNF
EAIKRGGANNGIDIYVEYTGHGLVDILGFPEPNVYITADKQKNGIKANFKIRHNVEDGQVADHYQQNT
PIGDGPVLLPDNHYLSTQSVLSKDPNEKRDHMLLEFVTAAGITLGMDELYKGGTGGSMSKGEELFTGVV
PILVELDGDVNGHKFSVRGEGEGDATNGKLTLLKFICTTGKLPVPWPTLVTTTLTYGVQCFSRYPDHMKQHD
FFKSAMPEGYVQERTISFKDDGTYKTRAEVKFEGDTLVNRIELKIDFKEDGNILGHKLEYNFPPATTD
PEGAYETVKKEYKRKWNIVWLKPLGFNNTYTLTVKDELAKQYNLKTFSDLAKISDKLILGATMFFLEGPD
GYPGLQKLYNFKFKHTKSMDMGIRYTAIDNNEVQVIDAWATDGLLVSHKLEDDKAFFPPYAAPIIR
QDVLDKHPELKDVLNLANQISLEEMQKLNKVDGEGQDPAKVAKVKEFLKEKGLILQVDEQKLISEEDLNA
VGQDTQEVIVVPHSLPFKVVVISAILALVVLTIISLIILIMLWQKKPR


FIG. 70F

Structure I





Wherein:



“” is a framework portion;



“” is a first signaling portion;

“” is an optional linker;

wherein the signaling portion is present at a site within the framework portion that undergoes a conformational change upon interaction of the framework portion with an analyte

FIG. 71

GENETICALLY ENCODED BIOSENSORS

CROSS REFERENCE TO RELATED APPLICATIONS

[0001] This application is a Divisional of, and claims the benefit of priority under 35 U.S.C. § 121 to, U.S. application Ser. No. 16/902,160 filed Jun. 15, 2020, now allowed, which is a Continuation of, and claims priority under 35 U.S.C. § 121 to, U.S. application Ser. No. 16/002,697 filed Jun. 7, 2018, which is a Continuation-In-Part of, and claims priority under 35 U.S.C. § 120 to, U.S. application Ser. No. 15/904,574 filed Feb. 26, 2018, which is a Divisional application of, and claims the benefit of priority under 35 U.S.C. § 121 to, U.S. application Ser. No. 15/664,326 filed Jul. 31, 2017, which is a Divisional application of, and claims the benefit of priority under 35 U.S.C. § 121 to, U.S. application Ser. No. 14/350,199 filed Nov. 18, 2014, which is a U.S. National Phase application of, and claims the benefit of priority under 35 U.S.C. 371 to, International Application No. PCT/US2012/059219 filed Oct. 8, 2012, which claims the benefit of priority under 35 U.S.C. § 119(e) to U.S. Application No. 61/544,867 filed Oct. 7, 2011.

SEQUENCE LISTING

[0002] The instant application contains a Sequence Listing that has been submitted in ASCII format via EFS-Web and is hereby incorporated by reference in its entirety. The ASCII copy, created on 25 Oct. 2021, is named 30872_0020003_SEQ.txt, and is 254553 bytes in size.

TECHNICAL FIELD

[0003] This disclosure relates to genetically encoded biosensors and methods for the design, production, and use of such biosensors.

BACKGROUND

[0004] Protein-based sensors that transduce microscopic binding events into macroscopically observable signals are available to allow real-time visualization of a variety of biological events and/or molecules (Frommer et al., *Chem. Soc. Rev.*, 38:2833-2841, 2009). Such sensors can be targeted and/or expressed in living cells, tissues, and organisms, and permit imaging with minimally invasive techniques (Okumoto, *Curr. Opin. Biotechnol.*, 21:45-54, 2010). Application of these sensors is limited by the narrow range of analytes that can be detected and/or by their inability to distinguish signal over noise.

SUMMARY

[0005] In one aspect, a recombinant peptide biosensor is provided that includes an analyte-binding framework portion and a signaling portion, wherein the signaling portion is present within the framework portion at a site or amino acid position that undergoes a conformational change upon interaction of the framework portion with a defined, specific, or selected analyte.

[0006] In one embodiment, the signaling portion is allosterically regulated by the framework portion such that signaling from the signaling portion is altered upon interaction of the framework portion with the analyte. In some embodiments, signaling by the signaling portion detectably increases upon interaction of the framework portion with the

analyte. In some embodiments, signaling by the signaling portion detectably decreases upon interaction of the framework portion with the analyte. In some embodiments, signaling by the signaling portion is proportional to the level of interaction between the framework portion and the analyte.

[0007] In some embodiments, the signaling portion is a superfolder (SF) fluorescent protein (see, for example, Pedelacq et al., 2006, *Nature Biotechnol.*, 24:79-88), a protein that exhibits robust folding, even when fused to a protein that folds poorly. In some embodiments, the SF protein is circularly permuted. In some embodiments, the SF protein is a green fluorescent protein, a yellow fluorescent protein, a red fluorescent protein, or a blue fluorescent protein.

[0008] In some embodiments, the framework portion has a first structure in the absence of an analyte and a second structure, that is detectably distinct from the first structure, in the presence of the analyte. In some embodiments, the conformational change between the first structure and the second structure allosterically regulates the signaling portion. In some embodiments, the framework portion is a periplasmic binding protein (PBP) or a variant of a PBP.

[0009] In some embodiments, the analyte-binding framework portion binds specifically to an analyte selected from the group consisting of glucose, maltose, phosphonate, glutamate, GABA, and ACh.

[0010] In another aspect, a recombinant peptide biosensor is provided that includes an amino acid sequence having at least 90% identity to a recombinant peptide biosensor selected from the group consisting of SEQ ID NOs: 176-182, wherein the recombinant peptide biosensor binds specifically to glutamate.

[0011] In one embodiment, the recombinant peptide biosensor includes a recombinant peptide biosensor selected from the group consisting of SEQ ID NOs: 176-182 comprising 10 or fewer conservative amino acid substitutions, wherein the recombinant peptide biosensor binds specifically to glutamate. In some embodiments, the recombinant peptide biosensor includes a recombinant peptide biosensor selected from the group consisting of SEQ ID NOs: 176-182.

[0012] In still another aspect, a recombinant peptide biosensor is provided that includes an amino acid sequence having at least 90% identity to a recombinant peptide biosensor having the sequence shown in SEQ ID NO: 183, wherein the recombinant peptide biosensor binds specifically to glucose.

[0013] In some embodiments, the recombinant peptide biosensor includes a recombinant peptide biosensor having the sequence shown in SEQ ID NO: 183 comprising 10 or fewer conservative amino acid substitutions, wherein the recombinant peptide biosensor binds specifically to glucose. In some embodiments, the recombinant peptide biosensor includes a recombinant peptide biosensor having the sequence shown in SEQ ID NO: 183.

[0014] In one aspect, a recombinant peptide biosensor is provided that includes an amino acid sequence having at least 90% identity to a recombinant peptide biosensor having the sequence shown in SEQ ID NO: 184, wherein the recombinant peptide biosensor binds specifically to GABA.

[0015] In one embodiment, the recombinant peptide biosensor includes a recombinant peptide biosensor having the sequence shown in SEQ ID NO: 184 comprising 10 or fewer conservative amino acid substitutions, wherein the recom-

binant peptide biosensor binds specifically to GABA. In one embodiment, the recombinant peptide biosensor includes a recombinant peptide biosensor having the sequence shown in SEQ ID NO: 184.

[0016] In another aspect, a recombinant peptide biosensor is provided that includes an amino acid sequence having at least 90% identity to a recombinant peptide biosensor having a sequence selected from the group consisting of SEQ ID NO: 185 and 186, wherein the recombinant peptide biosensor binds specifically to ACh.

[0017] In one embodiment, the recombinant peptide biosensor includes a recombinant peptide biosensor having a sequence selected from the group consisting of SEQ ID NO: 185 and 186 comprising 10 or fewer conservative amino acid substitutions, wherein the recombinant peptide biosensor binds specifically to ACh. In one embodiment, the recombinant peptide biosensor includes a recombinant peptide biosensor having a sequence selected from the group consisting of SEQ ID NO: 185 and 186.

[0018] In one aspect, a nucleic acid is provided that encodes a recombinant peptide biosensor as described herein.

[0019] In one aspect, a vector is provided that includes a nucleic acid as described herein.

[0020] In one aspect, a cell is provided that includes a nucleic acid as described herein.

[0021] In one aspect, a cell is provided that includes a vector as described herein.

[0022] In one aspect, a kit is provided that includes a recombinant peptide biosensor as described herein, a nucleic acid as described herein, a vector as described herein, and/or the cell as described herein.

[0023] In still another aspect, a method is provided for detecting glutamate, the method comprising detecting a level of fluorescence emitted by a recombinant peptide biosensor, the peptide biosensor having an amino acid sequence selected from the group consisting of SEQ ID NOs: 176-182, and correlating the level of fluorescence with the presence of glutamate.

[0024] In some embodiments, the recombinant peptide biosensor is expressed from a nucleic acid. In some embodiments, the method includes contacting the recombinant peptide biosensor with a sample comprising glutamate. In some embodiments, the method includes correlating the level of fluorescence with a concentration of glutamate. In some embodiments, the method includes comparing the level of fluorescence with a level of fluorescence emitted by the recombinant peptide biosensor in the presence of a sample comprising a known concentration or range of concentrations of glutamate. In some embodiments, the method is performed in vitro.

[0025] In some aspects, a method for detecting glucose is provided, the method comprising detecting a level of fluorescence emitted by a recombinant peptide biosensor, the peptide biosensor having an amino acid sequence shown in SEQ ID NO: 183, and correlating the level of fluorescence with the presence of glucose.

[0026] In some aspects, a method for detecting GABA is provided, the method comprising detecting a level of fluorescence emitted by a recombinant peptide biosensor, the peptide biosensor having an amino acid sequence shown in SEQ ID NO: 184, and correlating the level of fluorescence with the presence of GABA.

[0027] In some aspects, a method for detecting ACh is provided, the method comprising detecting a level of fluorescence emitted by a recombinant peptide biosensor, the peptide biosensor having an amino acid sequence selected from the group consisting of SEQ ID NOs: 185 and 186, and correlating the level of fluorescence with the presence of ACh.

[0028] In some aspects, a method for detecting a defined, selected, or specific analyte is provided, the method comprising detecting a level of fluorescence emitted by a recombinant peptide biosensor of claim 1; and correlating the level of fluorescence with the presence of a defined, selected, or specific analyte

[0029] In some embodiments, the recombinant peptide biosensor is expressed from a nucleic acid. In some embodiments, the method includes contacting the recombinant peptide biosensor with a sample comprising the analyte. In some embodiments, the method includes correlating the level of fluorescence with a concentration of the analyte. In some embodiments, the method includes comparing the level of fluorescence with a level of fluorescence emitted by the recombinant peptide biosensor in the presence of a sample comprising a known concentration or range of concentrations of the analyte. In some embodiments, the method is performed in vitro. In some embodiments, the analyte is selected from the group consisting of glutamate, glucose, GABA, and ACh.

[0030] The present disclosure provides genetically encoded recombinant peptides containing an analyte-binding framework portion linked (e.g., operably linked) to a signaling portion, wherein the signaling portion is allosterically regulated by the framework portion upon interaction of the framework portion with an analyte (e.g., a defined, selected, and/or specific analyte). These constructs can be used as biosensors, e.g., to transduce microscopic binding events into macroscopically observable signals.

[0031] The present disclosure provides, in part, recombinant peptides for use as biosensors (e.g., recombinant peptide biosensors) that include (e.g., comprise, consist essentially of, or consist of), e.g., include at least, an analyte-binding framework portion and a signaling portion. As described in further detail herein, such signaling portions are present within the framework portion at a site or amino acid position that undergoes a conformational change (e.g., a conformational change sufficient to alter a physical and/or functional characteristic of the signaling portion, e.g., a substantial conformational change) upon interaction of the framework portion with a defined, specific, or selected analyte (e.g. such as an analyte to which the framework portion or a region thereof, and/or the biosensor, specifically binds).

[0032] For example, in some instances, the signaling portion is allosterically regulated by the framework portion such that signaling from the signaling portion is altered (e.g. wherein a first level of signaling is altered or changed to a second level of signaling that can be distinguished using routine methods of detection from the first) upon interaction of the framework portion with the analyte. In some instances, signaling by the signaling portion can detectably increase or decrease upon interaction of the framework portion with the analyte. In some instances, signaling by the signaling portion upon interaction of the biosensor with a defined, specific, or selected analyte (e.g. such as an analyte to which the framework portion or a region thereof, and/or

the biosensor, specifically binds) can be proportional or can correlate with to the level of interaction between the framework portion and the analyte such that the level of interaction can be determined from the signaling or alteration thereof.

[0033] In some instances, framework portions of the biosensors disclosed herein have a first structure in the absence of an analyte and a second structure that is detectably distinct from the first structure in the presence of the analyte. In some instances, the conformational change between the first structure and the second structure allosterically regulates the signaling portion.

[0034] In some instances, framework portions of the biosensors disclosed herein can be, or can include (e.g., comprise, consist essentially of, or consist of), periplasmic binding proteins (PBP) or variants of a PBP. In some instances, exemplary PBPs or variants thereof can include, but are not limited to, peptides with at least 90% identity to a peptide selected from the group consisting of SEQ ID NO:105, SEQ ID NO: 106, SEQ ID NO:107, SEQ ID NO:108, SEQ ID NO:109, SEQ ID NO: 110, SEQ ID NO:111, SEQ ID NO:113, and SEQ ID NO:114. In some instances, exemplary PBPs or variants thereof can include, but are not limited to, peptides with at least 95% identity to a peptide selected from the group consisting of SEQ ID NO:105, SEQ ID NO: 106, SEQ ID NO:107, SEQ ID NO:108, SEQ ID NO:109, SEQ ID NO: 110, SEQ ID NO:111, SEQ ID NO:113, and SEQ ID NO:114. In some instances, exemplary PBPs or variants thereof can include, but are not limited to, peptides selected from the group consisting of SEQ ID NO:105, SEQ ID NO: 106, SEQ ID NO:107, SEQ ID NO:108, SEQ ID NO:109, SEQ ID NO: 110, SEQ ID NO:111, SEQ ID NO:113, and SEQ ID NO:114. In some instances, exemplary PBPs or variants thereof can include, but are not limited to, peptides selected from the group consisting of SEQ ID NO:105, SEQ ID NO: 106, SEQ ID NO:107, SEQ ID NO:108, SEQ ID NO:109, SEQ ID NO: 110, SEQ ID NO:111, SEQ ID NO:113, and SEQ ID NO:114 comprising 10 or fewer conservative amino acid substitutions. PBPs or variants thereof disclosed herein can be truncated.

[0035] In some instances, signaling portions of the biosensors disclosed herein can be or can include (e.g., comprise, consist essentially of, or consist of) one or more (e.g., one, two three, four, five, and less than ten) circularly permuted fluorescent proteins (cpFPs). Such cpFPs can be included but are not limited to, for example, green fluorescent proteins, yellow fluorescent proteins, red fluorescent proteins, and/or blue fluorescent proteins.

[0036] In some instances, biosensors disclosed herein, e.g., analyte-binding framework portions of biosensors disclosed herein, can bind (e.g., bind specifically) to glucose. Such sensors can be referred to as glucose binding biosensors or glucose biosensors.

[0037] In some instances, biosensors disclosed herein, e.g., analyte-binding framework portions of biosensors disclosed herein, can bind (e.g., bind specifically) to maltose. Such sensors can be referred to as maltose binding biosensors or maltose biosensors.

[0038] In some instances, biosensors disclosed herein, e.g., analyte-binding framework portions of biosensors disclosed herein, can bind (e.g., bind specifically) to phosphonate.

[0039] Such sensors can be referred to as phosphonate binding biosensors or phosphonate biosensors.

[0040] In some instances, biosensors disclosed herein, e.g., analyte-binding framework portions of biosensors disclosed herein, can bind (e.g., bind specifically) to glutamate. Such sensors can be referred to as glutamate binding biosensors or glutamate biosensors.

[0041] In some instances, biosensors disclosed herein can include (e.g., comprise, consist essentially of, or consist of): an amino acid sequence with at least 90% identity to a recombinant peptide biosensor selected from the group consisting of SEQ ID NO: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, and 53, wherein the recombinant peptide biosensor binds specifically to maltose; a recombinant peptide biosensor selected from the group consisting of SEQ ID NO: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, and 53 comprising 10 or fewer conservative amino acid substitutions, wherein the recombinant peptide biosensor binds specifically to maltose; and/or a recombinant peptide biosensor selected from the group consisting of SEQ ID NO: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, and 53.

[0042] In some instances, biosensors disclosed herein can include (e.g., comprise, consist essentially of, or consist of): an amino acid sequence with at least 90% identity to a recombinant peptide biosensor selected from the group consisting of SEQ ID NO: 62 and 63, wherein the recombinant peptide biosensor binds specifically to glutamate; a recombinant peptide biosensor selected from the group consisting of SEQ ID NO: 62 and 63 comprising 10 or fewer conservative amino acid substitutions, wherein the recombinant peptide biosensor binds specifically to glutamate; and/or a recombinant peptide biosensor selected from the group consisting of SEQ ID NO: 62 and 63.

[0043] In some instances, biosensors disclosed herein can include (e.g., comprise, consist essentially of, or consist of): an amino acid sequence with at least 90% identity to a recombinant peptide biosensor selected from the group consisting of SEQ ID NO: 77 and 78, wherein the recombinant peptide biosensor binds specifically to phosphonate; a recombinant peptide biosensor selected from the group consisting of SEQ ID NO: 77 and 78 comprising 10 or fewer conservative amino acid substitutions, wherein the recombinant peptide biosensor binds specifically to phosphonate; and/or a recombinant peptide biosensor selected from the group consisting of SEQ ID NO: 77 and 78.

[0044] In some instances, biosensors disclosed herein can include (e.g., comprise, consist essentially of, or consist of): an amino acid sequence with at least 90% identity to a recombinant peptide biosensor selected from the group consisting of SEQ ID NO: 91, 92, 93 and 94, wherein the recombinant peptide biosensor binds specifically to glucose; a recombinant peptide biosensor selected from the group consisting of SEQ ID NO: 91, 92, 93 and 94 comprising 10 or fewer conservative amino acid substitutions, wherein the recombinant peptide biosensor binds specifically to glucose; and/or a recombinant peptide biosensor selected from the group consisting of SEQ ID NO: 91, 92, 93 and 94.

[0045] In some instances, biosensors disclosed herein can include (e.g., comprise, consist essentially of, or consist of): SEQ ID NO:91; SEQ ID NO:92; SEQ ID NO:93; SEQ ID NO:95.

[0046] In some instances, any recombinant biosensor disclosed herein can be isolated and/or purified. The terms "isolated" or "purified," when applied to a biosensor dis-

closed herein includes nucleic acid proteins and peptides that are substantially free or free of other cellular material or culture medium when produced by recombinant techniques, or substantially free or free of precursors or other chemicals when chemically synthesized.

[0047] The disclosure also provides, in part, nucleic acids (e.g., isolated and/or purified nucleic acids) encoding any one or more of the recombinant peptide biosensors disclosed herein. For example, nucleic acids can encode: an amino acid sequence with at least 90% identity to a recombinant peptide biosensor selected from the group consisting of SEQ ID NO: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, and 53, wherein the recombinant peptide biosensor binds specifically to maltose; a recombinant peptide biosensor selected from the group consisting of SEQ ID NO: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, and 53 comprising 10 or fewer conservative amino acid substitutions, wherein the recombinant peptide biosensor binds specifically to maltose; a recombinant peptide biosensor selected from the group consisting of SEQ ID NO: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, and 53; an amino acid sequence with at least 90% identity to a recombinant peptide biosensor selected from the group consisting of SEQ ID NO: 62 and 63, wherein the recombinant peptide biosensor binds specifically to glutamate; a recombinant peptide biosensor selected from the group consisting of SEQ ID NO: 62 and 63 comprising 10 or fewer conservative amino acid substitutions, wherein the recombinant peptide biosensor binds specifically to glutamate; a recombinant peptide biosensor selected from the group consisting of SEQ ID NO: 62 and 63; an amino acid sequence with at least 90% identity to a recombinant peptide biosensor selected from the group consisting of SEQ ID NO: 77 and 78, wherein the recombinant peptide biosensor binds specifically to phosphonate; a recombinant peptide biosensor selected from the group consisting of SEQ ID NO: 77 and 78 comprising 10 or fewer conservative amino acid substitutions, wherein the recombinant peptide biosensor binds specifically to phosphonate; a recombinant peptide biosensor selected from the group consisting of SEQ ID NO: 77 and 78; an amino acid sequence with at least 90% identity to a recombinant peptide biosensor selected from the group consisting of SEQ ID NO: 91, 92, 93 and 94, wherein the recombinant peptide biosensor binds specifically to glucose; a recombinant peptide biosensor selected from the group consisting of SEQ ID NO: 91, 92, 93 and 94 comprising 10 or fewer conservative amino acid substitutions, wherein the recombinant peptide biosensor binds specifically to glucose; a recombinant peptide biosensor selected from the group consisting of SEQ ID NO: 91, 92, 93 and 94; and/or SEQ ID NO: 91; SEQ ID NO: 92; SEQ ID NO: 93; SEQ ID NO: 95.

[0048] In some instances, the disclosure includes vectors containing one or a plurality of the nucleic acids disclosed herein and cells containing such vectors. In some instances, the disclosure provides cells containing one or a plurality of nucleic acids disclosed herein.

[0049] In some instances, the disclosure includes kits related to the biosensors and nucleic acids disclosed herein. Such kits can include or contain, for example, a biosensor, a nucleic acid encoding a biosensor, vectors, and/or cells, provided herein.

[0050] In some instances, the disclosure provides methods related to the biosensors and nucleic acids disclosed herein. Such methods can include methods of making, using, and/or selling the biosensors and nucleic acids disclosed herein. For

example, methods can include methods for producing genetically encoded recombinant peptide biosensors. In such instances, methods can include, for example, selecting a framework portion that binds specifically to a target analyte and that undergoes a conformational change upon interacting binding to the target analyte, identifying a site or amino acid position within the selected framework portion where or around which the conformational change occurs, and inserting a signaling portion into the site or amino acid position. In some instances, framework portions include periplasmic binding proteins (PBPs) disclosed herein. Exemplary PBPs include PBPs that bind (e.g., bind specifically) to glucose.

[0051] In some instances, the present disclosure includes methods for detecting glucose, e.g., in a sample containing a level of glucose. Such methods can include, detecting a level of fluorescence emitted by a recombinant peptide biosensor, the peptide biosensor having an amino acid sequence selected from the group consisting of SEQ ID NO: 91, 92, 93 and 94, and correlating the level of fluorescence with the presence of glucose. In some instances, recombinant peptide biosensors used in the methods herein are expressed from nucleic acids. In some instances, methods include contacting the recombinant peptide biosensor with a test sample (e.g., a sample comprising glucose). In some instances, methods can include the level of fluorescence emitted by a biosensor (e.g., a biosensor bound to glucose) with a concentration glucose in the sample. Such correlation can include, for example, comparing the level of fluorescence with a level of fluorescence emitted by the recombinant peptide biosensor in the presence of a sample comprising a known concentration or range of concentrations of glucose. In some instance, the level of fluorescence emitted by the recombinant peptide biosensor in the presence (e.g., bound or bound specifically to) of a sample comprising a known concentration or range of concentrations of glucose is stored on an electronic database.

[0052] One of skill will appreciate that such methods can be adapted for any defined, specific, or selected analyte. For example, in some instances, the disclosure provides methods for detecting a defined, selected, or specific analyte. These methods can include detecting a level of fluorescence emitted by a recombinant peptide biosensor expressed from a nucleic acid and correlating the level of fluorescence with the presence the defined, selected, or specific analyte. In some instances, methods include contacting the recombinant peptide biosensor with a sample comprising the analyte. In some instances, methods include correlating the level of fluorescence with a concentration of the analyte. In some instances, methods include comparing the level of fluorescence with a level of fluorescence emitted by the recombinant peptide biosensor in the presence of a sample comprising a known concentration or range of concentrations of the analyte, wherein the level of fluorescence emitted by the recombinant peptide biosensor in the presence of a sample comprising a known concentration or range of concentrations of the analyte is stored on an electronic database.

[0053] In some instances, the present disclosure provides methods for detecting a defined, selected, or specific analyte, the method comprising detecting a level of fluorescence emitted by a recombinant peptide biosensor of any one of claims 1-36; and correlating the level of fluorescence with the presence of a defined, selected, or specific analyte. In some instances, recombinant peptide biosensors can be

expressed from a nucleic acid. In some instances, methods can include contacting the recombinant peptide biosensor with a sample comprising the analyte. In some instances, methods can include correlating the level of fluorescence with a concentration of the analyte and, optionally, comparing the level of fluorescence with a level of fluorescence emitted by the recombinant peptide biosensor in the presence of a sample comprising a known concentration or range of concentrations of the analyte. In some instances, the level of fluorescence emitted by the recombinant peptide biosensor in the presence of a sample comprising a known concentration or range of concentrations of the analyte is stored on an electronic database.

[0054] Methods herein can be performed in vitro.

[0055] In some instances, the present disclosure provides compositions containing any one or a plurality of the peptide biosensors and/or nucleic acids disclosed herein.

[0056] Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Methods and materials are described herein for use in the present invention; other, suitable methods and materials known in the art can also be used. The materials, methods, and examples are illustrative only and not intended to be limiting. All publications, patent applications, patents, sequences, database entries, and other references mentioned herein are incorporated by reference in their entirety. In case of conflict, the present specification, including definitions, will control.

[0057] Other features and advantages of the invention will be apparent from the following detailed description and figures, and from the claims.

DESCRIPTION OF DRAWINGS

[0058] FIG. 1 Cartoon representation showing ligand bound *Escherichia coli* malto-dextrin-binding protein (EcMBP) and potential circularly-permuted fluorescent protein (cpFP) insertion sites.

[0059] FIG. 2 Cartoon representation showing ligand bound *Pyrococcus furiosus* maltotriose binding protein (PfMBP) and potential cpFP insertion sites.

[0060] FIG. 3 Cartoon representation showing ligand bound *E. coli* glutamate-binding protein (EcYbeJ) and potential cpFP insertion sites.

[0061] FIG. 4 Cartoon representation showing ligand bound *E. coli* phosphonate-binding protein (EcPhnD) and potential cpFP insertion sites.

[0062] FIG. 5 Cartoon representation showing ligand bound *Thermus thermophilus* glucose binding protein (TtGBP) and potential cpFP insertion sites.

[0063] FIG. 6A-B Changes in EcMBP upon maltose binding and locations at which circularly-permuted fluorescent protein (cpFP) was inserted are shown as colored spheres at the C α positions. Yellow: 165-166, Green: 175-176, Cyan: 311-312, Violet: 317-318 (FIG. 6A). (FIG. 6B) shows backbone structural changes. The C α dihedral is calculated from the four atoms: C α i+2, C α i+1, C α i, C α i-1. Δ Dihedral is calculated as the difference in dihedrals between the closed (1ANF) and open (1OMP) states of MBP, and corrected to fall within a range of -180° to 180° . The regions near residues 175 and 311 are labeled. There is a crystallographic artifact at the N-terminus resulting in the appearance of significant structural changes.

[0064] FIG. 7A Amino acid sequence of MBP-165-cpGFP (SEQ ID NO:1).

[0065] FIG. 7B Amino acid sequence of MBP-165-cpGFP.PPYF (SEQ ID NO:2).

[0066] FIG. 7C Amino acid sequence of MBP-165-cpGFP.PCF (SEQ ID NO:3).

[0067] FIG. 8A Amino acid sequence of MBP-175-cpGFP (SEQ ID NO:4).

[0068] FIG. 8B Amino acid sequence of MBP-175-cpGFP.L1-HL (SEQ ID NO:5).

[0069] FIG. 9A Amino acid sequence of MBP-311-cpGFP (SEQ ID NO:6).

[0070] FIG. 9B Amino acid sequence of MBP-311-cpGFP.L2-NP (SEQ ID NO:7).

[0071] FIG. 10 Amino acid sequence of MBP-317-cpGFP (SEQ ID NO:8).

[0072] FIGS. 11A-11D Line charts showing EcMBP plot of $\Delta F/F$ for clarified lysate screen of cpGFP linker-screens at insertion points 165, 175, 311, and 317. The horizontal dashed line at zero indicates no fluorescence change. Standard deviations in $\Delta F/F$ are less than 10% of an average ΔF (repetitions for MBP165-cpGFP.PPYF yields $\Delta F/F$ values of 2.51, 2.63, and 2.54).

[0073] FIG. 12 Isothermal titration calorimetry (ITC) of MBP317-cpGFP with maltose.

[0074] FIG. 13 Graph showing EcMBP165-cpGFP.PPYF affinity variant binding maltose-binding curves. Binding curves for affinity variants of MBP165-cpGFP.PPYF. Data is fit to a single-binding site isotherm. Curve-fit affinities are: WT binding pocket, 5 μ M (●); W230A, 32 μ M (■); W62A, 375 μ M (▲); W340A, >1 mM (▼); 1329W, 11 μ M (□).

[0075] FIGS. 14A-14D Line graphs showing maltose and sucrose binding curves for wild-type and 5-7 variants of the EcMBP-cpGFP sensors. Maltose (black) and sucrose (red) binding curves for wild-type (filled, solid lines) and 5-7 variants (open, dashed lines) of the MBP-cpGFP sensors. MBP165-cpGFP.PPYF (FIG. 14A); MBP165-cpGFP.PCF (FIG. 14B); MBP175-cpGFP.L1-HL (FIG. 14C); MBP311-cpGFP.L2-NP (FIG. 14D).

[0076] FIGS. 15A-15D Line graphs showing emission spectra for colored variants of EcMBP sensors. Fluorescence emission spectra of the MBP165-Blue, Cyan, Green, and Yellow wild-type sensors (FIG. 15A) and the 5-7 variants (FIG. 15B) in the absence of ligand (dashed lines, open circles), with 10 mM maltose (solid lines, filled circles), or 10 mM sucrose (solid lines, filled squares). Sensors were excited at 383, 433, 485, and 485 nm, respectively. Titration of maltose and sucrose in the Blue, Cyan, Green, and Yellow MBP165 wild-type sensors (FIG. 15C) and for the 5-7 variants (FIG. 15D). Filled circles are titration of maltose, open circles are titration of sucrose. For the wild-type sensors, Kds for maltose binding are: Blue 3.3 μ M, Cyan 13 μ M, Green 4.5 μ M, Yellow 3.3 μ M. No sucrose binding is observed. For the 5-7 variants, Kd of Green is 2.4 mM (sucrose) and 7.1 mM (maltose). Kd of Yellow is 2.5 mM (sucrose) and 4.5 mM (maltose).

[0077] FIG. 16 Plot of $\Delta F/F$ for clarified lysate screen of MBP165-cpBFP linker-screen. The horizontal dashed line at zero indicates no fluorescence change.

[0078] FIGS. 17A-17B Line graphs showing maltose binding. Blue (wt binding pocket) has an affinity of 2.7 μ M. Green (W230A) has an affinity of 40 μ M. Yellow (W62A) has an affinity of 350 μ M. Cyan (W340A) has an affinity of

approximately 1.7 mM. Data is plotted at $\Delta F/F$ (FIG. 17A) or normalized to Fractional Saturation (FIG. 17B).

[0079] FIGS. 18A-18C Images bacterial cells expressing (FIG. 18A) EGFP, (FIG. 18B) PPYF, or (FIG. 18C) PPYF. T203V in the absence (top) and presence (bottom) of maltose.

[0080] FIGS. 19A-19B Line graphs showing EcMBP-cpGFP.PPYF.T203V 2-photon excitation spectra. MBP165-cpAzurite.L2-FE (FIG. 19A), -cpCFP.PCF (FIG. 19A), -cpGFP.PPYF (FIG. 19B), and -cpYFP.PPYF (FIG. 19B) were excited at the wavelengths indicated and emission measured through appropriate wavelength filters. Two graphs are shown to present different y-axis scales. Optimal $\Delta F/F$ values for 2-photon excitation of the spectral variants of MBP165 are: -cpAzurite, 1.1 (ex 760 nm); -cpCFP, 2.3 (ex 830-960 nm); -cpGFP, 10.0 (ex 940 nm); -cpYFP, 2.6 (ex 940 nm).

[0081] FIGS. 20A-20C Images showing EcMBP-cpGFP.PPYF.T203V expressing HEK cells. Images of individual HEK293 cells expressing membrane displayed PPYF. T203V in the absence of maltose (FIG. 20A), in the presence of 1 mM maltose (FIG. 20B), and after washout with maltose-free buffer (FIG. 20C). Scale bars are 10 μ m.

[0082] FIGS. 21A-21B Graphs showing quantification of fluorescence of EcMBP-cpGFP.PPYF.T203V when displayed on the surface of HEK cells. (FIG. 21A) Concentration dependence. (FIG. 21B) Observed fluorescence after a "puff" of HBSS solution containing 1 mM maltose and 2.5 nM Alexa Fluor® 568 (Invitrogen, Carlsbad, Calif.).

[0083] FIGS. 22A-22D Cartoon representations and close-up views of inter-domain linkers and selected amino acids of the cpGFP chromophore environment of the structure of MBP175-cpGFP.L1-HL (FIG. 22A and FIG. 22B) and MBP311-cpGFP.L2-NP (FIG. 22C and FIG. 22D) bound to maltose. The MBP domain is colored as in FIG. 1. The cpGFP domain is green and the inter-domain linkers are colored white. The cpGFP chromophore is displayed as sticks and the bound maltose as red and white spheres. Ordered water molecules are represented as red spheres. Selected hydrogen bonds are displayed as dashed black lines. β -strands 10 and 11 of cpGFP are displayed as semi-transparent for clarity. The 2Fo-Fc electron density map calculated with the displayed residues omitted from the model is shown as blue mesh.

[0084] FIGS. 23A-23D EcMBP-cpGFP: effect of T203V mutation on fluorescence. (FIG. 23A) Emission spectra of 1 μ M purified eGFP (filled circles), cpGFP (filled squares), MBP165-cpGFP.PPYF (open circles), and MBP165-cpGFP.PPYF+T203V (open squares) in the absence (dashed lines) or presence (solid lines) of 1 mM maltose. cpGFP is half as bright as eGFP, and the saturated MBP165-cpGFP.PPYF variants are about half as bright as cpGFP. (FIG. 23B) Titration of maltose for MBP165-cpGFP.PPYF (filled squares), and MBP165-cpGFP.PPYF+T203V (filled circles). Affinities for each protein are the same, but with different $\Delta F/F$. (FIG. 23C) Emission spectra of 1 μ M purified eGFP (filled circles), cpGFP (filled squares), MBP311-cpGFP.L2-NP (open circles), and MBP311-cpGFP.L2-NP+T203V (open squares) in the absence (dashed lines) or presence (solid lines) of 1 mM maltose. Note that mutation T203V decreases the fluorescence of both the apo-state and the saturated state of MBP311-cpGFP.L2-NP. (FIG. 23D) Titration of maltose for MBP311-cpGFP.L2-NP (filled squares), and MBP311-cpGFP.L2-NP+T203V (filled

circles). Affinities for each protein are the same, but with $\Delta F/F$ slightly increased for the T203V variant.

[0085] FIG. 24A Amino acid sequence of PfMBP171-cpGFP (SEQ ID NO:50)

[0086] FIG. 24B Amino acid sequence of PfMBP171cpGFP.L2-FE (SEQ ID NO:51)

[0087] FIG. 25A Amino acid sequence of PfMBP316-cpGFP (SEQ ID NO:52)

[0088] FIG. 25B Amino acid sequence of PfMBP316-cpGFP.L1-NP (SEQ ID NO:53)

[0089] FIG. 26A-26B Plot of $\Delta F/F$ for clarified lysate screen of cpGFP linker-screens at insertion points 171 (FIG. 26A) and 316 (FIG. 26B).

[0090] FIGS. 27A-27D Plot of Beta-sheet circular dichroism (CD) signal as a function of temperature.

[0091] FIGS. 28A-28B PfMBP Fluorescence vs. temperature. (FIG. 28A) Plot of fluorescence as a function of temperature in the presence (solid) or absence (dashed) of ligand. (FIG. 28B) Plot of $\Delta F/F$ as a function of temperature. Using the data from FIG. 27A, $\Delta F/F$ for each protein (Fbound-Fapo/Fapo) was calculated for each temperature.

[0092] FIGS. 28C-28E Line graphs showing the function of immobilized and soluble proteins.

[0093] FIG. 29A Amino acid sequence of EcYbeJ253-cpGFP (SEQ ID NO:62).

[0094] FIG. 29B Amino acid sequence of EcYbeJ253-cpGFP.L1LVL2NP (SEQ ID NO:63).

[0095] FIG. 30 EcYbeJ binding curves. Plot of $\Delta F/F$ as a function of [Glutamate], μ M. The first generation sensor, EcYbeJ253.L1-LV (with the A184V mutation (grey, solid) has an affinity for glutamate of about 100 μ M and a $\Delta F/F$ of 1.2. The reversion of that affinity mutation, V184A, in the L1-LV background increases affinity to 1 (grey dashed). The second generation sensor, with the L2-NP linker optimization and the A184V mutation, has a $\Delta F/F$ of at least 4 and an affinity for glutamate of about 100 μ M (black solid).

[0096] FIG. 31 EcYbeJ Hema/cMyc analysis. The effect of N- and C-terminal tags on $\Delta F/F$ and glutamate affinity were determined by expressing variously tagged versions of the EcYbeJ253.L1LVL2NP protein in bacteria. The presence of the pRSET leader sequence (black) has no effect on $\Delta F/F$ (~5) or affinity (~120 when compared to the version without a tag (grey). The addition of the cMyc tag to the C-terminus retains $\Delta F/F$ and increases affinity slightly, to 60 μ M. The addition of the N-terminal hemagglutinin tag, with (green) or without (orange) the cMyc tag, decreases $\Delta F/F$ substantially.

[0097] FIGS. 32A-32B EcYbeJ253-cpGFP.L1LVL2NP.pMinDis expressed in HEK293 cells. (FIG. 32A) Images of the sensor expressing HEK cells in the absence of glutamate (left), with 100 μ M glutamate (center), and re-imaged after wash-out of glutamate with buffer (right). (FIG. 32B) By measuring the equilibrium $\Delta F/F$ with different concentrations of glutamate in the buffer, an in situ binding affinity (black) can be obtained. The surface displayed sensor has a higher affinity (3 μ M) for glutamate than the soluble sensor (grey), which is about 90 μ M.

[0098] FIG. 33 EcYbeJ253-cpGFP.L1LVL2NP.pMinDis expressed in neuronal culture, and responds rapidly to added glutamate (green). Red shows signal of 2.5 nM Alexa Fluor® 568 (Invitrogen, Carlsbad, Calif.), also in pipette.

[0099] FIG. 34A Amino acid sequence of EcPhnD90-cpGFP (SEQ ID NO:77).

[0100] FIG. 34B Amino acid sequence of EcPhnD90-cpGFP.L1AD+L297R+L301R (SEQ ID NO: 78).

[0101] FIGS. 35A-35C EcPhnD90-cpGFP Binding Curves. For both the L1AD and the L1AD+L297R+L301R variants, binding was determined for (FIG. 35A) 2-aminoethylphosphonate (2AEP), (FIG. 35B) methylphosphonate (MP), and (FIG. 35C) ethylphosphonate (EP).

[0102] FIGS. 36A-36C The crystal structures of the ligand-free (FIG. 36A), open state (with H157A mutation to the binding pocket) and the ligand-bound (FIG. 36B), closed state of EcPhnD clearly shows a large conformational change. Residues in between which cpGFP is inserted in EcPhnD90-cpGFP are marked by red spheres, in the equatorial strand (red). (FIG. 36C) Analysis of the change in Δ Dihedral (Δ Dihedral) clearly shows that residues for which there is the greatest Δ Dihedral upon going from the open to the closed state are residues 88 (Δ Dihedral= -75°), 89 (Δ Dihedral= 123°), and 90 (Δ Dihedral= 52°).

[0103] FIG. 37A Amino acid sequence of TtGBP326-cpGFP (SEQ ID NO:91).

[0104] FIG. 37B Amino acid sequence of TtGBP326.L1PA (SEQ ID NO:92).

[0105] FIG. 37C Amino acid sequence of TtGBP326.H66A (SEQ ID NO:93).

[0106] FIG. 37D Amino acid sequence of TtGBP326.H348A (SEQ ID NO:94).

[0107] FIG. 38 TtGBP326-cpGFP Binding Curves. Plot of Δ F/F as a function of [Glucose], mM.

[0108] FIG. 39 An image showing TtGBP326-cpGFP expressed as a transgenic reporter of intracellular glucose in cultured human cells.

[0109] FIGS. 40A-40B Are line graphs showing that the addition of extracellular glucose increases TtGBP326-cpGFP fluorescence in human cells.

[0110] FIG. 41 Amino acid sequence of *Escherichia coli* maltodextrin-binding protein (EcMBP) (SEQ ID NO: 105).

[0111] FIG. 42 Amino acid sequence of *Pyrococcus furiosus* maltose-binding protein (PMBP) (SEQ ID NO: 106).

[0112] FIG. 43 Amino acid sequence of *E. coli* glutamate-binding protein (EcYbeJ) (SEQ ID NO:107).

[0113] FIG. 44 Amino acid sequence of *E. coli* phosphate-binding protein (EcPhnD) (SEQ ID NO:108).

[0114] FIG. 45 Amino acid sequence of *Thermus thermophilus* glucose-binding protein (TtGBP) (SEQ ID NO:109).

[0115] FIG. 46 Amino acid sequence of UniProt accession number Q92N37 (SEQ ID NO: 110).

[0116] FIG. 47 Amino acid sequence of UniProt accession number DOVWX8 (SEQ ID NO:111).

[0117] FIG. 48 Amino acid sequence of UniProt accession number Q7CX36 (SEQ ID NO:112).

[0118] FIG. 49 Amino acid sequence of UniProt accession number POAD96 (SEQ ID NO:113).

[0119] FIG. 50 Amino acid sequence of TtGBP326.L1PA.L2NP.H66A.H348A.L276V (SEQ ID NO:114).

[0120] FIG. 51 A line graph showing binding of TtGBP326.L1PA.L2NP.H66A.H348A.L276V to glucose.

[0121] FIG. 52 A line graph showing fluorescence increase upon addition of glucose to HEK293 cells expressing TtGBP326.L1PA.L2NP.H66A.H348A.L276V on their extracellular surface.

[0122] FIG. 53A-D SF-iGluSnFR.A184S shows larger responses to visual stimuli than SF-iGluSnFR.A184V. (FIG. 53A) Two-photon standard-deviation projection of SF-iGluSnFR.A184S and A184V expressed in ferret visual cortex

(A184S: 190 μ m, A184V: 175 μ m, scale bar 100 μ m). (FIG. 53B) Trial-averaged stimulus-evoked responses (shown for ROI 1) reveal robust orientation tuning and peak amplitudes of \sim 30% Δ F/F for A184S. Peak responses plotted as a function of stimulus orientation show robust selectivity with the A184S variant. For the A184V variant, stimulus-evoked fluctuations are too small (\sim 5% Δ F/F) to generate robust tuning plots. (FIG. 53C) Two-photon standard-deviation projection of an isolated dendritic segment with active spines revealed with SF-iGluSnFR.A184S. Individual dendritic spines are driven selectively and strongly by drifting gratings. Orientation tuning from peak responses shows large spine responses (30-50% Δ F/F) and, importantly, reveals that spines on a single dendritic branch can receive differently tuned excitatory input. (FIG. 53D) Same as in (FIG. 53C) for SF-iGluSnFR.A184V. Dendritic spine responses with A184V are weak and almost unresolvable.

[0123] FIG. 54A-D SF-iGluSnFR. S72A permits resolution of multiple glutamate release events in cultured mouse embryonic hippocampal neurons. (FIG. 54A) Single (dashed) and averaged (solid) traces of SF-iGluSnFR.S72A (blue) and SF-iGluSnFR.A184V (red) response to 20 Hz paired electrical stimuli. (FIG. 54B) Histogram showing intensity second pulse to first pulse response. (FIG. 54C) The faster off-rate of S72A can be used to observe vesicle release depression. Higher concentrations of extracellular calcium can increase vesicle release, leading to vesicle exhaustion as the train of field pulses progresses. (FIG. 54D) The slow decay of A184V obscures this depression.

[0124] FIG. 55A-D S72A variant shows faster bouton fluorescence signals resulting from single or trains of electrical stimulation mouse cerebellar brain slice. (FIG. 55A) Averaged response from single boutons expressing GCaMP6f (GC6f) at 2 mM $[Ca^{2+}]_{extracellular}$ (green), GC6f at 1.5 mM $[Ca^{2+}]_e$ (black), SF-iGluSnFR.A184V at 1.5 mM $[Ca^{2+}]_e$ (A184V, red) and SF-iGluSnFR.S72A at 1.5 mM $[Ca^{2+}]_e$ (S72A, blue), normalized to peak response. In parenthesis the number of trials used to calculate the average. Right, summary plots of Δ F/F₀, 10-90% rise time, 50% decay time and signal-to-noise-ratio (SNR). Multiple comparisons were performed with the Kruskal-Wallis test and the Dunn's multiple comparisons test. *P<0.05, **P<0.01, ****P<0.0001. (FIG. 55B) Two-photon fluorescent image of granule cells and parallel fibers expressing A184V in cerebellum slice (GL—granule layer, ML—molecular layer). Yellow arrows indicate labeled soma of granule cells, and circle indicate boutons from parallel fibers. Bottom, example of single trial A184V fluorescence responses to 20 Hz electrical stimulation (red) and the average of 10 trials (purple). (FIG. 55C) Population average fluorescence responses to 20 Hz stimulation (n boutons=5 GC6f; n=17, A184V; n=3, S72A). Traces are normalized to the peak of the first response. (FIG. 55D) Population average of response to 100 Hz electrical stimulation (n boutons=9 GC6f; n=9, A184V; n=7, S72A) normalized to the maximum amplitude (left) or to the peak of the first response (middle), and average response of all the boutons. n is number of boutons. Black arrows indicate time of electrical stimulation.

[0125] FIG. 56A-B High-speed two photon imaging (1016 Hz frame rate) of a neuron expressing SF-Venus-iGluSnFR. (FIG. 56A) RuBi-glutamate was uncaged for 10 msec. at each of two 5 μ m spots (red arrowheads) on the dendrites. Saturation denotes the glutamate transient amplitude. Yel-

low line indicates locations for traces shown in (FIG. 56B). (FIG. 56B) Recorded traces at nine pixels at various distances from the uncaging focus, along the yellow line in (FIG. 56A). The traces are approximate maximum likelihood solutions recovered with the FADE algorithm. (Kazempour et al., Proceedings of the 2017 Asilomar Conference on Signals, Systems, and Computers, October 29-November 1, Pacific Grove, Calif.), which incorporates dynamics having arbitrarily fast rise but slow decay. This recording is of a single uncaging event, without averaging.

[0126] FIG. 57 Circular dichroism of iGluSnFR and SF-iGluSnFR. 20 μM purified and dialyzed protein in 0.1 \times PBS was analyzed by circular dichroism (Chirascan, Applied Biophysics). Grey, iGluSnFR; black, SF-iGluSnFR; green, cpSFGFP; thick line, with 1 mM glutamate; thin line, no glutamate. Spectra were collected with a 1 sec. sampling time after equilibration for 2 min at each temperature. The first unfolding transition is shifted from about 50° C. to 55° C. by inclusion of the Superfolder mutations to cpGFP. Interestingly, the second transition, at about 75° C., which parallels the transition of cpSFGFP alone, is unchanged.

[0127] FIG. 58A-D Spectra of SF-iGluSnFR. (FIG. 58A) 2-photon cross-section of purified, soluble iGluSnFR (grey) and SF-iGluSnFR (black) in the ligand-free (dashed line) and glutamate-saturated (solid line) state. Excitation (FIG. 58B), emission (FIG. 58C), and absorption spectra (FIG. 58D) of iGluSnFR (grey), SF-iGluSnFR (black), and cpSFGFP (green) with glutamate (solid line) and without (dashed line).

[0128] FIG. 59A-E Representative images of (FIG. 59A) SF-iGluSnFR and (FIG. 59B) iGluSnFR in mouse somatosensory cortex taken at 0.9 $\mu\text{m}/\text{pixel}$, 0.126 nsec dwell time per μm , 80 mW power, prior to bleaching. 20 nl of AAV2/1.hSynapsin1.iGluSnFR or SF-iGluSnFR (identical virus titer, prepared by the same person) was injected three weeks before imaging. Contrast adjusted to 10 greyscale in both images to make original iGluSnFR observable. Mean signal-to-noise ratios ($n=2$ animals) are 66 vs. 14 (80 mW power) and 2.4 vs. 0.3 (5 mW power). (FIG. 59C) & (FIG. 59D) Representative images of SF-iGluSnFR and iGluSnFR taken with 5 mW power, which is more typical in live imaging conditions. (FIG. 59E) Bleaching of SF-iGluSnFR (black) and original iGluSnFR (grey) at 80 mW power and 10 \times zoom (0.09 $\mu\text{m}/\text{pixel}$, 1.26 nsec dwell time per μm).

[0129] FIG. 60A-B In vitro binding affinity. (FIG. 60A) Titration of bacterially expressed iGluSnFR and SF-iGluSnFR and variants. Affinities (K_d) for original iGluSnFR, SF-iGluSnFR.A184S, SF-iGluSnFR.A184V, and SF-iGluSnFR.S72A are 84 ± 7 μM , 7.5 ± 0.4 μM , 41 ± 7 μM , and 200 ± 5 μM respectively. (FIG. 60B) Kinetics of glutamate binding by stopped-flow fluorescence spectroscopy. Equal volumes of 1 μM SF-iGluSnFR (A184S, A184V, or S72A) and glutamate (variable concentration) were mixed in an SX.18MV stopped-flow spectrometer (Applied Photophysics, Surrey, UK). Representative traces shown. Pseudo-first order analysis indicates that the on-rate of binding for SF-iGluSnFR.A184S, A184V, S72A are 6, 5, and 0.6 $\mu\text{M}^{-1}\text{sec}^{-1}$, respectively. The off rates, as determined by the y-intercept, are 25 sec^{-1} , 52 sec^{-1} , and 108 sec^{-1} respectively. Error bars are standard deviation of three measurements.

[0130] FIG. 61 Binding affinity screening. Pellets of bacterially expressed A184X variants of iGluSnFR were washed 5 times in PBS to remove bound glutamate. After

freezing and thawing, pellets were clarified by centrifugation and titrated with glutamate to screen for their affinity for glutamate. There is a general trend of larger amino acids resulting in weaker affinity.

[0131] FIG. 62 Affinity of SF-iGluSnFR variants displayed on the surface of neurons. AAV2/1.hSynapsin1.SF-iGluSnFR variants (1 μl of 1E13 GC/ml) were used to infect rat hippocampal neuronal culture 3 days after culturing. After 10 days in vitro, fluorescence was monitored under continuous flow of buffer with varying concentrations of glutamate. Affinities (K_d) for SF-iGluSnFR.A184S, SF-iGluSnFR.A184V, and SF-iGluSnFR.S72A are 0.6, 2.1, and 34 μM respectively. Affinities for SF-Venus.A184V and SF-Azurite.A184V are 2.0 and 9 μM respectively. Bottom panel is zoom-in of top panel.

[0132] FIG. 63A-B Rise and decay of fluorescence signal resulting from a single field stimulation (1 msec., 90 mA) in rat hippocampal culture (10 DIV, 7 DPI) in non-flowing buffer (FIG. 63B). Traces in FIG. 63A are the average of three ROIs (bottom) and three trials (top); error bars are standard deviation of those nine measurements. The large error for GCaMP6f results from back propagating action potentials, which can be seen in differences from individual ROIs.

[0133] FIG. 64 Examples of individual responses for ROIs 1, 2, and 3 (top). Responses of individual Spines #2 and #3 (from FIG. 53) (middle). Histogram showing distribution of spine responses (bottom). Response amplitudes across individual trials were consistently greater for A184S than the A184V when examining all stimulus-evoked responses.

[0134] FIG. 65A-D Mouse neuronal culture images. The fluorescent labeling pattern and intensity of primary hippocampal neurons transduced with AAV2/1.hSynapsin1.SF-iGluSnFR.S72A or with AAV2/1.hSynapsin1-SF-iGluSnFR.A184V at DIV4 and imaged at DIV13 looked qualitatively similar for both variants and as expected for a membrane targeted protein. To resolve fast stimulus associated changes in fluorescence, a time series of 100 frames at 60 Hz during a paired-pulse stimulation paradigm was acquired. Basal fluorescence before stimulation was clearly stronger for A184V, the high affinity sensor, leading to a higher SNR (FIG. 65A and FIG. 65C). However, when dividing each frame by an average of the pre-stimulus images for both variants of SF-iGluSnFR localized spots where fluorescence increases was observed (FIG. 65B, FIG. 65D, arrows), likely representing synaptic release sites. ROIs were defined based on these spots, and fluorescence within these ROIs (background subtracted) was averaged for every image in the time series.

[0135] FIG. 66A-J Vesicle release sites can be localized by identifying the center of stimulus-evoked SF-iGluSnFR fluorescence changes. (FIGS. 66A and 66B) Representative images of SF-iGluSnFR.S72A and SF-iGluSnFR.A184V expression in primary neuron cultures. Markers indicate the centers of Gaussians fitted to fluorescence profiles calculated across identified release sites from consecutive stimulation trials (such as shown in (FIG. 66E)). Note that the scatter of the centers of the localized release sites is substantially larger for SF-iGluSnFR.A184V (16-25 stimulation trials per experiment with inter-stimulus intervals of 20-60 s, 20 frames before and 10 frames after stimulation were recorded). (FIGS. 66C and 66D) Spots of increased fluorescence as they occur immediately after electrical stimulation when neurons are expressing SF-iGluSnFR.S72A or

SF-iGluSnFR.A184V. 10 frames after the stimulus were averaged and divided by an average of 5 frames before stimulation. In this way, structures, which do not change fluorescence after simulation (background/inactive dendritic segments) will become 1. The lookup table of these images was adjusted to range from 1 to 1.5. (FIG. 66E) Line profiles calculated across the response sites shown in (FIG. 66C) and (FIG. 66D) (dashed lines) and superimposed Gaussian fits (lines). The width of the fitted Gaussian profiles were 0.57 and 1.11 μm for SF-iGluSnFR.S72A and SF-iGluSnFR.A184V, respectively. (FIG. 66F) Localization is more precise for SF-iGluSnFR.S72A. For each selected responding site ($n=28-53$), the mean deviation of the center of the Gaussians across the stimulation trials was calculated. These values were averaged and bar graphed for each SF-iGluSnFR variant. (FIGS. 66G and 66H) Width and amplitude of fitted Gaussian functions are significantly larger for the high affinity A184V sensor. (FIG. 66I) Gaussian fits to profiles obtained from individual (not averaged) frames after stimulus reveal the persistence of the SF-iGluSnFR.A184V variant. (FIG. 66J) Left: Gaussians fitted to the SF-iGluSnFR.A184V-mediated signal progressively broaden over time indicating that also sensor molecules remote to the site of release bind glutamate. Right: Same data as on left, but plotted as width² over 4*t. The data points can be approximated by a line consistent with a diffusional spread of glutamate. The slope of the fitted line estimates the apparent (A184V-slowed) diffusion coefficient (D_{app}) of synaptically released glutamate to be 4.3 $\mu\text{m}^2/\text{s}$ in vitro. This value is orders of magnitude smaller than the diffusion coefficient of free glutamate in solution ($\sim 600-700 \mu\text{m}^2/\text{s}$) indicating that A184V not only prolongs but also substantially localizes glutamate molecules at the sites of release.

[0136] FIG. 67A-C Annotated amino acid sequences of SF-iGluSnFR (FIG. 67A), SF-Azurite-iGluSnFR (FIG. 67C), and SF-Venus-iGluSnFR (FIG. 67B). Domains colored as indicated. Affinity modulating mutations S72A and A184V/S are indicated by orange arrow. Mutations from SF-iGluSnFR to SF-Venus-iGluSnFR and SF-Azurite-iGluSnFR indicated in red.

[0137] FIG. 68A-D Characterization of soluble, purified SF-Azurite-iGluSnFR. (FIG. 68A) Titration of SF-Azurite-iGluSnFR yields a K_d of $62 \pm 11 \mu\text{M}$, error bars are standard deviation of three measurements. Excitation (FIG. 67B), emission (FIG. 67C), and absorption (FIG. 68D) spectra of SF-Azurite-iGluSnFR (light blue) and Azurite (dark blue), with glutamate (solid line) and without (dashed line).

[0138] FIG. 69A-D Spectra of SF-Venus-iGluSnFR. Excitation (FIG. 69A), emission (FIG. 69B), and absorbance (FIG. 69C) spectra of SF-Venus-iGluSnFR (yellow) with (solid line) and without (dashed line) glutamate. (FIG. 69D) 2-photon spectrum with SF-iGluSnFR (black) and vertical 1030 nm markup included for reference.

[0139] FIG. 70A-F Annotated amino acid sequences of the SF biosensors disclosed herein. Affinity modulating mutations S72A and A184V/S are indicated with small case letters. For SEQ ID NOs: 176-182, each domain is indicated with underlining as follows: IgG secretion signal; GltI5-253; SF-GFP147-238; Linker; SF-GFP1-146; GltI254-279;

Myc epitope; PDGFR transmembrane domain 513-561. SF-iGluSnFR.A184V (SEQ ID NO: 179); SF-iGluSnFR.A184S (SEQ ID NO: 177); SF-iGluSnFR.S72A (SEQ ID NO: 178); SF-Venus-iGluSnFR.A184V (SEQ ID NO: 179; mutations at residues T203Y and Y65G to shift the color and at residues

F46L and S72A to increase chromophore maturation are shown in lower case); SF-Venus-iGluSnFR.A184S (SEQ ID NO: 180; mutations at residues T203Y and Y65G to shift the color and at residues F46L and S72A to increase chromophore maturation are shown in lower case); SF-Venus-iGluSnFR.S72A (SEQ ID NO: 181; mutations at residues T203Y and Y65G to shift the color and at residues F46L and S72A to increase chromophore maturation are shown in lower case); SF-Azurite-iGluSnFR (SEQ ID NO: 182; mutations at residues T65S and Y66H to shift the color and at residues V150I and V224R to improve maturation and brightness are shown in lower case); Linker1 mutations: GltI-cpSFGFP connection from PILVSHNV (SEQ ID NO: 187) to PILGYHNV (SEQ ID NO: 188); Linker2 mutations: cpSFGFP-GltI connection from YNFNNPLN (SEQ ID NO: 189) to YNFNEQLN (SEQ ID NO: 190)); iDexSnFR (or SF-GlucoseSensor) (SEQ ID NO: 183); iGABASnFR (SEQ ID NO: 184; cpSFGFP was inserted after D276 of the Pf622 starting sequence. Insertion of cpSFGFP is after residue D276 of Pf622. Residues RS near the N-terminus encode BgIII, and residues LQ at the C-terminus encode PstI. Mutations included in iGABASnFR include: affinity modulating hinge mutation: Pf622: F101L; Pf622-SFGFP interface: Pf622: N260A; Linker 1: SHNVY (SEQ ID NO: 191) of SFGFP to LAQVR (SEQ ID NO: 192) (SFGFP: S147L, H148A, N149Q, Y151R); Linker 2: SFGFP (SEQ ID NO: 193); F145W; Linker 2: SVLAP (SEQ ID NO: 194) of Pf622 to ANLAP (SEQ ID NO: 195) (Pf622: S277A, V278N); Binding site mutation: Pf622: F102G/Y. Underlining indicates the domain as follows: IgG secretion signal; Pf6222-276; SF-GFP147-238; Linker; SF-GFP1-146; Pf622277-320; Myc epitope; PDGFR transmembrane domain 513-561. Binding site mutation F102 indicated with a small case letter; and iAChSnFR (*E. coli* expression vector shown in SEQ ID NO: 185 with the domains indicated as follows: pHHM.His.tag; leader sequence; Thermoanaerobactersp.X513cholinebindingproteinsequence; Linkerregions; Circularlypermutedsuper-folderEGFP; Myc tag C-terminal sequence and mammalian expression vector shown in SEQ ID NO: 186 with the domains indicated as follows: IgG secretion sequence; leader; Thermoanaerobactersp.X513cholinebindingproteinsequence; Linkerregions; Circularlypermutedsuper-folderEGFP; PDGFR transmembrane sequence).

[0140] FIG. 71 A schematic of Structure I as described herein.

DETAILED DESCRIPTION

[0141] The present disclosure is based, at least in part, on the discovery of structures and methods related to and useful for genetically encoded biosensors. Specifically, the disclosure provides genetically encoded recombinant or chimeric peptides for use as biosensors and methods for the design, production, and use of such biosensors. As described below, these sensors can be employed (e.g., expressed) in biological systems to detect and/or monitor a wide range of target analytes (e.g., a defined, selected, and/or specific analytes) due, in part, to the signal change generated by the sensors upon binding to their respective analyte(s), which signal change allows bound and unbound sensors to be distinguished.

[0142] While the disclosure encompasses generic biosensors and methods related thereto, examples of particular

binding sensors, including biosensors for detecting maltose, sucrose, maltotriose, glutamate, phosphonate, and glucose are also disclosed.

Compositions

[0143] Provided herein are genetically encoded biosensors, i.e., nucleic acids encoding peptides, and/or the encoded peptides (e.g., isolated peptides), for use as biosensors. Biosensors herein include genetically encoded recombinant peptides containing an analyte-binding framework portion linked (e.g., operably linked) to at least one independent signaling portion, wherein the independent signaling portion is allosterically modulated or regulated by the framework portion upon interaction of the framework portion with an analyte (e.g., a defined, selected, and/or specific analyte), such that signaling from the signaling portions is altered upon interaction of the framework portion with the analyte.

[0144] In some instances, an independent signaling portion is present at a site within the framework portion that undergoes a conformational change upon interaction of the framework portion with an analyte such that the conformational change allosterically modulates or regulates signaling by the signaling portion. For example, biosensors herein can include structure I.

[0145] In some instances, signaling by the signaling portion is detectably altered upon interaction (e.g., binding) of the framework portion with an analyte. For example, signaling by the signaling portion can detectably increase or detectably decrease upon interaction (e.g., binding) of the framework portion with an analyte. In some cases, biosensors have a signal change upon binding (e.g., specific binding) to their respective analyte of at least about, for example, ± 0.5 , and/or an increase or decrease in signal of at least about, for example, 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90%, 100%, 250%, 500%, 750%, 1000%, or more than 1000%, e.g., relative to unbound biosensor. In some increases, the level of signal change is linked to background signal. Values represented here can be converted and/or expressed into any conventional units using ordinary skill. For example, units can be expressed as 'signal change' (as used above), $\Delta F/F$ and/or as signal-to-noise ratio (e.g., $\Delta F/F$ multiplied by the square root of the number of photons collected). In some instances, signaling by a biosensor can be intensity based.

[0146] In some instances, biosensors herein are distinguishable from Forster resonance energy transfer, also known as fluorescence resonance energy transfer (FRET)-based sensors, which require donor and acceptor chromophores, e.g., that function in concert, in that they include independently functioning or detectable signaling portions. For example, in some instances, signaling by a first signaling portion of a biosensor herein is independent of signaling by a second signaling portion within the same or a distinct biosensor. As noted above, signaling portions are allosterically regulated by the framework portion to which they are linked upon interaction of the framework portion with an analyte (e.g., a defined, selected, and/or specific analyte).

Framework Portions

[0147] Framework portions include genetically encoded macromolecules (e.g., proteins or peptides) that undergo conformational alteration (e.g., a structural change) upon

interaction (e.g., binding) with, or to, an analyte (e.g., an analyte-binding dependent conformational alteration). For example, genetically encoded framework portions can have a first structure in the absence of an analyte (e.g., in an unbound or open state) and a second structure, that is detectably distinct (e.g., differences in structures before and after a conformational change can be observed using methods known in the art) from the first structure, in the presence of an analyte (e.g., in a bound or closed state), e.g., under physiologic conditions. In some instances, the conformational change that occurs upon interaction with an analyte (e.g., an analyte-binding dependent conformational alteration) is detectably distinct (e.g., can be observed using methods known in the art) from a conformational change that may occur for the same protein or peptide under other physiological conditions (e.g., a change in conformation induced by altered temperature, pH, voltage, ion concentration, phosphorylation).

[0148] Methods for identifying proteins or peptides that exhibit suitable conformational characteristics and/or for observing differences in structure between structures or before and after a conformational change are known in the art and/or are described herein. Such methods can include, for example, one or more of structural analysis, crystallography, NMR, EPR using Spin label techniques, Circular Dichroism (CD), Hydrogen Exchange surface Plasmon resonance, calorimetry, and/or FRET.

[0149] In some instances, framework portions can have a first structure in the absence of an analyte (e.g., in an unbound or open state) and a second structure, that is detectably distinct (e.g., can be observed using methods known in the art) from the first structure, in the presence of an analyte (e.g., in a bound or closed state), e.g., under physiologic conditions, wherein the structural change between the open and closed state can allosterically modulate an independent signaling portion recombinantly (e.g., artificially introduced) present within the framework portion (see, e.g., Structure I).

[0150] Framework portions can also interact (e.g., bind) with at least one analyte (e.g., at least one defined, specific, and/or selected analyte). In some instances, a framework portion can interact specifically with one analyte (e.g., at least one defined, specific, and/or selected analyte). In such cases, affinity of binding between the framework binding peptide and the analyte can be high or can be controlled (e.g., with millimolar, micromolar, nanomolar, or picomolar affinity). Alternatively, the single framework binding protein can bind two or more analytes (e.g., two or more defined, specific, and/or selected analytes). In such cases, affinity of binding to the two or more analytes can be the same or distinct. For example, the affinity of binding can be greater for one analyte than it is for a second or third, etc., analyte. In some instances, binding between a framework portion and an analyte (e.g., at least one defined, specific, and/or selected analyte) have an affinity of for example, 10 mM to 1 pM.

[0151] As used herein, the term "analyte" can include naturally occurring and/or synthetic sugars, amino acids, proteins (e.g., proteins, peptides, and/or antibodies), hormones, ligands, chemicals (e.g., small molecules), pharmaceuticals, nucleic acids, cells, tissues, and combinations thereof.

[0152] In some instances, biosensors can include one, two, or more framework binding portions that bind (e.g., binds

specifically) a single analyte (e.g., a single defined, specific, and/or selected analyte) or distinct analytes (e.g., two or more distinct defined, specific and/or selected analytes). Alternatively or in addition, the framework portion can be chimeric. In such cases, a first part of the framework portion can be a first peptide or can be derived from a first peptide, and a second part of the framework portion can be a second peptide or can be derived from a second peptide, wherein the first a second peptides are combined to result in a single peptide.

[0153] Accordingly, framework portions can include macromolecules that undergo a conformational change upon interaction with an analyte. One non-limiting example of a suitable macromolecule is Calmodulin (CaM). CaM is in an extended shape in the absence of Ca^{2+} and in a condensed conformation in the presence of Ca^{2+} (Kuboniwa et al., Nat. Struc. Biol., 2:768-776, 1996 and Fallon and Quioco, Structure, 11:1303-1307, 2003).

[0154] In some instances, a framework binding portion can be a bacterial protein or can be derived from a bacterial protein. Suitable bacterial proteins can include, but are not limited to, for example, periplasmic binding proteins (PBPs).

[0155] PBPs from bacteria are generally useful in the biosensors herein at least because they undergo dramatic conformational changes upon ligand binding (Ouioco et al. Mol. Microbiol., 20:17-225, 1996). X-ray crystal structures of the apo (open) and bound (closed) forms of various PBPs reveal that these proteins have two (typically, although some have more) domains that undergo a large hinge-twist movement relative to each other in a Venus flytrap manner (Dwyer and Hellinga, Curr. Opin. Struc. Biol., 14:495-504, 2004). This conformational change has been exploited to create a number of FRET-based genetically encoded sensors (see, e.g., Deuschle et al., Pro. Sci, 14:2304-2314, 2005; Deuschle et al., Cytometry, 64:3-9, 2005; Okumoto et al., Proc. Natl. Acad. Sci. USA., 102:8740-8745, 2005; Bogner and Ludewig, J. Fluoresc., 17:350-360, 2007; and Gu et al., FEBS Letters, 580:5885-5893, 2006). In addition, the ligand-binding diversity of the PBP superfamily is large (Dwyer and Hellinga, Curr. Opin. Struc. Biol., 14:495-504, 2004).

[0156] In some instances, framework portions can include, for example, one or more of: arabinose-binding protein(s), glucose/galactose-binding protein(s), histidine-binding protein(s), maltose-binding protein(s), glutamine-binding protein(s), maltotriose-binding protein(s), RBP, ribose-binding protein(s), acetylcholine binding protein(s), choline binding protein(s), lysine binding protein(s), arginine binding protein(s), gamma aminobutyric acid (GABA) binding protein (s), ion-binding protein(s), peptide-binding protein(s), lactate-binding protein(s), histamine-binding protein(s), and/or Leucine/Isoleucine/Valine binding protein(s), including full length proteins, fragments, and/or variants thereof.

[0157] In some instances, exemplary framework portions can include: SEQ ID NO:105, which is *Escherichia coli* maltodextrin-binding protein (EcMBP) (UniProt accession number POAEX9); SEQ ID NO: 106, which is *Pyrococcus Furiosus* maltotriose-binding protein (PfMBP) (UniProt accession number P58300); SEQ ID NO:107, which is *E. coli* glutamate-binding protein (EcYbeJ) (UniProt accession number Q1R3F7); SEQ ID NO:108, which is *E. coli* phosphate-binding protein (EcPhnD) (UniProt accession number P37902); and/or SEQ ID NO:109, which is *Thermus*

thermophilus glucose-binding protein (TtGBP) (UniProt accession number Q72KX2, including full length proteins, fragments, and/or variants thereof.

[0158] In some instances, exemplary framework portions can include SEQ ID NO: 110 (UniProt accession number Q92N37); SEQ ID NO:111 (UniProt accession number D0VWx8, SEQ ID NO:112 (UniProt accession number Q7CX36), and/or SEQ ID NO:113 (UniProt accession number POAD96, including full length proteins, fragments, and/or variants thereof.

[0159] In some embodiments, exemplary framework portions can include residues 24-272 and 517-542 of SEQ ID NO: 176 (SF-iGluSnFR.A184V); residues 24-272 and 517-542 of SEQ ID NO: 177 (SF-iGluSnFR.A184S); residues 24-272 and 517-542 of SEQ ID NO: 178 (SF-iGluSnFR.S72A); residues 24-272 and 517-542 of SEQ ID NO: 179 (SF-Venus-iGluSnFR.A184V); residues 24-272 and 517-542 of SEQ ID NO: 180 (SF-Venus-iGluSnFR.A184S); residues 24-272 and 517-542 of SEQ ID NO: 181 (SF-Venus-iGluSnFR.S72A); residues 24-271 and 519-541 of SEQ ID NO: 182 (SF-Azurite-iGluSnFR); residues 24-350 and 595-664 of SEQ ID NO: 183 (iDexSnFR or SF-GlucoseSensor); residues 24-298 and 543-586 of SEQ ID NO: 184 (iGABASnFR); residues 25-99 and 348-545 of SEQ ID NO: 185 (iAChSnFR *E. coli* expression sequence); or residues 24-98 and 347-544 of SEQ ID NO: 186 (iAChSnFR mammalian expression sequence).

[0160] In some instances, framework portions, or biosensors, do not include signal peptides, or portions of signal peptides, that would otherwise be present in the peptide from which the framework portion is derived.

Signaling Portions

[0161] Biosensors herein include one or more genetically encoded signaling portions (e.g., independent signaling portions) within the amino acid sequence of a framework portion at a site(s) within the framework portion that undergo(es) a conformational change upon interaction of the framework portion with an analyte (e.g., a defined, specific, and/or selected analyte).

[0162] Signaling portions (e.g., independent signaling portions) include genetically encoded molecules (e.g., peptides or proteins) that can be allosterically induced to emit a detectable signal (e.g., an analyte-binding dependent signal).

[0163] In some instances, the detectable signal is detectably distinct (e.g., can be distinguished using methods known in the art and/or disclosed herein) from a signal emitted by the molecule prior to allosteric inducement (e.g., signaling portions can emit a detectable signal in two detectably distinct states. For example, first signal can be emitted in unbound state and a second signal can be emitted in bound state). As noted above, in some instances, the detectable signal is proportional to the degree of allosteric inducement. In some instances, if two or more signaling portions are present in a biosensor, then two or more detectably distinct signals can be emitted by the biosensor.

[0164] In some instances, a genetically encoded independent signaling portion is a genetically encoded fluorescent protein (FP), e.g., a macromolecule containing a functional group (e.g., a fluorophore) that absorbs energy of a specific wavelength and re-emits energy at a different (but equally specific) wavelength, including, for example, circularly permuted FP (cpFP). In some instances, a signaling portion is

a “superfolder” FP (e.g., Pedelacq et al., 2006, Nat. Biotech., 24:79-88), e.g., a circularly permuted SF FP.

[0165] As used herein, the term “fluorophore” relates to a functional group in a molecule which will absorb energy of a specific wavelength and re-emit energy at a different (but equally specific) wavelength. In some instances, fluorophore containing molecules include fluorescent proteins. The fluorophore in green fluorescent protein (GFP) includes Ser-Tyr-Gly sequence (i.e., Ser65-dehydroTyr66-Gly67), which is post-translationally modified to a 4-(p-hydroxybenzylidene)-imidazolidin-5. Exemplary genetically encoded fluorescent proteins include, but are not limited to, fluorescent proteins from coelenterate marine organisms, e.g., *Aequorea victoria*, *Trachyphyllia geoffroyi*, coral of the *Discosoma* genus, *Rennilla mulleri*, *Anemonia sulcata*, *Heteractis crispa*, *Entacmaea quadricolor*, and/or GFP (including the variants S65T and EGFP, *Rennilla mulleri* GFP), cyan fluorescent protein (CFP), including Cerulean, and mCerulean3 (described by Markwardt et al., PLoS ONE, 6(3) e17896.doi:10.1371/journal.pone.0017896), CGFP (CFP with Thr203Tyr: Has an excitation and emission wavelength that is intermediate between CFP and EGFP), yellow fluorescent protein (YFP, e.g., GFP-Ser65Gly/Ser72Ala/Thr203Tyr; YFP (e.g., GFP-Ser65Gly/Ser72Ala/Thr203Tyr) with Val168Leu/Gln69Lys); Citrine (i.e., YFP-Val168Leu/Gln69Met), Venus (i.e., YFP-Phe46Leu/Phe64Leu/Met153Thr/Val163Ala/Ser175Gly), PA-GFP (i.e., GFP-Val/163Ala/Thr203His), Kaede), red fluorescent protein (RFP, e.g., long wavelength fluorescent protein, e.g., DsRed (DsRed1, DsRed2, DsRed-Express, mRFP1, drFP583, dsFP593, asFP595), eqFP611, and/or other fluorescent proteins known in the art (see, e.g., Zhang et al., Nature Reviews, Molecular and Cellular Biology, 3:906-908, 2002).

[0166] As set forth above, in some instances, fluorophore containing molecules include fluorescent proteins that can be or that are circularly permuted. Circular permutation methods are known in the art (see, e.g., Baird et al., Proc. Natl. Acad. Sci., 96:11241-11246, 1999; Topell and Glockshuber, Methods in Molecular Biology, 183:31-48, 2002) as are “superfolder” (SF) proteins (e.g., Pedelacq et al., 2006, Nat. Biotech., 24:79-88) (e.g., circularly permuted SF proteins).

[0167] In some instances, single-FP sensors have a number of advantages: they preserve spectral bandwidth for multi-analyte imaging; their saturated states may be nearly as bright as the parental FP, and their ligand-free states may be arbitrarily dim, providing large theoretical fluorescence increases. This allows for much greater changes in fluorescence and thus increased signal-to-noise ratios and greater resistance to photobleaching artifacts (Tian et al., Nat. Methods, 6:875-881, 2009).

[0168] In some instances, issues arising from long-term effects such as gene regulation and protein expression and degradation can be identified by simply fusing the intensity-based sensor to a another fluorescent protein of different color, to serve as a reference channel.

[0169] In some instances, biosensors can include circularly permuted YFP (cpYFP) as a cpFP. cpYFP has been used as a reporter element in the creation of sensors for H₂O₂ (HyPer) (Belousov et al., Nat. Methods, 3:281-286, 2006), cGMP (FlnG) (Nausch et al., Proc. Natl. Acad. Sci. USA., 105:365-370, 2008), ATP:ADP ratio (Perceval) (Berg et al., Nat. Methods., 105:365-370, 2008), and calcium ions

(Nakai et al., Nat. Biotechnol., 19:137-141, 2001), including full length, fragments, and/or variants thereof.

[0170] In some embodiments, exemplary sensor portions can include residues 273-516 of SEQ ID NO: 176 (SF-iGluSnFR.A184V); residues 273-516 of SEQ ID NO: 177 (SF-iGluSnFR.A184S); residues 273-516 of SEQ ID NO: 178 (SF-iGluSnFR.S72A); residues 273-516 of SEQ ID NO: 179 (SF-Venus-iGluSnFR.A184V); residues 273-516 of SEQ ID NO: 180 (SF-Venus-iGluSnFR.A184S); residues 273-516 of SEQ ID NO: 181 (SF-Venus-iGluSnFR.S72A); residues 272-518 of SEQ ID NO: 182 (SF-Azurite-iGluSnFR); residues 351-594 of SEQ ID NO: 183 (iDexSnFR or SF-GlucoseSensor); residues 299-544 of SEQ ID NO: 184 (iGABASnFR); residues 104-343 of SEQ ID NO: 185 (iAChSnFR *E. coli* expression sequence); or residues 103-342 of SEQ ID NO: 186 (iAChSnFR mammalian expression sequence).

Linker Portions

[0171] As shown in Structure I, biosensors herein can optionally include one or more genetically encoded linkers positioned between or operably linking the framework portion and the signaling portion. Linker portions can include at least one naturally occurring or synthetic amino acid (discussed below) as exemplified by SEQ ID NOs: 9-49, 54-61, 64-76, 79-90, 95-104. In some instances, linker can include one or more of SEQ ID NOs: 9-49, 54-61, 64-76, 79-90, 95-104, and/or portions of SEQ ID NOs: 9-49, 54-61, 64-76, 79-90, 95-104. For example, linkers can include, but are not limited to, one or more of: PxSHNVY (SEQ ID NO:114), xPSHNVY (SEQ ID NO:115), xxSHNVY (SEQ ID NO:116), xxSHNVF (SEQ ID NO:117), PxSHNVF (SEQ ID NO:118), PxSYNVF (SEQ ID NO:119), xxSYNVF (SEQ ID NO:120), PxSYNVF (SEQ ID NO:121), xxSYNVF (SEQ ID NO:122), PxSxNVY (SEQ ID NO:123), PxSHxVY (SEQ ID NO:124), PxSHNxY (SEQ ID NO:125), PxSHNVx (SEQ ID NO:126), FNxxY (SEQ ID NO:127), FNxY (SEQ ID NO:128), FNY (SEQ ID NO:129), FxY (SEQ ID NO:130), xxY (SEQ ID NO:131), WxY (SEQ ID NO:132), xKY, (SEQ ID NO:133), FNPxY (SEQ ID NO:134), FNxPY (SEQ ID NO:135), HNS (SEQ ID NO:136), GGS (SEQ ID NO:137), xxS (SEQ ID NO:138), xxK (SEQ ID NO:139), GGK (SEQ ID NO:140), PXS (SEQ ID NO:141), xPS (SEQ ID NO:142), Px (SEQ ID NO:143), xP (SEQ ID NO:144), IxxS (SEQ ID NO:145), NxPK (SEQ ID NO:146), NPcK (SEQ ID NO:147), PPxSH (SEQ ID NO:148), PPxxSH (SEQ ID NO:149), PPPxSH (SEQ ID NO:150), PPxPSH (SEQ ID NO:151), xxSH (SEQ ID NO:152), PPxx (SEQ ID NO:153), FNxxKN (SEQ ID NO:154), FNxxKN (SEQ ID NO:155), FNxxPKN (SEQ ID NO:156), FNPxxKN (SEQ ID NO:157), FNxx (SEQ ID NO:158), N, ADGSSH (SEQ ID NO:159), ADxxSH (SEQ ID NO:160), ADxPSH (SEQ ID NO:161), ADPxxSH (SEQ ID NO:162), ADxx (SEQ ID NO:163), ADxxSH (SEQ ID NO:164), FNPG (SEQ ID NO:165), FNxxPG (SEQ ID NO:166), xxPG (SEQ ID NO:167), FNxx (SEQ ID NO:168), FNPx (SEQ ID NO:169), KYxxSH (SEQ ID NO:170), KYPxSH (SEQ ID NO:171), KYxPSH (SEQ ID NO:172), FxxP (SEQ ID NO:173), FNxP (SEQ ID NO:174), and/or FNPx (SEQ ID NO:175), where “x” indicates any amino acid.

[0172] In some embodiments, exemplary linker portions can include residues 365-370 of SEQ ID NO: 176 (SF-iGluSnFR.A184V); residues 365-370 of SEQ ID NO: 177

(SF-iGluSnFR.A184S); residues 365-370 of SEQ ID NO: 178 (SF-iGluSnFR.S72A); residues 365-370 of SEQ ID NO: 179 (SF-Venus-iGluSnFR.A184V); residues 365-370 of SEQ ID NO: 180 (SF-Venus-iGluSnFR.A184S); residues 365-370 of SEQ ID NO: 181 (SF-Venus-iGluSnFR.S72A); residues 365-370 of SEQ ID NO: 182 (SF-Azurite-iGluSnFR); residues 443-448 of SEQ ID NO: 183 (iDexSnFR or SF-GlucoseSensor); residues 391-396 of SEQ ID NO: 184 (iGABASnFR); residues 100-103 and 344-347 of SEQ ID NO: 185 (iAChSnFR *E. coli* expression sequence); or residues 99-102 and 343-346 of SEQ ID NO: 186 (iAChSnFR mammalian expression sequence).

Exemplary Biosensor Constructs

[0173] As noted above, biosensors herein include genetically encoded biosensors, i.e., nucleic acids encoding biosensors, and/or the encoded biosensors (e.g., isolated biosensors), for use as biosensors. In some instances, nucleic acids encoding biosensors include isolated nucleic acids. In some instances, the portion of a nucleic acid encoding a biosensor can include a single reading frame encoding the biosensor. For example, a biosensor can be encoded by a portion of a nucleic acid that falls within a start codon and a stop codon. In some instances, biosensors are isolated (e.g., biosensors are substantially free of contaminating and/or non-biosensor components).

[0174] In some instances, biosensors can include, for example, one or more framework portions selected from the group consisting of: arabinose-binding protein(s), glucose/galactose-binding protein(s), histidine-binding protein(s), maltose-binding protein(s), maltotriose-binding protein(s), glutamine-binding protein(s), RBP, ribose-binding protein(s), acetylcholine binding protein(s), choline binding protein(s), lysine binding protein(s), arginine binding protein(s), gamma aminobutyric acid (GABA) binding protein(s), ion-binding protein(s), peptide-binding protein(s), lactate-binding protein(s), histamine-binding protein(s), and/or Leucine/Isoleucine/Valine binding protein(s), including full length proteins, fragments, and/or variants thereof, including full length proteins, fragments and/or variants thereof, and at least one independent signaling portion present at a site within the framework portion that undergoes a conformational change upon interaction of the framework portion with an analyte.

[0175] In some instances, biosensors can include, for example, one or more framework portions selected from the group consisting of: SEQ ID NO:105, which is *Escherichia coli* maltodextrin-binding protein (EcMBP) (UniProt accession number POAEX9); SEQ ID NO: 106, which is *Pyrococcus Furiosus* maltose-binding protein (PfMBP) (UniProt accession number P58300); SEQ ID NO:107, which is *E. coli* glutamate-binding protein (EcYbeJ) (UniProt accession number Q1R3F7); SEQ ID NO:108, which is *E. coli* phosphonate-binding protein (EcPhnD) (UniProt accession number P37902); and/or SEQ ID NO:109, which is *Thermus thermophilus* glucose-binding protein (TtGBP) (UniProt accession number Q72KX2), including full length proteins, fragments and/or variants thereof, and at least one independent signaling portion present at a site within the framework portion that undergoes a conformational change upon interaction of the framework portion with an analyte.

[0176] In some instances, biosensors can include, for example, one or more framework portions selected from the group consisting of: SEQ ID NO: 110 (UniProt accession

number Q92N37); SEQ ID NO:111 (UniProt accession number DOVWx8, SEQ ID NO:112 (UniProt accession number Q7CX36), and/or SEQ ID NO:113 (UniProt accession number POAD96), including full length proteins, fragments and/or variants thereof, and at least one independent signaling portion present at a site within the framework portion that undergoes a conformational change upon interaction of the framework portion with an analyte.

[0177] In some instances, biosensors include any one or more:

[0178] Maltose biosensors SEQ ID NOs: 1-8 (e.g., *Escherichia coli* maltodextrin-binding protein (EcMBP)) or SEQ ID NOs: 50-53 (e.g., *Pyrococcus furiosus* maltose-binding protein (PfMBP)), including full length proteins, fragments and/or variants thereof;

[0179] Glutamate biosensors SEQ ID NOs: 62-63 (e.g., *E. coli* glutamate-binding protein (EcYbeJ)) or SEQ ID NOs: 176-182, including full length proteins, fragments and/or variants thereof;

[0180] Phosphonate biosensors SEQ ID NOs: 77-78 (e.g., *E. coli* phosphonate-binding protein (EcPhnD)), including full length proteins, fragments and/or variants thereof;

[0181] Glucose biosensors SEQ ID NOs: 91-94 (e.g., *Thermus thermophilus* glucose-binding protein (TtGBP)) and SEQ ID NO: 183, including full length proteins, fragments and/or variants thereof;

[0182] GABA biosensors SEQ ID NO: 184, including full length proteins, fragments and/or variants thereof; and/or

[0183] ACh biosensors SEQ ID NOs: 185 & 186, including full length proteins, fragments and/or variants thereof.

[0184] In some instances, nucleic acids encoding, and/or amino acid sequences of, any of the framework portions, signaling portions, linker portions, or the entire biosensor sequence (e.g., SEQ ID NOs: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, 53, 62, 63, 77, 78, 91, 92, 93, 94, and/or 176-186) (e.g., any amino acid sequence) disclosed herein can be modified to generate fragments (e.g., truncated peptides) and/or variants (e.g., peptides with a defined sequence homology to the peptides disclosed herein). Variants can include framework portions, signaling portions, linker portions, or biosensors with amino acid sequences with homology to the framework portions, signaling portions, linker portions, or biosensors disclosed herein and/or truncated forms of the framework portions, signaling portions, linker portions, or biosensors herein. In some instances, truncated forms of the framework portions, signaling portions, linker portions, or biosensors herein can include 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 50-100, 101-150, fewer amino acids than the framework portions, signaling portions, linker portions, and/or biosensors herein, e.g., wherein the truncated biosensor variants retain at least a portion of the binding and/or signaling properties of same biosensor without truncation (e.g., at least 50%, 60%, 70%, 80%, 90%, or 100% of the binding and/or signaling properties of the same biosensor without truncation). In addition, truncations can be made at the amino-terminus, the carboxy-terminus, and/or within the body of the framework portions, signaling portions, linker portions, and/or biosensors herein.

[0185] While variants are generally observed and discussed at the amino acid level, the actual modifications are typically introduced or performed at the nucleic acid level. For example, variants with 95%, 96%, 97%, 98, or 99%

sequence identity to SEQ ID NOs: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, 53, 62, 63, 77, 78, 91, 92, 93, 94, and/or 176-186 can be generated by modifying the nucleic acids encoding SEQ ID NOs: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, 53, 62, 63, 77, 78, 91, 92, 93, 94, and/or 176-186 using techniques (e.g., cloning techniques) known in the art and/or that are disclosed herein.

[0186] As with all peptides, polypeptides, and proteins, including fragments thereof, it is understood that modifications to the amino acid sequence can occur that do not alter the nature or function of the peptides, polypeptides, or proteins. Such modifications include conservative amino acids substitutions and are discussed in greater detail below.

[0187] The peptides, polypeptides, and proteins, including fragments thereof, provided herein are biosensors whose activity can be tested or verified, for example, using the in vitro and/or in vivo assays described herein.

[0188] In some instances, any of the framework portions, signaling portions, or the biosensor sequence (e.g., SEQ ID NOs: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, 53, 62, 63, 77, 78, 91, 92, 93, 94, and/or 176-186) (e.g., any amino acid sequence) described herein can be modified and varied so long as their desired function is maintained. For example, the polypeptides can be modified as long as the resulting variant polypeptides have the same or better characteristics as the polypeptide from which they derived. For example, the variants can have the same or better affinity for their respective analyte.

[0189] In some instances, the interacting face of a modified peptide can be the same (e.g., substantially the same) as an unmodified peptide (methods for identifying the interacting face of a peptide are known in the art (Gong et al., *BMC: Bioinformatics*, 6:1471-2105 (2007); Andrade and Wei et al., *Pure and Appl. Chem.*, 64(11):1777-1781 (1992); Choi et al., *Proteins: Structure, Function, and Bioinformatics*, 77(1):14-25 (2009); Park et al., *BMC: and Bioinformatics*, 10:1471-2105 (2009)), e.g., to maintain binding to an analyte. Alternatively, amino acids within the interacting face can be modified, e.g., to decrease binding to an analyte and/or to change analyte specificity.

[0190] The interacting face of a peptide is the region of the peptide that interacts or associates with other molecules (e.g., other proteins). Generally, amino acids within the interacting face are naturally more highly conserved than those amino acids located outside the interacting face or interface regions of a protein. In some instances, an amino acid within the interacting face region of any of the framework portions or the biosensor sequence (e.g., SEQ ID NOs: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, 53, 62, 63, 77, 78, 91, 92, 93, 94, and/or 176-186) (e.g., any amino acid sequence) disclosed herein can be the same as the amino acid shown in any of the framework portions or the biosensor sequence (e.g., SEQ ID NOs: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, 53, 62, 63, 77, 78, 91, 92, 93, 94, and/or 176-186) (e.g., any amino acid sequence) disclosed herein or can include conservative amino acid substitutions. In some instances, an amino acid within the interacting face region any of the framework portions or the biosensor sequence (e.g., SEQ ID NOs: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, 53, 62, 63, 77, 78, 91, 92, 93, 94, and/or 176-186) (e.g., any amino acid sequence) disclosed herein can be substituted with an amino acid that increases the interaction between the framework portion or the biosensor sequence (e.g., SEQ ID NOs: 1, 2, 3, 4, 5, 6, 7, 8, 50,

51, 52, 53, 62, 63, 77, 78, 91, 92, 93, 94, and/or 176-186) (e.g., any amino acid sequence) and an analyte.

[0191] In some instances, genetically encoded biosensors can include peptides that have at least 80, 85, 90, 95, 96, 97, 98, 99 percent identity to the framework portions, signaling portions, or the biosensor sequence (e.g., SEQ ID NOs: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, 53, 62, 63, 77, 78, 91, 92, 93, 94, and/or 176-186) (e.g., any amino acid sequence) described herein. Those of skill in the art readily understand how to determine the identity of two polypeptides. For example, the identity can be calculated after aligning the two sequences so that the identity is at its highest level.

[0192] Another way of calculating identity can be performed by published algorithms. Optimal alignment of sequences for comparison may be conducted by the local identity algorithm of Smith and Waterman, *Adv. Appl. Math.*, 2:482 (1981), by the identity alignment algorithm of Needleman and Wunsch, *J. Mol. Biol.* 48:443 (1970), by the search for similarity method of Pearson and Lipman, *Proc. Natl. Acad. Sci. USA* 85:2444 (1988), by computerized implementations of these algorithms (GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group, 575 Science Dr., Madison, Wis.), or by inspection.

[0193] The same types of identity can be obtained for nucleic acids by, for example, the algorithms disclosed in Zuker, *Science* 244:48-52 (1989); Jaeger et al., *Proc. Natl. Acad. Sci. USA* 86:7706-10 (1989); Jaeger et al., *Methods Enzymol.* 183:281-306 (1989), which are herein incorporated by reference for at least material related to nucleic acid alignment. It is understood that any of the methods typically can be used and that in certain instances the results of these various methods may differ, but the skilled artisan understands if identity is found with at least one of these methods, the sequences would be said to have the stated identity and to be disclosed herein.

[0194] Amino acid sequence modifications typically fall into one or more of three classes: substitutional, insertional, or deletional modifications. Insertions include amino and/or terminal fusions as well as intra-sequence insertions of single or multiple amino acid residues. Insertions ordinarily will be smaller insertions than those of amino or carboxyl terminal fusions, for example, on the order of one to four residues. Deletions are characterized by the removal of one or more amino acid residues from the protein sequence. Typically, no more than about from 2 to 6 residues are deleted at any one site within the protein molecule. Amino acid substitutions are typically of single residues, but can occur at a number of different locations at once; insertions usually will be on the order of about from 1 to 10 amino acid residues; and deletions will range about from 1 to 30 residues. Deletions or insertions can be made in adjacent pairs, i.e., a deletion of 2 residues or insertion of 2 residues. Substitutions, deletions, insertions or any combination thereof may be combined to arrive at a final construct. The mutations must not place the sequence out of reading frame and preferably will not create complementary regions that could produce secondary mRNA structure. Substitutional modifications are those in which at least one residue has been removed and a different residue inserted in its place. In some instances, substitutions can be conservative amino acid substitutions. In some instances, variants herein can include one or more conservative amino acid substitutions. For example, variants can include 1, 2, 3, 4, 5, 6, 7, 8, 9, 10,

11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 20-30, 30-40, or 40-50 conservative amino acid substitutions. Alternatively, variants can include 50 or fewer, 40 or fewer, 30 or fewer, 20 or fewer, 10 or fewer, 9 or fewer, 8 or fewer, 7 or fewer, 6 or fewer, 5 or fewer, 4 or fewer, 3 or fewer, or 2 or fewer conservative amino acid substitutions. Such substitutions generally are made in accordance with the following Table 1 and are referred to as conservative substitutions. Methods for predicting tolerance to protein modification are known in the art (see, e.g., Guo et al., Proc. Natl. Acad. Sci., USA, 101(25):9205-9210 (2004)).

TABLE 1

Conservative Amino Acid Substitutions	
Amino Acid	Substitutions (others are known in the art)
Ala	Ser, Gly, Cys
Arg	Lys, Gln, His
Asn	Gln, His, Glu, Asp
...	
Asp	Glu, Asn, Gln
Cys	Ser, Met, Thr
Gln	Asn, Lys, Glu, Asp, Arg
Glu	Asp, Asn, Gln
Gly	Pro, Ala, Ser
His	Asn, Gln, Lys
Ile	Leu, Val, Met, Ala
Leu	Ile, Val, Met, Ala
Lys	Arg, Gln, His
Met	Leu, Ile, Val, Ala, Phe
Phe	Met, Leu, Tyr, Trp, His
Ser	Thr, Cys, Ala
Thr	Ser, Val, Ala
Trp	Tyr, Phe
Tyr	Trp, Phe, His
Val	Ile, Leu, Met, Ala, Thr

[0195] In some instances, substitutions are not conservative. For example, an amino acid can be replaced with an amino acid that can alter some property or aspect of the peptide. In some instances, non-conservative amino acid substitutions can be made, e.g., to change the structure of a peptide, to change the binding properties of a peptide (e.g., to increase or decrease the affinity of binding of the peptide to an analyte and/or to alter increase or decrease the binding specificity of the peptide).

[0196] Modifications, including the specific amino acid substitutions, are made by known methods. By way of example, modifications are made by site-specific mutagenesis of nucleotides in the DNA encoding the protein, thereby producing DNA encoding the modification, and thereafter expressing the DNA in recombinant cell culture. Techniques for making substitution mutations at predetermined sites in DNA having a known sequence are well known, for example M13 primer mutagenesis and PCR mutagenesis.

Nucleic Acids

[0197] The disclosure also features nucleic acids encoding the biosensors (e.g., SEQ ID NOs: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, 53, 62, 63, 77, 78, 91, 92, 93, 94, and/or 176-186) described herein, including variants and/or fragments of the biosensors (e.g., variants and/or fragments of SEQ ID NOs: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, 53, 62, 63, 77, 78, 91, 92, 93, 94, and/or 176-186). These sequences include all degenerate sequences related to the specific polypeptide sequence, i.e., all nucleic acids having a sequence that encodes one

particular polypeptide sequence as well as all nucleic acids, including degenerate nucleic acids, encoding the disclosed variants and derivatives of the polypeptide sequences. Thus, while each particular nucleic acid sequence may not be written out herein, it is understood that each and every sequence is in fact disclosed and described herein through the disclosed polypeptide sequences.

[0198] In some instances, nucleic acids can encode biosensors with 95, 96, 97, 98, or 99 identity to SEQ ID NOs: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, 53, 62, 63, 77, 78, 91, 92, 93, 94, and/or 176-186.

[0199] In some instances, nucleic acids can encode SEQ ID NOs: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, 53, 62, 63, 77, 78, 91, 92, 93, 94, and/or 176-186 containing 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 20-30, 30-40, or 40-50 conservative amino acid substitutions.

[0200] In some instances, nucleic acids can encode SEQ ID NOs: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, 53, 62, 63, 77, 78, 91, 92, 93, 94, and/or 176-186 containing 50 or fewer, 40 or fewer, 30 or fewer, 20 or fewer, 10 or fewer, 9 or fewer, 8 or fewer, 7 or fewer, 6 or fewer, 5 or fewer, 4 or fewer, 3 or fewer, or 2 or fewer conservative amino acid substitutions.

[0201] Also provided herein are vectors comprising the biosensors (e.g., SEQ ID NOs: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, 53, 62, 63, 77, 78, 91, 92, 93, 94, and/or 176-186) described herein, including variants and/or fragments of the biosensors (e.g., SEQ ID NOs: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, 53, 62, 63, 77, 78, 91, 92, 93, 94, and/or 176-186). For example:

[0202] Vectors can include nucleic acids that encode biosensors with 95, 96, 97, 98, or 99 identity to SEQ ID NOs: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, 53, 62, 63, 77, 78, 91, 92, 93, 94, and/or 176-186.

[0203] Vectors can include nucleic acids that encode SEQ ID NOs: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, 53, 62, 63, 77, 78, 91, 92, 93, 94, and/or 176-186 containing 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 20-30, 30-40, or 40-50 conservative amino acid substitutions.

[0204] Vectors can include nucleic acids that encode SEQ ID NOs: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, 53, 62, 63, 77, 78, 91, 92, 93, 94, and/or 176-186 containing 50 or fewer, 40 or fewer, 30 or fewer, 20 or fewer, 10 or fewer, 9 or fewer, 8 or fewer, 7 or fewer, 6 or fewer, 5 or fewer, 4 or fewer, 3 or fewer, or 2 or fewer conservative amino acid substitutions.

[0205] Examples of suitable vectors include, but are not limited to, plasmids, artificial chromosomes, such as BACs, YACs, or PACs, and viral vectors. As used herein, vectors are agents that transport the disclosed nucleic acids into a cell without degradation and, optionally, include a promoter yielding expression of the nucleic acid molecule in the cells into which it is delivered.

[0206] Viral vectors can include, for example, Adenovirus, Adeno-associated virus, herpes virus, Vaccinia virus, Polio virus, Sindbis, and other RNA viruses, including these viruses with the HIV backbone. Any viral families which share the properties of these viruses which make them suitable for use as vectors are suitable. Retroviral vectors, in general are described by Coffin et al., Retroviruses, Cold Spring Harbor Laboratory Press (1997), which is incorporated by reference herein for the vectors and methods of making them. The construction of replication-defective adenoviruses has been described (Berkner et al., J. Virology 61:1213-20 (1987); Massie et al., Mol. Cell. Biol. 6:2872-83 (1986); Haj-Ahmad et al., J. Virology 57:267-74 (1986);

Davidson et al., *J. Virology* 61:1226-39 (1987); Zhang et al., *BioTechniques* 15:868-72 (1993)). Recombinant adenoviruses have been shown to achieve high efficiency after direct, in vivo delivery to airway epithelium, hepatocytes, vascular endothelium, CNS parenchyma, and a number of other tissue sites. Other useful systems include, for example, replicating and host-restricted non-replicating Vaccinia virus vectors.

[0207] Non-viral based vectors can include expression vectors comprising nucleic acid molecules and nucleic acid sequences encoding polypeptides, wherein the nucleic acids are operably linked to an expression control sequence. Suitable vector backbones include, for example, those routinely used in the art such as plasmids, artificial chromosomes, BACs, YACs, or PACs. Numerous vectors and expression systems are commercially available from such corporations as Novagen (Madison, Wis.), Clontech (Pal Alto, Calif.), Stratagene (La Jolla, Calif.), and Invitrogen/Life Technologies (Carlsbad, Calif.). Vectors typically contain one or more regulatory regions. Regulatory regions include, without limitation, promoter sequences, enhancer sequences, response elements, protein recognition sites, inducible elements, protein binding sequences, 5' and 3' untranslated regions (UTRs), transcriptional start sites, termination sequences, polyadenylation sequences, and introns.

[0208] Promoters controlling transcription from vectors in mammalian host cells may be obtained from various sources, for example, the genomes of viruses such as polyoma,

[0209] Simian Virus 40 (SV40), adenovirus, retroviruses, hepatitis B virus, and most preferably cytomegalovirus (CMV), or from heterologous mammalian promoters, e.g. β -actin promoter or EF1 α promoter, or from hybrid or chimeric promoters (e.g., CMV promoter fused to the β -actin promoter). Of course, promoters from the host cell or related species are also useful herein.

[0210] Enhancer generally refers to a sequence of DNA that functions at no fixed distance from the transcription start site and can be either 5' or 3' to the transcription unit. Furthermore, enhancers can be within an intron as well as within the coding sequence itself. They are usually between 10 and 300 base pairs in length, and they function in cis. Enhancers usually function to increase transcription from nearby promoters. Enhancers can also contain response elements that mediate the regulation of transcription. While many enhancer sequences are known from mammalian genes (globin, elastase, albumin, fetoprotein, and insulin), enhancers derived from a eukaryotic cell viruses can be used. Examples of such can include the SV40 enhancer on the late side of the replication origin, the cytomegalovirus early promoter enhancer, the polyoma enhancer on the late side of the replication origin, and adenovirus enhancers.

[0211] The promoter and/or the enhancer can be inducible (e.g. chemically or physically regulated). A chemically regulated promoter and/or enhancer can, for example, be regulated by the presence of alcohol, tetracycline, a steroid, or a metal. A physically regulated promoter and/or enhancer can, for example, be regulated by environmental factors, such as temperature and light. Optionally, the promoter and/or enhancer region can act as a constitutive promoter and/or enhancer to maximize the expression of the region of the transcription unit to be transcribed. In certain vectors, the promoter and/or enhancer region can be active in a cell type

specific manner. Optionally, in certain vectors, the promoter and/or enhancer region can be active in all eukaryotic cells, independent of cell type. Promoters of this type can include the CMV promoter, the SV40 promoter, the β -actin promoter, the EF1 α promoter, and the retroviral long terminal repeat (LTR).

[0212] The provided vectors also can include, for example, origins of replication and/or markers. A marker gene can confer a selectable phenotype, e.g., antibiotic resistance, on a cell. The marker product is used to determine if the vector has been delivered to the cell and once delivered is being expressed. Examples of selectable markers for mammalian cells are dihydrofolate reductase (DHFR), thymidine kinase, neomycin, neomycin analog G418, hygromycin, puromycin, and blasticidin. When such selectable markers are successfully transferred into a mammalian host cell, the transformed mammalian host cell can survive if placed under selective pressure. Examples of other markers include, for example, the *E. coli* lacZ gene, green fluorescent protein (GFP), and luciferase. In addition, an expression vector can include a tag sequence designed to facilitate manipulation or detection (e.g., purification or localization) of the expressed polypeptide. Tag sequences, such as GFP, glutathione S-transferase (GST), polyhistidine, c-myc, hemagglutinin, or FLAGTM tag (Kodak; New Haven, Conn.) sequences typically are expressed as a fusion with the encoded polypeptide. Such tags can be inserted anywhere within the polypeptide including at either the carboxyl or amino terminus.

[0213] The disclosure further provides cells comprising the biosensors (e.g., SEQ ID NOs: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, 53, 62, 63, 77, 78, 91, 92, 93, 94, and/or 176-186) described herein, including variants and/or fragments of the biosensors (e.g., SEQ ID NOs: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, 53, 62, 63, 77, 78, 91, 92, 93, 94, and/or 176-186). Cells can include, for example, eukaryotic and/or prokaryotic cells. For example, cells can include, but are not limited to cells of *E. coli*, *Pseudomonas*, *Bacillus*, *Streptomyces*; fungi cells such as yeasts (*Saccharomyces*, and methylotrophic yeast such as *Pichia*, *Candida*, *Hansenula*, and *Torulopsis*); and animal cells, such as CHO, R1.1, B-W and LM cells, African Green Monkey kidney cells (for example, COS 1, COS 7, BSC1, BSC40, and BMT10), insect cells (for example, Sf9), human cells and plant cells. Suitable human cells can include, for example, HeLa cells or human embryonic kidney (HEK) cells. In general, cells that can be used herein are commercially available from, for example, the American Type Culture Collection (ATCC), P.O. Box 1549, Manassas, Va. 20108. See also F. Ausubel et al., *Current Protocols in Molecular Biology*, John Wiley & Sons, New York, N.Y., (1998).

[0214] Optionally, the biosensors (e.g., SEQ ID NOs: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, 53, 62, 63, 77, 78, 91, 92, 93, 94, and/or 176-186) described herein, including variants and/or fragments of the biosensors (e.g., SEQ ID NOs: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, 53, 62, 63, 77, 78, 91, 92, 93, 94, and/or 176-186) can be located in the genome of the cell (e.g., can be stably expressed in the cell) or can be transiently expressed in the cell.

[0215] Methods of making the provided cells are known and the method of transformation and choice of expression vector will depend on the host system selected. Transformation and transfection methods are described, e.g., in F. Ausubel et al., *Current Protocols in Molecular Biology*, John

Wiley & Sons, New York, N.Y., (1998), and, as described above, expression vectors may be chosen from examples known in the art.

[0216] There are a number of compositions and methods which can be used to deliver the nucleic acid molecules and/or polypeptides to cells, either *in vitro* or *in vivo* via, for example, expression vectors. These methods and compositions can largely be broken down into two classes: viral based delivery systems and non-viral based delivery systems. Such methods are well known in the art and readily adaptable for use with the compositions and methods described herein.

[0217] By way of example, the provided polypeptides and/or nucleic acid molecules can be delivered via virus like particles. Virus like particles (VLPs) consist of viral protein (s) derived from the structural proteins of a virus. Methods for making and using virus like particles are described in, for example, Garcea and Gissmann, *Current Opinion in Biotechnology* 15:513-7 (2004). The provided polypeptides can be delivered by subviral dense bodies (DBs). DBs transport proteins into target cells by membrane fusion. Methods for making and using DBs are described in, for example, Pepperl-Klindworth et al., *Gene Therapy* 10:278-84 (2003). The provided polypeptides can be delivered by tegument aggregates. Methods for making and using tegument aggregates are described in International Publication No. WO 2006/110728.

[0218] Also provided are transgenic animals comprising one or more cells the biosensors (e.g., SEQ ID NOs: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, 53, 62, 63, 77, 78, 91, 92, 93, 94, and/or 176-186) described herein, including variants and/or fragments of the biosensors (e.g., SEQ ID NOs: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, 53, 62, 63, 77, 78, 91, 92, 93, 94, and/or 176-186). As used herein, the term animal refers to non-human animals, including, mammals, amphibians and birds. Specifically, examples include sheep, feline, bovines, ovines, pigs, horses, rabbits, guinea pigs, mice, hamsters, rats, non-human primates, and the like. As used herein, transgenic animal refers to any animal, in which one or more of the cells of the animal contain a heterologous nucleic acid. The heterologous nucleic acid can be introduced using known transgenic techniques. The nucleic acid is introduced into the cell, directly or indirectly. For example, the nucleic acid can be introduced into a precursor of the cell or by way of deliberate genetic manipulation, such as by microinjection or by infection with a recombinant virus. The nucleic acid may be integrated within a chromosome, or it may be an extrachromosomally replicating DNA.

[0219] Methods for making transgenic animals using a variety of transgenes have been described in Wagner et al. (1981) *Proc. Nat. Acad. Sci. USA*, 78:5016-5020; Stewart et al. (1982) *Science*, 217:1046-1048; Constantini et al. (1981) *Nature*, 294:92-94; Lacy et al. (1983) *Cell*, 34:343-358; McKnight et al. (1983) *Cell*, 34:335-341; Brinstar et al. (1983) *Nature*, 306:332-336; Palmiter et al. (1982) *Nature*, 300:611-615; Palmiter et al. (1982) *Cell*, 29:701-710; and Palmiter et al. (1983) *Science*, 222:809-814. Such methods are also described in U.S. Pat. Nos. 6,175,057; 6,180,849; and 6,133,502.

[0220] By way of example, the transgenic animal can be created by introducing a nucleic acid into, for example, an embryonic stem cell, an unfertilized egg, a fertilized egg, a spermatozoon or a germinal cell containing a primordial germinal cell thereof, preferably in the embryogenic stage in

the development of a non-human mammal (more preferably in the single-cell or fertilized cell stage and generally before the 8-cell phase). The nucleic acid can be introduced by known means, including, for example, the calcium phosphate method, the electric pulse method, the lipofection method, the agglutination method, the microinjection method, the particle gun method, the DEAE-dextran method and other such method. Optionally, the nucleic acid is introduced into a somatic cell, a living organ, a tissue cell or other cell by gene transformation methods. Cells including the nucleic acid may be fused with the above-described germinal cell by a commonly known cell fusion method to create a transgenic animal.

[0221] For embryonic stem (ES) cells, an ES cell line may be employed, or embryonic cells may be obtained freshly from a host, e.g., mouse, rat, guinea pig, and the like. Such cells are grown on an appropriate fibroblast-feeder layer or grown in the presence of appropriate growth factors, such as leukemia inhibiting factor (LIF). When ES cells have been transformed, they may be used to produce transgenic animals. After transformation, the cells are plated onto a feeder layer in an appropriate medium. Cells containing the construct may be detected by employing a selective medium. After sufficient time for colonies to grow, they are picked and analyzed for the occurrence of homologous recombination or integration of the nucleic acid. Those colonies that are positive may then be used for embryo manipulation and blastocyst injection. Blastocysts are obtained from 4 to 6 week old superovulated females. The ES cells are trypsinized, and the modified cells are injected into the blastocoel of the blastocyst. After injection, the blastocysts are returned to each uterine horn of pseudopregnant females. Females are then allowed to go to term and the resulting litters screened for mutant cells having the construct. By providing for a different phenotype of the blastocyst and the ES cells, chimeric progeny can be readily detected. The chimeric animals are screened for the presence of the nucleic acid, and males and females having the modification are mated to produce homozygous progeny transgenic animals.

[0222] Kits comprising one or more containers and the nucleic acid sequences, polypeptides, vectors, cells, provided herein, or combinations thereof, are also provided. For example, provided is a kit comprising (i) a nucleic acid sequence encoding a biosensor described herein (e.g., one or more of SEQ ID NOs: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, 53, 62, 63, 77, 78, 91, 92, 93, 94, and/or 176-186), including variants and/or fragments of the biosensor (e.g., variants or fragments of one or more of SEQ ID NOs: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, 53, 62, 63, 77, 78, 91, 92, 93, 94, and/or 176-186), (ii) a polypeptide comprising a biosensor described herein (e.g., one or more of SEQ ID NOs: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, 53, 62, 63, 77, 78, 91, 92, 93, 94, and/or 176-186), including variants and/or fragments of the biosensor (e.g., variants or fragments of one or more of SEQ ID NOs: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, 53, 62, 63, 77, 78, 91, 92, 93, 94, and/or 176-186), (iii) a vector comprising the nucleic acid of (i), (iv) a cell comprising the nucleic acid or (i) and/or the polypeptide of (ii), (v) a cell comprising the vector of (iii). The kit can comprise any combination of (i)-(v). Optionally, the kit further comprises reagents for using the nucleic acid or peptide biosensors, vectors, and/or cells. For example, if the kit comprises cells, the kit may also comprise cell culture medium. Optionally, the kit further

comprises instructions for use. Optionally, the kit further comprises a GPCR, a GPCR-encoding nucleic acid sequence.

Design and Production/Manufacture Methods

[0223] Using the methods described herein, it is possible to design, produce, and/or adapt genetically encoded biosensors to assays for a variety of classes of analytes. The provided materials and methods facilitate the discovery of new compounds targeting a wide array of protein targets, including but not limited to: endogenous targets responsible for disease state progression, targets on pathogens for treating infectious diseases, and endogenous targets to be avoided (thus screening early for potential drug side effects and toxicity).

[0224] Methods herein provide systematic and generic approaches for the design and production of genetically encoded recombinant peptides containing an analyte-binding framework portion linked (e.g., operably linked) to a signaling portion, wherein the signaling portion is allosterically modulated or regulated by the framework portion upon interaction of the framework portion with an analyte. Generally, methods include: (i) selecting one or more target analytes; (ii) selecting a framework portion (e.g., a PBP) that interacts with (e.g., interacts specifically with) or binds to (e.g., binds specifically to) the target analyte and that undergoes a conformational change upon interacting with or binding to the analyte; (iii) identifying sites or amino acid positions within the framework portion (e.g., the PBP) where the conformational change occurs; and (iv) inserting or cloning a signaling portion into the site or amino acid position identified in (iii). Methods can, optionally, further include: (v) modifying or optimizing linker sequences between the framework portion and the signaling portion, for example, by genetic manipulation (e.g., by point mutation); (vi) modifying or optimizing analyte binding; (vii) modifying the signal generated by the biosensor; and/or (viii) cloning the biosensor into a suitable vector.

[0225] In some instances: (iii) includes identification of insertion sites by analysis of the structure (e.g., crystal structure) of the selected framework portion (e.g., the selected PBP) in one or both of its open and closed states to determine amino acid positions at which analyte-binding dependent structural changes occur. In instances where structures for both open and closed states are not available, analysis can be conducted by analogy to a structurally similar framework portion (e.g., PBP); (iv) includes cloning a signaling portion (e.g., a cpFP) at the site identified in (iii) such that the analyte-binding dependent structural change observed in (iii) will result in a conformational change in the signaling portion (e.g., the cpFP) and allosteric modulation of the signaling portion; (v) includes generating a library of mutants of biosensors with distinct linker sequences (e.g., by point mutation), screening the library of mutants to identify mutants with enhanced properties (e.g., improved signal-to-noise ratio), and selecting mutants with enhanced properties (e.g., improved signal-to-noise ratio); (vi) includes increasing or decreasing binding or affinity of the framework portion to the analyte, e.g., by modifying amino acids in the interacting face of the framework portion or regions within the framework portion that are critical for analyte binding; (vii) includes increasing or decreasing signal emission by the signaling portion and/or changing the color of the signal

where the signaling portion is a FP (e.g., a cpFP). Methods including (i)-(viii) are exemplified in the Examples section herein.

Methods of Use

[0226] The disclosure further provides methods for using the biosensors disclosed herein (e.g., one or more of SEQ ID NOs: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, 53, 62, 63, 77, 78, 91, 92, 93, 94, and/or 176-186), including variants and/or fragments of the biosensor (e.g., variants or fragments of one or more of SEQ ID NOs: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, 53, 62, 63, 77, 78, 91, 92, 93, 94, and/or 176-186) to detect analytes, e.g., in biological systems. Such methods can include, for example:

[0227] Use of a maltose biosensor disclosed herein (e.g., one or more of SEQ ID NOs: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, and/or 53 including variants and/or fragments of SEQ ID NOs: 1, 2, 3, 4, 5, 6, 7, 8, 50, 51, 52, and/or 53) to detect maltose, e.g., in a biological system;

[0228] Use of a glutamate biosensor disclosed herein (e.g., one or more of SEQ ID NOs: 62, 63, and/or 176-182 including variants and/or fragments of SEQ ID NOs: 62, 63, and/or 176-182) to detect glutamate, e.g., in a biological system;

[0229] Use of a phosphonate biosensor disclosed herein (e.g., one or more of SEQ ID NOs: 77 and/or 78 including variants and/or fragments of SEQ ID NOs: 77 and/or 78) to detect phosphonate, e.g., in a biological system; and/or

[0230] Use of a glucose biosensor disclosed herein (e.g., one or more of SEQ ID NOs: 91, 92, 93, 94, and/or 183 including variants and/or fragments of SEQ ID NOs: 91, 92, 93, 94, and/or 183) to detect glucose, e.g., in a biological system.

[0231] Use of a GABA biosensor disclosed herein (e.g., SEQ ID NO: 184 including variants and/or fragments of SEQ ID NO: 184) to detect GABA, e.g., in a biological system.

[0232] Use of an ACh biosensor disclosed herein (e.g., one or more of SEQ ID NOs: 185 and/or 186 including variants and/or fragments of SEQ ID NOs: 185 and/or 186) to detect ACh, e.g., in a biological system.

[0233] Techniques for performing such methods are known in the art and/or are exemplified herein. For example, methods can include introducing one or more biosensors into a biological system (e.g., a cell); expressing the one or more biosensors in the biological system (e.g., the cell); monitoring the signal emitted by the expressed biosensor in the biological system; and correlating the signal emitted by the expressed biosensor in the biological system with a level of the analyte in the biological system.

EXAMPLES

[0234] The invention is further described in the following examples, which do not limit the scope of the invention described in the claims.

[0235] Example 1: Maltose Indicators

[0236] Genetically encoded maltose indicators were generated using *Escherichia coli* maltodextrin-binding protein (EcMBP) as a framework and either circularly permuted β -lactamase (cpBla) or circularly permuted fluorescent protein (cpFP) as a signal. Data describe below suggest that cpBla and cpFP are not interchangeable.

[0237] Allosteric coupling of ligand binding to fluorescence was hypothesized to require:

[0238] i) that the site in into which cpGFP is inserted have the capacity to transduce the global conformational change the scaffold protein (EcMBP in this example) to the local environment of the chromophore in cpGFP; and

[0239] ii) that the local environment of the chromophore (e.g., linkers) be optimized to maximize the difference in emission between unbound (apo) and the bound (in this example maltose-bound) states.

Example 1A: Identification of cpGFP Insertion Sites in EcMBP

[0240] Potential insertion sites were identified using the crystal structures of the maltose-bound, closed form of EcMBP (Ouiicho et al., Structure, 5:997-1015, 1997) and the ligand-free, open form of EcMBP shown in FIG. 1 (Sharff et al., Biochemistry, 31:10657-10663, 1992) to guide rational design of EcMBP-cpGFP fusions that would result in maltose-dependent GFP fluorescence.

[0241] For (i), the change in dihedral angle (defined by the C α atoms spanning four residues) was analyzed to identify maltose-dependent structural changes in sequentially adjacent residues (FIG. 6); this analysis showed that the C α chain is “torqued” around residues 175 (Δ Dihedral=)+41° and 311 (Δ Dihedral=)-22° upon ligand binding. This sequential conformational change was predicted to be coupled to structural changes of an inserted cpGFP, resulting in maltose-dependent fluorescence for the fusion protein.

[0242] Previous studies using randomly digested and reassembled circularly permuted β -lactamase (cpBla) and EcMBP showed maltose-dependent β -lactamase activity in proteins with insertions of cpBla at EcMBP residues 165 and 317 (Guntas et al., Chem. Biol., 11:1483-1487, 2004; Guntas and Ostermeier, J. Mol. Biol., 336:263-273, 2004).

[0243] Since the Δ Dihedral of EcMBP165 is +11° (moderate change) and EcMBP317 is +2° (no real change), four EcMBP-cpGFP templates were constructed by inserting cpGFP into EcMBP at sites 165, 175 (identified herein), 311 (identified herein), and 317 to test our predictive method and the interchangeability of cpBla and cpGFP at sites identified from the EcMBP-cpBla screen. These constructs were named MBP165-cpGFP, MBP175-cpGFP, MBP311-cpGFP, and MBP317-cpGFP (names were modified to encompass variants (e.g., with modified linker sequences). The cpGFP used is cpGFP146 described in Baird et al. (Proc. Natl. Acad. Sci., USA, 96:11241-11246, 1999). PCR assembly was used to construct fusion proteins with GlyGly-linkers between EcMBP and each terminus of cpGFP. The amino acid sequence of each construct is shown in FIGS. 6-9. The sequences of SEQ ID NOs:1-3 shown in FIGS. 7A-7C (i.e., MBP165-cpGFP) differ in the linker sequence between MBP 1-165 and cpGFP 147-238 (linker 1: see the line ending in amino acid 240)). The sequences of SEQ ID NOs: 4-5 shown in FIGS. 8A-8B (i.e., MBP175-cpGFP) differ in the sequence between MBP 1-175 and cpGFP 147-238 (linker 1: see the line ending in amino acid 240)). The sequences of SEQ ID NOs: 6-7 shown in FIGS. 9A-9B (i.e., MBP311-cpGFP) differ in the sequence between cpGFP 1-146 and MBP 312-370 (linker 2: see the line ending in amino acid 640)). Each construct includes 3 linkers: A linker between the C-terminus of the C-terminal portion of MBP and the N-terminus of cpGFP (i.e., linker 2), a linker

between the N-terminus of cpGFP and C-terminus of the N-terminal portion of MBP, and a linker in cpGFP (i.e., linker 3).

Example 1B: Linker Optimization

[0244] Libraries of variants of SEQ ID NOs: 1-8 were generated with randomized linkers by single-stranded uracil template mutagenesis (see Kunkel et al., Methods Enzymol., 204:125-139, 1991) using the primers listed below:

165 Linker 1 Primers:	(SEQ ID NO: 9)
PLIAADGxxNVYIM	
	(SEQ ID NO: 10)
PLIAADxxNVYIM	
	(SEQ ID NO 11)
PLIAADGGxxNVYIM	
	(SEQ ID NO: 12)
PLIAADGxPNVYIMG	
	(SEQ ID NO: 13)
PLIAADGIxNVYIMG	
	(SEQ ID NO: 14)
PLIAADPxSHNVYIM	
	(SEQ ID NO: 15)
PLIAADxPSHNVYIM	
	(SEQ ID NO: 16)
PLIAADxxSHNVYIM	
	(SEQ ID NO: 17)
PLIAADxxSHNVFIM	
	(SEQ ID NO: 18)
PLIAADPxSHNVFIM	
	(SEQ ID NO: 19)
PLIAADPxSYNVFIM	
	(SEQ ID NO: 20)
PLIAADxxSYNVFIM	
	(SEQ ID NO: 21)
PLIAADPxSYNVFIM	
	(SEQ ID NO: 22)
PLIAADxxSYNVFIM	
	(SEQ ID NO: 23)
PLIAADPxSxNVYIM	
	(SEQ ID NO: 24)
PLIAADPxSHxVYIM	
	(SEQ ID NO: 25)
PLIAADPxSHNxYIM	
	(SEQ ID NO: 26)
PLIAADPxSHNVxIM	
165 Linker 2 Primers:	(SEQ ID NO: 27)
KLEYNFNxxYAFKYEN	
	(SEQ ID NO: 28)
KLEYNFNxYAFKYEN	
	(SEQ ID NO: 29)
KLEYNFNyAFKYEN	

-continued

KLEYNFxxYAFKYEN	(SEQ ID NO: 30)
KLEYNxxxYAFKYEN	(SEQ ID NO: 31)
KLEYNWxxYAFKYEN	(SEQ ID NO: 32)
KLEYNxKYAFKYEN	(SEQ ID NO: 33)
KLEYNFNPxxYAFKYEN	(SEQ ID NO: 34)
KLEYNFNPxPYAFKYEN	(SEQ ID NO: 35)
175 Linker 1 Primers:	
AFKYENxxSHNVYIM	(SEQ ID NO: 36)
175 Linker 2 Primers:	
KLEYNFNxxKYDIKDV	(SEQ ID NO: 37)
311 Linker 1 Primers:	
KSYEELxxSHNVYIM	(SEQ ID NO: 38)
KSYEELPxSHNVYIM	(SEQ ID NO: 39)
KSYEELxPSHNVYIM	(SEQ ID NO: 40)
311 Linker 2 Primers:	
KLEYNFNxxAKDPRIA	(SEQ ID NO: 41)
KLEYNFNPxxAKDPRIA	(SEQ ID NO: 42)
KLEYNFNxPAKDPRIA	(SEQ ID NO: 43)
317 Linker 1 Primers:	
ELAKDPRxxSHNVYIM	(SEQ ID NO: 44)
ELAKDPRxxxSHNVYIM	(SEQ ID NO: 45)
ELAKDPRxxxSHNVYIM	(SEQ ID NO: 46)
317 Linker 2 Primers:	
KLEYNFNxAATMENA	(SEQ ID NO: 47)
KLEYNFNxxxAATMENA	(SEQ ID NO: 48)
KLEYNFNxxxxAATMENA	(SEQ ID NO: 49)

[0245] Where “x” indicates that a degenerate primer (with DNA sequence “NNS”) was used to encode all 20 possible amino acids.

[0246] About 400 variants were screened in semi-high-throughput fashion, measuring fluorescence intensity of clarified cell lysate in the absence and presence of 10 mM maltose.

[0247] Insertion of cpGFP as MBP317, a site previously reported for cpBla, did not show maltose-dependent fluorescence (FIG. 11) even though the framework protein still

bound maltose, as determined by isothermal titration calorimetry (FIG. 12). These data demonstrate that identification of insertion sites by a method other than insertion of cpGFP (such as insertion of cpBla) is not sufficient to identify sites that transduce ligand binding to changes in fluorescence intensity

[0248] Insertion of cpGFP at residue 165 of EcMBP (EcMBP165-cpGFP), another position reported in cpBla studies (Guntas and Ostermeier, supra) with -GlyGly-linkers flanking the cpGFP resulted in a protein in which fluorescence increased 20% ($\Delta F/F=0.2$) upon addition of saturating maltose.

[0249] Screening a fully-degenerate, length-two library (“XX”) at either the EcMBP-cpGFP linker (linker 1) or the cpGFP-EcMBP linker (linker 2) yielded proteins with maltose-dependent fluorescent increases >300% or decreases >50% (FIG. 11). Many of the variants with increased $\Delta F/F$ values had linkers containing proline(s). Subsequent libraries constructed from oligonucleotides encoding XP or PX and randomization of the residues in GFP from residue 146 to 150 were screened, yielding a final variant with: a two-proline EcMBP-cpGFP linker, a two-glycine cpGFP-EcMBP linker, GFP-H148Y, and GFP-Y151F. This variant, called “EcMBP165-cpGFP.PPYF” (abbreviated PPYF (SEQ ID NO:2)) has a $\Delta F/F=2.5$, a K_d for maltose of 3 μ M. Screens also identified variant EcMBP311-cpGFP.L2-NP (-AsnPro- at linker 2 (SEQ ID NO:7)), which has a $\Delta F/F$ of 1.0 and a K_d for maltose of 2 μ M. This variant has an inferior maltose-dependent fluorescence increase than PPYF, but demonstrates generality of the cpFP insertion method.

[0250] EcMBP175-cpGFP was also screened with XX linkers, and a few variants with $\Delta F/F \approx 1$ were identified (FIG. 11). One mutant, with the first linker encoding HL (EcMBP175-cpGFP.L1-HL (SEQ ID NO:5)), has a $\Delta F/F=0.5$ and a K_d for maltose of 1.3 μ M.

[0251] These data support that choice of insertion site by structural analysis is preferable to random insertion.

Example 1C: Modifying Ligand Binding and/or Fluorescent Properties of Sensors

[0252] One objective in the development of generic biosensors is for the framework to permit independent optimization of binding and signaling properties. Analysis of whether biosensors herein permit such optimization was tested using the high-SNR sensor PPYF, by: (i) rationally altering maltose-binding affinity; (ii) changing the ligand-binding specificity from maltose to sucrose, and (iii) creating a family of sensors in multiple colors.

[0253] As a first step, the impacts of mutations of three tryptophan side-chains in the maltose-binding pocket (W230, W62, and W340) were tested. These sites have previously been shown to lower the affinity of EcMBP for maltose by one, two, or three orders of magnitude, respectively, when mutated to alanine (Martineau et al., J. Mol. Biol., 214:337-352, 1990). A mutation to the hinge region, I329W, was also made to PPYF, as this has been shown to increase maltose affinity by about 2-fold in both wild-type EcMBP (Marvin and Hellinga, Nat. Struc. Biol., 8:795-798, 2001) and in the EcMBP-cpBla switches (Guntas et al., Chem. Biol., 11:1483-1487, 2004; Kim and Ostermeier, Arch. Biochem. Biophys., 446:44-51, 2006). As shown in FIG. 13, for the PPYF sensor, the three tryptophan-to-alanine binding-pocket mutations behaved as expected, low-

ering affinity by between one and three orders of magnitude. In contrast, the I329W mutation did not increase affinity as expected, but rather decreased it. $\Delta F/F$ also decreased. This data suggests that the mechanism of fluorescence change in this sensor is dependent on subtle interactions between EcMBP and cpGFP that are linked to the I329W mutation.

[0254] As an alternative test for changing the ligand-binding specificity of the sensor while preserving fluorescence signaling, “5-7” mutations (D14L, K15E, W62Y, E111Y), previously shown to confer EcMBP with an affinity for sucrose (Guntas and Mansell, Proc. Natl. Acad. Sci., 102:11224-11229, 2005), were made to PPYF. As shown in FIG. 14A, the mutations conferred to the sensor about 2 mM affinity for sucrose and ~ 3 mM affinity for maltose. To address a discrepancy between expected (micromolar) and observed (millimolar) affinity for disaccharides, the 5-7 mutations were made to sensors with cpGFP inserted at different positions in EcMBP, and with different linker compositions. In the context of EcMBP165-cpGFP.PCF, the 5-7 mutations conferred very low (but observable) binding preference for sucrose over maltose (FIG. 14B). The trend of higher (but still weak) affinity for sucrose (~ 0.6 mM) over maltose (~ 6 mM) continued when the 5-7 mutations are made in the context of EcMBP175-cpGFP.L1-HL (FIG. 14C). In the context of EcMBP311-cpGFP.L2-NP, the 5-7 mutations appeared to eliminate all binding (FIG. 14D). The preference for sucrose over maltose of the 5-7 variants of the sensors is consistent with the binding properties of the 5-7 variants of EcMBP alone and EcMBP-cpBla (Guntas and Mansell, Proc. Natl. Acad. Sci., 102:11224-11229, 2005). The lower affinity for both ligands of the 5-7 variants of the sensors may be the consequence of the inserted cpGFP shifting the open and closed equilibrium.

[0255] These data suggest that ligand binding and fluorescent properties of biosensors can be independently modified.

Example 1D: Modifying Sensor Color

[0256] The color of GFP can be altered by changing the amino acids that either comprise or interact with the chromophore (see Shaner et al., J. Cell. Sci. 120:4247-4260, 2007, for a review).

[0257] Using PPYF as a template, mutations Y66W (to yield a cyan variant, “cpCFP”), L64F+T65G+V68L+T203Y (yellow, “cpYFP”), and Y66H (blue, “cpBFP”) mutations were made (see Cubitt et al., Trends Biochem., 20:448-455, 1995, for exemplary methods). As shown in FIG. 15, the variants exhibit fluorescence emission spectra consistent with their respective intended designs.

[0258] The $\Delta F/F$ of the color variants in response to maltose is different (in each case inferior) from the $\Delta F/F$ of 2.5 observed in PPYF-green. The EcMBP165-cpYFP.PPYF sensor, which has the same covalent chromophore structure as PPYF, has the greatest $\Delta F/F$ of the three spectral variants (FIG. 15A). EcMBP165-cpCFP.PPYF has a lower $\Delta F/F$ than the green and yellow variants, but by incorporating previously identified mutations, (L1-PC+GFP-Y151F; the resulting protein is called EcMBP165.cpCFP.PCF), a variant with $\Delta F/F=0.8$ was obtained (FIG. 15A).

[0259] The EcMBP165-cpBFP.PPYF variant, while dimly fluorescent, is not a sensor, and a screen of 800 linker variants failed to produce any variant with $\Delta F/F > 0.2$ (FIG. 16).

[0260] Since EcMBP165-cpBFP.PPYF was very dim, Azurite mutations T65S+V150I+V224R were included to increase brightness and stability, and make EcMBP165-cpAzurite a good template for linker screening. Using oligonucleotides encoding XX amino acid linkers, a variant was obtained, EcMBP165-cpAzurite.L2-FE, that had $\Delta F/F=0.8$ (FIG. 15).

Example 1E: Modifying Sensor Color and Ligand Specificity/Affinity

[0261] The four sucrose-binding “5-7” mutations described above that conferred weak sucrose affinity in the green sensor (EcMBP165-cpGFP.PPYF) were converted to blue, cyan, and yellow maltose sensors (EcMBP165-cpAzurite.L2-FE, EcMBP165-cpCFP.PCF, and EcMBP165-cpYFP.PPYF). The green and yellow sensors showed increased fluorescence upon addition of 10 mM sucrose, but the cyan and blue proteins did not (FIG. 15A). Like the green variant, the yellow variant had no detectable sucrose affinity with the wild type binding pocket (FIG. 15C) and millimolar affinity for both sugars, with preference for sucrose over maltose (FIG. 15D).

[0262] As seen in FIG. 17, as maltose concentration increased, the blue sensor increased in fluorescence first ($K_d \sim 2.7$ μ M), then the green ($K_d \sim 40$ μ M), then the yellow ($K_d \sim 350$ μ M), and at high maltose concentrations, the cyan variant began to increase its fluorescence ($K_d \sim 1.7$ mM).

Example 1F: Imaging Bacteria

[0263] The ultimate value of genetically encoded fluorescent sensors is in their utility for observing analyte flux in living cells and organisms. In a simple proof-of-principle experiment, *Escherichia coli* expressing PPYF or PPYF.T203V (see “Second-generation maltose sensors” below) were imaged in the green fluorescence channel in the absence of maltose, and then re-imaged after addition of saturating maltose to the media.

[0264] As shown in FIG. 18, bacteria expressing the sensors clearly became brighter, while control bacteria expressing EGFP appeared unchanged. Increased fluorescence was quantified by measuring the peak (gray-value) pixel intensity of each bacterium. Those expressing PPYF undergo an approximate doubling of fluorescence (bacterium-averaged $\Delta F/F=1.1 \pm 0.4$), those expressing PPYF.T203V have slightly increased $\Delta F/F$ ($\Delta F/F=1.29 \pm 0.2$), while those expressing EGFP have no change in fluorescence ($\Delta F/F=-0.01 \pm 0.05$).

Example 1F: 2-Photon Imaging of Mammalian Cells

[0265] Multi-photon microscopy opened new frontiers for in vivo fluorescence imaging, in particular for neuronal activity visualization through the use of genetically encoded calcium indicators (Tian et al., Nat. Methods, 3:281-286, 2009; Denk et al., Science, 248:73-76, 1990; Denk and Svoboda, Neuron, 18:351-357, 1997).

[0266] To demonstrate that the maltose sensors described herein have the potential to be used for 2-photon imaging, fluorescence excitation spectra were collected. As shown in FIG. 19, with a 535 nm bandpass emission filter (50 nm bp), EcMBP165-cpGFP.PPYF showed a 10-fold maltose-dependent increase in fluorescence when excited at 940 nm. All

four spectral variants showed a significant maltose-dependent increase in 2-photon fluorescence.

Example 1G: Sub-Cloning Maltose Sensors

[0267] EcMBP165-cpGFP.PPYF.T203V (see “Second-generation maltose sensors” below) were cloned into a modified version of the pDisplay vector (Invitrogen) for extracellular display on the surface of transiently transfected human embryonic kidney (HEK293) cells.

[0268] As shown in FIG. 20, the sensor localized to the plasma membrane and increased in brightness in a concentration-dependent manner when perfused with buffers of varying maltose concentration. The $\Delta F/F$ is 5.8-fold, very close to that of the soluble protein produced in *E. coli*, with the mid-point of the maltose-dependent fluorescence increase being 6.5 μM (FIG. 21A), very similar to the affinity determined on purified protein (5 μM). Furthermore, the surface displayed sensor responded rapidly to a pulse of 1 mM maltose (FIG. 21A), indicating that the time course for its action is useful for transient events.

Example 1H: Crystal Structure Analysis of Maltose Sensors

[0269] High-resolution structures of several of the maltose sensors described above were generated. Crystallization trials were performed with EcMBP165-cpGFP.PPYF, EcMBP175-cpGFP.L1-HL, and EcMBP311-cpGFP.L2-NP in the presence and absence of excess maltose, from which both EcMBP175-cpGFP.L1-HL and EcMBP311-cpGFP.L2-NP crystallized in the presence of maltose. X-ray structures were solved to 1.9 and 2.0 Å resolution, respectively, by molecular replacement (FIGS. 22A-22C).

[0270] The structures of the cpGFP and EcMBP domains of the sensors are superimposable with published crystal structures of cpGFP (from GCaMP2; RMSD=0.36 and 0.38 Å, respectively, for comparing 221 common C α atoms) and EcMBP-maltose (RMSD=0.43 and 0.37 Å, 370 C α). The structure of EcMBP is largely unperturbed by insertion of the cpGFP domain; only residues around the 175 and 311 insertion sites showed any significant displacement.

[0271] GFP-H148, which H-bonds the GFP chromophore in the structure of native GFP, also directly H-bonded to the chromophore in the EcMBP175-cpGFP.L1-HL-maltose structure (FIG. 22B), although a different rotamer was observed. In the EcMBP311-cpGFP.L2-NP-maltose structure, GFP-H148 is pulled away from the chromophore and is largely replaced by the Asn from linker 2, which makes H-bond interactions to both strand 8 of the GFP barrel and the chromophore phenolate oxygen (through a water molecule, FIG. 22D). GFP-H148, meanwhile, seemed to stabilize the conformation of linker 2 of EcMBP311-cpGFP.L2-NP by H-bonding the backbone carbonyl of the linker 2 Asn. There is some solvent access to the cpGFP chromophore through the hole in the GFP barrel created by circular permutation, although the inter-domain linkers block much of the opening in both structures. Relatively few contacts are made between the cpGFP and EcMBP domains.

[0272] Based on the structures of two maltose-bound sensors, the sensing mechanism likely involves a shift in the relative position of linker 1 and linker 2 induced by the conformational change in the EcMBP domain associated with maltose binding (FIG. 5). The register shift of interactions between the two linkers could alter the proximity of

linker 2 and nearby side-chains to the cpGFP chromophore and change the water structure in the cpGFP opening, leading to a shift in the chromophore protonation equilibrium. This might explain why rigid proline is preferred in either linker, since conformational changes upon ligand binding might be better propagated through the rigid linkers to the cpGFP chromophore environment.

Example 1I: Generation of Second-Generation Maltose Sensors

[0273] In an attempt to increase brightness and $\Delta F/F$ of GCaMP, the local environment of the chromophore was altered by randomizing residues within cpGFP, and screening for improved variants (Tian et al., *nat. Methods*, 6:875-881, 2009).

[0274] As shown in FIG. 23, in the context of EcMBP165-cpGFP.PPYF, the T203V mutation decreases the fluorescence emission of the apo-state by half (FIG. 23A), while saturated fluorescence and affinity are unchanged (FIG. 23B), increasing $\Delta F/F$ to 6.5. In the maltose-saturated state, PPYF itself has about a quarter the brightness of EGFP, and half the brightness of cpGFP.

[0275] In the context of EcMBP311-cpGFP.L2-NP, the T203V mutation decreases the brightness of both the apo-state and the saturated-state equally, resulting in no significant change in $\Delta F/F$ (FIGS. 23C and D).

[0276] These results indicate that the benefits of the T203V mutation are not universally transferable, and that cpGFP-based fluorescent sensors need to be optimized individually.

Example 2: Maltotriose Indicators

[0277] Genetically encoded maltotriose indicators were created using *Pyrococcus furiosus* maltotriose binding protein. As described below, only the structure of the ligand-bound state *P. furiosus* maltotriose binding protein (PfMBP) is available. As shown in FIGS. 1 and 2, PfMBP is homologous to EcMBP (compare FIGS. 1 and 2). Two sensors were made, PfMBP171 and PfMBP316, the insertion points for which were selected based on homology to EcMBP165 and EcMBP311, respectively. Linkers were optimized. PfMBP sensors have a $\Delta F/F$ of ~1.2.

[0278] *Pyrococcus furiosus* is a thermophilic organism. Proteins from thermophiles have been shown to be more amenable to mutation than those from mesophiles (Bloom et al., *Proc. Natl. Acad. Sci.*, 103:5869-5874, 2006). As an alternative to developing new sensors by inserting cpGFP into PBPs, it should also be possible to generate new sensors by changing the ligand-binding specificity of an existing PBP-based sensor.

[0279] It has previously been shown that the binding sites of PBPs can be reengineered to accommodate novel ligands (Looger et al., *Nature*, 423:185-190, 2003). However, those re-design efforts used framework proteins from mesophiles and suffered from poor stability. We hypothesized that PfMBP, which is intrinsically more stable than EcMBP, is more tolerant of mutations. To test this hypothesis, we characterized and compared the stability of PfMBP to EcMBP, PfMBP-cpGFP sensors to EcMBP-cpGFP sensors, PfMBP binding site mutants to EcMBP binding site mutants, and PfMBP-cpGFP sensor binding site mutants to EcMBP-cpGFP sensor binding site mutants. Conclusively, the PfMBP variants were more stable than the EcMBP variants.

Finally, we demonstrate that the increased thermo-stability of the PfMBP-cpGFP sensors is useful for the measurement of maltotriose at temperatures as high as 60° C., whereas the EcMBP-cpGFP sensors are only useful for the measurement of maltose at temperatures as high as 40° C.

Example 2A: Identification of cpGFP Insertion Sites in PfMBP

[0280] The ligand-bound (closed) structure of PfMBP is available (Evdokimov et al., *J. Mol. Biol.*, 305:891-904, 2001), but the unbound structure is not. Accordingly, insertion sites for the PfMBP-cpGFP sensors were identified by homology to EcMBP.

[0281] Sites were selected based on the structural similarities between PfMBP and EcMBP. Two sites were selected. One of these sites is EcMBP311, which is homologous to PfMBP316. This site is at juncture between the end of the cluster of helices (Helices 8a, 8b, 8c) and the start of the “equatorial” spanning helix (Helix 9). Another site that was made into a sensor in EcMBP was EcMBP165, which is homologous to PfMBP171. cpGFP was inserted into PfMBP at each of these sites. The sequences of the resulting constructs, PfMBP171-cpGFP and PfMBP316-cpGFP, are shown in FIGS. 24 and 25, respectively.

Example 2B: Linker Optimization

[0282] Libraries of variants of SEQ ID NOs: 50-53 were generated with randomized linkers by single-stranded uracil template mutagenesis using the primers listed below:

175 Linker 1 Primers: (SEQ ID NO: 54)
AIAQAFxxSHNVYIMA

(SEQ ID NO: 55)
AIAQAFPxSHNVYIMA

171 Linker 2 Primers: (SEQ ID NO: 56)
KLEYNFNxxYYFDDKTE

316 Linker1 Primers (SEQ ID NO: 57)
VLDDPExxHNVYIM

(SEQ ID NO: 58)
VLDDPEIxxSHNVYIM

316 Linker2 Primers (SEQ ID NO: 59)
KLEYNFPxxNDPVIY

(SEQ ID NO: 60)
KLEYNFPNxxKNDPVIY

(SEQ ID NO: 61)
KLEYNFPNxxKNDPVIY

[0283] Where “x” indicates that a degenerate primer (with DNA sequence “NNS”) was used to encode all 20 possible amino acids.

[0284] Several thousand variants were screened in semi-high-throughput fashion, measuring fluorescence intensity of clarified cell lysate in the absence and presence of 1 mM maltotriose.

[0285] Screening a fully-degenerate, length-two library (“XX”) at either the PfMBP171-cpGFP linker (linker 1) or the cpGFP-PfMBP linker (linker 2) yielded proteins with

maltotriose-dependent fluorescent increases >100% or decreases >20% (FIG. 26A). A variant from this group with a GlyGly PfMBP-cpGFP linker and a PheGlu cpGFP-PfMBP linker was selected for further characterization. This variant, called “PfMBP171-cpGFP.L2FE” has a $\Delta F/F=1.2$, a Kd for maltotriose of <1 μ M.

[0286] Screening a fully-degenerate, length-two library (“XX”) at either the PfMBP316-cpGFP linker (linker 1) or the cpGFP-PfMBP linker (linker 2) also yielded proteins with maltotriose-dependent fluorescent increases >100% or decreases >20% (FIG. 26B). A variant from this group with a GlyGly PfMBP-cpGFP linker and a PheGlu cpGFP-PfMBP linker was selected for further characterization. This variant, called “PfMBP316-cpGFP.L1-NP” has a $\Delta F/F=1.2$, a Kd for maltotriose of 40 μ M.

[0287] These data support that structurally homologous frameworks can be compared to identify insertion sites for cpGFP.

Example 2C: Characterization of the Thermostability of the PfMBP and PfMBP-cpGFP Compared to EcMBP and EcMBP-cpGFP

[0288] Thermal stability of PfMBP171-cpGFP.L2FE was measured using circular-dichroism (CD) and compared to the original EcMBP and PfMBP binding proteins, along with cpGFP. Following the changes by means of CD allowed determination of whether different transitions happened in alpha, beta, or both kinds of structures.

[0289] Given that cpGFP is a beta barrel, strong transitions in the beta signal alone were associated with changes in this kind of structure. In the same way, transitions in both kinds of signals were associated with the binding protein structure. As shown in FIG. 27A, PfMBP is significantly more thermostable than EcMBP. In fact, while EcMBP denatured at about 50° C., PfMBP did not denature at temperatures less than 80° C. Also, the addition of maltose to EcMBP stabilized the protein by about 10° C.

[0290] As shown in FIG. 27B, the stability of the EcMBP component of the EcMBP165-cpGFP.PPYF sensor decreased from 50° C. to 45° C. with insertion of cpGFP, while the intrinsic stability of cpGFP in the sensor remained unchanged. There was little change in the stability of the PfMBP component of the PfMBP171-cpGFP.L2FE sensor with insertion of cpGFP (FIG. 27B). Moreover, PfMBP seemed to exert a small stabilizing effect over the inserted cpGFP, as shown by the change in the steepness and melting point of the curve of the soluble form and the PfMBP171-cpGFP.L2FE sensor. All the associations made between transitions and domain unfolding were supported by CD spectra taken at the beginning and the end of each temperature ramp.

[0291] Analysis of whether the PfMBP scaffold was more tolerant of mutation than the EcMBP scaffold was also performed. Proof-of-principle mutations were made to the ligand-binding sites of EcMBP and PfMBP, and their respective sensors. In EcMBP, Asn12 was mutated to Trp to result in steric clashes with the surrounding residues, and backbone, of the binding pocket. The homologous mutation in PfMBP is Ala13Trp, which would be expected to have the same effect.

[0292] As shown in FIG. 27C, N12W decreased the Tm of EcMBP from 50° C. to 40° C., while the corresponding mutation in PfMBP, A13W, had no noticeable effect. This data confirms that the thermo-philic protein is more tolerant

of mutations to the binding site. Furthermore, in the context of the sensors, the N12W mutation to EcMBP165-cpGFP.PPYF completely destabilized the binding protein component of the sensor (FIG. 27D), while the A13W mutation in PfMBP171-cpGFP.L2FE had no effect on stability (FIG. 27D).

Example 2D: Tolerance of PfMBP Sensor to Increased Temperature

[0293] Fluorescence of the protein in the apo and ligand-bound states at was measured at different temperatures.

[0294] As shown in FIG. 28A, fluorescence of the EcMBP165-cpGFP.PPYF sensor in the bound state was higher than it is in the apo-state at lower temperatures, by about 4-fold. However, at around 55° C. (the unfolding transition of the EcMBP component) the fluorescence of the EcMBP165-cpGFP.PPYF sensor dropped precipitously. As a result, EcMBP165-cpGFP.PPYF is unsuitable for detection of maltose at temperatures greater than 50° C. (FIG. 28B). In contrast, PfMBP171-cpGFP.L2FE retained its maltotriose binding capabilities at high temperatures (FIGS. 28A and 28B), and is limited only by the intrinsic fluorescence of the cpGFP component, which decays at about 80° C. (FIG. 28A).

Example 2E: Measurement of Maltodextrins in Hot Liquids

[0295] To demonstrate that the soluble and immobilized sensors function similarly, PfMBP171-cpGFP.L2FE, PfMBP316-cpGFPL1XXX, and EcMBP165-cpGFP.PPYF.T203V were immobilized via their N-terminal poly-histidine tags on to the surface of Ni-NTA coated glass. In a fluorescence plate reader, the immobilized proteins performed similarly to their soluble counterparts (see FIGS. 28C, 28D, and 28F).

[0296] Next, a prototype device was constructed, with a light guide providing the excitation light and returning the fluorescent emitted light back to the photodetector, the bio-sensor protein immobilized to Ni-NTA coated coverslips, and the coverslip attached to the end of the light guide. The “wand” of the detector was dipped into different compositions of solutions, each with varying concentrations of maltose or maltotriose. Experiments were performed at different temperatures. PfMBP-cpGFP sensor performed better at higher temperatures (as high as 60° C.) than the EcMBP-cpGFP sensor.

Example 3: Glutamate Indicators

[0297] Glutamate indicators were created from *Escherichia coli* glutamate-binding protein (EcYbeJ). As with PfMBP in Example 2, only the structure of the ligand-bound EcYbeJ is available. EcYbeJ is homologous to EcMBP, but to a lesser degree. The best homology match between a site in EcYbeJ and a site in a binding protein for which an intensity-based sensor has already been created is EcYbeJ253 and EcMBP311 (described herein). As shown in FIG. 3, both sites are at the junction of “Rising Helix 8” and the “Equatorial Helix/Coil.” The amino acid composition of the cpGFP and EcYbeJ junction was made the same as that of the EcMBP311-cpGFP sensor (Linker 2=NP). The amino acid composition of the EcYbeJ junction and cpGFP was optimized to LV (Linker 1=LV). The variant has a $\Delta F/F$ of 5.

Example 3A: Identification of cpGFP Insertion Sites

[0298] The ligand-bound (closed) structure of *Shigella flexneri* glutamate binding protein is available (Fan et al., Protein Pept. Lett., 13:513-516, 2006). This protein has only 4 amino acid mutations relative to EcYbeJ, and is thus an appropriate model.

[0299] Insertion sites for the EcYbeJ-cpGFP sensors were identified by homology to EcMBP. Based on the topology map (FIG. 3), position 311 in EcMBP was identified as an acceptable insertion site for EcYbeJ. EcMBP311 is equivalent to EcYbeJ253. EcYbeJ253 is at juncture between the end of the cluster of helices (Helices 8a, 8b, 8c) and the start of the “equatorial” spanning helix (Helix 9). In YbeJ, the structure that is homologous to the equatorial helix is the equatorial coil (depicted in red, to match the red coloring of Helix 9).

[0300] Intrinsic affinity of wild-type YbeJ for glutamate (~1 μ M) was too high to permit high-throughput screening of linker libraries. Endogenous glutamate (from the growth media) saturates the sensor, making measurement of the unbound state technically challenging. A mutation to YbeJ (A184V), in the “hinge” of the protein were made. Mutation of this residue to Trp or Arg have previously been shown to decrease affinity in FRET-based sensors (see Okumoto et al., Proc. Natl. Acad. Sci., 102:8740-8745, 2005). EcYbeJ253 (A184V)-cpGFP has an affinity for glutamate of about 100 μ M. All references to EcYbeJ253-cpGFP, unless otherwise noted, refer to the A184V variant. The sequences of the EcYbeJ constructs are shown in FIG. 29.

Example 3B: Linker Optimization

[0301] Libraries of variants of SEQ ID NOs: 62-63 were generated with randomized linkers by single-stranded uracil template mutagenesis using the primers listed below:

253 Linker 1 Primers:

FKNPIPPxSHNVYIMA	(SEQ ID NO: 64)
FKNPIPPxxSHNVYIMA	(SEQ ID NO: 65)
FKNPIPPPxSHNVYIMA	(SEQ ID NO: 66)
FKNPIPPxPSHNVYIMA	(SEQ ID NO: 67)
KWFKNPIxxSHNVYIMA	(SEQ ID NO: 68)
FKNPIPPxxNVYIMAD	(SEQ ID NO: 69)
KWFKNPIxxNVYIMAD	(SEQ ID NO: 70)
253 Linker 2 Primers:	
KLEYNFNxKNLNMNF	(SEQ ID NO: 71)
KLEYNFNxxKNLNMNF	(SEQ ID NO: 72)
KLEYNFNxPKNLNMNF	(SEQ ID NO: 73)

-continued

(SEQ ID NO: 74)
KLEYNFNPxxKNLNMNF

(SEQ ID NO: 75)
GHKLEYNxxLNMNF

(SEQ ID NO: 76)
KLEYNFNxxLNMNF

[0302] Where “x” indicates that a degenerate primer (with DNA sequence “NNS”) was used to encode all 20 possible amino acids.

[0303] Several thousand variants were screened in semi-high-throughput fashion, measuring fluorescence intensity of clarified cell lysate in the absence and presence of 10 mM glutamate.

[0304] Screening a fully-degenerate, length-two library (“XX”) at the EcYbeJ253-cpGFP linker (linker 1) identified a sensor with glutamate-dependent fluorescent increases of ~100%. This variant has a LeuVal EcYbeJ-cpGFP linker (L1-LV) and was used as the framework for optimization of the cpGFP-EcYbeJ253 linker (linker 2). The results of that screen yielded a protein with glutamate-dependent fluorescent increase of ~500% and a linker 2 composition of AsnPro. As shown in FIG. 30, this variant, called “EcYbeJ253-cpGFP.L1LVL2NP” has a $\Delta F/F=5$, a Kd for glutamate of 100 μ M. Interestingly, the composition of the second linker, AsnPro, is the same as the linker composition of EcMBP311-cpGFP.L2NP.

Example 3C: Detection of Extracellular Glutamate

[0305] EcYbeJ253-cpGFP.L1LVL2NP was cloned into the pDisplay™ vector to allow targeting and anchoring of the sensor to the plasma membrane. The resulting construct was transfected into cultured mammalian cells (HEK293) to visualize the addition of glutamate to extracellular media. Constructs were also generated in a bacterial expression vector with the epitope tags individually and in combination.

[0306] As shown in FIG. 31, the hemagglutinin tag interferes with the fluorescence change. EcYbeJ253-cpGFP.L1LVL2NP was re-cloned into a derivative of the pDisplay™ vector, lacking the hemagglutinin tag, called pMinDis (for Minimal Display). This new construct, when expressed in HEK293 cells, shows a change in fluorescence intensity under 2-photon excitation that is approximately the same as the soluble protein (see FIG. 32) with higher affinity, of about 1 μ M (see FIG. 32).

[0307] To demonstrate that the sensor is functional in neurons, and not just cultured HEK cells, the gene from EcYbeJ253-cpGFP.L1LVL2NP was cloned into an adeno-associated virus vector (AAV) under control of the synapsin promoter. Virus particles were generated and used to infect cultured primary hippocampus neurons from rats 7 days after culturing. 14 days after culturing (and 7 days after infection), the infected neurons were imaged under 2-photon microscopy (FIG. 33).

Example 4: Phosphonate Indicators

[0308] An indicator for phosphonate compounds was created from *Escherichia coli* phosphonate-binding protein (EcPhnD). In this instance, only the structure of the ligand-bound state was available at the time the sensor was conceived. EcPhnD is homologous to EcMBP to a lesser degree and to EcYbeJ to a greater degree. The best homology match

between a site in EcPhnD and a site in a binding protein for which an intensity-based sensor has already been created is EcPhnD90 and EcYbeJ253. There is no “Rising Helix 8” in EcPhnD, but there is an “Equatorial Helix/Coil” (FIG. 4). cpGFP was inserted at the Equatorial Helix/Coil and linkers were optimized to yield a sensor with $\Delta F/F$ of 1.2. EcPhnD is a dimmer, so, a pair of mutations (L297R+L301R) were made to convert it to a monomer. The monomer variant has a $\Delta F/F$ of 1.6.

Example 4A: Identification of cpGFP Insertion Sites in EcPhnD

[0309] Insertion sites for the EcPhnD-cpGFP sensors were identified using the ligand-bound (closed) structure of EcPhnD by homology to EcMBP. Based on the topology map (FIG. 4), position 311 in EcMBP was identified as an acceptable insertion site in EcPhnD. EcMBP311 corresponds to EcPhnD90. This site is at the point where the rising strand (Strand D) of EcPhnD has a small bend in it that runs equatorial to the rest of the sheets in the protein. Even though it is topologically different from the “equatorial” spanning helix (Helix 9) of EcMBP its equatorial alignment is similar, and with just the closed structure at the time, in an environment that was expected to undergo significant dihedral change upon binding ligand. Sequences of EcPhnD constructs are shown in FIG. 34.

Example 4B: Linker Optimization

[0310] Libraries of variants of SEQ ID NOs: 77-78 were generated with randomized linkers by single-stranded uracil template mutagenesis using the primers listed below:

90 Linker 1 Primers:

(SEQ ID NO: 79)
QTVAADGSSHNVIYMA

(SEQ ID NO: 80)
QTVAADxxSHNVIYMA

(SEQ ID NO: 81)
QTVAADxPSHNVIYMA

(SEQ ID NO: 82)
QTVAADPxSHNVIYMA

(SEQ ID NO: 83)
QTVAADxxNVIYMA

(SEQ ID NO: 84)
QTVAADxxSHNVIYMA

(SEQ ID NO: 85)
VFQTVAXxSHNVIYMA

90 Linker 2 Primers:

(SEQ ID NO: 86)
HKLEYNFNPGYWSVLI

(SEQ ID NO: 87)
HKLEYNFNxxPGYWSVLI

(SEQ ID NO: 88)
HKLEYNxxPGYWSVLI

(SEQ ID NO: 89)
HKLEYNFNxxYWSVLI

(SEQ ID NO: 90)
HKLEYNFNPxYWSVLI

[0311] Where “x” indicates that a degenerate primer (with DNA sequence “NNS”) was used to encode all 20 possible amino acids.

[0312] Several thousand variants were screened in semi-high-throughput fashion, measuring fluorescence intensity of clarified cell lysate in the absence and presence of 100 μ M 2AEP.

[0313] Screening a number of fully-degenerate libraries at the EcPhnD90-cpGFP linker (linker 1) yielded a protein with 2AEP-dependent fluorescent increases of >100%. This variant has a AlaAsp EcPhnD-cpGFP linker (L1-AD) and a $\Delta F/F$ of 1.2. The variant came from a linker that also deleted two residues, effectively making the insertion point of cpGFP occur after residue D88, and then skipping to residue P91 at the cpGFP-EcPhnD linker.

[0314] It was observed from the crystal structure that EcPhnD forms a dimer. To disrupt the dimer inter-face and potentially simplify the observable binding behavior of the EcPhnD protein, two mutations, L297R and L301R, were introduced into the dimerization helices. These mutations were expected, by charge repulsion, to disrupt the dimer interface. As shown in FIG. 35, incorporation of L279R and L301R mutations into EcPhnD90-cpGFP.L1AD caused $\Delta F/F$ to increase to 1.6 in response to 2AEP.

[0315] Further attempts to crystallize the open, ligand-unbound form of the protein were successful after making a mutation to the binding site, H157A, that substantially decreased affinity for phosphonate compounds. This mutant was crystallized in the absence of ligand, and the open state of the protein solved. The Δ Dihedral analysis (FIG. 36) showed that the region of greatest dihedral change was the group of residues from 88-90, just one amino acid away from the site chosen by homology to the equatorial helix.

[0316] These data further indicate that Δ Dihedral metric is sufficient for identifying sites in PBPs into which cpGFP can be inserted and result in intensity-based fluorescent sensors.

Example 5: Glucose Indicators

[0317] Glucose indicators were created from *Thermus thermophilus* glucose binding protein (TtGBP). In this instance, only the structure of the ligand-bound state is available. TtGBP is very homologous to EcMBP and PfMBP (compare FIG. 5 with FIGS. 1 and 2). The insertion point (TtGBP326) was chosen by homology to EcMBP311 and PfMBP316. The amino acid composition of the cpGFP and TtGBP junction was made the same as that of the EcMBP311-cpGFP and EcYbeJ253 sensors (Linker 2=NP). Linker 1 was optimized (Linker 1=PA) and the TtGBP326 sensor have a $\Delta F/F$ of ~2.5. To improve its utility for the measuring glucose concentrations in human blood, the affinity was weakened from its native ~1 μ M to 1.5 mM by mutation of two residues in the binding pocket (H66A+H348A).

Example 5A: Identification of cpGFP Insertion Sites in TtGBP

[0318] The ligand-bound (closed) structure of TtGBP is available (Cuneo et al., J. Mol. Biol., 362:259-270, 2006). Accordingly, insertion sites for the TtGBP-cpGFP sensors were identified by homology to EcMBP and PfMBP. Based on the topology map (FIG. 5), it is apparent that TtGBP, PfMBP, and EcMBP are structurally similar in the closed, ligand-bound state. Positions in EcMBP determined by the

dihedral analysis (see above) were predicted to be acceptable insertion sites in TtGBP. EcMBP311 is homologous to TtGBP326. This site is at juncture between the end of the cluster of helices (Helices 8a, 8b, 8c) and the start of the “equatorial” spanning helix (Helix 9). The amino acid sequence of the TtGBP construct is shown in FIG. 37.

Example 5B: Linker Optimization

[0319] Libraries of variants of SEQ ID NO:91 were generated with randomized linkers by single-stranded uracil template mutagenesis using the primers listed below:

326 Linker 1 Primers:

DSDPSKYxxSHNVYIM (SEQ ID NO: 95)

DSDPSKYPxxSHNVYIM (SEQ ID NO: 96)

DSDPSKYxPSHNVYIM (SEQ ID NO: 97)

RLSDSPSxxSHNVYIM (SEQ ID NO: 98)

DSDPSKYxxNVYIM (SEQ ID NO: 99)

326 Linker 2 Primers:

KLEYNFPNxxNAYGQSA (SEQ ID NO: 100)

KLEYNFxxPNAYGQSA (SEQ ID NO: 101)

GHKLEYNxxNAYGQSA (SEQ ID NO: 102)

KLEYNFPNxxPNAYGQSA (SEQ ID NO: 103)

KLEYNFPNxxNAYGQSA (SEQ ID NO: 104)

[0320] Where “x” indicates that a degenerate primer (with DNA sequence “NNS”) was used to encode all 20 possible amino acids.

[0321] Several hundred variants were screened in semi-high-throughput fashion, measuring fluorescence intensity of clarified cell lysate in the absence and presence of 10 mM glucose.

[0322] Linker 1 was optimized (Linker 1=PA) and the TtGBP326-cpGFP.L1PAL2NP sensor has a $\Delta F/F$ of ~2.5 (see FIG. 38). Additionally, the TtGBP sensor was tested with and without the N-terminal pRSET tag and no difference was observed. Specifically, both sensors exhibited an affinity for glucose of about 1.5 mM and a $\Delta F/F$ of 2.5.

[0323] Data showing that it was possible to construct a glucose sensor by replacing the EcMBP or PfMBP with TtGBP, retaining the composition of linker 2, and optimizing the composition of linker 1, indicates that the methods for generating sensors disclosed herein can be used to generate sensors using any suitable framework.

Example 5C: Detecting Changes in Glucose Concentration in Vivo

[0324] The TtGBP326-cpGFP.L1PAL2NP sensor was cloned into a variant of the pDisplay™ vector lacking the N-terminal secretion sequence, the N-terminal hemaggluti-

nin tag, the C-terminal cMyc tag, and the C-terminal PDGFR membrane anchoring domain.

[0325] The TtGBP sensor was cloned into a mammalian expression vector (based on the pDisplay™ vector described in Example 3 above) with the secretion, epitope, and transmembrane anchoring peptides removed, thus resulting in cytosolic expression of the TtGBP326-cpGFP.L1PAL2NP+H66A+H348A sensor. The construct was transfected into HEK293 cells. As shown in FIG. 39, the TtGBP sensor was expressed in the cytosol.

[0326] As shown in FIG. 40, addition of 10 mM glucose to the media increases fluorescence.

[0327] The TtGBP326-cpGFP.L1PAL2NP+H66A+H348A sensor was further modified by L276V mutation to produce TtGBP326.L1PA.L2NP.H66A.H348A.L276V (see FIG. 50). As shown in FIG. 51, this construct has an affinity for glucose of 6.5 mM.

[0328] Additionally, the TtGBP326.L1P1.L2NP.G66A.H348A.L276V was cloned into the pMinDis derivative of the pDisplay vector and expressed on the extracellular surface of HEK293 cells. After exchanging the HEK293 cell media for PBS, addition of glucose to the PBS led to an increase in fluorescence (see FIG. 52).

[0329] These data indicate, in part, that the pRSET tag is not essential to the function of the sensor and that the TtGBP326-cpGFP.L1PAL2NP sensor is capable of detecting changes in the concentration of glucose inside or on the external surface of human cells.

Example 6: Stability, Affinity and Chromatic Variants of the Glutamate Sensor iGluSnFR

Example 6A: In Vivo Assessment of iGluSnFR Brightness in Apical Dendrites in Mouse Somatosensory Cortex

[0330] Wildtype C57/B6 mice were purchased from the Jackson Laboratory and group housed in the Janelia animal facility. Mice were injected at 8 weeks of age with AAV2/1.hSynapsin1.iGluSnFR.A184S or SF-iGluSnFR.A184S, at identical titers (1×10^{13} genomic copies per milliliter, GC/ml), volumes (20 nl), and locations (3 mm lateral to midline, 1.4 mm caudal to bregma, and 0.3 mm below the cortical surface). After viral injection, a craniotomy (3 mm diameter) was made over the injection site, and the skull was replaced with a #1.5 Schott glass and fixed in place with dental acrylic (Lang Dental Manufacturing), which also secured a titanium head bar to the skull for head-mounting during imaging experiments.

[0331] In vivo two-photon imaging experiments were performed during a state of 'quite wakefulness', after having been habituated to head fixation the prior 2-3 days. Period water rewards were given to keep animals hydrated and passive. For comparisons of intensity and bleaching, a custom two-photon microscope emitting 960 nm light from a Coherent Chameleon ultrafast laser was used. All experiments were performed using a 25 \times , 1.5 NA Olympus objective immersed in water. Image acquisition was performed with ScanImage (Vidrio) software and analyzed post hoc using ImageJ (NIH). Images were acquired at a variety of speeds/zooms, and powers in order to assess the impact of pulse energy and dwell time on bleaching and intensity. Images at each setting were acquired for 5 seconds. To analyze the data, images were averaged and thresholded to create a signal (above threshold) and background mask.

Signals in these masks were then averaged, and SNR was calculated from these as (signal-background)/(standard deviation of background). Bleaching percentage was calculated as the average intensity in the first 25% of the trace, divided by the last 25% of the trace.

Example 6B: Ferret Visual Cortex Assessment of SF-iGluSnFR.A184S and A184V

[0332] All procedures were approved by the Max Planck Florida Institute for Neuroscience Institutional Animal Care and Use Committee and adhered to the standards of the National Institutes of Health. Juvenile female ferrets (*Mustela putorius furo*, Marshall Farms) were used. Animals were housed in a vivarium under 16 hour light/8 hour dark cycle. The full methodological details for functional two-photon imaging of ferret visual cortex is previous described in detail (Wilson et al., 2016, Nat. Neurosci., 19:1003-9).

[0333] Briefly, juvenile female ferrets (*Mustela putorius furo*, Marshall Farms) aged P21-22 (n=2) were anesthetized with ketamine (50 mg/kg, IM) and isoflurane (1-3%) delivered in O₂, then intubated and artificially respired. Atropine (0.2 mg/kg, SC) and a 1:1 mixture of lidocaine and bupivacaine administered subcutaneously in the scalp. Animals were kept at 37° C. A small craniotomy (0.8 mm) was made over the visual cortex 7-8 mm lateral and 2-3 mm anterior to lambda. AAV2/1.hSynapsin1.Cre (Penn Vector Core) was diluted in phosphate-buffered saline (Sigma) and mixed with AAV2/1.hSynapsin-FLEX.SF-iGluSnFR.A184S or A184V for expression in layer 2/3 cortical neurons. Beveled glass micropipettes were lowered into the brain and 400-500 nl of virus were injected over 5 minutes at multiple depths below the pia. Following, the craniotomy was filled with 1% w/v agarose.

[0334] After four weeks, ferrets were anesthetized with 50 mg/kg ketamine and 1-3% isoflurane. Atropine (0.2 mg/kg, SQ) and bupivacaine were administered. Animals were kept at 37 to 38° C., artificially respired, and given intravenous fluids. Isoflurane (1-2%) was used throughout the surgical procedure to maintain a surgical plane of anesthesia. ECG, endtidal CO₂, external temperature, and internal temperature were continuously monitored. A custom titanium headplate was implanted on the skull at the viral injection site and the dura retracted to reveal the cortex. A custom insert with a single 4 mm coverglass (0.17 mm thickness) was placed onto the brain to gently compress the underlying cortex and dampen biological motion during imaging. The cranial window was hermetically sealed using a stainless steel retaining ring and Vetbond. Tropicamide Ophthalmic Solution and Phenylephrine Hydrochloride Ophthalmic Solution were applied and contact lenses were inserted into both eyes. Upon completion of the surgical procedure, Isoflurane was gradually reduced and pancuronium (2 mg/kg/hour) was delivered IV to immobilize the animal.

[0335] The animal was placed under the microscope 25 cm from the stimulus monitor, with the monitor subtending 130 degrees in azimuth and 74 degrees in elevation. Imaging was performed using a Bergamo II (Thorlabs) running ScanImage 5 or ScanImage 2015¹⁹ (Vidrio Technologies) with dispersion compensated 950 nm excitation provided by an Insight DS+ (Spectraphysics). Average excitation power after the exit pupil of the objective (16 \times , CFI75, Nikon Instruments) ranged from 25 to 40 mW. Two-photon frame triggers from ScanImage were synchronized with stimulus information using Spike2 (CED). Visual stimuli were gen-

erated using PsychoPy (Peirce, 2007, *J. Neurosci. Methods*, 162:8-13). Full-field drifting square-wave gratings (16 directions, 100% contrast, 0.1 cycles/°, 4 cycles/sec., 3 sec. stimulus period followed by 2-3 sec. ISI, plus a blank) were presented to the contralateral eye in a pseudorandom sequence for 8 trials.

[0336] Images were corrected for in-plane motion using a correlation-based approach (MATLAB). ROI drawing was performed in ImageJ (Schindelin et al., 2012, *Nat. Methods*, 9:676-82). Fluorescence time-courses were computed as the mean of all pixels within the ROI at each time point and were extracted as described in Sage et al. (2012, ImageJ User developer Conference 1:1). Fluorescence time courses were then synchronized with stimulus information, and visually evoked responses were computed as changes in fluorescence relative to the baseline fluorescence. Peak $\Delta F/F$ responses for field ROIs and dendritic spines ROIs were computed using the Fourier analysis to calculate mean and modulation amplitudes for each stimulus presentation, which were summed together.

Example 6C: Mouse Neuronal Culture Analysis

[0337] Primary Hippocampal Neuron Cultures

[0338] Primary hippocampal neuron cultures were prepared from embryonic mice (E16) as described previously (Woitecki et al., 2016, *J. Neurosci.*, 36:2561-70). Hippocampi were rinsed 3-5 times in Hank's Balanced Salt Solution (HBSS, Life technologies) and digested with trypsin (25 mg/ml, Life Technologies) for 20 min at 37° C. followed by DNase I (1 mg/ml; Roche). Subsequently, the tissue was dissociated using cannulas (three times 0.9 mm×40 mm; three times 0.45 mm×23 mm) and the solution was passed through a Nylon cell strainer (100 µm; BD Biosciences). The mesh was rinsed with 4-10 ml basal medium eagle (BME, Life technologies) supplemented with 0.5% glucose (Sigma-Aldrich), 10% fetal calf serum (FCS), 2% B-27, and 0.5 mM L-glutamine (all Life Technologies) to collect all cells. After counting, the cells were plated on cover slips in a 24-well cell culture plate at a density of 70,000 cells per 24-well and cultured in a humidified incubator at 37° C. and 5% CO₂.

[0339] Viral Vector Production

[0340] Recombinant AAV2/1 genomes were generated by large scale triple transfection of HEK293 cells as described previously (Marvin et al., 2013, *Nat. Methods*, 10:162-70). The adeno-associated virus (AAV) plasmid coding for SF-iGluSnFR.S72A or SF-iGluSnFR.A184V, helper plasmids encoding rep and cap genes (pRV1 and pH21), and adenoviral helper pFA6 (Stratagene) were transfected using the calcium phosphate transfection method. Cells were harvested ~72 h after transfection. To purify the virus, cell pellets were lysed in the presence of 0.5% sodium deoxycholate (Sigma) and 50 units/ml Benzonase endonuclease (Sigma). rAAV viral particles were purified from the cell lysate by HiTrap heparin HP column purification (GE Healthcare) and then concentrated using Amicon Ultra Centrifugal Filters (Millipore) until a final stock volume of 500 µl was reached.

[0341] Viral Transduction and Image Acquisition

[0342] Primary hippocampal neurons were transduced with AAV2/1.hSynapsin1.SF-iGluSnFR.S72A or with AAV2/1.hSynapsin1.SF-iGluSnFR.A184V on DIV4 and imaged on DIV13. A low amplitude field stimulation (1 msec, 20 mA, platinum bar electrodes) was applied to recruit

a small fraction (~20%) of neurons. Images were acquired with an EM-CCD camera (frame time 5-50 msec) and a stabilized LED light source of cultures visualized through a coverslip with high NA objective. All experiments were performed in Tyrode's solution (1 ml/min) at RT. Low and high affinity versions of SF-iGluSnFR were expressed in a comparable manner.

[0343] Glutamate Release Site Localization

[0344] Primary hippocampal neurons were transduced with rAAV-SF-iGluSnFR.S72A or with rAAV-SF-iGluSnFR.A184V on DIV3-5 and used for experiments on DIV13-18. A low amplitude electrical field stimulation (1 msec., 20 mA, platinum bar electrodes) was applied to activate a small fraction (~20%) of neurons only. Per experiment, stimuli were applied 16-25 times at an inter-stimulus interval of 20-60 sec. Images were acquired with an EM-CCD camera (Hamamatsu ImagEM X1, 8 ms exposure, 125 Hz acquisition rate) attached to an inverted microscope (Nikon T1 Eclipse) using a triggered, stabilized LED light source (Cairn OptoLED with 470 nm excitation wavelength, 470/40 emission filter and 525/50 excitation filter). Cells were imaged through a coverslip with a high NA objective (Zeiss, 63×, 1.4 NA, water). All experiments were performed in saline (1 ml/min, as described above) at room temperature.

[0345] In each experiment, 30 images were acquired per stimulation trial (20 before and 10 after stimulation). Each of the 30 images was registered with StackReg Plugin in ImageJ to the first image. The image series was then normalized to the average of 5 frames before stimulation to distinguish responding sites (>1) and non-responding structures (~1). For selection of responding sites to be included in the analysis, 10 normalized images subsequent to the stimulus in the first trial were averaged. All spots of increased fluorescence (FIG. 64c,d) that reached at least 50% of the $\Delta F/F$ value of the brightest spot in the image were defined as responding sites and used for further analysis. The spatial extent of glutamate release sites was quantified by extracting a brightness profile based on a line (length: 12-30 pixels, width: 3 pixels) drawn along the underlying neurite. These profiles were calculated for each stimulation trial and each responding site in an experiment and fitted by Gaussians with Igor Pro 6.3 (Wavemetrics).

[0346] In each experiment (n=6 and 8 for S72A and A184V, respectively, each consisting of 16-25 trials) the mean deviation of the center (X₀ position), the average width and the average amplitude of the fitted Gaussians were calculated per response site and averaged across all experiments and statistically compared by an unpaired Mann-Whitney test, n=28 and 53 for S72A and A184V, respectively).

Example 6D: Cerebellar Parallel Fiber Analysis

[0347] Stereotaxic Injections.

[0348] To fluorescently label boutons of parallel fibers, stereotaxic injections of viral vectors expressing SF-iGluSnFR or GCaMP6f into cerebellar vermis were performed. The following vectors were used: AAV-DJ.hSynapsin.SF-iGluSnFR (1.9×10¹³ GC/ml), AAV2/1.hSynapsin.SF-iGluSnFR.S72A (2.6×10¹³ GC/ml), AAV-DJ.CAGFLEX.SF-iGluSnFR.S72A (6.3×10¹² GC/ml) or AAV-DJ.hSynapsin.GCaMP6f (1.2×10¹³ GC/ml). Mice between 30 and 60 days old were deeply anesthetized before surgery with a mixture of hypnotic (ketamine 1.5%, Merial) and analgesic (xylazine

0.05%, Bayer) anesthetics mixed in NaCl and injected in the peritoneum. A local anesthetic (xylocaine 2% gel, Newpharma) was applied on top of the location of the cranial incision. The anesthetized mouse was then placed on a stereotaxic frame adaptor comprising adjustable ear bars and tooth holder. The skull was then perforated at the injection site with a surgical drill. The vermis was identified using the Paxinos and Franklin mouse brain atlas. The injection of viral constructs in the vermis (100 nl; 6.5 mm caudal to bregma, lateral 0.2 mm, ventral 3.6 mm and 3.4 mm) was performed by slow infusion (100 nl/min) with steel needles (26G×50 mm and 36G×70 mm, Phymep) connected to a pump via a catheter and a Hamilton syringe. Injected mice were then kept 2 to 4 weeks to allow transgene expression.

[0349] Slice Preparation

[0350] All protocols were approved by the ethics committee CEEA-Paris1. Cerebellar acute slices were prepared from adult CB6F1 mice (F1 cross of BalbC and C57Bl/6J) or Gabra6 mice (B6; 129P2-Gabra6^{tm2(cre)W^{wis}/Mmucd}) of postnatal day 41 to 123. The mice were killed by rapid decapitation, after which the brains were quickly removed and placed in an ice-cold solution containing (in mM): 2.5 KCl, 0.5 CaCl₂, 4 MgCl₂, 1.25 NaH₂PO₄, 24 NaHCO₃, 25 glucose, 230 sucrose, and 0.5 ascorbic acid bubbled with 95% O₂ and 5% CO₂. Coronal slices were cut from the dissected cerebellar vermis using a vibratome (Leica VT1200S). After preparation, the slices were incubated at 32° C. for 30 minutes in the following solution (in mM): 85 NaCl, 2.5 KCl, 0.5 CaCl₂, 4 MgCl₂, 1.25 NaH₂PO₄, 24 NaHCO₃, 25 glucose, 75 sucrose and 0.5 ascorbic acid. Slices were then transferred to an external recording solution containing (in mM): 125 NaCl, 2.5 KCl, 1.5 CaCl₂, 1.5 MgCl₂, 1.25 NaH₂PO₄, 25 NaHCO₃, 25 glucose and 0.5 ascorbic acid, and maintained at room temperature for up to 6 hours. All slice recordings were performed at 36-38° C.

[0351] Transmitted Light and Fluorescence Imaging

[0352] Parallel fiber and boutons expressing SF-iGluSnFR or GCaMP6f were identified using an Ultima two-photon scanning scanhead (Bruker Nano Surfaces Division, Middleton, Wis., USA) that was mounted on an Olympus BX61W1 microscope, equipped with a water-immersion objective (60×, 1.1 NA, Olympus Optical, Tokyo, Japan) and infrared DotD-gradient contrast. Two-photon excitation was performed with a pulsed Ti: Sapphire laser (DeepSee, Spectra-Physics, France) tuned to 920 nm for imaging morphology, glutamate and Ca²⁺ fluorescence detection.

[0353] Boutons from parallel fibers were identified by increase fluorescence as response to 100 or 300 Hz trains. The probe response was evoked with 60 μs voltage pulses 5-15 V above threshold (Digitimer Ltd, Letchworth Garden City, UK) using a patch pipette (typically with a tip resistance of 4-6 MΩ) filled with ACSF and placed in the molecular layer adjacent to labelled parallel fibers. Activation of boutons was routinely confirmed by verifying increase in fluorescence in response to 100 or 300 Hz trains of stimulation. Line-scan imaging through boutons was performed at dwell time of 0.8 μsec per pixel, for 300 to 800 msec. Individual traces were background subtracted and averaged with no smoothing or filtration for single events for SF-iGluSnFR, or background subtracted and averaged with smoothing for GCaMP6f, 20 Hz and 100 Hz trains. SNR was calculated from the peak of the fit to the fluorescent events

divided by the average SD of a 20 msec baseline window. Data were analyzed and presented using custom-written macros in Igor Pro.

Example 6E: Fast Imaging of SF-Venus-iGluSnFR

[0354] Primary Rat Hippocampal Neuron Cultures

[0355] A mixed cell culture (neurons and glia) was prepared from Sprague-Dawley rat pups (Charles River Laboratories). Briefly, P0 pups were decapitated, and the brains were dissected into ice-cold neural dissection solution (NDS, 10 mM HEPES (Sigma) in HBSS (Invitrogen), pH 7.4). Hippocampi were dissected and cut into small pieces to facilitate enzyme digestion. Hippocampi pieces were transferred using a large bore pipette into a 15 ml conical tube and incubated with enzyme digest solution (Papain, Worthington Biologicals) at 37° C. for 30 min. After 30 min., the enzyme solution was removed, and Plating Media (MEM media containing 10% FBS) was added and tissue pieces were triturated resulting in mostly single cells. The cell suspension was filtered using a 45 μm filter. The filtered cell suspension was centrifuged, and the resulting cell pellet was re-suspended with Plating Media and counted.

[0356] For electroporation, 1 μg of DNA was mixed with 1×10⁶ cells using the Amaxa Nucleofector II instrument. Cells were plated onto coverslips coated with Poly-D-Lysine (Sigma) and kept at 37° C., 5% CO₂ in PM for ~24 hours and then in NbActiv4 (BrainBits) was added for the duration with medium exchanges every 4 days.

[0357] Glutamate Uncaging and Imaging

[0358] Rat hippocampal culture was imaged on DIV19 at room temperature in HEPES buffered Tyrode's solution (145 mM NaCl, 2.5 mM KCl, 10 mM glucose, 10 mM HEPES, 2 mM CaCl₂, 1 mM MgCl₂, pH 7.4).

[0359] Excitation was with a 1030 nm, 5 MHz, 190 fsec laser (Menlo Systems, model: Bluecut). Average power was 39 mW at the sample. Fluorescence collected at 560/80 nm with a Hamamatsu MPPC detector. The field of view is a 256 μm diameter circle, 1280 pixels across. The bath contained HEPES buffered Tyrode's solution plus 10 μM NBQX and 150 μM RuBi-Glutamate (Tocris). Glutamate uncaging was performed with 420 nm fiber-coupled LEDs (Thorlabs M420F2). The tips of the fibers were imaged onto the sample plane through the same objective used for activity imaging.

Example 6F: Summary of Results

[0360] The intensity-based glutamate-sensing fluorescent reporter (iGluSnFR) (Marvin et al., 2013, Nat. Methods, 10:162-70) has become an invaluable tool for studying glutamate dynamics in diverse systems, including retina (Park et al., 2014, J. Neurosci., 34:3976-81; Borghuis et al., 2013, J. Neurosci., 33:10972-85), mouse olfactory bulb (Brunert et al., 2016, J. Neurosci., 36:6820-35) and cat visual cortex (O'Herron et al., 2016, Nature, 534:378-82). Beyond specific circuits, iGluSnFR also allows mesoscale "functional connectomic" mapping (Xie et al., 2016, J. Neurosci., 36:1261-72) and mechanistic studies of Huntington's disease (Jiang et al., 2016, J. Neurosci., 36:3453-70), synaptic spillover (Rosa et al., 2015, eLife, 4:728), cortical spreading depression (Enger et al., 2015, Cerebral Cortex, 25:4469-76) and exocytotic vesicle fusion (Bao et al., 2016, Nat. Struct. Biol., 23:67-73). However, iGluSnFR is insufficient for some applications due to poor expression (in some brain regions), and kinetics that do not match the time

courses of some observations. Here, we describe variants that are functionally brighter (due to increased expression on cell membrane), have tighter or weaker affinity (resulting from slower or faster off-rates), and fluoresce blue, green, or yellow.

[0361] Replacement of circularly permuted eGFP with circularly permuted “superfolder” GFP (Pedelacq et al., 2006, *Nat. Biotech.*, 24:79-88) (SF-iGluSnFR) yielded 5-fold higher soluble-protein expression levels in bacteria (0.5 $\mu\text{mol/l}$ L growth vs. 0.1 $\mu\text{mol/l}$ L). Circular dichroism indicates an increase in melting temperature transition (T_m) of $\sim 5^\circ$ C. (FIG. 57). The 2-photon cross-section and excitation, emission, and absorption spectra of SF-iGluSnFR are similar to the original (FIG. 58a-d). Head-to-head comparison of SF-iGluSnFR with original iGluSnFR in mouse somatosensory cortex shows substantially more robust expression by the former (FIG. 59a,b). Under typical imaging conditions (<20 mW, 130-nanosecond dwell time per pixel), SF-iGluSnFR is bright enough for repeated imaging, while original iGluSnFR is too dim (FIG. 59c,d). While a faster 2-photon in vivo photobleaching rate was observed for SF-iGluSnFR in somatosensory cortex (FIG. 59e), partially-bleached SF-iGluSnFR was still brighter than iGluSnFR. Thus, SF-iGluSnFR will have superior expression in vivo, where the quantity of deliverable DNA can be limiting.

[0362] While the affinity of membrane-displayed iGluSnFR (4 μM) is adequate for some in vivo applications, tighter variants are needed for circumstances of limiting glutamate concentrations, such as at sparsely-firing synapses. Additionally, measuring glutamate release events with raster scanning microscopes requires variants with slower off-rates so that the decay time from glutamate binding is long enough to be sufficiently sampled at the operating frame rate for most experiments (typically <100 Hz). Replacement of eGFP with superfolder GFP increases the in vitro affinity of soluble SF-iGluSnFR two-fold compared to original iGluSnFR (40 μM vs. 80 FIG. 60a). To further modulate affinity, the conformational coupling between the open-closed equilibrium of bacterial periplasmic binding proteins (PBPs, e.g. the glutamate-binding protein in iGluSnFR) and their ligand-binding affinity (Marvin et al., 2001, *Nat. Struct. Biol.*, 8:795-8) was exploited. Briefly, mutation of residues in the “hinge” of PBPs can allosterically alter affinity, without compromising the stereochemical integrity of the ligand-binding site. In a bacterial lysate assay, an A184X library of the iGluSnFR glutamate-binding domain (mutated to valine in the original iGluSnFR) was screened. Reversion to alanine or other small amino acids tightened affinity, while larger side chains weakened affinity (FIG. 61).

[0363] A184S was introduced into SF-iGluSnFR to generate a tighter variant. (Reversion A184A had a low $\Delta F/F$.) Affinities of purified soluble protein were 7 μM and 40 μM for the A184S and A184V (unmutated from iGluSnFR) SF-iGluSnFR variants, respectively (FIG. 60a). The tighter affinity of the A184S variant arises from a slower off-rate (FIG. 60b). The affinity variants were re-cloned into an AAV vector containing an IgG secretion signal and a PDGFR transmembrane domain. Viral expression on cultured rat hippocampal neurons (AAV2/1.hSynapsin1.SF-iGluSnFR) yields glutamate affinities about an order of magnitude tighter than the soluble form (0.7 μM and 2 μM for A184S and A184V, respectively; FIG. 62). A similar increase in affinity upon membrane tethering was seen with the original

sensor (Marvin et al., 2013, *Nat. Methods*, 10:162-70). Whole-field stimulation (50 Hz) of these cultures shows that their relative half-times of fluorescence decay parallel their in vitro kinetics, with all variants having faster decay than GCaMP6f (FIG. 63).

[0364] In vivo, the tighter/slower SF-iGluSnFR.A184S variant shows improved detection of stimulus-evoked glutamate release in the ferret visual cortex in response to presented drifting gratings (FIG. 53a,b). Peak amplitudes reached 30% $\Delta F/F$ for SF-iGluSnFR.A184S but only 5% $\Delta F/F$ for SF-iGluSnFR.A184V when imaged at 30 Hz. Greater $\Delta F/F$ of SF-iGluSnFR.A184S allows extraction of robust orientation tuning curves compared to SF-iGluSnFR.A184V. Enhanced sensitivity of the A184S variant also allowed orientation-selective responses to be resolved in individual dendritic spines (FIG. 53c,d). Synaptic glutamate release as measured with SF-iGluSnFR.A184S was not only strongly selective for visual stimuli, but response amplitudes across individual trials were consistently greater than the A184V variant when examining all stimulus-evoked responses (A184S median $\Delta F/F=16\%$, $n=72$ spines; A184V median $\Delta F/F=9\%$, $n=22$ spines; $p=2e-115$, Wilcoxon rank-sum test) or only preferred stimuli (A184S median $\Delta F/F=27\%$, $n=72$ spines; A184V median $\Delta F/F=14\%$, $n=22$ spines; $p=9e-23$, Wilcoxon rank-sum test) (FIG. 64).

[0365] While slow off-rate variants of SF-iGluSnFR are better for detecting individual synaptic events by temporal summation of fluorescence, faster off-rate variants are needed for temporal resolution spiking dynamics and at large synapses where glutamate clearance is limiting. A weaker variant of SF-iGluSnFR (S72A) was made by removing a hydrogen bond between the protein and glutamate. Soluble SF-iGluSnFR.S72A has 200 μM affinity for glutamate (FIG. 60a), arising from a combination of both slower on-rate and faster off-rate (FIG. 60b). In neuronal culture, S72A has an affinity of 35 μM , an order of magnitude weaker than its parent, A184V (FIG. 62).

[0366] In rat neuronal culture, without buffer perfusion, fluorescence of the culture (not localized to specific structures) returns to baseline within 100 msec. of a single electrical stimulation for S72A, faster than A184V, A184S, or GCaMP6f (FIG. 63). In mouse neuronal culture (FIG. 65), the substantially faster off-rate of S72A provides enhanced temporal resolution of paired (20 Hz) electrical stimuli over the A184V variant (FIG. 54a,b), making it useful for assessing short-term synaptic plasticity. A train of 6 electrical pulses (20 Hz) in 1 mM extracellular Ca^{2+} can be resolved as equal, individual release events by observation with S72A, while A184V yields an integrated signal (FIG. 54c,d). In 3.5 mM extracellular Ca^{2+} , vesicles are released with higher probability during the initial stimulation (Dodge et al., 1967, *J. Physiol.*, 193:419-32). This can be observed by S72A, as reported by a reduction in fluorescence response as the train of field pulses progresses (FIG. 54c), while these differences are obscured by the slower decay of A184V (FIG. 54d). Thus, while S72A has a lower $\Delta F/F$ in response to the same amount of glutamate being released (due to weaker affinity), its faster kinetics provides enhanced temporal resolution of synaptic activity. Similarly, S72A provides enhanced spatial resolution of glutamate release over A184V (FIG. 66).

[0367] With fast rise and decay times, it was examined whether SF-iGluSnFR could be used as an alternative to GCaMP6f for monitoring neuronal activity in mouse cer-

ebellar brain slice. Single cerebellar granule cell bouton responses to single action potentials (APs) could indeed be resolved using fast linescan detection (<1 ms per line; FIG. 55a), and were much faster than GCaMP6f rise and decay times at both 2 mM and 1.5 mM extracellular calcium. The S72A variant had by far the fastest response (S72 half decay 7.9 ± 1.0 ms, A184V 28.1 ± 1.6 ms, GCaMP6f 1.5 mM $[Ca^{2+}]_e$ 37.9 ± 3.9 ms, GCaMP6f 1.5 mM $[Ca^{2+}]_e$ 108.6 ± 8.8 ms). The signal-to-noise-ratios (SNRs) were best for A184V, but even S72A produced better SNRs than GCaMP6f under physiological extracellular calcium concentrations (1.5 mM). The superior SNR of A184V showed putative single vesicle release events in single trials (FIG. 55b). However, if many bouton responses are pooled and averaged for each trial, single spike detection at 20 Hz is feasible (see average trace, FIG. 55b). For 20 Hz stimuli, both the A184V and S72A variants produced little accumulation of bouton fluorescence after 10 stimuli as compared to GCaMP6f (FIG. 55c), similar to the dendritic responses in culture (FIG. 54). For 100 Hz train stimuli, discrete release events could be detected, in contrast to GCaMP6f (FIG. 55d). Note the poor temporal precision of the train response, in contrast to A184V and S72A. Thus both A184V and S72A enable a larger dynamic range of reported firing frequencies, with S72A providing the largest range due to its low affinity. Moreover, the fast kinetics of SF-iGluSnFR.A184V and SF-iGluSnFR.S72A could be used for a more reliable estimate of spike times (versus GCaMP6f), and are much better suited to high-frequency spike detection (>100 Hz) which is necessary for the high instantaneous firing rates of cerebellar granule cells (van Beugen et al., 2013, *Frontiers in Neural Circuits*, 7:95).

[0368] Introduction of chromophore mutations from GFP variants Azurite (Mena et al., 2006, *Nat. Biotech.*, 24:1569-71) or Venus (Nagai et al., 2002, *Nat. Biotech.*, 20:87-90) to SF-iGluSnFR led to functional blue and yellow versions, respectively. The former required re-optimization of the

residues that link the FP with the glutamate-binding protein. The latter was a straightforward modular replacement. (Annotated amino acid sequences are given in FIG. 67). SF-Azurite-iGluSnFR has significantly lower $\Delta F/F$ (FIG. 68), perhaps a result of intrinsic differences in chromophore structure. SF-Venus-iGluSnFR has similar affinity and maximum fluorescence response to glutamate as SF-iGluSnFR, but with red-shifted excitation and emission spectra (FIG. 69). Importantly, its 2-photon excitation spectrum is sufficiently red-shifted to allow strong excitation at 1030 nm (FIG. 69), compatible with relatively inexpensive, powerful femtosecond fiber lasers (Tang et al., 2009, *J. Biomed. Optics*, 14:030508). These powerful lasers enable simultaneous excitation of many foci, enabling very fast (1016 Hz) large-area imaging by recording projections of the sample and computationally reconstructing images (Kazempour, et al., 2018). In neuronal culture, two near-simultaneous pulses of glutamate uncaging can be resolved with both high spatial and temporal resolution by measuring fluorescence changes in a neuron expressing SF-Venus-iGluSnFR.A184V (FIG. 56).

[0369] The iGluSnFR variants described here increase the power of genetically encoded glutamate imaging. Affinity variants with altered kinetics broaden the range of observable glutamate release events. Chromatic mutants allow fast imaging with cheap lasers, and potential utility in multi-color imaging. Improved membrane targeting and photostability will be valuable in all applications.

OTHER EMBODIMENTS

[0370] It is to be understood that while the invention has been described in conjunction with the detailed description thereof, the foregoing description is intended to illustrate and not limit the scope of the invention, which is defined by the scope of the appended claims. Other aspects, advantages, and modifications are within the scope of the following claims.

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      20             25             30

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 35             40             45

Gly Asp Lys Gly Tyr Asn Gly Leu Ala Glu Val Gly Lys Lys Phe Glu
 50             55             60

Lys Asp Thr Gly Ile Lys Val Thr Val Glu His Pro Asp Lys Leu Glu
 65             70             75             80

Glu Lys Phe Pro Gln Val Ala Ala Thr Gly Asp Gly Pro Asp Ile Ile
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Leu	Ser	Thr	Gln	Ser	Lys	Leu	Ser	Lys	Asp	Pro	Asn	Glu	Lys	Arg	Asp
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Glu	Asp	Gly	Asn	Ile	Leu	Gly	His	Lys	Leu	Glu	Tyr	Asn	Phe	Asn	Gly
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Met	Ala	Asp	Lys	Gln	Lys	Asn	Gly	Ile	Lys	Ala	Asn	Phe	Lys	Ile	Arg
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His	Met	Val	Leu	Leu	Glu	Phe	Val	Thr	Ala	Ala	Gly	Ile	Thr	Leu	Gly
		275					280					285			
Met	Asp	Glu	Leu	Tyr	Lys	Gly	Gly	Thr	Gly	Gly	Ser	Met	Val	Ser	Lys
	290					295					300				
Gly	Glu	Glu	Leu	Phe	Thr	Gly	Val	Val	Pro	Ile	Leu	Val	Glu	Leu	Asp
	305					310				315					320
Gly	Asp	Val	Asn	Gly	His	Lys	Phe	Ser	Val	Ser	Gly	Glu	Gly	Glu	Gly
				325					330						335
Asp	Ala	Thr	Tyr	Gly	Lys	Leu	Thr	Leu	Lys	Phe	Ile	Cys	Thr	Thr	Gly
			340					345					350		
Lys	Leu	Pro	Val	Pro	Trp	Pro	Thr	Leu	Val	Thr	Thr	Leu	Thr	Tyr	Gly
		355					360					365			
Val	Gln	Cys	Phe	Ser	Arg	Tyr	Pro	Asp	His	Met	Lys	Gln	His	Asp	Phe
	370					375					380				
Phe	Lys	Ser	Ala	Met	Pro	Glu	Gly	Tyr	Ile	Gln	Glu	Arg	Thr	Ile	Phe
	385				390					395					400
Phe	Lys	Asp	Asp	Gly	Asn	Tyr	Lys	Thr	Arg	Ala	Glu	Val	Lys	Phe	Glu
				405					410						415
Gly	Asp	Thr	Leu	Val	Asn	Arg	Ile	Glu	Leu	Lys	Gly	Ile	Asp	Phe	Lys
			420					425					430		
Glu	Asp	Gly	Asn	Ile	Leu	Gly	His	Lys	Leu	Glu	Tyr	Asn	Phe	Asn	Gly
		435					440					445			
Gly	Tyr	Ala	Phe	Lys	Tyr	Glu	Asn	Gly	Lys	Tyr	Asp	Ile	Lys	Asp	Val
	450					455					460				
Gly	Val	Asp	Asn	Ala	Gly	Ala	Lys	Ala	Gly	Leu	Thr	Phe	Leu	Val	Asp
	465				470					475					480
Leu	Ile	Lys	Asn	Lys	His	Met	Asn	Ala	Asp	Thr	Asp	Tyr	Ser	Ile	Ala
				485					490						495
Glu	Ala	Ala	Phe	Asn	Lys	Gly	Glu	Thr	Ala	Met	Thr	Ile	Asn	Gly	Pro
			500						505					510	
Trp	Ala	Trp	Ser	Asn	Ile	Asp	Thr	Ser	Lys	Val	Asn	Tyr	Gly	Val	Thr
		515					520						525		
Val	Leu	Pro	Thr	Phe	Lys	Gly	Gln	Pro	Ser	Lys	Pro	Phe	Val	Gly	Val
	530					535						540			
Leu	Ser	Ala	Gly	Ile	Asn	Ala	Ala	Ser	Pro	Asn	Lys	Glu	Leu	Ala	Lys
	545				550					555					560
Glu	Phe	Leu	Glu	Asn	Tyr	Leu	Leu	Thr	Asp	Glu	Gly	Leu	Glu	Ala	Val
				565					570						575

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245					250					255					
Leu	Ser	Thr	Gln	Ser	Lys	Leu	Ser	Lys	Asp	Pro	Asn	Glu	Lys	Arg	Asp
			260					265					270		
His	Met	Val	Leu	Leu	Glu	Phe	Val	Thr	Ala	Ala	Gly	Ile	Thr	Leu	Gly
		275					280					285			
Met	Asp	Glu	Leu	Tyr	Lys	Gly	Gly	Ser	Met	Val	Ser	Lys	Gly	Glu	Glu
	290				295							300			
Leu	Phe	Thr	Gly	Val	Val	Pro	Ile	Leu	Val	Glu	Leu	Asp	Gly	Asp	Val
305					310					315					320
Asn	Gly	His	Lys	Phe	Ser	Val	Ser	Gly	Glu	Gly	Glu	Gly	Asp	Ala	Thr
				325					330					335	
Tyr	Gly	Lys	Leu	Thr	Leu	Lys	Phe	Ile	Cys	Thr	Thr	Gly	Lys	Leu	Pro
			340						345					350	
Val	Pro	Trp	Pro	Thr	Leu	Val	Thr	Thr	Leu	Thr	Tyr	Gly	Val	Gln	Cys
		355						360					365		
Phe	Ser	Arg	Tyr	Pro	Asp	His	Met	Lys	Gln	His	Asp	Phe	Phe	Lys	Ser
370							375						380		
Ala	Met	Pro	Glu	Gly	Tyr	Ile	Gln	Glu	Arg	Thr	Ile	Phe	Phe	Lys	Asp
385					390					395					400
Asp	Gly	Asn	Tyr	Lys	Thr	Arg	Ala	Glu	Val	Lys	Phe	Glu	Gly	Asp	Thr
				405					410						415
Leu	Val	Asn	Arg	Ile	Glu	Leu	Lys	Gly	Ile	Asp	Phe	Lys	Glu	Asp	Gly
			420					425						430	
Asn	Ile	Leu	Gly	His	Lys	Leu	Glu	Tyr	Asn	Phe	Asn	Gly	Gly	Tyr	Ala
		435						440					445		
Phe	Lys	Tyr	Glu	Asn	Gly	Lys	Tyr	Asp	Ile	Lys	Asp	Val	Gly	Val	Asp
450							455						460		
Asn	Ala	Gly	Ala	Lys	Ala	Gly	Leu	Thr	Phe	Leu	Val	Asp	Leu	Ile	Lys
465					470					475					480
Asn	Lys	His	Met	Asn	Ala	Asp	Thr	Asp	Tyr	Ser	Ile	Ala	Glu	Ala	Ala
				485					490						495
Phe	Asn	Lys	Gly	Glu	Thr	Ala	Met	Thr	Ile	Asn	Gly	Pro	Trp	Ala	Trp
			500						505					510	
Ser	Asn	Ile	Asp	Thr	Ser	Lys	Val	Asn	Tyr	Gly	Val	Thr	Val	Leu	Pro
		515						520					525		
Thr	Phe	Lys	Gly	Gln	Pro	Ser	Lys	Pro	Phe	Val	Gly	Val	Leu	Ser	Ala
530							535						540		
Gly	Ile	Asn	Ala	Ala	Ser	Pro	Asn	Lys	Glu	Leu	Ala	Lys	Glu	Phe	Leu
545					550					555					560
Glu	Asn	Tyr	Leu	Leu	Thr	Asp	Glu	Gly	Leu	Glu	Ala	Val	Asn	Lys	Asp
				565					570						575
Lys	Pro	Leu	Gly	Ala	Val	Ala	Leu	Lys	Ser	Tyr	Glu	Glu	Glu	Leu	Val
			580						585					590	
Asp	Lys	Pro	Arg	Ile	Ala	Ala	Thr	Met	Glu	Asn	Ala	Gln	Lys	Gly	Glu
		595						600					605		
Ile	Met	Pro	Asn	Ile	Pro	Gln	Met	Ser	Ala	Phe	Trp	Tyr	Ala	Val	Arg
610							615						620		
Thr	Ala	Val	Ile	Asn	Ala	Ala	Ser	Gly	Arg	Gln	Thr	Val	Asp	Glu	Asp
625					630					635					640
Leu	Lys	Asp	Ala	Gln	Thr	Arg	Ile	Thr	Lys	Gly	Ser	His	His	His	His
				645					650						655

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His His Gly

<210> SEQ ID NO 4
 <211> LENGTH: 661
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic recombinant peptide biosensor

<400> SEQUENCE: 4

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Met Arg Gly Ser His His His His His His Gly Met Ala Ser Met Thr
1          5          10          15
Gly Gly Gln Gln Met Gly Arg Asp Leu Tyr Asp Asp Asp Asp Lys Asp
20          25          30
Arg Trp Gly Ser Lys Ile Glu Glu Gly Lys Leu Val Ile Trp Ile Asn
35          40          45
Gly Asp Lys Gly Tyr Asn Gly Leu Ala Glu Val Gly Lys Lys Phe Glu
50          55          60
Lys Asp Thr Gly Ile Lys Val Thr Val Glu His Pro Asp Lys Leu Glu
65          70          75          80
Glu Lys Phe Pro Gln Val Ala Ala Thr Gly Asp Gly Pro Asp Ile Ile
85          90          95
Phe Trp Ala His Asp Arg Phe Gly Gly Tyr Ala Gln Ser Gly Leu Leu
100         105         110
Ala Glu Ile Thr Pro Asp Lys Ala Phe Gln Asp Lys Leu Tyr Pro Phe
115         120         125
Thr Trp Asp Ala Val Arg Tyr Asn Gly Lys Leu Ile Ala Tyr Pro Ile
130         135         140
Ala Val Glu Ala Leu Ser Leu Ile Tyr Asn Lys Asp Leu Leu Pro Asn
145         150         155         160
Pro Pro Lys Thr Trp Glu Glu Ile Pro Ala Leu Asp Lys Glu Leu Lys
165         170         175
Ala Lys Gly Lys Ser Ala Leu Met Phe Asn Leu Gln Glu Pro Tyr Phe
180         185         190
Thr Trp Pro Leu Ile Ala Ala Asp Gly Gly Tyr Ala Phe Lys Tyr Glu
195         200         205
Asn Gly Gly Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn
210         215         220
Gly Ile Lys Ala Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Gly
225         230         235         240
Val Gln Leu Ala Tyr His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly
245         250         255
Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Lys Leu
260         265         270
Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe
275         280         285
Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Gly
290         295         300
Gly Thr Gly Gly Ser Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly
305         310         315         320
Val Val Pro Ile Leu Val Glu Leu Asp Gly Asp Val Asn Gly His Lys
325         330         335

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Phe Ser Val Ser Gly Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu
 340 345 350
 Thr Leu Lys Phe Ile Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro
 355 360 365
 Thr Leu Val Thr Thr Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr
 370 375 380
 Pro Asp His Met Lys Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu
 385 390 395 400
 Gly Tyr Ile Gln Glu Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr
 405 410 415
 Lys Thr Arg Ala Glu Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg
 420 425 430
 Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly
 435 440 445
 His Lys Leu Glu Tyr Asn Phe Asn Gly Gly Lys Tyr Asp Ile Lys Asp
 450 455 460
 Val Gly Asp Asn Ala Gly Ala Lys Ala Gly Leu Thr Phe Leu Val Asp
 465 470 475 480
 Leu Ile Lys Asn Lys His Met Asn Ala Asp Thr Asp Tyr Ser Ile Ala
 485 490 495
 Glu Ala Ala Phe Asn Lys Gly Glu Thr Ala Met Thr Ile Asn Gly Pro
 500 505 510
 Trp Ala Trp Ser Asn Ile Asp Thr Ser Lys Val Asn Tyr Gly Val Thr
 515 520 525
 Val Leu Pro Thr Phe Lys Gly Gln Pro Ser Lys Pro Phe Val Gly Val
 530 535 540
 Leu Ser Ala Gly Ile Asn Ala Ala Ser Pro Asn Lys Glu Leu Ala Lys
 545 550 555 560
 Glu Phe Leu Glu Asn Tyr Leu Leu Thr Asp Glu Gly Leu Glu Ala Val
 565 570 575
 Asn Lys Asp Lys Pro Leu Gly Ala Val Ala Leu Lys Ser Tyr Glu Glu
 580 585 590
 Glu Leu Val Asp Lys Pro Arg Ile Ala Ala Thr Met Glu Asn Ala Gln
 595 600 605
 Lys Gly Glu Ile Met Pro Asn Ile Pro Gln Met Ser Ala Phe Trp Tyr
 610 615 620
 Ala Val Arg Thr Ala Val Ile Asn Ala Ala Ser Gly Gln Thr Val Asp
 625 630 635 640
 Glu Asp Leu Lys Asp Ala Gln Thr Arg Ile Thr Lys Gly Ser His His
 645 650 655
 His His His His Gly
 660

<210> SEQ ID NO 5
 <211> LENGTH: 663
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
 recombinant peptide biosensor
 <400> SEQUENCE: 5

Met Arg Gly Ser His His His His His His Gly Met Ala Ser Met Thr

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1	5	10	15
Gly Gly Gln Gln Met Gly Arg Asp Leu Tyr Asp Asp Asp Asp Lys Asp	20	25	30
Arg Trp Gly Ser Lys Ile Glu Glu Gly Lys Leu Val Ile Trp Ile Asn	35	40	45
Gly Asp Lys Gly Tyr Asn Gly Leu Ala Glu Val Gly Lys Lys Phe Glu	50	55	60
Lys Asp Thr Gly Ile Lys Val Thr Val Glu His Pro Asp Lys Leu Glu	65	70	75
Glu Lys Phe Pro Gln Val Ala Ala Thr Gly Asp Gly Pro Asp Ile Ile	85	90	95
Phe Trp Ala His Asp Arg Phe Gly Gly Tyr Ala Gln Ser Gly Leu Leu	100	105	110
Ala Glu Ile Thr Pro Asp Lys Ala Phe Gln Asp Lys Leu Tyr Pro Phe	115	120	125
Thr Trp Asp Ala Val Arg Tyr Asn Gly Lys Leu Ile Ala Tyr Pro Ile	130	135	140
Ala Val Glu Ala Leu Ser Leu Ile Tyr Asn Lys Asp Leu Leu Pro Asn	145	150	155
Pro Pro Lys Thr Trp Glu Glu Ile Pro Ala Leu Asp Lys Glu Leu Lys	165	170	175
Ala Lys Gly Lys Ser Ala Leu Met Phe Asn Leu Gln Glu Pro Tyr Phe	180	185	190
Thr Trp Pro Leu Ile Ala Ala Asp Gly Gly Tyr Ala Phe Lys Tyr Glu	195	200	205
Asn His Leu Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn	210	215	220
Gly Ile Lys Ala Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Gly	225	230	235
Val Gln Leu Ala Tyr His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly	245	250	255
Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Lys Leu	260	265	270
Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe	275	280	285
Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Gly	290	295	300
Gly Thr Gly Gly Ser Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly	305	310	315
Val Val Pro Ile Leu Val Glu Leu Asp Gly Asp Val Asn Gly His Lys	325	330	335
Phe Ser Val Ser Gly Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu	340	345	350
Thr Leu Lys Phe Ile Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro	355	360	365
Thr Leu Val Thr Thr Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr	370	375	380
Pro Asp His Met Lys Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu	385	390	395
Gly Tyr Ile Gln Glu Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr	405	410	415

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Lys Thr Arg Ala Glu Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg
 420 425 430
 Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly
 435 440 445
 His Lys Leu Glu Tyr Asn Phe Asn Gly Gly Lys Tyr Asp Ile Lys Asp
 450 455 460
 Val Gly Val Asp Asn Ala Gly Ala Lys Ala Gly Leu Thr Phe Leu Val
 465 470 475 480
 Asp Leu Ile Lys Asn Lys His Met Asn Ala Asp Thr Asp Tyr Ser Ile
 485 490 495
 Ala Glu Ala Ala Phe Asn Lys Gly Glu Thr Ala Met Thr Ile Asn Gly
 500 505 510
 Pro Trp Ala Trp Ser Asn Ile Asp Thr Ser Lys Val Asn Tyr Gly Val
 515 520 525
 Thr Val Leu Pro Thr Phe Lys Gly Gln Pro Ser Lys Pro Phe Val Gly
 530 535 540
 Val Leu Ser Ala Gly Ile Asn Ala Ala Ser Pro Asn Lys Glu Leu Ala
 545 550 555 560
 Lys Glu Phe Leu Glu Asn Tyr Leu Leu Thr Asp Glu Gly Leu Glu Ala
 565 570 575
 Val Asn Lys Asp Lys Pro Leu Gly Ala Val Ala Leu Lys Ser Tyr Glu
 580 585 590
 Glu Glu Leu Val Lys Asp Pro Arg Ile Ala Ala Thr Met Glu Asn Ala
 595 600 605
 Gln Lys Gly Glu Ile Met Pro Asn Ile Pro Gln Met Ser Ala Phe Trp
 610 615 620
 Tyr Ala Val Arg Thr Ala Val Ile Asn Ala Ala Ser Gly Arg Gln Thr
 625 630 635 640
 Val Asp Glu Asp Leu Lys Asp Ala Gln Thr Arg Ile Thr Lys Gly Ser
 645 650 655
 His His His His His His Gly
 660

<210> SEQ ID NO 6

<211> LENGTH: 655

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic recombinant peptide biosensor

<400> SEQUENCE: 6

Met Arg Gly Ser His His His His His Gly Met Ala Ser Met Thr
 1 5 10 15
 Gly Gly Gln Gln Met Gly Arg Asp Leu Tyr Asp Asp Asp Lys Asp
 20 25 30
 Arg Trp Gly Ser Lys Ile Glu Glu Gly Lys Leu Val Ile Trp Ile Asn
 35 40 45
 Gly Asp Lys Gly Tyr Asn Gly Leu Ala Glu Val Gly Lys Lys Phe Glu
 50 55 60
 Lys Asp Thr Gly Ile Lys Val Thr Val Glu His Pro Asp Lys Leu Glu
 65 70 75 80
 Glu Lys Phe Pro Gln Val Ala Ala Thr Gly Asp Gly Pro Asp Ile Ile

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85					90					95					
Phe	Trp	Ala	His	Asp	Arg	Phe	Gly	Gly	Tyr	Ala	Gln	Ser	Gly	Leu	Leu
		100						105					110		
Ala	Glu	Ile	Thr	Pro	Asp	Lys	Ala	Phe	Gln	Asp	Lys	Leu	Tyr	Pro	Phe
		115					120					125			
Thr	Trp	Asp	Ala	Val	Arg	Tyr	Asn	Gly	Lys	Leu	Ile	Ala	Tyr	Pro	Ile
	130					135					140				
Ala	Val	Glu	Ala	Leu	Ser	Leu	Ile	Tyr	Asn	Lys	Asp	Leu	Leu	Pro	Asn
145					150					155					160
Pro	Pro	Lys	Thr	Trp	Glu	Glu	Ile	Pro	Ala	Leu	Asp	Lys	Glu	Leu	Lys
			165						170						175
Ala	Lys	Gly	Lys	Ser	Ala	Leu	Met	Phe	Asn	Leu	Gln	Glu	Pro	Tyr	Phe
			180					185					190		
Thr	Trp	Pro	Leu	Ile	Ala	Ala	Asp	Gly	Gly	Tyr	Ala	Phe	Lys	Tyr	Glu
		195					200					205			
Asn	Gly	Lys	Tyr	Asp	Ile	Lys	Asp	Val	Gly	Val	Asp	Asn	Ala	Gly	Ala
210						215						220			
Lys	Ala	Gly	Leu	Thr	Phe	Leu	Val	Asp	Leu	Ile	Lys	Asn	Lys	His	Met
225					230						235				240
Asn	Ala	Asp	Thr	Asp	Tyr	Ser	Ile	Ala	Glu	Ala	Ala	Phe	Asn	Lys	Gly
			245						250						255
Glu	Thr	Ala	Met	Thr	Ile	Asn	Gly	Pro	Trp	Ala	Trp	Ser	Asn	Ile	Asp
			260					265						270	
Thr	Ser	Lys	Val	Asn	Tyr	Gly	Val	Thr	Val	Leu	Pro	Thr	Phe	Lys	Gly
		275					280						285		
Gln	Pro	Ser	Lys	Pro	Phe	Val	Gly	Val	Leu	Ser	Ala	Gly	Ile	Asn	Ala
290						295					300				
Ala	Ser	Pro	Asn	Lys	Glu	Leu	Ala	Lys	Glu	Phe	Leu	Glu	Asn	Tyr	Leu
305					310					315					320
Leu	Thr	Asp	Glu	Gly	Leu	Glu	Ala	Val	Asn	Lys	Asp	Lys	Pro	Leu	Gly
			325						330						335
Ala	Val	Ala	Leu	Lys	Ser	Tyr	Glu	Glu	Glu	Leu	Gly	Gly	Ser	His	Asn
			340						345					350	
Val	Tyr	Ile	Met	Ala	Asp	Lys	Gln	Arg	Asn	Gly	Ile	Lys	Ala	Asn	Phe
		355					360						365		
Lys	Ile	Arg	His	Asn	Ile	Glu	Asp	Gly	Gly	Val	Gln	Leu	Ala	Tyr	His
370						375					380				
Tyr	Gln	Gln	Asn	Thr	Pro	Ile	Gly	Asp	Gly	Pro	Val	Leu	Leu	Pro	Asp
385					390					395					400
Asn	His	Tyr	Leu	Ser	Thr	Gln	Ser	Lys	Leu	Ser	Lys	Asp	Pro	Asn	Glu
			405						410						415
Lys	Arg	Asp	His	Met	Val	Leu	Leu	Glu	Phe	Val	Thr	Ala	Ala	Gly	Ile
			420						425					430	
Thr	Leu	Gly	Met	Asp	Glu	Leu	Tyr	Lys	Gly	Gly	Thr	Gly	Gly	Ser	Met
		435					440						445		
Val	Ser	Lys	Gly	Glu	Glu	Leu	Phe	Thr	Gly	Val	Val	Pro	Ile	Leu	Val
		450					455					460			
Glu	Leu	Asp	Gly	Asp	Val	Asn	Gly	His	Lys	Phe	Ser	Val	Ser	Gly	Glu
465					470					475					480
Gly	Glu	Gly	Asp	Ala	Thr	Tyr	Gly	Lys	Leu	Thr	Leu	Lys	Phe	Ile	Cys
				485					490						495

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Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu
      500                               505                               510

Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys Gln
      515                               520                               525

His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Ile Gln Glu Arg
      530                               535                               540

Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val
      545                               550                               555                               560

Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile
      565                               570                               575

Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn
      580                               585                               590

Phe Asn Gly Gly Ala Lys Asp Pro Arg Ile Ala Ala Thr Met Glu Asn
      595                               600                               605

Ala Gln Lys Gly Glu Ile Met Pro Asn Ile Pro Gln Met Ser Ala Phe
      610                               615                               620

Trp Tyr Ala Val Arg Thr Ala Val Ile Asn Ala Ala Ser Gly Arg Gln
      625                               630                               635                               640

Thr Val Asp Glu Asp Leu Lys Asp Ala Gln Thr Arg Ile Thr Lys
      645                               650                               655

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<210> SEQ ID NO 7

<211> LENGTH: 655

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic recombinant peptide biosensor

<400> SEQUENCE: 7

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Met Arg Gly Ser His His His His His Gly Met Ala Ser Met Thr
1      5      10      15

Gly Gly Gln Gln Met Gly Arg Asp Leu Tyr Asp Asp Asp Lys Asp
20     25     30

Arg Trp Gly Ser Lys Ile Glu Glu Gly Lys Leu Val Ile Trp Ile Asn
35     40     45

Gly Asp Lys Gly Tyr Asn Gly Leu Ala Glu Val Gly Lys Lys Phe Glu
50     55     60

Lys Asp Thr Gly Ile Lys Val Thr Val Glu His Pro Asp Lys Leu Glu
65     70     75     80

Glu Lys Phe Pro Gln Val Ala Ala Thr Gly Asp Gly Pro Asp Ile Ile
85     90     95

Phe Trp Ala His Asp Arg Phe Gly Gly Tyr Ala Gln Ser Gly Leu Leu
100    105    110

Ala Glu Ile Thr Pro Asp Lys Ala Phe Gln Asp Lys Leu Tyr Pro Phe
115    120    125

Thr Trp Asp Ala Val Arg Tyr Asn Gly Lys Leu Ile Ala Tyr Pro Ile
130    135    140

Ala Val Glu Ala Leu Ser Leu Ile Tyr Asn Lys Asp Leu Leu Pro Asn
145    150    155    160

Pro Pro Lys Thr Trp Glu Glu Ile Pro Ala Leu Asp Lys Glu Leu Lys
165    170    175

Ala Lys Gly Lys Ser Ala Leu Met Phe Asn Leu Gln Glu Pro Tyr Phe

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180				185				190							
Thr	Trp	Pro	Leu	Ile	Ala	Ala	Asp	Gly	Gly	Tyr	Ala	Phe	Lys	Tyr	Glu
		195					200							205	
Asn	Gly	Lys	Tyr	Asp	Ile	Lys	Asp	Val	Gly	Val	Asp	Asn	Ala	Gly	Ala
	210					215					220				
Lys	Ala	Gly	Leu	Thr	Phe	Leu	Val	Asp	Leu	Ile	Lys	Asn	Lys	His	Met
	225				230					235					240
Asn	Ala	Asp	Thr	Asp	Tyr	Ser	Ile	Ala	Glu	Ala	Ala	Phe	Asn	Lys	Gly
			245							250					255
Glu	Thr	Ala	Met	Thr	Ile	Asn	Gly	Pro	Trp	Ala	Trp	Ser	Asn	Ile	Asp
			260						265						270
Thr	Ser	Lys	Val	Asn	Tyr	Gly	Val	Thr	Val	Leu	Pro	Thr	Phe	Lys	Gly
		275					280							285	
Gln	Pro	Ser	Lys	Pro	Phe	Val	Gly	Val	Leu	Ser	Ala	Gly	Ile	Asn	Ala
	290					295					300				
Ala	Ser	Pro	Asn	Lys	Glu	Leu	Ala	Lys	Glu	Phe	Leu	Glu	Asn	Tyr	Leu
	305				310					315					320
Leu	Thr	Asp	Glu	Gly	Leu	Glu	Ala	Val	Asn	Lys	Asp	Lys	Pro	Leu	Gly
			325							330					335
Ala	Val	Ala	Leu	Lys	Ser	Tyr	Glu	Glu	Glu	Leu	Gly	Gly	Ser	His	Asn
			340						345						350
Val	Tyr	Ile	Met	Ala	Asp	Lys	Gln	Arg	Asn	Gly	Ile	Lys	Ala	Asn	Phe
		355					360							365	
Lys	Ile	Arg	His	Asn	Ile	Glu	Asp	Gly	Gly	Val	Gln	Leu	Ala	Tyr	His
	370					375					380				
Tyr	Gln	Gln	Asn	Thr	Pro	Ile	Gly	Asp	Gly	Pro	Val	Leu	Leu	Pro	Asp
	385				390					395					400
Asn	His	Tyr	Leu	Ser	Thr	Gln	Ser	Lys	Leu	Ser	Lys	Asp	Pro	Asn	Glu
			405							410					415
Lys	Arg	Asp	His	Met	Val	Leu	Leu	Glu	Phe	Val	Thr	Ala	Ala	Gly	Ile
			420						425					430	
Thr	Leu	Gly	Met	Asp	Glu	Leu	Tyr	Lys	Gly	Gly	Thr	Gly	Gly	Ser	Met
		435					440							445	
Val	Ser	Lys	Gly	Glu	Glu	Leu	Phe	Thr	Gly	Val	Val	Pro	Ile	Leu	Val
	450					455					460				
Glu	Leu	Asp	Gly	Asp	Val	Asn	Gly	His	Lys	Phe	Ser	Val	Ser	Gly	Glu
	465				470					475					480
Gly	Glu	Gly	Asp	Ala	Thr	Tyr	Gly	Lys	Leu	Thr	Leu	Lys	Phe	Ile	Cys
			485							490					495
Thr	Thr	Gly	Lys	Leu	Pro	Val	Pro	Trp	Pro	Thr	Leu	Val	Thr	Thr	Leu
			500						505						510
Thr	Tyr	Gly	Val	Gln	Cys	Phe	Ser	Arg	Tyr	Pro	Asp	His	Met	Lys	Gln
		515					520							525	
His	Asp	Phe	Phe	Lys	Ser	Ala	Met	Pro	Glu	Gly	Tyr	Ile	Gln	Glu	Arg
	530					535					540				
Thr	Ile	Phe	Phe	Lys	Asp	Asp	Gly	Asn	Tyr	Lys	Thr	Arg	Ala	Glu	Val
	545				550					555					560
Lys	Phe	Glu	Gly	Asp	Thr	Leu	Val	Asn	Arg	Ile	Glu	Leu	Lys	Gly	Ile
			565							570					575
Asp	Phe	Lys	Glu	Asp	Gly	Asn	Ile	Leu	Gly	His	Lys	Leu	Glu	Tyr	Asn
			580						585						590

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Phe Asn Asn Pro Ala Lys Asp Pro Arg Ile Ala Ala Thr Met Glu Asn
 595 600 605

Ala Gln Lys Gly Glu Ile Met Pro Asn Ile Pro Gln Met Ser Ala Phe
 610 615 620

Trp Tyr Ala Val Arg Thr Ala Val Ile Asn Ala Ala Ser Gly Arg Gln
 625 630 635 640

Thr Val Asp Glu Asp Leu Lys Asp Ala Gln Thr Arg Ile Thr Lys
 645 650 655

<210> SEQ ID NO 8
 <211> LENGTH: 659
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
 recombinant peptide biosensor

<400> SEQUENCE: 8

Met Arg Gly Ser His His His His His Gly Met Ala Ser Met Thr
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Gly Gly Gln Gln Met Gly Arg Asp Leu Tyr Asp Asp Asp Lys Asp
 20 25 30

Arg Trp Gly Ser Lys Ile Glu Glu Gly Lys Leu Val Ile Trp Ile Asn
 35 40 45

Gly Asp Lys Gly Tyr Asn Gly Leu Ala Glu Val Gly Lys Lys Phe Glu
 50 55 60

Lys Asp Thr Gly Ile Lys Val Thr Val Glu His Pro Asp Lys Leu Glu
 65 70 75 80

Glu Lys Phe Pro Gln Val Ala Ala Thr Gly Asp Gly Pro Asp Ile Ile
 85 90 95

Phe Trp Ala His Asp Arg Phe Gly Gly Tyr Ala Gln Ser Gly Leu Leu
 100 105 110

Ala Glu Ile Thr Pro Asp Lys Ala Phe Gln Asp Lys Leu Tyr Pro Phe
 115 120 125

Thr Trp Asp Ala Val Arg Tyr Asn Gly Lys Leu Ile Ala Tyr Pro Ile
 130 135 140

Ala Val Glu Ala Leu Ser Leu Ile Tyr Asn Lys Asp Leu Leu Pro Asn
 145 150 155 160

Pro Pro Lys Thr Trp Glu Glu Ile Pro Ala Leu Asp Lys Glu Leu Lys
 165 170 175

Ala Lys Gly Lys Ser Ala Leu Met Phe Asn Leu Gln Glu Pro Tyr Phe
 180 185 190

Thr Trp Pro Leu Ile Ala Ala Asp Gly Gly Tyr Ala Phe Lys Tyr Glu
 195 200 205

Asn Gly Lys Tyr Asp Ile Lys Asp Val Gly Val Asp Asn Ala Gly Ala
 210 215 220

Lys Ala Gly Leu Thr Phe Leu Val Asp Leu Ile Lys Asn Lys His Met
 225 230 235 240

Asn Ala Asp Thr Asp Tyr Ser Ile Ala Glu Ala Ala Phe Asn Lys Gly
 245 250 255

Glu Thr Ala Met Thr Ile Asn Gly Pro Trp Ala Trp Ser Asn Ile Asp
 260 265 270

Thr Ser Lys Val Asn Tyr Gly Val Thr Val Leu Pro Thr Phe Lys Gly

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275				280				285							
Gln	Pro	Ser	Lys	Pro	Phe	Val	Gly	Val	Leu	Ser	Ala	Gly	Ile	Asn	Ala
290						295					300				
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305					310					315					320
Leu	Thr	Asp	Glu	Gly	Leu	Glu	Ala	Val	Asn	Lys	Asp	Lys	Pro	Leu	Gly
					325				330					335	
Ala	Val	Ala	Leu	Lys	Ser	Tyr	Glu	Glu	Glu	Leu	Val	Lys	Asp	Pro	Arg
			340						345				350		
Ser	His	Asn	Val	Tyr	Ile	Met	Ala	Asp	Lys	Gln	Lys	Asn	Gly	Ile	Lys
		355					360						365		
Ala	Asn	Phe	Lys	Ile	Arg	His	Asn	Ile	Glu	Asp	Gly	Gly	Val	Gln	Leu
	370					375					380				
Ala	Tyr	His	Tyr	Gln	Gln	Asn	Thr	Pro	Ile	Gly	Asp	Gly	Pro	Val	Leu
	385				390					395					400
Leu	Pro	Asp	Asn	His	Tyr	Leu	Ser	Thr	Gln	Ser	Lys	Leu	Ser	Lys	Asp
				405					410					415	
Pro	Asn	Glu	Lys	Arg	Asp	His	Met	Val	Leu	Leu	Glu	Phe	Val	Thr	Ala
			420						425				430		
Ala	Gly	Ile	Thr	Leu	Gly	Met	Asp	Glu	Leu	Tyr	Lys	Gly	Gly	Thr	Gly
		435				440						445			
Gly	Ser	Met	Val	Ser	Lys	Gly	Glu	Glu	Leu	Phe	Thr	Gly	Val	Val	Pro
	450				455						460				
Ile	Leu	Val	Glu	Leu	Asp	Gly	Asp	Val	Asn	Gly	His	Lys	Phe	Ser	Val
	465				470					475					480
Ser	Gly	Glu	Gly	Glu	Gly	Asp	Ala	Thr	Tyr	Gly	Lys	Leu	Thr	Leu	Lys
				485					490					495	
Phe	Ile	Cys	Thr	Thr	Gly	Lys	Leu	Pro	Val	Pro	Trp	Pro	Thr	Leu	Val
			500						505				510		
Thr	Thr	Leu	Thr	Tyr	Gly	Val	Gln	Cys	Phe	Ser	Arg	Tyr	Pro	Asp	His
		515					520						525		
Met	Lys	Gln	His	Asp	Phe	Phe	Lys	Ser	Ala	Met	Pro	Glu	Gly	Tyr	Ile
	530				535						540				
Gln	Glu	Arg	Thr	Ile	Phe	Phe	Lys	Asp	Asp	Gly	Asn	Tyr	Lys	Thr	Arg
	545				550					555					560
Ala	Glu	Val	Lys	Phe	Glu	Gly	Asp	Thr	Leu	Val	Asn	Arg	Ile	Glu	Leu
				565					570					575	
Lys	Gly	Ile	Asp	Phe	Lys	Glu	Asp	Gly	Asn	Ile	Leu	Gly	His	Lys	Leu
			580						585				590		
Glu	Tyr	Asn	Phe	Asn	Ala	Ala	Thr	Met	Glu	Asn	Ala	Gln	Lys	Gly	Glu
		595					600						605		
Ile	Met	Pro	Asn	Ile	Pro	Gln	Met	Ser	Ala	Phe	Trp	Tyr	Ala	Val	Arg
	610				615						620				
Thr	Ala	Val	Ile	Asn	Ala	Ala	Ser	Gly	Arg	Gln	Thr	Val	Asp	Glu	Asp
	625				630					635				640	
Leu	Lys	Asp	Ala	Gln	Thr	Arg	Ile	Thr	Lys	Gly	Ser	His	His	His	His
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His	His	Gly													

<210> SEQ ID NO 9

<211> LENGTH: 14

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<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (8)..(9)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 9

Pro Leu Ile Ala Ala Asp Gly Xaa Xaa Asn Val Tyr Ile Met
1 5 10

<210> SEQ ID NO 10
<211> LENGTH: 13
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (7)..(8)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 10

Pro Leu Ile Ala Ala Asp Xaa Xaa Asn Val Tyr Ile Met
1 5 10

<210> SEQ ID NO 11
<211> LENGTH: 15
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (9)..(10)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 11

Pro Leu Ile Ala Ala Asp Gly Gly Xaa Xaa Asn Val Tyr Ile Met
1 5 10 15

<210> SEQ ID NO 12
<211> LENGTH: 15
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (8)..(8)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 12

Pro Leu Ile Ala Ala Asp Gly Xaa Pro Asn Val Tyr Ile Met Gly
1 5 10 15

<210> SEQ ID NO 13
<211> LENGTH: 15
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic

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oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (9)..(9)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 13

Pro Leu Ile Ala Ala Asp Gly Ile Xaa Asn Val Tyr Ile Met Gly
1 5 10 15

<210> SEQ ID NO 14
<211> LENGTH: 15
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (8)..(8)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 14

Pro Leu Ile Ala Ala Asp Pro Xaa Ser His Asn Val Tyr Ile Met
1 5 10 15

<210> SEQ ID NO 15
<211> LENGTH: 15
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (7)..(7)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 15

Pro Leu Ile Ala Ala Asp Xaa Pro Ser His Asn Val Tyr Ile Met
1 5 10 15

<210> SEQ ID NO 16
<211> LENGTH: 15
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (7)..(8)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 16

Pro Leu Ile Ala Ala Asp Xaa Xaa Ser His Asn Val Tyr Ile Met
1 5 10 15

<210> SEQ ID NO 17
<211> LENGTH: 15
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (7)..(8)

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<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 17

Pro Leu Ile Ala Ala Asp Xaa Xaa Ser His Asn Val Phe Ile Met
1 5 10 15

<210> SEQ ID NO 18

<211> LENGTH: 15

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide

<220> FEATURE:

<221> NAME/KEY: MOD_RES

<222> LOCATION: (8)..(8)

<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 18

Pro Leu Ile Ala Ala Asp Pro Xaa Ser His Asn Val Phe Ile Met
1 5 10 15

<210> SEQ ID NO 19

<211> LENGTH: 15

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide

<220> FEATURE:

<221> NAME/KEY: MOD_RES

<222> LOCATION: (8)..(8)

<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 19

Pro Leu Ile Ala Ala Asp Pro Xaa Ser Tyr Asn Val Phe Ile Met
1 5 10 15

<210> SEQ ID NO 20

<211> LENGTH: 15

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide

<220> FEATURE:

<221> NAME/KEY: MOD_RES

<222> LOCATION: (7)..(8)

<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 20

Pro Leu Ile Ala Ala Asp Xaa Xaa Ser Tyr Asn Val Phe Ile Met
1 5 10 15

<210> SEQ ID NO 21

<211> LENGTH: 15

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide

<220> FEATURE:

<221> NAME/KEY: MOD_RES

<222> LOCATION: (8)..(8)

<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 21

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Pro Leu Ile Ala Ala Asp Pro Xaa Ser Tyr Asn Val Phe Ile Met
1           5           10           15
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<210> SEQ ID NO 22
<211> LENGTH: 15
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
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<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (7)..(8)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 22
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Pro Leu Ile Ala Ala Asp Xaa Xaa Ser Tyr Asn Val Phe Ile Met
1           5           10           15
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<210> SEQ ID NO 23
<211> LENGTH: 15
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
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<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (8)..(8)
<223> OTHER INFORMATION: Any amino acid
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (10)..(10)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 23
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Pro Leu Ile Ala Ala Asp Pro Xaa Ser Xaa Asn Val Tyr Ile Met
1           5           10           15
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<210> SEQ ID NO 24
<211> LENGTH: 15
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
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<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (8)..(8)
<223> OTHER INFORMATION: Any amino acid
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (11)..(11)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 24
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Pro Leu Ile Ala Ala Asp Pro Xaa Ser His Xaa Val Tyr Ile Met
1           5           10           15
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<210> SEQ ID NO 25
<211> LENGTH: 15
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
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<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (8)..(8)
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<223> OTHER INFORMATION: Any amino acid
 <220> FEATURE:
 <221> NAME/KEY: MOD_RES
 <222> LOCATION: (12)..(12)
 <223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 25

Pro Leu Ile Ala Ala Asp Pro Xaa Ser His Asn Xaa Tyr Ile Met
 1 5 10 15

<210> SEQ ID NO 26
 <211> LENGTH: 15
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
 oligopeptide
 <220> FEATURE:
 <221> NAME/KEY: MOD_RES
 <222> LOCATION: (8)..(8)
 <223> OTHER INFORMATION: Any amino acid
 <220> FEATURE:
 <221> NAME/KEY: MOD_RES
 <222> LOCATION: (13)..(13)
 <223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 26

Pro Leu Ile Ala Ala Asp Pro Xaa Ser His Asn Val Xaa Ile Met
 1 5 10 15

<210> SEQ ID NO 27
 <211> LENGTH: 16
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
 oligopeptide
 <220> FEATURE:
 <221> NAME/KEY: MOD_RES
 <222> LOCATION: (8)..(9)
 <223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 27

Lys Leu Glu Tyr Asn Phe Asn Xaa Xaa Tyr Ala Phe Lys Tyr Glu Asn
 1 5 10 15

<210> SEQ ID NO 28
 <211> LENGTH: 15
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
 oligopeptide
 <220> FEATURE:
 <221> NAME/KEY: MOD_RES
 <222> LOCATION: (8)..(8)
 <223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 28

Lys Leu Glu Tyr Asn Phe Asn Xaa Tyr Ala Phe Lys Tyr Glu Asn
 1 5 10 15

<210> SEQ ID NO 29
 <211> LENGTH: 14
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic

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oligopeptide

<400> SEQUENCE: 29

Lys Leu Glu Tyr Asn Phe Asn Tyr Ala Phe Lys Tyr Glu Asn
1 5 10

<210> SEQ ID NO 30

<211> LENGTH: 14

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide

<220> FEATURE:

<221> NAME/KEY: MOD_RES

<222> LOCATION: (7)..(7)

<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 30

Lys Leu Glu Tyr Asn Phe Xaa Tyr Ala Phe Lys Tyr Glu Asn
1 5 10

<210> SEQ ID NO 31

<211> LENGTH: 14

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide

<220> FEATURE:

<221> NAME/KEY: MOD_RES

<222> LOCATION: (6)..(7)

<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 31

Lys Leu Glu Tyr Asn Xaa Xaa Tyr Ala Phe Lys Tyr Glu Asn
1 5 10

<210> SEQ ID NO 32

<211> LENGTH: 14

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide

<220> FEATURE:

<221> NAME/KEY: MOD_RES

<222> LOCATION: (7)..(7)

<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 32

Lys Leu Glu Tyr Asn Trp Xaa Tyr Ala Phe Lys Tyr Glu Asn
1 5 10

<210> SEQ ID NO 33

<211> LENGTH: 14

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide

<220> FEATURE:

<221> NAME/KEY: MOD_RES

<222> LOCATION: (6)..(6)

<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 33

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Lys Leu Glu Tyr Asn Xaa Lys Tyr Ala Phe Lys Tyr Glu Asn
1 5 10

<210> SEQ ID NO 34
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 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
 oligopeptide
 <220> FEATURE:
 <221> NAME/KEY: MOD_RES
 <222> LOCATION: (9)..(9)
 <223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 34

Lys Leu Glu Tyr Asn Phe Asn Pro Xaa Tyr Ala Phe Lys Tyr Glu Asn
1 5 10 15

<210> SEQ ID NO 35
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 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
 oligopeptide
 <220> FEATURE:
 <221> NAME/KEY: MOD_RES
 <222> LOCATION: (8)..(8)
 <223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 35

Lys Leu Glu Tyr Asn Phe Asn Xaa Pro Tyr Ala Phe Lys Tyr Glu Asn
1 5 10 15

<210> SEQ ID NO 36
 <211> LENGTH: 15
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
 oligopeptide
 <220> FEATURE:
 <221> NAME/KEY: MOD_RES
 <222> LOCATION: (7)..(8)
 <223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 36

Ala Phe Lys Tyr Glu Asn Xaa Xaa Ser His Asn Val Tyr Ile Met
1 5 10 15

<210> SEQ ID NO 37
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 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
 oligopeptide
 <220> FEATURE:
 <221> NAME/KEY: MOD_RES
 <222> LOCATION: (8)..(9)
 <223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 37

Lys Leu Glu Tyr Asn Phe Asn Xaa Xaa Lys Tyr Asp Ile Lys Asp Val
1 5 10 15

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<210> SEQ ID NO 38
<211> LENGTH: 15
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (7)..(8)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 38

Lys Ser Tyr Glu Glu Leu Xaa Xaa Ser His Asn Val Tyr Ile Met
1 5 10 15

<210> SEQ ID NO 39
<211> LENGTH: 15
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (8)..(8)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 39

Lys Ser Tyr Glu Glu Leu Pro Xaa Ser His Asn Val Tyr Ile Met
1 5 10 15

<210> SEQ ID NO 40
<211> LENGTH: 15
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (7)..(7)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 40

Lys Ser Tyr Glu Glu Leu Xaa Pro Ser His Asn Val Tyr Ile Met
1 5 10 15

<210> SEQ ID NO 41
<211> LENGTH: 16
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (8)..(9)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 41

Lys Leu Glu Tyr Asn Phe Asn Xaa Xaa Ala Lys Asp Pro Arg Ile Ala
1 5 10 15

<210> SEQ ID NO 42
<211> LENGTH: 16
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence

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<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (9)..(9)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 42

Lys Leu Glu Tyr Asn Phe Asn Pro Xaa Ala Lys Asp Pro Arg Ile Ala
1 5 10 15

<210> SEQ ID NO 43
<211> LENGTH: 16
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (8)..(8)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 43

Lys Leu Glu Tyr Asn Phe Asn Xaa Pro Ala Lys Asp Pro Arg Ile Ala
1 5 10 15

<210> SEQ ID NO 44
<211> LENGTH: 15
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (8)..(8)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 44

Glu Leu Ala Lys Asp Pro Arg Xaa Ser His Asn Val Tyr Ile Met
1 5 10 15

<210> SEQ ID NO 45
<211> LENGTH: 16
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (8)..(9)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 45

Glu Leu Ala Lys Asp Pro Arg Xaa Xaa Ser His Asn Val Tyr Ile Met
1 5 10 15

<210> SEQ ID NO 46
<211> LENGTH: 17
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:

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<221> NAME/KEY: MOD_RES
<222> LOCATION: (8)..(10)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 46

Glu Leu Ala Lys Asp Pro Arg Xaa Xaa Xaa Ser His Asn Val Tyr Ile
1 5 10 15

Met

<210> SEQ ID NO 47
<211> LENGTH: 15
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (8)..(8)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 47

Lys Leu Glu Tyr Asn Phe Asn Xaa Ala Ala Thr Met Glu Asn Ala
1 5 10 15

<210> SEQ ID NO 48
<211> LENGTH: 16
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (8)..(9)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 48

Lys Leu Glu Tyr Asn Phe Asn Xaa Xaa Ala Ala Thr Met Glu Asn Ala
1 5 10 15

<210> SEQ ID NO 49
<211> LENGTH: 17
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (8)..(10)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 49

Lys Leu Glu Tyr Asn Phe Asn Xaa Xaa Xaa Ala Ala Thr Met Glu Asn
1 5 10 15

Ala

<210> SEQ ID NO 50
<211> LENGTH: 671
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
recombinant peptide biosensor

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<400> SEQUENCE: 50

Met	Arg	Gly	Ser	His	His	His	His	His	His	Gly	Met	Ala	Ser	Met	Thr
1				5						10				15	
Gly	Gly	Gln	Gln	Met	Gly	Arg	Asp	Leu	Tyr	Asp	Asp	Asp	Asp	Lys	Asp
			20					25						30	
Arg	Trp	Gly	Ser	Lys	Ile	Glu	Glu	Gly	Lys	Val	Val	Ile	Trp	His	Ala
		35						40					45		
Met	Gln	Pro	Asn	Glu	Leu	Glu	Val	Phe	Gln	Ser	Leu	Ala	Glu	Glu	Tyr
	50					55						60			
Met	Ala	Leu	Cys	Pro	Glu	Val	Glu	Ile	Val	Phe	Glu	Gln	Lys	Pro	Asn
	65					70				75					80
Leu	Glu	Asp	Ala	Leu	Lys	Ala	Ala	Ile	Pro	Thr	Gly	Gln	Gly	Pro	Asp
				85					90					95	
Leu	Phe	Ile	Trp	Ala	His	Asp	Trp	Ile	Gly	Lys	Phe	Ala	Glu	Ala	Gly
			100					105						110	
Leu	Leu	Glu	Pro	Ile	Asp	Glu	Tyr	Val	Thr	Glu	Asp	Leu	Leu	Asn	Glu
		115						120					125		
Phe	Ala	Pro	Met	Ala	Gln	Asp	Ala	Met	Gln	Tyr	Lys	Gly	His	Tyr	Tyr
	130					135						140			
Ala	Leu	Pro	Phe	Ala	Ala	Glu	Thr	Val	Ala	Ile	Ile	Tyr	Ser	Lys	Glu
	145				150					155					160
Met	Val	Ser	Glu	Pro	Pro	Lys	Thr	Phe	Asp	Glu	Met	Lys	Ala	Ile	Met
				165					170					175	
Glu	Lys	Tyr	Tyr	Asp	Pro	Ala	Asn	Glu	Lys	Tyr	Gly	Ile	Ala	Trp	Pro
			180					185						190	
Ile	Asn	Ala	Tyr	Phe	Ile	Ser	Ala	Ile	Ala	Gln	Ala	Phe	Gly	Gly	Ser
	195						200						205		
His	Asn	Val	Tyr	Ile	Met	Ala	Asp	Lys	Gln	Lys	Asn	Gly	Ile	Lys	Ala
	210					215						220			
Asn	Phe	Lys	Ile	Arg	His	Asn	Ile	Glu	Asp	Gly	Gly	Val	Gln	Leu	Ala
	225				230					235					240
Tyr	His	Tyr	Gln	Gln	Asn	Thr	Pro	Ile	Gly	Asp	Gly	Pro	Val	Leu	Leu
				245					250					255	
Pro	Asp	Asn	His	Tyr	Leu	Ser	Thr	Gln	Ser	Lys	Leu	Ser	Lys	Asp	Pro
			260					265						270	
Asn	Glu	Lys	Arg	Asp	His	Met	Val	Leu	Leu	Glu	Phe	Val	Thr	Ala	Ala
		275					280						285		
Gly	Ile	Thr	Leu	Gly	Met	Asp	Glu	Leu	Tyr	Lys	Gly	Gly	Thr	Gly	Gly
	290					295					300				
Ser	Met	Val	Ser	Lys	Gly	Glu	Glu	Leu	Phe	Thr	Gly	Val	Val	Pro	Ile
	305				310						315				320
Leu	Val	Glu	Leu	Asp	Gly	Asp	Val	Asn	Gly	His	Lys	Phe	Ser	Val	Ser
				325					330					335	
Gly	Glu	Gly	Glu	Gly	Asp	Ala	Thr	Tyr	Gly	Lys	Leu	Thr	Leu	Lys	Phe
			340					345						350	
Ile	Cys	Thr	Thr	Gly	Lys	Leu	Pro	Val	Pro	Trp	Pro	Thr	Leu	Val	Thr
		355					360						365		
Thr	Leu	Thr	Tyr	Gly	Val	Gln	Cys	Phe	Ser	Arg	Tyr	Pro	Asp	His	Met
	370					375						380			
Lys	Gln	His	Asp	Phe	Phe	Lys	Ser	Ala	Met	Pro	Glu	Gly	Tyr	Ile	Gln
	385				390						395				400

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Glu Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala
405 410 415

Glu Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys
420 425 430

Gly Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu
435 440 445

Tyr Asn Phe Asn Gly Gly Tyr Tyr Phe Asp Asp Lys Thr Glu Gln Pro
450 455 460

Gly Leu Asp Lys Pro Glu Thr Ile Glu Gly Phe Lys Phe Phe Phe Thr
465 470 475 480

Glu Ile Trp Pro Tyr Met Ala Pro Thr Gly Asp Tyr Asn Thr Gln Gln
485 490 495

Ser Ile Phe Leu Glu Gly Arg Ala Pro Met Met Val Asn Gly Pro Trp
500 505 510

Ser Ile Asn Asp Val Lys Lys Ala Gly Ile Asn Phe Gly Val Val Pro
515 520 525

Leu Pro Pro Ile Ile Lys Asp Gly Lys Glu Tyr Trp Pro Arg Pro Tyr
530 535 540

Gly Gly Val Lys Leu Ile Tyr Phe Ala Ala Gly Ile Lys Asn Lys Asp
545 550 555 560

Ala Ala Trp Lys Phe Ala Lys Trp Leu Thr Thr Ser Glu Glu Ser Ile
565 570 575

Lys Thr Leu Ala Leu Glu Leu Gly Tyr Ile Pro Val Leu Thr Lys Val
580 585 590

Leu Asp Asp Pro Glu Ile Lys Asn Asp Pro Val Ile Tyr Gly Phe Gly
595 600 605

Gln Ala Val Gln His Ala Tyr Leu Met Pro Lys Ser Pro Lys Met Ser
610 615 620

Ala Val Trp Gly Gly Val Asp Gly Ala Ile Asn Glu Ile Leu Gln Asp
625 630 635 640

Pro Gln Asn Ala Asp Ile Glu Gly Ile Leu Lys Lys Tyr Gln Gln Glu
645 650 655

Ile Leu Asn Asn Met Gln Gly Ser His His His His His His Gly
660 665 670

<210> SEQ ID NO 51

<211> LENGTH: 671

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic recombinant peptide biosensor

<400> SEQUENCE: 51

Met Arg Gly Ser His His His His His Gly Met Ala Ser Met Thr
1 5 10 15

Gly Gly Gln Gln Met Gly Arg Asp Leu Tyr Asp Asp Asp Lys Asp
20 25 30

Arg Trp Gly Ser Lys Ile Glu Glu Gly Lys Val Val Ile Trp His Ala
35 40 45

Met Gln Pro Asn Glu Leu Glu Val Phe Gln Ser Leu Ala Glu Glu Tyr
50 55 60

Met Ala Leu Cys Pro Glu Val Glu Ile Val Phe Glu Gln Lys Pro Asn

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65	70	75	80
Leu Glu Asp Ala Leu Lys Ala Ala Ile Pro Thr Gly Gln Gly Pro Asp 85 90 95			
Leu Phe Ile Trp Ala His Asp Trp Ile Gly Lys Phe Ala Glu Ala Gly 100 105 110			
Leu Leu Glu Pro Ile Asp Glu Tyr Val Thr Glu Asp Leu Leu Asn Glu 115 120 125			
Phe Ala Pro Met Ala Gln Asp Ala Met Gln Tyr Lys Gly His Tyr Tyr 130 135 140			
Ala Leu Pro Phe Ala Ala Glu Thr Val Ala Ile Ile Tyr Ser Lys Glu 145 150 155 160			
Met Val Ser Glu Pro Pro Lys Thr Phe Asp Glu Met Lys Ala Ile Met 165 170 175			
Glu Lys Tyr Tyr Asp Pro Ala Asn Glu Lys Tyr Gly Ile Ala Trp Pro 180 185 190			
Ile Asn Ala Tyr Phe Ile Ser Ala Ile Ala Gln Ala Phe Gly Gly Ser 195 200 205			
His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn Gly Ile Lys Ala 210 215 220			
Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Gly Val Gln Leu Ala 225 230 235 240			
Tyr His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu Leu 245 250 255			
Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Lys Leu Ser Lys Asp Pro 260 265 270			
Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val Thr Ala Ala 275 280 285			
Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Gly Gly Thr Gly Gly 290 295 300			
Ser Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile 305 310 315 320			
Leu Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser 325 330 335			
Gly Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe 340 345 350			
Ile Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr 355 360 365			
Thr Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met 370 375 380			
Lys Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Ile Gln 385 390 395 400			
Glu Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala 405 410 415			
Glu Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys 420 425 430			
Gly Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu 435 440 445			
Tyr Asn Phe Asn Phe Glu Tyr Tyr Phe Asp Asp Lys Thr Glu Gln Pro 450 455 460			
Gly Leu Asp Lys Pro Glu Thr Ile Glu Gly Phe Lys Phe Phe Phe Thr 465 470 475 480			

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Glu Ile Trp Pro Tyr Met Ala Pro Thr Gly Asp Tyr Asn Thr Gln Gln
485 490 495
Ser Ile Phe Leu Glu Gly Arg Ala Pro Met Met Val Asn Gly Pro Trp
500 505 510
Ser Ile Asn Asp Val Lys Lys Ala Gly Ile Asn Phe Gly Val Val Pro
515 520 525
Leu Pro Pro Ile Ile Lys Asp Gly Lys Glu Tyr Trp Pro Arg Pro Tyr
530 535 540
Gly Gly Val Lys Leu Ile Tyr Phe Ala Ala Gly Ile Lys Asn Lys Asp
545 550 555 560
Ala Ala Trp Lys Phe Ala Lys Trp Leu Thr Thr Ser Glu Glu Ser Ile
565 570 575
Lys Thr Leu Ala Leu Glu Leu Gly Tyr Ile Pro Val Leu Thr Lys Val
580 585 590
Leu Asp Asp Pro Glu Ile Lys Asn Asp Pro Val Ile Tyr Gly Phe Gly
595 600 605
Gln Ala Val Gln His Ala Tyr Leu Met Pro Lys Ser Pro Lys Met Ser
610 615 620
Ala Val Trp Gly Gly Val Asp Gly Ala Ile Asn Glu Ile Leu Gln Asp
625 630 635 640
Pro Gln Asn Ala Asp Ile Glu Gly Ile Leu Lys Lys Tyr Gln Gln Glu
645 650 655
Ile Leu Asn Asn Met Gln Gly Ser His His His His His His Gly
660 665 670

<210> SEQ ID NO 52
<211> LENGTH: 662
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
recombinant peptide biosensor

<400> SEQUENCE: 52

Met Arg Gly Ser His His His His His His Gly Met Ala Ser Met Thr
1 5 10 15
Gly Gly Gln Gln Met Gly Arg Asp Leu Tyr Asp Asp Asp Asp Lys Asp
20 25 30
Arg Trp Gly Ser Lys Ile Glu Glu Gly Lys Val Val Ile Trp His Ala
35 40 45
Met Gln Pro Asn Glu Leu Glu Val Phe Gln Ser Leu Ala Glu Glu Tyr
50 55 60
Met Ala Leu Cys Pro Glu Val Glu Ile Val Phe Glu Gln Lys Pro Asn
65 70 75 80
Leu Glu Asp Ala Leu Lys Ala Ala Ile Pro Thr Gly Gln Gly Pro Asp
85 90 95
Leu Phe Ile Trp Ala His Asp Trp Ile Gly Lys Phe Ala Glu Ala Gly
100 105 110
Leu Leu Glu Pro Ile Asp Glu Tyr Val Thr Glu Asp Leu Leu Asn Glu
115 120 125
Phe Ala Pro Met Ala Gln Asp Ala Met Gln Tyr Lys Gly His Tyr Tyr
130 135 140
Ala Leu Pro Phe Ala Ala Glu Thr Val Ala Ile Ile Tyr Ser Lys Glu

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145					150						155					160
Met	Val	Ser	Glu	Pro	Pro	Lys	Thr	Phe	Asp	Glu	Met	Lys	Ala	Ile	Met	
				165					170						175	
Glu	Lys	Tyr	Tyr	Asp	Pro	Ala	Asn	Glu	Lys	Tyr	Gly	Ile	Ala	Trp	Pro	
			180					185						190		
Ile	Asn	Ala	Tyr	Phe	Ile	Ser	Ala	Ile	Ala	Gln	Ala	Phe	Gly	Gly	Tyr	
		195					200					205				
Tyr	Phe	Asp	Asp	Lys	Thr	Glu	Gln	Pro	Gly	Leu	Asp	Lys	Pro	Glu	Thr	
	210					215					220					
Ile	Glu	Gly	Phe	Lys	Phe	Phe	Phe	Thr	Glu	Ile	Trp	Pro	Tyr	Met	Ala	
	225				230						235				240	
Pro	Thr	Gly	Asp	Tyr	Asn	Thr	Gln	Gln	Ser	Ile	Phe	Leu	Glu	Gly	Arg	
				245					250						255	
Ala	Pro	Met	Met	Val	Asn	Gly	Pro	Trp	Ser	Ile	Asn	Asp	Val	Lys	Lys	
			260					265						270		
Ala	Gly	Ile	Asn	Phe	Gly	Val	Val	Pro	Leu	Pro	Pro	Ile	Ile	Lys	Asp	
		275					280							285		
Gly	Lys	Glu	Tyr	Trp	Pro	Arg	Pro	Tyr	Gly	Gly	Val	Lys	Leu	Ile	Tyr	
	290					295					300					
Phe	Ala	Ala	Gly	Ile	Lys	Asn	Lys	Asp	Ala	Ala	Trp	Lys	Phe	Ala	Lys	
	305				310					315					320	
Trp	Leu	Thr	Thr	Ser	Glu	Glu	Ser	Ile	Lys	Thr	Leu	Ala	Leu	Glu	Leu	
				325					330						335	
Gly	Tyr	Ile	Pro	Val	Leu	Thr	Lys	Val	Leu	Asp	Asp	Pro	Glu	Ile	Ser	
			340						345					350		
His	Asn	Val	Tyr	Ile	Met	Ala	Asp	Lys	Gln	Lys	Asn	Gly	Ile	Lys	Ala	
		355					360							365		
Asn	Phe	Lys	Ile	Arg	His	Asn	Ile	Glu	Asp	Gly	Gly	Val	Gln	Leu	Ala	
	370					375					380					
Tyr	His	Tyr	Gln	Gln	Asn	Thr	Pro	Ile	Gly	Asp	Gly	Pro	Val	Leu	Leu	
	385				390					395					400	
Pro	Asp	Asn	His	Tyr	Leu	Ser	Thr	Gln	Ser	Lys	Leu	Ser	Lys	Asp	Pro	
				405						410					415	
Asn	Glu	Lys	Arg	Asp	His	Met	Val	Leu	Leu	Glu	Phe	Val	Thr	Ala	Ala	
			420					425						430		
Gly	Ile	Thr	Leu	Gly	Met	Asp	Glu	Leu	Tyr	Lys	Gly	Gly	Thr	Gly	Gly	
		435					440							445		
Ser	Met	Val	Ser	Lys	Gly	Glu	Glu	Leu	Phe	Thr	Gly	Val	Val	Pro	Ile	
	450					455					460					
Leu	Val	Glu	Leu	Asp	Gly	Asp	Val	Asn	Gly	His	Lys	Phe	Ser	Val	Ser	
	465				470					475					480	
Gly	Glu	Gly	Glu	Gly	Asp	Ala	Thr	Tyr	Gly	Lys	Leu	Thr	Leu	Lys	Phe	
				485					490						495	
Ile	Cys	Thr	Thr	Gly	Lys	Leu	Pro	Val	Pro	Trp	Pro	Thr	Leu	Val	Thr	
			500						505					510		
Thr	Leu	Thr	Tyr	Gly	Val	Gln	Cys	Phe	Ser	Arg	Tyr	Pro	Asp	His	Met	
			515					520					525			
Lys	Gln	His	Asp	Phe	Phe	Lys	Ser	Ala	Met	Pro	Glu	Gly	Tyr	Ile	Gln	
	530					535							540			
Glu	Arg	Thr	Ile	Phe	Phe	Lys	Asp	Asp	Gly	Asn	Tyr	Lys	Thr	Arg	Ala	
	545					550				555					560	

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Glu Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys
565 570 575

Gly Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu
580 585 590

Tyr Asn Phe Asn Lys Asn Asp Pro Val Ile Tyr Gly Phe Gly Gln Ala
595 600 605

Val Gln His Ala Tyr Leu Met Pro Lys Ser Pro Lys Met Ser Ala Val
610 615 620

Trp Gly Gly Val Asp Gly Ala Ile Asn Glu Ile Leu Gln Asp Pro Gln
625 630 635 640

Asn Ala Asp Ile Glu Gly Ile Leu Lys Lys Tyr Gln Gln Glu Ile Leu
645 650 655

Asn Asn Met Gln Gly Ser
660

<210> SEQ ID NO 53

<211> LENGTH: 664

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic recombinant peptide biosensor

<400> SEQUENCE: 53

Met Arg Gly Ser His His His His His Gly Met Ala Ser Met Thr
1 5 10 15

Gly Gly Gln Gln Met Gly Arg Asp Leu Tyr Asp Asp Asp Lys Asp
20 25 30

Arg Trp Gly Ser Lys Ile Glu Glu Gly Lys Val Val Ile Trp His Ala
35 40 45

Met Gln Pro Asn Glu Leu Glu Val Phe Gln Ser Leu Ala Glu Glu Tyr
50 55 60

Met Ala Leu Cys Pro Glu Val Glu Ile Val Phe Glu Gln Lys Pro Asn
65 70 75 80

Leu Glu Asp Ala Leu Lys Ala Ala Ile Pro Thr Gly Gln Gly Pro Asp
85 90 95

Leu Phe Ile Trp Ala His Asp Trp Ile Gly Lys Phe Ala Glu Ala Gly
100 105 110

Leu Leu Glu Pro Ile Asp Glu Tyr Val Thr Glu Asp Leu Leu Asn Glu
115 120 125

Phe Ala Pro Met Ala Gln Asp Ala Met Gln Tyr Lys Gly His Tyr Tyr
130 135 140

Ala Leu Pro Phe Ala Ala Glu Thr Val Ala Ile Ile Tyr Ser Lys Glu
145 150 155 160

Met Val Ser Glu Pro Pro Lys Thr Phe Asp Glu Met Lys Ala Ile Met
165 170 175

Glu Lys Tyr Tyr Asp Pro Ala Asn Glu Lys Tyr Gly Ile Ala Trp Pro
180 185 190

Ile Asn Ala Tyr Phe Ile Ser Ala Ile Ala Gln Ala Phe Gly Gly Tyr
195 200 205

Tyr Phe Asp Asp Lys Thr Glu Gln Pro Gly Leu Asp Lys Pro Glu Thr
210 215 220

Ile Glu Gly Phe Lys Phe Phe Phe Thr Glu Ile Trp Pro Tyr Met Ala

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225	230					235					240				
Pro Thr Gly Asp Tyr	Asn Thr Gln Gln Ser Ile Phe Leu Glu Gly Arg	245						250	255						
Ala Pro Met Met Val Asn Gly Pro Trp Ser Ile Asn Asp Val Lys Lys	260					265					270				
Ala Gly Ile Asn Phe Gly Val Val Pro Leu Pro Pro Ile Ile Lys Asp	275					280					285				
Gly Lys Glu Tyr Trp Pro Arg Pro Tyr Gly Gly Val Lys Leu Ile Tyr	290					295					300				
Phe Ala Ala Gly Ile Lys Asn Lys Asp Ala Ala Trp Lys Phe Ala Lys	305					310					315				
Trp Leu Thr Thr Ser Glu Glu Ser Ile Lys Thr Leu Ala Leu Glu Leu	325					330					335				
Gly Tyr Ile Pro Val Leu Thr Lys Val Leu Asp Asp Pro Glu Ile Pro	340					345					350				
Pro Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn Gly Ile	355					360					365				
Lys Ala Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Gly Val Gln	370					375					380				
Leu Ala Tyr His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val	385					390					395				
Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Lys Leu Ser Lys	405					410					415				
Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val Thr	420					425					430				
Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Gly Gly Thr	435					440					445				
Gly Gly Ser Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val	450					455					460				
Pro Ile Leu Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser	465					470					475				
Val Ser Gly Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu	485					490					495				
Lys Phe Ile Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu	500					505					510				
Val Thr Thr Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp	515					520					525				
His Met Lys Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr	530					535					540				
Ile Gln Glu Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr	545					550					555				
Arg Ala Glu Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu	565					570					575				
Leu Lys Gly Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys	580					585					590				
Leu Glu Tyr Asn Phe Asn Lys Asn Asp Pro Val Ile Tyr Gly Phe Gly	595					600					605				
Gln Ala Val Gln His Ala Tyr Leu Met Pro Lys Ser Pro Lys Met Ser	610					615					620				
Ala Val Trp Gly Gly Val Asp Gly Ala Ile Asn Glu Ile Leu Gln Asp	625					630					635				
										640					

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Pro Gln Asn Ala Asp Ile Glu Gly Ile Leu Lys Lys Tyr Gln Gln Glu
 645 650 655

Ile Leu Asn Asn Met Gln Gly Ser
 660

<210> SEQ ID NO 54
 <211> LENGTH: 16
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
 oligopeptide
 <220> FEATURE:
 <221> NAME/KEY: MOD_RES
 <222> LOCATION: (7)..(8)
 <223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 54

Ala Ile Ala Gln Ala Phe Xaa Xaa Ser His Asn Val Tyr Ile Met Ala
 1 5 10 15

<210> SEQ ID NO 55
 <211> LENGTH: 16
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
 oligopeptide
 <220> FEATURE:
 <221> NAME/KEY: MOD_RES
 <222> LOCATION: (8)..(8)
 <223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 55

Ala Ile Ala Gln Ala Phe Pro Xaa Ser His Asn Val Tyr Ile Met Ala
 1 5 10 15

<210> SEQ ID NO 56
 <211> LENGTH: 17
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
 oligopeptide
 <220> FEATURE:
 <221> NAME/KEY: MOD_RES
 <222> LOCATION: (8)..(9)
 <223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 56

Lys Leu Glu Tyr Asn Phe Asn Xaa Xaa Tyr Tyr Phe Asp Asp Lys Thr
 1 5 10 15

Glu

<210> SEQ ID NO 57
 <211> LENGTH: 14
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
 oligopeptide
 <220> FEATURE:
 <221> NAME/KEY: MOD_RES
 <222> LOCATION: (7)..(8)
 <223> OTHER INFORMATION: Any amino acid

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<400> SEQUENCE: 57

Val Leu Asp Asp Pro Glu Xaa Xaa His Asn Val Tyr Ile Met
1 5 10

<210> SEQ ID NO 58

<211> LENGTH: 16

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide

<220> FEATURE:

<221> NAME/KEY: MOD_RES

<222> LOCATION: (8)..(9)

<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 58

Val Leu Asp Asp Pro Glu Ile Xaa Xaa Ser His Asn Val Tyr Ile Met
1 5 10 15

<210> SEQ ID NO 59

<211> LENGTH: 14

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide

<220> FEATURE:

<221> NAME/KEY: MOD_RES

<222> LOCATION: (7)..(8)

<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 59

Lys Leu Glu Tyr Asn Phe Xaa Xaa Asn Asp Pro Val Ile Tyr
1 5 10

<210> SEQ ID NO 60

<211> LENGTH: 16

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide

<220> FEATURE:

<221> NAME/KEY: MOD_RES

<222> LOCATION: (8)..(8)

<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 60

Lys Leu Glu Tyr Asn Phe Asn Xaa Pro Lys Asn Asp Pro Val Ile Tyr
1 5 10 15

<210> SEQ ID NO 61

<211> LENGTH: 16

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide

<220> FEATURE:

<221> NAME/KEY: MOD_RES

<222> LOCATION: (9)..(9)

<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 61

Lys Leu Glu Tyr Asn Phe Asn Pro Xaa Lys Asn Asp Pro Val Ile Tyr
1 5 10 15

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<210> SEQ ID NO 62
 <211> LENGTH: 556
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
 recombinant peptide biosensor

<400> SEQUENCE: 62

Met Arg Gly Ser His His His His His Gly Met Ala Ser Met Thr
 1 5 10 15
 Gly Gly Gln Gln Met Gly Arg Asp Leu Tyr Asp Asp Asp Lys Asp
 20 25 30
 Arg Trp Gly Ser Ala Ala Gly Ser Thr Leu Asp Lys Ile Ala Lys Asn
 35 40 45
 Gly Val Ile Val Val Gly His Arg Glu Ser Ser Val Pro Phe Ser Tyr
 50 55 60
 Tyr Asp Asn Gln Gln Lys Val Val Gly Tyr Ser Gln Asp Tyr Ser Asn
 65 70 75 80
 Ala Ile Val Glu Ala Val Lys Lys Lys Leu Asn Lys Pro Asp Leu Gln
 85 90 95
 Val Lys Leu Ile Pro Ile Thr Ser Gln Asn Arg Ile Pro Leu Leu Gln
 100 105 110
 Asn Gly Thr Phe Asp Phe Glu Cys Gly Ser Thr Thr Asn Asn Val Glu
 115 120 125
 Arg Gln Lys Gln Ala Ala Phe Ser Asp Thr Ile Phe Val Val Gly Thr
 130 135 140
 Arg Leu Leu Thr Lys Lys Gly Gly Asp Ile Lys Asp Phe Ala Asn Leu
 145 150 155 160
 Lys Asp Lys Ala Val Val Val Thr Ser Gly Thr Thr Ser Glu Val Leu
 165 170 175
 Leu Asn Lys Leu Asn Glu Glu Gln Lys Met Asn Met Arg Ile Ile Ser
 180 185 190
 Ala Lys Asp His Gly Asp Ser Phe Arg Thr Leu Glu Ser Gly Arg Ala
 195 200 205
 Val Ala Phe Met Met Asp Asp Val Leu Leu Ala Gly Glu Arg Ala Lys
 210 215 220
 Ala Lys Lys Pro Asp Asn Trp Glu Ile Val Gly Lys Pro Gln Ser Gln
 225 230 235 240
 Glu Ala Tyr Gly Cys Met Leu Arg Lys Asp Asp Pro Gln Phe Lys Lys
 245 250 255
 Leu Met Asp Asp Thr Ile Ala Gln Val Gln Thr Ser Gly Glu Ala Glu
 260 265 270
 Lys Trp Phe Asp Lys Trp Phe Lys Asn Pro Ile Leu Val Ser His Asn
 275 280 285
 Val Tyr Ile Met Ala Asp Lys Gln Lys Asn Gly Ile Lys Ala Asn Phe
 290 295 300
 Lys Ile Arg His Asn Ile Glu Asp Gly Gly Val Gln Leu Ala Tyr His
 305 310 315 320
 Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro Asp
 325 330 335
 Asn His Tyr Leu Ser Thr Gln Ser Lys Leu Ser Lys Asp Pro Asn Glu

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Arg	Gln	Lys	Gln	Ala	Ala	Phe	Ser	Asp	Thr	Ile	Phe	Val	Val	Gly	Thr	130	135	140	
Arg	Leu	Leu	Thr	Lys	Lys	Gly	Gly	Asp	Ile	Lys	Asp	Phe	Ala	Asn	Leu	145	150	155	160
Lys	Asp	Lys	Ala	Val	Val	Val	Thr	Ser	Gly	Thr	Thr	Ser	Glu	Val	Leu	165	170	175	
Leu	Asn	Lys	Leu	Asn	Glu	Glu	Gln	Lys	Met	Asn	Met	Arg	Ile	Ile	Ser	180	185	190	
Ala	Lys	Asp	His	Gly	Asp	Ser	Phe	Arg	Thr	Leu	Glu	Ser	Gly	Arg	Ala	195	200	205	
Val	Ala	Phe	Met	Met	Asp	Asp	Val	Leu	Leu	Ala	Gly	Glu	Arg	Ala	Lys	210	215	220	
Ala	Lys	Lys	Pro	Asp	Asn	Trp	Glu	Ile	Val	Gly	Lys	Pro	Gln	Ser	Gln	225	230	235	240
Glu	Ala	Tyr	Gly	Cys	Met	Leu	Arg	Lys	Asp	Asp	Pro	Gln	Phe	Lys	Lys	245	250	255	
Leu	Met	Asp	Asp	Thr	Ile	Ala	Gln	Val	Gln	Thr	Ser	Gly	Glu	Ala	Glu	260	265	270	
Lys	Trp	Phe	Asp	Lys	Trp	Phe	Lys	Asn	Pro	Ile	Leu	Val	Ser	His	Asn	275	280	285	
Val	Tyr	Ile	Met	Ala	Asp	Lys	Gln	Lys	Asn	Gly	Ile	Lys	Ala	Asn	Phe	290	295	300	
Lys	Ile	Arg	His	Asn	Ile	Glu	Asp	Gly	Gly	Val	Gln	Leu	Ala	Tyr	His	305	310	315	320
Tyr	Gln	Gln	Asn	Thr	Pro	Ile	Gly	Asp	Gly	Pro	Val	Leu	Leu	Pro	Asp	325	330	335	
Asn	His	Tyr	Leu	Ser	Thr	Gln	Ser	Lys	Leu	Ser	Lys	Asp	Pro	Asn	Glu	340	345	350	
Lys	Arg	Asp	His	Met	Val	Leu	Leu	Glu	Phe	Val	Thr	Ala	Ala	Gly	Ile	355	360	365	
Thr	Leu	Gly	Met	Asp	Glu	Leu	Tyr	Lys	Gly	Gly	Thr	Gly	Gly	Ser	Met	370	375	380	
Val	Ser	Lys	Gly	Glu	Glu	Leu	Phe	Thr	Gly	Val	Val	Pro	Ile	Leu	Val	385	390	395	400
Glu	Leu	Asp	Gly	Asp	Val	Asn	Gly	His	Lys	Phe	Ser	Val	Ser	Gly	Glu	405	410	415	
Gly	Glu	Gly	Asp	Ala	Thr	Tyr	Gly	Lys	Leu	Thr	Leu	Lys	Phe	Ile	Cys	420	425	430	
Thr	Thr	Gly	Lys	Leu	Pro	Val	Pro	Trp	Pro	Thr	Leu	Val	Thr	Thr	Leu	435	440	445	
Thr	Tyr	Gly	Val	Gln	Cys	Phe	Ser	Arg	Tyr	Pro	Asp	His	Met	Lys	Gln	450	455	460	
His	Asp	Phe	Phe	Lys	Ser	Ala	Met	Pro	Glu	Gly	Tyr	Ile	Gln	Glu	Arg	465	470	475	480
Thr	Ile	Phe	Phe	Lys	Asp	Asp	Gly	Asn	Tyr	Lys	Thr	Arg	Ala	Glu	Val	485	490	495	
Lys	Phe	Glu	Gly	Asp	Thr	Leu	Val	Asn	Arg	Ile	Glu	Leu	Lys	Gly	Ile	500	505	510	
Asp	Phe	Lys	Glu	Asp	Gly	Asn	Ile	Leu	Gly	His	Lys	Leu	Glu	Tyr	Asn	515	520	525	
Phe	Asn	Asn	Pro	Leu	Asn	Met	Asn	Phe	Glu	Leu	Ser	Asp	Glu	Met	Lys				

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530          535          540
Ala Leu Phe Lys Glu Pro Asn Asp Lys Ala Leu Lys
545          550          555

<210> SEQ ID NO 64
<211> LENGTH: 16
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
recombinant peptide biosensor
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (8)..(8)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 64

Phe Lys Asn Pro Ile Pro Pro Xaa Ser His Asn Val Tyr Ile Met Ala
1         5         10        15

<210> SEQ ID NO 65
<211> LENGTH: 17
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
recombinant peptide biosensor
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (8)..(9)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 65

Phe Lys Asn Pro Ile Pro Pro Xaa Xaa Ser His Asn Val Tyr Ile Met
1         5         10        15

Ala

<210> SEQ ID NO 66
<211> LENGTH: 17
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
recombinant peptide biosensor
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (9)..(9)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 66

Phe Lys Asn Pro Ile Pro Pro Pro Xaa Ser His Asn Val Tyr Ile Met
1         5         10        15

Ala

<210> SEQ ID NO 67
<211> LENGTH: 17
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
recombinant peptide biosensor
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (8)..(8)
<223> OTHER INFORMATION: Any amino acid

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<400> SEQUENCE: 67

Phe Lys Asn Pro Ile Pro Pro Xaa Pro Ser His Asn Val Tyr Ile Met
1 5 10 15

Ala

<210> SEQ ID NO 68

<211> LENGTH: 17

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
recombinant peptide biosensor

<220> FEATURE:

<221> NAME/KEY: MOD_RES

<222> LOCATION: (8)..(9)

<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 68

Lys Trp Phe Lys Asn Pro Ile Xaa Xaa Ser His Asn Val Tyr Ile Met
1 5 10 15

Ala

<210> SEQ ID NO 69

<211> LENGTH: 16

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
recombinant peptide biosensor

<220> FEATURE:

<221> NAME/KEY: MOD_RES

<222> LOCATION: (8)..(9)

<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 69

Phe Lys Asn Pro Ile Pro Pro Xaa Xaa Asn Val Tyr Ile Met Ala Asp
1 5 10 15

<210> SEQ ID NO 70

<211> LENGTH: 16

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
recombinant peptide biosensor

<220> FEATURE:

<221> NAME/KEY: MOD_RES

<222> LOCATION: (8)..(9)

<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 70

Lys Trp Phe Lys Asn Pro Ile Xaa Xaa Asn Val Tyr Ile Met Ala Asp
1 5 10 15

<210> SEQ ID NO 71

<211> LENGTH: 15

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
recombinant peptide biosensor

<220> FEATURE:

<221> NAME/KEY: MOD_RES

<222> LOCATION: (8)..(8)

<223> OTHER INFORMATION: Any amino acid

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<400> SEQUENCE: 71

Lys Leu Glu Tyr Asn Phe Asn Xaa Lys Asn Leu Asn Met Asn Phe
1 5 10 15

<210> SEQ ID NO 72

<211> LENGTH: 16

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
recombinant peptide biosensor

<220> FEATURE:

<221> NAME/KEY: MOD_RES

<222> LOCATION: (8)..(9)

<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 72

Lys Leu Glu Tyr Asn Phe Asn Xaa Xaa Lys Asn Leu Asn Met Asn Phe
1 5 10 15

<210> SEQ ID NO 73

<211> LENGTH: 16

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
recombinant peptide biosensor

<220> FEATURE:

<221> NAME/KEY: MOD_RES

<222> LOCATION: (8)..(8)

<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 73

Lys Leu Glu Tyr Asn Phe Asn Xaa Pro Lys Asn Leu Asn Met Asn Phe
1 5 10 15

<210> SEQ ID NO 74

<211> LENGTH: 16

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
recombinant peptide biosensor

<220> FEATURE:

<221> NAME/KEY: MOD_RES

<222> LOCATION: (9)..(9)

<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 74

Lys Leu Glu Tyr Asn Phe Asn Pro Xaa Lys Asn Leu Asn Met Asn Phe
1 5 10 15

<210> SEQ ID NO 75

<211> LENGTH: 14

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
recombinant peptide biosensor

<220> FEATURE:

<221> NAME/KEY: MOD_RES

<222> LOCATION: (8)..(9)

<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 75

Gly His Lys Leu Glu Tyr Asn Xaa Xaa Leu Asn Met Asn Phe
1 5 10

-continued

<210> SEQ ID NO 76
 <211> LENGTH: 14
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
 recombinant peptide biosensor
 <220> FEATURE:
 <221> NAME/KEY: MOD_RES
 <222> LOCATION: (8)..(9)
 <223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 76

Lys Leu Glu Tyr Asn Phe Asn Xaa Xaa Leu Asn Met Asn Phe
 1 5 10

<210> SEQ ID NO 77
 <211> LENGTH: 567
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
 recombinant peptide biosensor

<400> SEQUENCE: 77

Met His His His His His His Gly Ser Glu Glu Gln Glu Lys Ala Leu
 1 5 10 15

Asn Phe Gly Ile Ile Ser Thr Glu Ser Gln Gln Asn Leu Lys Pro Gln
 20 25 30

Trp Thr Pro Phe Leu Gln Asp Met Glu Lys Lys Leu Gly Val Lys Val
 35 40 45

Asn Ala Phe Phe Ala Pro Asp Tyr Ala Gly Ile Ile Gln Gly Met Arg
 50 55 60

Phe Asn Lys Val Asp Ile Ala Trp Tyr Gly Asn Leu Ser Ala Met Glu
 65 70 75 80

Ala Val Asp Arg Ala Asn Gly Gln Val Phe Ala Gln Thr Val Ala Ala
 85 90 95

Asp Gly Ser Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Arg Asn
 100 105 110

Gly Ile Lys Ala Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Gly
 115 120 125

Val Gln Leu Ala Tyr His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly
 130 135 140

Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Lys Leu
 145 150 155 160

Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe
 165 170 175

Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Gly
 180 185 190

Gly Thr Gly Gly Ser Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly
 195 200 205

Val Val Pro Ile Leu Val Glu Leu Asp Gly Asp Val Asn Gly His Lys
 210 215 220

Phe Ser Val Ser Gly Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu
 225 230 235 240

Thr Leu Lys Phe Ile Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro

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		245						250						255					
Thr	Leu	Val	Thr	Thr	Leu	Thr	Tyr	Gly	Val	Gln	Cys	Phe	Ser	Arg	Tyr				
		260						265						270					
Pro	Asp	His	Met	Lys	Gln	His	Asp	Phe	Phe	Lys	Ser	Ala	Met	Pro	Glu				
		275					280					285							
Gly	Tyr	Ile	Gln	Glu	Arg	Thr	Ile	Phe	Phe	Lys	Asp	Asp	Gly	Asn	Tyr				
	290					295					300								
Lys	Thr	Arg	Ala	Glu	Val	Lys	Phe	Glu	Gly	Asp	Thr	Leu	Val	Asn	Arg				
305					310					315					320				
Ile	Glu	Leu	Lys	Gly	Ile	Asp	Phe	Lys	Glu	Asp	Gly	Asn	Ile	Leu	Gly				
			325						330					335					
His	Lys	Leu	Glu	Tyr	Asn	Phe	Asn	Pro	Gly	Tyr	Trp	Ser	Val	Leu	Ile				
		340					345						350						
Val	Asn	Lys	Asp	Ser	Pro	Ile	Asn	Asn	Leu	Asn	Asp	Leu	Leu	Ala	Lys				
		355					360					365							
Arg	Lys	Asp	Leu	Thr	Phe	Gly	Asn	Gly	Asp	Pro	Asn	Ser	Thr	Ser	Gly				
370						375				380									
Phe	Leu	Val	Pro	Gly	Tyr	Tyr	Val	Phe	Ala	Lys	Asn	Ile	Asn	Ile	Ser				
385					390					395					400				
Ala	Ser	Asp	Phe	Lys	Arg	Thr	Val	Asn	Ala	Gly	His	Glu	Thr	Asn	Ala				
			405						410					415					
Leu	Ala	Val	Ala	Asn	Lys	Gln	Val	Asp	Val	Ala	Thr	Asn	Asn	Thr	Glu				
		420					425						430						
Asn	Leu	Asp	Lys	Leu	Lys	Thr	Ser	Ala	Pro	Glu	Lys	Leu	Lys	Glu	Leu				
		435				440						445							
Lys	Val	Ile	Trp	Lys	Ser	Pro	Leu	Ile	Pro	Gly	Asp	Pro	Ile	Val	Trp				
450					455					460									
Arg	Lys	Asn	Leu	Ser	Glu	Thr	Thr	Lys	Asp	Lys	Ile	Tyr	Asp	Phe	Phe				
465				470						475					480				
Met	Asn	Tyr	Gly	Lys	Thr	Pro	Glu	Glu	Lys	Ala	Val	Leu	Glu	Arg	Leu				
			485						490					495					
Gly	Trp	Ala	Pro	Phe	Arg	Ala	Ser	Ser	Asp	Leu	Gln	Leu	Val	Pro	Ile				
		500						505						510					
Arg	Gln	Leu	Ala	Leu	Phe	Lys	Glu	Met	Gln	Ser	Val	Lys	Asp	Asn	Lys				
		515					520						525						
Gly	Leu	Asn	Glu	Gln	Asp	Lys	Leu	Ala	Lys	Thr	Thr	Ala	Ile	Gln	Ala				
	530					535						540							
Gln	Leu	Asp	Asp	Leu	Asp	Arg	Leu	Asn	Asn	Ala	Leu	Ser	Ala	Met	Ser				
545					550					555					560				
Ser	Val	Ser	Lys	Ala	Val	Gln													
			565																

<210> SEQ ID NO 78
<211> LENGTH: 566
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic recombinant peptide biosensor

<400> SEQUENCE: 78

Met His His His His His His Gly Ser Glu Glu Gln Glu Lys Ala Leu
1 5 10 15

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Asn	Phe	Gly	Ile	Ile	Ser	Thr	Glu	Ser	Gln	Gln	Asn	Leu	Lys	Pro	Gln
		20						25					30		
Trp	Thr	Pro	Phe	Leu	Gln	Asp	Met	Glu	Lys	Lys	Leu	Gly	Val	Lys	Val
		35					40					45			
Asn	Ala	Phe	Phe	Ala	Pro	Asp	Tyr	Ala	Gly	Ile	Ile	Gln	Gly	Met	Arg
	50					55					60				
Phe	Asn	Lys	Val	Asp	Ile	Ala	Trp	Tyr	Gly	Asn	Leu	Ser	Ala	Met	Glu
65					70					75					80
Ala	Val	Asp	Arg	Ala	Asn	Gly	Gln	Val	Phe	Ala	Gln	Thr	Val	Ala	Ala
				85					90					95	
Asp	Ala	Asp	Asn	Val	Tyr	Ile	Met	Ala	Asp	Lys	Gln	Arg	Asn	Gly	Ile
			100					105					110		
Lys	Ala	Asn	Phe	Lys	Ile	Arg	His	Asn	Ile	Glu	Asp	Gly	Gly	Val	Gln
		115					120					125			
Leu	Ala	Tyr	His	Tyr	Gln	Gln	Asn	Thr	Pro	Ile	Gly	Asp	Gly	Pro	Val
	130					135					140				
Leu	Leu	Pro	Asp	Asn	His	Tyr	Leu	Ser	Thr	Gln	Ser	Lys	Leu	Ser	Lys
145					150					155					160
Ser	Lys	Asp	Pro	Asn	Glu	Lys	Arg	Asp	His	Met	Val	Leu	Leu	Glu	Phe
				165					170					175	
Val	Thr	Ala	Ala	Gly	Ile	Thr	Leu	Gly	Met	Asp	Glu	Leu	Tyr	Lys	Gly
			180					185					190		
Gly	Thr	Gly	Gly	Ser	Met	Val	Ser	Lys	Gly	Glu	Glu	Leu	Phe	Thr	Gly
		195					200					205			
Val	Val	Pro	Ile	Leu	Val	Glu	Leu	Asp	Gly	Asp	Val	Asn	Gly	His	Lys
	210					215					220				
Phe	Ser	Val	Ser	Gly	Glu	Gly	Glu	Gly	Asp	Ala	Thr	Tyr	Gly	Lys	Leu
225					230					235					240
Thr	Leu	Lys	Phe	Ile	Cys	Thr	Thr	Gly	Lys	Leu	Pro	Val	Pro	Trp	Pro
				245					250					255	
Thr	Leu	Val	Thr	Thr	Leu	Thr	Tyr	Gly	Val	Gln	Cys	Phe	Ser	Arg	Tyr
			260					265					270		
Pro	Asp	His	Met	Lys	Gln	His	Asp	Phe	Phe	Lys	Ser	Ala	Met	Pro	Glu
		275					280					285			
Gly	Tyr	Ile	Gln	Glu	Arg	Thr	Ile	Phe	Phe	Lys	Asp	Asp	Gly	Asn	Tyr
	290					295					300				
Lys	Thr	Arg	Ala	Glu	Val	Lys	Phe	Glu	Gly	Asp	Thr	Leu	Val	Asn	Arg
305					310					315					320
Ile	Glu	Leu	Lys	Gly	Ile	Asp	Phe	Lys	Glu	Asp	Gly	Asn	Ile	Leu	Gly
				325					330					335	
His	Lys	Leu	Glu	Tyr	Asn	Phe	Asn	Pro	Gly	Tyr	Trp	Ser	Val	Leu	Ile
			340					345					350		
Val	Asn	Lys	Asp	Ser	Pro	Ile	Asn	Asn	Leu	Asn	Asp	Leu	Leu	Ala	Lys
		355					360					365			
Arg	Lys	Asp	Leu	Thr	Phe	Gly	Asn	Gly	Asp	Pro	Asn	Ser	Thr	Ser	Gly
	370					375					380				
Phe	Leu	Val	Pro	Gly	Tyr	Tyr	Val	Phe	Ala	Lys	Asn	Asn	Ile	Ser	Ala
385					390					395					400
Gly	Lys	Thr	Pro	Glu	Glu	Lys	Ala	Val	Leu	Glu	Arg	Leu	Gly	Trp	Ala
				405					410					415	
Pro	Phe	Arg	Ala	Ser	Ser	Asp	Leu	Gln	Leu	Val	Pro	Ile	Arg	Gln	Leu

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420	425	430	
Ala Leu Phe Lys Glu Met Gln Ser Val Lys Asp Asn Lys Gly Leu Asn 435	440	445	
Glu Gln Asp Lys Leu Ala Lys Thr Thr Ala Ile Gln Ala Gln Leu Asp 450	455	460	
Asp Leu Asp Arg Arg Asn Asn Ala Arg Ser Ala Met Ser Ser Val Ser 465	470	475	480
Asn Tyr Gly Lys Thr Pro Glu Glu Lys Ala Val Leu Glu Arg Leu Gly 485	490	495	
Trp Ala Pro Phe Arg Ala Ser Ser Asp Leu Gln Leu Val Pro Ile Arg 500	505	510	
Gln Leu Ala Leu Phe Lys Glu Met Gln Ser Val Lys Asp Asn Lys Gly 515	520	525	
Leu Asn Glu Gln Asp Lys Leu Ala Lys Thr Thr Ala Ile Gln Ala Gln 530	535	540	
Leu Asp Asp Leu Asp Arg Leu Asn Asn Ala Leu Ser Ala Met Ser Ser 545	550	555	560
Val Ser Lys Ala Val Gln 565			

<210> SEQ ID NO 79
 <211> LENGTH: 16
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic oligopeptide

<400> SEQUENCE: 79

Gln Thr Val Ala Ala Asp Gly Ser Ser His Asn Val Tyr Ile Met Ala			
1	5	10	15

<210> SEQ ID NO 80
 <211> LENGTH: 16
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic oligopeptide
 <220> FEATURE:
 <221> NAME/KEY: MOD_RES
 <222> LOCATION: (7)..(8)
 <223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 80

Gln Thr Val Ala Ala Asp Xaa Xaa Ser His Asn Val Tyr Ile Met Ala			
1	5	10	15

<210> SEQ ID NO 81
 <211> LENGTH: 16
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic oligopeptide
 <220> FEATURE:
 <221> NAME/KEY: MOD_RES
 <222> LOCATION: (7)..(7)
 <223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 81

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Gln	Thr	Val	Ala	Ala	Asp	Xaa	Pro	Ser	His	Asn	Val	Tyr	Ile	Met	Ala
1				5					10					15	

<210> SEQ ID NO 82
 <211> LENGTH: 16
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
 oligopeptide
 <220> FEATURE:
 <221> NAME/KEY: MOD_RES
 <222> LOCATION: (8)..(8)
 <223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 82

Gln	Thr	Val	Ala	Ala	Asp	Pro	Xaa	Ser	His	Asn	Val	Tyr	Ile	Met	Ala
1				5					10					15	

<210> SEQ ID NO 83
 <211> LENGTH: 14
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
 oligopeptide
 <220> FEATURE:
 <221> NAME/KEY: MOD_RES
 <222> LOCATION: (7)..(8)
 <223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 83

Gln	Thr	Val	Ala	Ala	Asp	Xaa	Xaa	Asn	Val	Tyr	Ile	Met	Ala
1				5				10					

<210> SEQ ID NO 84
 <211> LENGTH: 16
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
 oligopeptide
 <220> FEATURE:
 <221> NAME/KEY: MOD_RES
 <222> LOCATION: (7)..(8)
 <223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 84

Gln	Thr	Val	Ala	Ala	Asp	Xaa	Xaa	Ser	His	Asn	Val	Tyr	Ile	Met	Ala
1				5				10						15	

<210> SEQ ID NO 85
 <211> LENGTH: 16
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
 oligopeptide
 <220> FEATURE:
 <221> NAME/KEY: MOD_RES
 <222> LOCATION: (7)..(8)
 <223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 85

Val	Phe	Gln	Thr	Val	Ala	Xaa	Xaa	Ser	His	Asn	Val	Tyr	Ile	Met	Ala
1				5				10						15	

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<210> SEQ ID NO 86
<211> LENGTH: 16
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide

<400> SEQUENCE: 86

His Lys Leu Glu Tyr Asn Phe Asn Pro Gly Tyr Trp Ser Val Leu Ile
1 5 10 15

<210> SEQ ID NO 87
<211> LENGTH: 18
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (9)..(10)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 87

His Lys Leu Glu Tyr Asn Phe Asn Xaa Xaa Pro Gly Tyr Trp Ser Val
1 5 10 15

Leu Ile

<210> SEQ ID NO 88
<211> LENGTH: 16
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (7)..(8)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 88

His Lys Leu Glu Tyr Asn Xaa Xaa Pro Gly Tyr Trp Ser Val Leu Ile
1 5 10 15

<210> SEQ ID NO 89
<211> LENGTH: 16
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (9)..(10)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 89

His Lys Leu Glu Tyr Asn Phe Asn Xaa Xaa Tyr Trp Ser Val Leu Ile
1 5 10 15

<210> SEQ ID NO 90
<211> LENGTH: 16
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic

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oligopeptide
 <220> FEATURE:
 <221> NAME/KEY: MOD_RES
 <222> LOCATION: (10)..(10)
 <223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 90

His Lys Leu Glu Tyr Asn Phe Asn Pro Xaa Tyr Trp Ser Val Leu Ile
 1 5 10 15

<210> SEQ ID NO 91
 <211> LENGTH: 678
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
 recombinant peptide biosensor

<400> SEQUENCE: 91

Met Arg Gly Ser His His His His His His Gly Met Ala Ser Met Thr
 1 5 10 15

Gly Gly Gln Gln Met Gly Arg Asp Leu Tyr Asp Asp Asp Lys Asp
 20 25 30

Arg Trp Gly Ser Lys Leu Glu Ile Phe Ser Trp Trp Ala Gly Asp Glu
 35 40 45

Gly Pro Ala Leu Glu Ala Leu Ile Arg Leu Tyr Lys Gln Lys Tyr Pro
 50 55 60

Gly Val Glu Val Ile Asn Ala Thr Val Thr Gly Gly Ala Gly Val Asn
 65 70 75 80

Ala Arg Ala Val Leu Lys Thr Arg Met Leu Gly Gly Asp Pro Pro Asp
 85 90 95

Thr Phe Gln Val His Ala Gly Met Glu Leu Ile Gly Thr Trp Val Val
 100 105 110

Ala Asn Arg Met Glu Asp Leu Ser Ala Leu Phe Arg Gln Glu Gly Trp
 115 120 125

Leu Gln Ala Phe Pro Lys Gly Leu Ile Asp Leu Ile Ser Tyr Lys Gly
 130 135 140

Gly Ile Trp Ser Val Pro Val Asn Ile His Arg Ser Asn Val Met Trp
 145 150 155 160

Tyr Leu Pro Ala Lys Leu Lys Glu Trp Gly Val Asn Pro Pro Arg Thr
 165 170 175

Trp Asp Glu Phe Leu Ala Thr Cys Gln Thr Leu Lys Gln Lys Gly Leu
 180 185 190

Glu Ala Pro Leu Ala Leu Gly Glu Asn Trp Thr Gln Gln His Leu Trp
 195 200 205

Glu Ser Val Ala Leu Ala Val Leu Gly Pro Asp Asp Trp Asn Asn Leu
 210 215 220

Trp Asn Gly Lys Leu Lys Phe Thr Asp Pro Lys Ala Val Arg Ala Trp
 225 230 235 240

Glu Val Phe Gly Arg Val Leu Asp Cys Ala Asn Lys Asp Ala Ala Gly
 245 250 255

Leu Ser Trp Gln Gln Ala Val Asp Arg Val Val Gln Gly Lys Ala Ala
 260 265 270

Phe Asn Val Met Gly Asp Trp Ala Ala Gly Tyr Met Thr Thr Thr Leu
 275 280 285

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Lys Leu Lys Pro Gly Thr Asp Phe Ala Trp Ala Pro Ser Pro Gly Thr
 290 295 300

Gln Gly Val Phe Met Met Leu Ser Asp Ser Phe Gly Leu Pro Lys Gly
 305 310 315 320

Ala Lys Asn Arg Gln Asn Ala Ile Asn Trp Leu Arg Leu Val Gly Ser
 325 330 335

Lys Glu Gly Gln Asp Thr Phe Asn Pro Leu Lys Gly Ser Ile Ala Ala
 340 345 350

Arg Leu Asp Ser Asp Pro Ser Lys Tyr Gly Gly Ser His Asn Val Tyr
 355 360 365

Ile Met Ala Asp Lys Gln Arg Asn Gly Ile Lys Ala Asn Phe Lys Ile
 370 375 380

Arg His Asn Ile Glu Asp Gly Gly Val Gln Leu Ala Tyr His Tyr Gln
 385 390 395 400

Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro Asp Asn His
 405 410 415

Tyr Leu Ser Thr Gln Ser Lys Leu Ser Lys Asp Pro Asn Glu Lys Arg
 420 425 430

Asp His Met Val Leu Leu Glu Phe Val Thr Ala Ala Gly Ile Thr Leu
 435 440 445

Gly Met Asp Glu Leu Tyr Lys Gly Gly Thr Gly Gly Ser Met Val Ser
 450 455 460

Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val Glu Leu
 465 470 475 480

Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly Glu Gly Glu
 485 490 495

Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile Cys Thr Thr
 500 505 510

Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu Thr Tyr
 515 520 525

Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys Gln His Asp
 530 535 540

Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Ile Gln Glu Arg Thr Ile
 545 550 555 560

Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe
 565 570 575

Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe
 580 585 590

Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn Phe Asn
 595 600 605

Asn Pro Asn Ala Tyr Gly Gln Ser Ala Met Arg Asp Trp Arg Ser Asn
 610 615 620

Arg Ile Val Gly Ser Leu Val His Gly Ala Val Ala Pro Glu Ser Phe
 625 630 635 640

Met Ser Gln Phe Gly Thr Val Met Glu Ile Phe Leu Gln Thr Arg Asn
 645 650 655

Pro Gln Ala Ala Ala Asn Ala Ala Gln Ala Ile Ala Asp Gln Val Gly
 660 665 670

Leu Gly Arg Leu Gly Gln
 675

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<210> SEQ ID NO 92
<211> LENGTH: 678
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
      recombinant peptide biosensor

<400> SEQUENCE: 92

Met Arg Gly Ser His His His His His His Gly Met Ala Ser Met Thr
 1           5           10
Gly Gly Gln Gln Met Gly Arg Asp Leu Tyr Asp Asp Asp Lys Asp
 20           25           30
Arg Trp Gly Ser Lys Leu Glu Ile Phe Ser Trp Trp Ala Gly Asp Glu
 35           40           45
Gly Pro Ala Leu Glu Ala Leu Ile Arg Leu Tyr Lys Gln Lys Tyr Pro
 50           55           60
Gly Val Glu Val Ile Asn Ala Thr Val Thr Gly Gly Ala Gly Val Asn
 65           70           75           80
Ala Arg Ala Val Leu Lys Thr Arg Met Leu Gly Gly Asp Pro Pro Asp
 85           90           95
Thr Phe Gln Val His Ala Gly Met Glu Leu Ile Gly Thr Trp Val Val
 100          105          110
Ala Asn Arg Met Glu Asp Leu Ser Ala Leu Phe Arg Gln Glu Gly Trp
 115          120          125
Leu Gln Ala Phe Pro Lys Gly Leu Ile Asp Leu Ile Ser Tyr Lys Gly
 130          135          140
Gly Ile Trp Ser Val Pro Val Asn Ile His Arg Ser Asn Val Met Trp
 145          150          155          160
Tyr Leu Pro Ala Lys Leu Lys Glu Trp Gly Val Asn Pro Pro Arg Thr
 165          170          175
Trp Asp Glu Phe Leu Ala Thr Cys Gln Thr Leu Lys Gln Lys Gly Leu
 180          185          190
Glu Ala Pro Leu Ala Leu Gly Glu Asn Trp Thr Gln Gln His Leu Trp
 195          200          205
Glu Ser Val Ala Leu Ala Val Leu Gly Pro Asp Asp Trp Asn Asn Leu
 210          215          220
Trp Asn Gly Lys Leu Lys Phe Thr Asp Pro Lys Ala Val Arg Ala Trp
 225          230          235          240
Glu Val Phe Gly Arg Val Leu Asp Cys Ala Asn Lys Asp Ala Ala Gly
 245          250          255
Leu Ser Trp Gln Gln Ala Val Asp Arg Val Val Gln Gly Lys Ala Ala
 260          265          270
Phe Asn Val Met Gly Asp Trp Ala Ala Gly Tyr Met Thr Thr Thr Leu
 275          280          285
Lys Leu Lys Pro Gly Thr Asp Phe Ala Trp Ala Pro Ser Pro Gly Thr
 290          295          300
Gln Gly Val Phe Met Met Leu Ser Asp Ser Phe Gly Leu Pro Lys Gly
 305          310          315          320
Ala Lys Asn Arg Gln Asn Ala Ile Asn Trp Leu Arg Leu Val Gly Ser
 325          330          335
Lys Glu Gly Gln Asp Thr Phe Asn Pro Leu Lys Gly Ser Ile Ala Ala
 340          345          350

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Arg Leu Asp Ser Asp Pro Ser Lys Tyr Pro Ala Ser His Asn Val Tyr
 355 360 365

Ile Met Ala Asp Lys Gln Arg Asn Gly Ile Lys Ala Asn Phe Lys Ile
 370 375 380

Arg His Asn Ile Glu Asp Gly Gly Val Gln Leu Ala Tyr His Tyr Gln
 385 390 395 400

Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro Asp Asn His
 405 410 415

Tyr Leu Ser Thr Gln Ser Lys Leu Ser Lys Asp Pro Asn Glu Lys Arg
 420 425 430

Asp His Met Val Leu Leu Glu Phe Val Thr Ala Ala Gly Ile Thr Leu
 435 440 445

Gly Met Asp Glu Leu Tyr Lys Gly Gly Thr Gly Gly Ser Met Val Ser
 450 455 460

Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val Glu Leu
 465 470 475 480

Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly Glu Gly Glu
 485 490 495

Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile Cys Thr Thr
 500 505 510

Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu Thr Tyr
 515 520 525

Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys Gln His Asp
 530 535 540

Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Ile Gln Glu Arg Thr Ile
 545 550 555 560

Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe
 565 570 575

Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe
 580 585 590

Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn Phe Asn
 595 600 605

Asn Pro Asn Ala Tyr Gly Gln Ser Ala Met Arg Asp Trp Arg Ser Asn
 610 615 620

Arg Ile Val Gly Ser Leu Val His Gly Ala Val Ala Pro Glu Ser Phe
 625 630 635 640

Met Ser Gln Phe Gly Thr Val Met Glu Ile Phe Leu Gln Thr Arg Asn
 645 650 655

Pro Gln Ala Ala Ala Asn Ala Ala Gln Ala Ile Ala Asp Gln Val Gly
 660 665 670

Leu Gly Arg Leu Gly Gln
 675

<210> SEQ ID NO 93
 <211> LENGTH: 678
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
 recombinant peptide biosensor
 <400> SEQUENCE: 93

Met Arg Gly Ser His His His His His His Gly Met Ala Ser Met Thr
 1 5 10 15

-continued

Gly Gly Gln Gln Met Gly Arg Asp Leu Tyr Asp Asp Asp Asp Lys Asp
 20 25 30
 Arg Trp Gly Ser Lys Leu Glu Ile Phe Ser Trp Trp Ala Gly Asp Glu
 35 40 45
 Gly Pro Ala Leu Glu Ala Leu Ile Arg Leu Tyr Lys Gln Lys Tyr Pro
 50 55 60
 Gly Val Glu Val Ile Asn Ala Thr Val Thr Gly Gly Ala Gly Val Asn
 65 70 75 80
 Ala Arg Ala Val Leu Lys Thr Arg Met Leu Gly Gly Asp Pro Pro Asp
 85 90 95
 Thr Phe Gln Val His Ala Gly Met Glu Leu Ile Gly Thr Trp Val Val
 100 105 110
 Ala Asn Arg Met Glu Asp Leu Ser Ala Leu Phe Arg Gln Glu Gly Trp
 115 120 125
 Leu Gln Ala Phe Pro Lys Gly Leu Ile Asp Leu Ile Ser Tyr Lys Gly
 130 135 140
 Gly Ile Trp Ser Val Pro Val Asn Ile His Arg Ser Asn Val Met Trp
 145 150 155 160
 Tyr Leu Pro Ala Lys Leu Lys Glu Trp Gly Val Asn Pro Pro Arg Thr
 165 170 175
 Trp Asp Glu Phe Leu Ala Thr Cys Gln Thr Leu Lys Gln Lys Gly Leu
 180 185 190
 Glu Ala Pro Leu Ala Leu Gly Glu Asn Trp Thr Gln Gln His Leu Trp
 195 200 205
 Glu Ser Val Ala Leu Ala Val Leu Gly Pro Asp Asp Trp Asn Asn Leu
 210 215 220
 Trp Asn Gly Lys Leu Lys Phe Thr Asp Pro Lys Ala Val Arg Ala Trp
 225 230 235 240
 Ala Arg Ala Val Leu Lys Thr Arg Met Leu Gly Gly Asp Pro Pro Asp
 245 250 255
 Thr Phe Gln Val Ala Ala Gly Met Glu Leu Ile Gly Thr Trp Val Val
 260 265 270
 Ala Asn Arg Met Glu Asp Leu Ser Ala Leu Phe Arg Gln Glu Gly Trp
 275 280 285
 Leu Gln Ala Phe Pro Lys Gly Leu Ile Asp Leu Ile Ser Tyr Lys Gly
 290 295 300
 Gly Ile Trp Ser Val Pro Val Asn Ile His Arg Ser Asn Val Met Trp
 305 310 315 320
 Ala Lys Asn Arg Gln Asn Ala Ile Asn Trp Leu Arg Leu Val Gly Ser
 325 330 335
 Lys Glu Gly Gln Asp Thr Phe Asn Pro Leu Lys Gly Ser Ile Ala Ala
 340 345 350
 Arg Leu Asp Ser Asp Pro Ser Lys Tyr Gly Gly Ser His Asn Val Tyr
 355 360 365
 Ile Met Ala Asp Lys Gln Arg Asn Gly Ile Lys Ala Asn Phe Lys Ile
 370 375 380
 Arg His Asn Ile Glu Asp Gly Gly Val Gln Leu Ala Tyr His Tyr Gln
 385 390 395 400
 Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro Asp Asn His
 405 410 415

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Tyr Leu Ser Thr Gln Ser Lys Leu Ser Lys Asp Pro Asn Glu Lys Arg
 420 425 430

Asp His Met Val Leu Leu Glu Phe Val Thr Ala Ala Gly Ile Thr Leu
 435 440 445

Gly Met Asp Glu Leu Tyr Lys Gly Gly Thr Gly Gly Ser Met Val Ser
 450 455 460

Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val Glu Leu
 465 470 475 480

Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly Glu Gly Glu
 485 490 495

Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile Cys Thr Thr
 500 505 510

Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu Thr Tyr
 515 520 525

Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys Gln His Asp
 530 535 540

Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Ile Gln Glu Arg Thr Ile
 545 550 555 560

Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe
 565 570 575

Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe
 580 585 590

Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn Phe Asn
 595 600 605

Asn Pro Asn Ala Tyr Gly Gln Ser Ala Met Arg Asp Trp Arg Ser Asn
 610 615 620

Arg Ile Val Gly Ser Leu Val His Gly Ala Val Ala Pro Glu Ser Phe
 625 630 635 640

Met Ser Gln Phe Gly Thr Val Met Glu Ile Phe Leu Gln Thr Arg Asn
 645 650 655

Pro Gln Ala Ala Ala Asn Ala Ala Gln Ala Ile Ala Asp Gln Val Gly
 660 665 670

Leu Gly Arg Leu Gly Gln
 675

<210> SEQ ID NO 94
 <211> LENGTH: 678
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
 recombinant peptide biosensor

<400> SEQUENCE: 94

Met Arg Gly Ser His His His His His His Gly Met Ala Ser Met Thr
 1 5 10 15

Gly Gly Gln Gln Met Gly Arg Asp Leu Tyr Asp Asp Asp Lys Asp
 20 25 30

Arg Trp Gly Ser Lys Leu Glu Ile Phe Ser Trp Trp Ala Gly Asp Glu
 35 40 45

Gly Pro Ala Leu Glu Ala Leu Ile Arg Leu Tyr Lys Gln Lys Tyr Pro
 50 55 60

Gly Val Glu Val Ile Asn Ala Thr Val Thr Gly Gly Ala Gly Val Asn
 65 70 75 80

-continued

Ala Arg Ala Val Leu Lys Thr Arg Met Leu Gly Gly Asp Pro Pro Asp
85 90 95

Thr Phe Gln Val His Ala Gly Met Glu Leu Ile Gly Thr Trp Val Val
100 105 110

Ala Asn Arg Met Glu Asp Leu Ser Ala Leu Phe Arg Gln Glu Gly Trp
115 120 125

Leu Gln Ala Phe Pro Lys Gly Leu Ile Asp Leu Ile Ser Tyr Lys Gly
130 135 140

Gly Ile Trp Ser Val Pro Val Asn Ile His Arg Ser Asn Val Met Trp
145 150 155 160

Tyr Leu Pro Ala Lys Leu Lys Glu Trp Gly Val Asn Pro Pro Arg Thr
165 170 175

Trp Asp Glu Phe Leu Ala Thr Cys Gln Thr Leu Lys Gln Lys Gly Leu
180 185 190

Glu Ala Pro Leu Ala Leu Gly Glu Asn Trp Thr Gln Gln His Leu Trp
195 200 205

Glu Ser Val Ala Leu Ala Val Leu Gly Pro Asp Asp Trp Asn Asn Leu
210 215 220

Trp Asn Gly Lys Leu Lys Phe Thr Asp Pro Lys Ala Val Arg Ala Trp
225 230 235 240

Glu Val Phe Gly Arg Val Leu Asp Cys Ala Asn Lys Asp Ala Ala Gly
245 250 255

Leu Ser Trp Gln Gln Ala Val Asp Arg Val Val Gln Gly Lys Ala Ala
260 265 270

Phe Asn Val Met Gly Asp Trp Ala Ala Gly Tyr Met Thr Thr Thr Leu
275 280 285

Lys Leu Lys Pro Gly Thr Asp Phe Ala Trp Ala Pro Ser Pro Gly Thr
290 295 300

Gln Gly Val Phe Met Met Leu Ser Asp Ser Phe Gly Leu Pro Lys Gly
305 310 315 320

Ala Lys Asn Arg Gln Asn Ala Ile Asn Trp Leu Arg Leu Val Gly Ser
325 330 335

Lys Glu Gly Gln Asp Thr Phe Asn Pro Leu Lys Gly Ser Ile Ala Ala
340 345 350

Arg Leu Asp Ser Asp Pro Ser Lys Tyr Gly Gly Ser His Asn Val Tyr
355 360 365

Ile Met Ala Asp Lys Gln Arg Asn Gly Ile Lys Ala Asn Phe Lys Ile
370 375 380

Arg His Asn Ile Glu Asp Gly Gly Val Gln Leu Ala Tyr His Tyr Gln
385 390 395 400

Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro Asp Asn His
405 410 415

Tyr Leu Ser Thr Gln Ser Lys Leu Ser Lys Asp Pro Asn Glu Lys Arg
420 425 430

Asp His Met Val Leu Leu Glu Phe Val Thr Ala Ala Gly Ile Thr Leu
435 440 445

Gly Met Asp Glu Leu Tyr Lys Gly Gly Thr Gly Gly Ser Met Val Ser
450 455 460

Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val Glu Leu
465 470 475 480

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Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly Glu Gly Glu
 485 490 495

Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile Cys Thr Thr
 500 505 510

Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu Thr Tyr
 515 520 525

Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys Gln His Asp
 530 535 540

Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Ile Gln Glu Arg Thr Ile
 545 550 555 560

Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe
 565 570 575

Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe
 580 585 590

Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn Phe Asn
 595 600 605

Asn Pro Asn Ala Tyr Gly Gln Ser Ala Met Arg Asp Trp Arg Ser Asn
 610 615 620

Arg Ile Val Gly Ser Leu Val Ala Gly Ala Val Ala Pro Glu Ser Phe
 625 630 635 640

Met Ser Gln Phe Gly Thr Val Met Glu Ile Phe Leu Gln Thr Arg Asn
 645 650 655

Pro Gln Ala Ala Ala Asn Ala Ala Gln Ala Ile Ala Asp Gln Val Gly
 660 665 670

Leu Gly Arg Leu Gly Gln
 675

<210> SEQ ID NO 95
 <211> LENGTH: 16
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
 oligopeptide
 <220> FEATURE:
 <221> NAME/KEY: MOD_RES
 <222> LOCATION: (8)..(9)
 <223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 95

Asp Ser Asp Pro Ser Lys Tyr Xaa Xaa Ser His Asn Val Tyr Ile Met
 1 5 10 15

<210> SEQ ID NO 96
 <211> LENGTH: 16
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
 oligopeptide
 <220> FEATURE:
 <221> NAME/KEY: MOD_RES
 <222> LOCATION: (9)..(9)
 <223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 96

Asp Ser Asp Pro Ser Lys Tyr Pro Xaa Ser His Asn Val Tyr Ile Met
 1 5 10 15

-continued

<210> SEQ ID NO 97
<211> LENGTH: 16
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (8)..(8)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 97

Asp Ser Asp Pro Ser Lys Tyr Xaa Pro Ser His Asn Val Tyr Ile Met
1 5 10 15

<210> SEQ ID NO 98
<211> LENGTH: 16
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (8)..(9)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 98

Arg Leu Asp Ser Asp Pro Ser Xaa Xaa Ser His Asn Val Tyr Ile Met
1 5 10 15

<210> SEQ ID NO 99
<211> LENGTH: 14
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (8)..(9)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 99

Asp Ser Asp Pro Ser Lys Tyr Xaa Xaa Asn Val Tyr Ile Met
1 5 10

<210> SEQ ID NO 100
<211> LENGTH: 16
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (8)..(9)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 100

Lys Leu Glu Tyr Asn Phe Asn Xaa Xaa Asn Ala Tyr Gly Gln Ser Ala
1 5 10 15

<210> SEQ ID NO 101
<211> LENGTH: 16
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence

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<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (7)..(8)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 101

Lys Leu Glu Tyr Asn Phe Xaa Xaa Pro Asn Ala Tyr Gly Gln Ser Ala
1 5 10 15

<210> SEQ ID NO 102
<211> LENGTH: 16
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (8)..(9)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 102

Gly His Lys Leu Glu Tyr Asn Xaa Xaa Asn Ala Tyr Gly Gln Ser Ala
1 5 10 15

<210> SEQ ID NO 103
<211> LENGTH: 16
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (8)..(8)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 103

Lys Leu Glu Tyr Asn Phe Asn Xaa Pro Asn Ala Tyr Gly Gln Ser Ala
1 5 10 15

<210> SEQ ID NO 104
<211> LENGTH: 16
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (9)..(9)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 104

Lys Leu Glu Tyr Asn Phe Asn Pro Xaa Asn Ala Tyr Gly Gln Ser Ala
1 5 10 15

<210> SEQ ID NO 105
<211> LENGTH: 396
<212> TYPE: PRT
<213> ORGANISM: Escherichia coli

<400> SEQUENCE: 105

Met Lys Ile Lys Thr Gly Ala Arg Ile Leu Ala Leu Ser Ala Leu Thr

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1	5	10	15
Thr Met Met Phe Ser Ala Ser Ala Leu Ala Lys Ile Glu Glu Gly Lys	20	25	30
Leu Val Ile Trp Ile Asn Gly Asp Lys Gly Tyr Asn Gly Leu Ala Glu	35	40	45
Val Gly Lys Lys Phe Glu Lys Asp Thr Gly Ile Lys Val Thr Val Glu	50	55	60
His Pro Asp Lys Leu Glu Glu Lys Phe Pro Gln Val Ala Ala Thr Gly	65	70	80
Asp Gly Pro Asp Ile Ile Phe Trp Ala His Asp Arg Phe Gly Gly Tyr	85	90	95
Ala Gln Ser Gly Leu Leu Ala Glu Ile Thr Pro Asp Lys Ala Phe Gln	100	105	110
Asp Lys Leu Tyr Pro Phe Thr Trp Asp Ala Val Arg Tyr Asn Gly Lys	115	120	125
Leu Ile Ala Tyr Pro Ile Ala Val Glu Ala Leu Ser Leu Ile Tyr Asn	130	135	140
Lys Asp Leu Leu Pro Asn Pro Pro Lys Thr Trp Glu Glu Ile Pro Ala	145	150	160
Leu Asp Lys Glu Leu Lys Ala Lys Gly Lys Ser Ala Leu Met Phe Asn	165	170	175
Leu Gln Glu Pro Tyr Phe Thr Trp Pro Leu Ile Ala Ala Asp Gly Gly	180	185	190
Tyr Ala Phe Lys Tyr Glu Asn Gly Lys Tyr Asp Ile Lys Asp Val Gly	195	200	205
Val Asp Asn Ala Gly Ala Lys Ala Gly Leu Thr Phe Leu Val Asp Leu	210	215	220
Ile Lys Asn Lys His Met Asn Ala Asp Thr Asp Tyr Ser Ile Ala Glu	225	230	240
Ala Ala Phe Asn Lys Gly Glu Thr Ala Met Thr Ile Asn Gly Pro Trp	245	250	255
Ala Trp Ser Asn Ile Asp Thr Ser Lys Val Asn Tyr Gly Val Thr Val	260	265	270
Leu Pro Thr Phe Lys Gly Gln Pro Ser Lys Pro Phe Val Gly Val Leu	275	280	285
Ser Ala Gly Ile Asn Ala Ala Ser Pro Asn Lys Glu Leu Ala Lys Glu	290	295	300
Phe Leu Glu Asn Tyr Leu Leu Thr Asp Glu Gly Leu Glu Ala Val Asn	305	310	320
Lys Asp Lys Pro Leu Gly Ala Val Ala Leu Lys Ser Tyr Glu Glu Glu	325	330	335
Leu Ala Lys Asp Pro Arg Ile Ala Ala Thr Met Glu Asn Ala Gln Lys	340	345	350
Gly Glu Ile Met Pro Asn Ile Pro Gln Met Ser Ala Phe Trp Tyr Ala	355	360	365
Val Arg Thr Ala Val Ile Asn Ala Ala Ser Gly Arg Gln Thr Val Asp	370	375	380
Glu Ala Leu Lys Asp Ala Gln Thr Arg Ile Thr Lys	385	390	395

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<211> LENGTH: 434

<212> TYPE: PRT

<213> ORGANISM: *Pyrococcus furiosus*

<400> SEQUENCE: 106

Met Arg Arg Ala Thr Tyr Ala Phe Ala Leu Leu Ala Ile Leu Val Leu
 1 5 10 15
 Gly Val Val Ala Ser Gly Cys Ile Gly Gly Gly Thr Thr Thr Pro Thr
 20 25 30
 Gln Thr Ser Pro Ala Thr Gln Pro Thr Thr Thr Gln Thr Pro Thr Gln
 35 40 45
 Thr Glu Thr Gln Ala Val Glu Cys Gly Ser Gly Lys Val Val Ile Trp
 50 55 60
 His Ala Met Gln Pro Asn Glu Leu Glu Val Phe Gln Ser Leu Ala Glu
 65 70 75 80
 Glu Tyr Met Ala Leu Cys Pro Glu Val Glu Ile Val Phe Glu Gln Lys
 85 90 95
 Pro Asn Leu Glu Asp Ala Leu Lys Ala Ala Ile Pro Thr Gly Gln Gly
 100 105 110
 Pro Asp Leu Phe Ile Trp Ala His Asp Trp Ile Gly Lys Phe Ala Glu
 115 120 125
 Ala Gly Leu Leu Glu Pro Ile Asp Glu Tyr Val Thr Glu Asp Leu Leu
 130 135 140
 Asn Glu Phe Ala Pro Met Ala Gln Asp Ala Met Gln Tyr Lys Gly His
 145 150 155 160
 Tyr Tyr Ala Leu Pro Phe Ala Ala Glu Thr Val Ala Ile Ile Tyr Asn
 165 170 175
 Lys Glu Met Val Ser Glu Pro Pro Lys Thr Phe Asp Glu Met Lys Ala
 180 185 190
 Ile Met Glu Lys Tyr Tyr Asp Pro Ala Asn Glu Lys Tyr Gly Ile Ala
 195 200 205
 Trp Pro Ile Asn Ala Tyr Phe Ile Ser Ala Ile Ala Gln Ala Phe Gly
 210 215 220
 Gly Tyr Tyr Phe Asp Asp Lys Thr Glu Gln Pro Gly Leu Asp Lys Pro
 225 230 235 240
 Glu Thr Ile Glu Gly Phe Lys Phe Phe Phe Thr Glu Ile Trp Pro Tyr
 245 250 255
 Met Ala Pro Thr Gly Asp Tyr Asn Thr Gln Gln Ser Ile Phe Leu Glu
 260 265 270
 Gly Arg Ala Pro Met Met Val Asn Gly Pro Trp Ser Ile Asn Asp Val
 275 280 285
 Lys Lys Ala Gly Ile Asn Phe Gly Val Val Pro Leu Pro Pro Ile Ile
 290 295 300
 Lys Asp Gly Lys Glu Tyr Trp Pro Arg Pro Tyr Gly Gly Val Lys Leu
 305 310 315 320
 Ile Tyr Phe Ala Ala Gly Ile Lys Asn Lys Asp Ala Ala Trp Lys Phe
 325 330 335
 Ala Lys Trp Leu Thr Thr Ser Glu Glu Ser Ile Lys Thr Leu Ala Leu
 340 345 350
 Glu Leu Gly Tyr Ile Pro Val Leu Thr Lys Val Leu Asp Asp Pro Glu
 355 360 365
 Ile Lys Asn Asp Pro Val Ile Tyr Gly Phe Gly Gln Ala Val Gln His

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290		295		300											
Asp	Lys	Leu	Ala	Lys	Thr	Thr	Ala	Ile	Gln	Ala	Gln	Leu	Asp	Asp	Leu
305					310					315					320
Asp	Arg	Leu	Asn	Asn	Ala	Leu	Ser	Ala	Met	Ser	Ser	Val	Ser	Lys	Ala
			325						330					335	
Val Gln															
<210> SEQ ID NO 108															
<211> LENGTH: 302															
<212> TYPE: PRT															
<213> ORGANISM: Escherichia coli															
<400> SEQUENCE: 108															
Met	Gln	Leu	Arg	Lys	Pro	Ala	Thr	Ala	Ile	Leu	Ala	Leu	Ala	Leu	Ser
1				5					10						15
Ala	Gly	Leu	Ala	Gln	Ala	Asp	Asp	Ala	Ala	Pro	Ala	Ala	Gly	Ser	Thr
		20						25					30		
Leu	Asp	Lys	Ile	Ala	Lys	Asn	Gly	Val	Ile	Val	Val	Gly	His	Arg	Glu
		35					40					45			
Ser	Ser	Val	Pro	Phe	Ser	Tyr	Tyr	Asp	Asn	Gln	Gln	Lys	Val	Val	Gly
	50					55					60				
Tyr	Ser	Gln	Asp	Tyr	Ser	Asn	Ala	Ile	Val	Glu	Ala	Val	Lys	Lys	Lys
65					70					75					80
Leu	Asn	Lys	Pro	Asp	Leu	Gln	Val	Lys	Leu	Ile	Pro	Ile	Thr	Ser	Gln
				85					90					95	
Asn	Arg	Ile	Pro	Leu	Leu	Gln	Asn	Gly	Thr	Phe	Asp	Phe	Glu	Cys	Gly
			100					105					110		
Ser	Thr	Thr	Asn	Asn	Val	Glu	Arg	Gln	Lys	Gln	Ala	Ala	Phe	Ser	Asp
		115						120					125		
Thr	Ile	Phe	Val	Val	Gly	Thr	Arg	Leu	Leu	Thr	Lys	Lys	Gly	Gly	Asp
	130					135					140				
Ile	Lys	Asp	Phe	Ala	Asn	Leu	Lys	Asp	Lys	Ala	Val	Val	Val	Thr	Ser
145					150					155					160
Gly	Thr	Thr	Ser	Glu	Val	Leu	Leu	Asn	Lys	Leu	Asn	Glu	Glu	Gln	Lys
				165					170					175	
Met	Asn	Met	Arg	Ile	Ile	Ser	Ala	Lys	Asp	His	Gly	Asp	Ser	Phe	Arg
			180					185					190		
Thr	Leu	Glu	Ser	Gly	Arg	Ala	Val	Ala	Phe	Met	Met	Asp	Asp	Ala	Leu
		195					200					205			
Leu	Ala	Gly	Glu	Arg	Ala	Lys	Ala	Lys	Lys	Pro	Asp	Asn	Trp	Glu	Ile
	210					215					220				
Val	Gly	Lys	Pro	Gln	Ser	Gln	Glu	Ala	Tyr	Gly	Cys	Met	Leu	Arg	Lys
225					230					235					240
Asp	Asp	Pro	Gln	Phe	Lys	Lys	Leu	Met	Asp	Asp	Thr	Ile	Ala	Gln	Val
				245					250					255	
Gln	Thr	Ser	Gly	Glu	Ala	Glu	Lys	Trp	Phe	Asp	Lys	Trp	Phe	Lys	Asn
			260					265					270		
Pro	Ile	Pro	Pro	Lys	Asn	Leu	Asn	Met	Asn	Phe	Glu	Leu	Ser	Asp	Glu
		275				280						285			
Met	Lys	Ala	Leu	Phe	Lys	Glu	Pro	Asn	Asp	Lys	Ala	Leu	Asn		
290					295							300			

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<210> SEQ ID NO 109
<211> LENGTH: 414
<212> TYPE: PRT
<213> ORGANISM: Thermus thermophilus

<400> SEQUENCE: 109

Met Arg Lys Trp Leu Leu Ala Ile Gly Met Val Leu Gly Leu Ser Ala
 1           5           10           15

Leu Ala Gln Gly Gly Lys Leu Glu Ile Phe Ser Trp Trp Ala Gly Asp
 20           25           30

Glu Gly Pro Ala Leu Glu Ala Leu Ile Arg Leu Tyr Lys Gln Lys Tyr
 35           40           45

Pro Gly Val Glu Val Ile Asn Ala Thr Val Thr Gly Gly Ala Gly Val
 50           55           60

Asn Ala Arg Ala Val Leu Lys Thr Arg Met Leu Gly Gly Asp Pro Pro
 65           70           75           80

Asp Thr Phe Gln Val His Ala Gly Met Glu Leu Ile Gly Thr Trp Val
 85           90           95

Val Ala Asn Arg Met Glu Asp Leu Ser Ala Leu Phe Arg Gln Glu Gly
 100          105          110

Trp Leu Gln Ala Phe Pro Lys Gly Leu Ile Asp Leu Ile Ser Tyr Lys
 115          120          125

Gly Gly Ile Trp Ser Val Pro Val Asn Ile His Arg Ser Asn Val Met
 130          135          140

Trp Tyr Leu Pro Ala Lys Leu Lys Glu Trp Gly Val Asn Pro Pro Arg
 145          150          155          160

Thr Trp Asp Glu Phe Leu Ala Thr Cys Gln Thr Leu Lys Gln Lys Gly
 165          170          175

Leu Glu Ala Pro Leu Ala Leu Gly Glu Asn Trp Thr Gln Gln His Leu
 180          185          190

Trp Glu Ser Val Ala Leu Ala Val Leu Gly Pro Asp Asp Trp Asn Asn
 195          200          205

Leu Trp Asn Gly Lys Leu Lys Phe Thr Asp Pro Lys Ala Val Arg Ala
 210          215          220

Trp Glu Val Phe Gly Arg Val Leu Asp Cys Ala Asn Lys Asp Ala Ala
 225          230          235          240

Gly Leu Ser Trp Gln Gln Ala Val Asp Arg Val Val Gln Gly Lys Ala
 245          250          255

Ala Phe Asn Val Met Gly Asp Trp Ala Ala Gly Tyr Met Thr Thr Thr
 260          265          270

Leu Lys Leu Lys Pro Gly Thr Asp Phe Ala Trp Ala Pro Ser Pro Gly
 275          280          285

Thr Gln Gly Val Phe Met Met Leu Ser Asp Ser Phe Gly Leu Pro Lys
 290          295          300

Gly Ala Lys Asn Arg Gln Asn Ala Ile Asn Trp Leu Arg Leu Val Gly
 305          310          315          320

Ser Lys Glu Gly Gln Asp Thr Phe Asn Pro Leu Lys Gly Ser Ile Ala
 325          330          335

Ala Arg Leu Asp Ser Asp Pro Ser Lys Tyr Asn Ala Tyr Gly Gln Ser
 340          345          350

Ala Met Arg Asp Trp Arg Ser Asn Arg Ile Val Gly Ser Leu Val His
 355          360          365

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Gly Ala Val Ala Pro Glu Ser Phe Met Ser Gln Phe Gly Thr Val Met
 370 375 380

Glu Ile Phe Leu Gln Thr Arg Asn Pro Gln Ala Ala Ala Asn Ala Ala
 385 390 395 400

Gln Ala Ile Ala Asp Gln Val Gly Leu Gly Arg Leu Gly Gln
 405 410

<210> SEQ ID NO 110
 <211> LENGTH: 318
 <212> TYPE: PRT
 <213> ORGANISM: Rhizobium meliloti

<400> SEQUENCE: 110

Met Ile Arg Thr Leu Ser Leu Lys Phe Met Leu Ala Gly Ala Val Cys
 1 5 10 15

Met Ala Thr Leu Thr Ala Gly Ser Ala Phe Ala Ala Glu Pro Glu Ser
 20 25 30

Cys Gly Thr Val Arg Phe Ser Asp Val Gly Trp Thr Asp Ile Thr Ala
 35 40 45

Thr Thr Ala Thr Ala Thr Thr Ile Leu Glu Ala Leu Gly Tyr Glu Thr
 50 55 60

Asp Val Lys Val Leu Ser Val Pro Val Thr Tyr Thr Ser Leu Lys Asn
 65 70 75 80

Lys Asp Ile Asp Val Phe Leu Gly Asn Trp Met Pro Thr Met Glu Ala
 85 90 95

Asp Ile Ala Pro Tyr Arg Glu Asp Lys Ser Val Glu Thr Val Arg Glu
 100 105 110

Asn Leu Ala Gly Ala Lys Tyr Thr Leu Ala Thr Asn Ala Lys Gly Ala
 115 120 125

Glu Leu Gly Ile Lys Asp Phe Lys Asp Ile Ala Ala His Lys Asp Glu
 130 135 140

Leu Asp Gly Lys Ile Tyr Gly Ile Glu Pro Gly Asn Asp Gly Asn Arg
 145 150 155 160

Leu Ile Ile Asp Met Val Glu Lys Gly Thr Phe Asp Leu Lys Gly Phe
 165 170 175

Glu Val Val Glu Ser Ser Glu Gln Gly Met Leu Ala Gln Val Ala Arg
 180 185 190

Ala Glu Lys Ser Gly Asp Pro Ile Val Phe Leu Gly Trp Glu Pro His
 195 200 205

Pro Met Asn Ala Asn Phe Lys Leu Thr Tyr Leu Ser Gly Gly Asp Asp
 210 215 220

Val Phe Gly Pro Asn Tyr Gly Gly Ala Thr Val His Thr Asn Val Arg
 225 230 235 240

Ala Gly Tyr Thr Thr Glu Cys Pro Asn Val Gly Lys Leu Leu Gln Asn
 245 250 255

Leu Ser Phe Ser Leu Gln Met Glu Asn Glu Ile Met Gly Lys Ile Leu
 260 265 270

Asn Asp Gly Glu Asp Pro Glu Lys Ala Ala Ala Ala Trp Leu Lys Asp
 275 280 285

Asn Pro Gln Ser Ile Glu Pro Trp Leu Ser Gly Val Ala Thr Lys Asp
 290 295 300

Gly Gly Asp Gly Leu Ala Ala Val Lys Ala Ala Leu Gly Leu
 305 310 315

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<210> SEQ ID NO 111
 <211> LENGTH: 249
 <212> TYPE: PRT
 <213> ORGANISM: Geobacillus stearothermophilus

<400> SEQUENCE: 111

Met Gly Gly Gly Arg Ser Thr Glu Thr Ser Ser Ser Ser Gly Gly Asp
 1 5 10 15
 Gly Gly Ala Thr Lys Lys Lys Val Val Val Gly Thr Asp Ala Ala Phe
 20 25 30
 Ala Pro Phe Glu Tyr Met Gln Lys Gly Lys Ile Val Gly Phe Asp Val
 35 40 45
 Asp Leu Leu Asp Ala Val Met Lys Ala Ala Gly Leu Asp Tyr Glu Leu
 50 55 60
 Lys Asn Ile Gly Trp Asp Pro Leu Phe Ala Ser Leu Gln Ser Lys Glu
 65 70 75 80
 Val Asp Met Gly Ile Ser Gly Ile Thr Ile Thr Asp Glu Arg Lys Gln
 85 90 95
 Ser Tyr Asp Phe Ser Asp Pro Tyr Phe Glu Ala Thr Gln Val Ile Leu
 100 105 110
 Val Lys Gln Gly Ser Pro Val Lys Asn Ala Leu Asp Leu Lys Gly Thr
 115 120 125
 Ile Gly Val Gln Asn Ala Thr Thr Gly Gln Glu Ala Ala Glu Lys Leu
 130 135 140
 Phe Gly Lys Gly Pro His Ile Lys Lys Phe Glu Thr Thr Val Val Ala
 145 150 155 160
 Ile Met Glu Leu Leu Asn Gly Gly Val Asp Ala Val Ile Thr Asp Asn
 165 170 175
 Ala Val Ala Asn Glu Tyr Val Lys Asn Asn Pro Asn Lys Lys Leu Gln
 180 185 190
 Val Ile Glu Asp Pro Lys Asn Phe Ala Ser Glu Tyr Tyr Gly Met Ile
 195 200 205
 Phe Pro Lys Asn Ser Glu Leu Lys Ala Lys Val Asp Glu Ala Leu Lys
 210 215 220
 Asn Val Ile Asn Ser Gly Lys Tyr Thr Glu Ile Tyr Lys Lys Trp Phe
 225 230 235 240
 Gly Lys Glu Pro Lys Leu Asp Arg Leu
 245

<210> SEQ ID NO 112
 <211> LENGTH: 370
 <212> TYPE: PRT
 <213> ORGANISM: Agrobacterium tumefaciens

<400> SEQUENCE: 112

Met Lys Lys Ser Leu Leu Ser Ala Val Ala Leu Thr Ala Met Val Ala
 1 5 10 15
 Phe Gly Gly Ser Ala Trp Ala Asp Val Val Ile Ala Val Gly Ala Pro
 20 25 30
 Leu Thr Gly Pro Asn Ala Ala Phe Gly Ala Gln Ile Gln Lys Gly Ala
 35 40 45
 Glu Gln Ala Ala Lys Asp Ile Asn Ala Ala Gly Gly Ile Asn Gly Glu
 50 55 60

-continued

Gln Ile Lys Ile Val Leu Gly Asp Asp Val Ser Asp Pro Lys Gln Gly
 65 70 75 80
 Ile Ser Val Ala Asn Lys Phe Val Ala Asp Gly Val Lys Phe Val Val
 85 90 95
 Gly His Phe Asn Ser Gly Val Ser Ile Pro Ala Ser Glu Val Tyr Ala
 100 105 110
 Glu Asn Gly Ile Leu Glu Ile Thr Pro Ala Ala Thr Asn Pro Val Phe
 115 120 125
 Thr Glu Arg Gly Leu Trp Asn Thr Phe Arg Thr Cys Gly Arg Asp Asp
 130 135 140
 Gln Gln Gly Gly Ile Ala Gly Lys Tyr Leu Ala Asp His Phe Lys Asp
 145 150 155 160
 Ala Lys Val Ala Ile Ile His Asp Lys Thr Pro Tyr Gly Gln Gly Leu
 165 170 175
 Ala Asp Glu Thr Lys Lys Ala Ala Asn Ala Ala Gly Val Thr Glu Val
 180 185 190
 Met Tyr Glu Gly Val Asn Val Gly Asp Lys Asp Phe Ser Ala Leu Ile
 195 200 205
 Ser Lys Met Lys Glu Ala Gly Val Ser Ile Ile Tyr Trp Gly Gly Leu
 210 215 220
 His Thr Glu Ala Gly Leu Ile Ile Arg Gln Ala Ala Asp Gln Gly Leu
 225 230 235 240
 Lys Ala Lys Leu Val Ser Gly Asp Gly Ile Val Ser Asn Glu Leu Ala
 245 250 255
 Ser Ile Ala Gly Asp Ala Val Glu Gly Thr Leu Asn Thr Phe Gly Pro
 260 265 270
 Asp Pro Thr Leu Arg Pro Glu Asn Lys Glu Leu Val Glu Lys Phe Lys
 275 280 285
 Ala Ala Gly Phe Asn Pro Glu Ala Tyr Thr Leu Tyr Ser Tyr Ala Ala
 290 295 300
 Met Gln Ala Ile Ala Gly Ala Ala Lys Ala Ala Gly Ser Val Glu Pro
 305 310 315 320
 Glu Lys Val Ala Glu Ala Leu Lys Lys Gly Ser Phe Pro Thr Ala Leu
 325 330 335
 Gly Glu Ile Ser Phe Asp Glu Lys Gly Asp Pro Lys Leu Pro Gly Tyr
 340 345 350
 Val Met Tyr Glu Trp Lys Lys Gly Pro Asp Gly Lys Phe Thr Tyr Ile
 355 360 365
 Gln Gln
 370

<210> SEQ ID NO 113

<211> LENGTH: 367

<212> TYPE: PRT

<213> ORGANISM: Escherichia coli

<400> SEQUENCE: 113

Met Asn Ile Lys Gly Lys Ala Leu Leu Ala Gly Cys Ile Ala Leu Ala
 1 5 10 15
 Phe Ser Asn Met Ala Leu Ala Glu Asp Ile Lys Val Ala Val Val Gly
 20 25 30
 Ala Met Ser Gly Pro Val Ala Gln Tyr Gly Asp Gln Glu Phe Thr Gly

-continued

35	40	45
Ala Glu Gln Ala Val Ala Asp Ile Asn Ala Lys Gly Gly Ile Lys Gly		
50	55	60
Asn Lys Leu Gln Ile Val Lys Tyr Asp Asp Ala Cys Asp Pro Lys Gln		
65	70	75 80
Ala Val Ala Val Ala Asn Lys Val Val Asn Asp Gly Ile Lys Tyr Val		
	85	90 95
Ile Gly His Leu Cys Ser Ser Ser Thr Gln Pro Ala Ser Asp Ile Tyr		
	100	105 110
Glu Asp Glu Gly Ile Leu Met Ile Thr Pro Ala Ala Thr Ala Pro Glu		
	115	120 125
Leu Thr Ala Arg Gly Tyr Gln Leu Ile Leu Arg Thr Thr Gly Leu Asp		
	130	135 140
Ser Asp Gln Gly Pro Thr Ala Ala Lys Tyr Ile Leu Glu Lys Val Lys		
145	150	155 160
Pro Gln Arg Ile Ala Ile Val His Asp Lys Gln Gln Tyr Gly Glu Gly		
	165	170 175
Leu Ala Arg Ala Val Gln Asp Gly Leu Lys Lys Gly Asn Ala Asn Val		
	180	185 190
Val Phe Phe Asp Gly Ile Thr Ala Gly Glu Lys Asp Phe Ser Thr Leu		
	195	200 205
Val Ala Arg Leu Lys Lys Glu Asn Ile Asp Phe Val Tyr Tyr Gly Gly		
	210	215 220
Tyr His Pro Glu Met Gly Gln Ile Leu Arg Gln Ala Arg Ala Ala Gly		
225	230	235 240
Leu Lys Thr Gln Phe Met Gly Pro Glu Gly Val Ala Asn Val Ser Leu		
	245	250 255
Ser Asn Ile Ala Gly Glu Ser Ala Glu Gly Leu Leu Val Thr Lys Pro		
	260	265 270
Lys Asn Tyr Asp Gln Val Pro Ala Asn Lys Pro Ile Val Asp Ala Ile		
	275	280 285
Lys Ala Lys Lys Gln Asp Pro Ser Gly Ala Phe Val Trp Thr Thr Tyr		
	290	295 300
Ala Ala Leu Gln Ser Leu Gln Ala Gly Leu Asn Gln Ser Asp Asp Pro		
305	310	315 320
Ala Glu Ile Ala Lys Tyr Leu Lys Ala Asn Ser Val Asp Thr Val Met		
	325	330 335
Gly Pro Leu Thr Trp Asp Glu Lys Gly Asp Leu Lys Gly Phe Glu Phe		
	340	345 350
Gly Val Phe Asp Trp His Ala Asn Gly Thr Ala Thr Asp Ala Lys		
	355	360 365

<210> SEQ ID NO 114
 <211> LENGTH: 677
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic recombinant peptide biosensor

<400> SEQUENCE: 114

Met Arg Gly Ser His His His His His Gly Met Ala Ser Met Thr
1 5 10 15

-continued

Gly	Gly	Gln	Gln	Met	Gly	Arg	Asp	Leu	Tyr	Asp	Asp	Asp	Asp	Lys	Asp
		20						25					30		
Arg	Trp	Gly	Ser	Lys	Leu	Glu	Ile	Phe	Ser	Trp	Trp	Ala	Gly	Asp	Glu
		35					40					45			
Gly	Pro	Ala	Leu	Glu	Ala	Leu	Ile	Arg	Leu	Tyr	Lys	Gln	Lys	Tyr	Pro
	50					55					60				
Gly	Val	Glu	Val	Ile	Asn	Ala	Thr	Val	Thr	Gly	Gly	Ala	Gly	Val	Asn
	65				70					75					80
Ala	Arg	Ala	Val	Leu	Lys	Thr	Arg	Met	Leu	Gly	Gly	Asp	Pro	Pro	Asp
				85					90					95	
Thr	Phe	Gln	Val	Ala	Ala	Gly	Met	Glu	Leu	Ile	Gly	Thr	Trp	Val	Val
		100						105					110		
Ala	Asn	Arg	Met	Glu	Asp	Leu	Ser	Ala	Leu	Phe	Arg	Gln	Glu	Gly	Trp
		115					120					125			
Leu	Gln	Ala	Phe	Pro	Lys	Gly	Leu	Ile	Asp	Leu	Ile	Ser	Tyr	Lys	Gly
	130					135					140				
Gly	Ile	Trp	Ser	Val	Pro	Val	Asn	Ile	His	Arg	Ser	Asn	Val	Met	Trp
	145				150					155					160
Tyr	Leu	Pro	Ala	Lys	Leu	Lys	Glu	Trp	Gly	Val	Asn	Pro	Pro	Arg	Thr
				165					170					175	
Trp	Glu	Phe	Leu	Ala	Thr	Cys	Gln	Thr	Leu	Lys	Gln	Lys	Gly	Leu	Glu
		180						185					190		
Ala	Pro	Leu	Ala	Leu	Gly	Glu	Asn	Trp	Thr	Gln	Gln	His	Leu	Trp	Glu
		195					200					205			
Ser	Val	Ala	Leu	Ala	Val	Leu	Gly	Pro	Asp	Asp	Trp	Asn	Asn	Leu	Trp
	210					215					220				
Asn	Gly	Lys	Leu	Lys	Phe	Thr	Asp	Pro	Lys	Ala	Val	Arg	Ala	Trp	Glu
	225				230					235					240
Val	Phe	Gly	Arg	Val	Leu	Asp	Cys	Ala	Asn	Lys	Asp	Ala	Ala	Gly	Leu
				245					250					255	
Ser	Trp	Gln	Gln	Ala	Val	Asp	Arg	Val	Val	Gln	Gly	Lys	Ala	Ala	Phe
			260					265					270		
Asn	Val	Met	Gly	Asp	Trp	Ala	Ala	Gly	Tyr	Met	Thr	Thr	Thr	Leu	Lys
		275					280					285			
Leu	Lys	Pro	Gly	Thr	Asp	Phe	Ala	Trp	Ala	Pro	Ser	Pro	Gly	Thr	Gln
	290					295					300				
Gly	Val	Phe	Met	Met	Val	Ser	Asp	Ser	Phe	Gly	Leu	Pro	Lys	Gly	Ala
	305				310					315					320
Lys	Asn	Arg	Gln	Asn	Ala	Ile	Asn	Trp	Leu	Arg	Leu	Val	Gly	Ser	Lys
				325					330					335	
Glu	Gly	Gln	Asp	Thr	Phe	Asn	Pro	Leu	Lys	Gly	Ser	Ile	Ala	Ala	Arg
			340					345					350		
Leu	Asp	Ser	Asp	Pro	Ser	Lys	Tyr	Pro	Ala	Ser	His	Asn	Val	Tyr	Ile
		355					360					365			
Met	Ala	Asp	Lys	Gln	Arg	Asn	Gly	Ile	Lys	Ala	Asn	Phe	Lys	Ile	Arg
	370					375					380				
His	Asn	Ile	Glu	Asp	Gly	Gly	Val	Gln	Leu	Ala	Tyr	His	Tyr	Gln	Gln
	385				390					395					400
Asn	Thr	Pro	Ile	Gly	Asp	Gly	Pro	Val	Leu	Leu	Pro	Asp	Asn	His	Tyr
				405					410					415	
Leu	Ser	Thr	Gln	Ser	Lys	Leu	Ser	Lys	Asp	Pro	Asn	Glu	Lys	Arg	Asp

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420					425					430					
His	Met	Val	Leu	Leu	Glu	Phe	Val	Thr	Ala	Ala	Gly	Ile	Thr	Leu	Gly
		435					440					445			
Met	Asp	Glu	Leu	Tyr	Lys	Gly	Gly	Thr	Gly	Gly	Ser	Met	Val	Ser	Lys
	450					455					460				
Gly	Glu	Glu	Leu	Phe	Thr	Gly	Val	Val	Pro	Ile	Leu	Val	Glu	Leu	Asp
465					470					475					480
Gly	Asp	Val	Asn	Gly	His	Lys	Phe	Ser	Val	Ser	Gly	Glu	Gly	Glu	Gly
			485						490						495
Asp	Ala	Thr	Tyr	Gly	Lys	Leu	Thr	Leu	Lys	Phe	Ile	Cys	Thr	Thr	Gly
			500						505						510
Lys	Leu	Pro	Val	Pro	Trp	Pro	Thr	Leu	Val	Thr	Thr	Leu	Thr	Tyr	Gly
		515						520							525
Val	Gln	Cys	Phe	Ser	Arg	Tyr	Pro	Asp	His	Met	Lys	Gln	His	Asp	Phe
	530					535					540				
Phe	Lys	Ser	Ala	Met	Pro	Glu	Gly	Tyr	Ile	Gln	Glu	Arg	Thr	Ile	Phe
545					550					555					560
Phe	Lys	Asp	Asp	Gly	Asn	Tyr	Lys	Thr	Arg	Ala	Glu	Val	Lys	Phe	Glu
				565					570						575
Gly	Asp	Thr	Leu	Val	Asn	Arg	Ile	Glu	Leu	Lys	Gly	Ile	Asp	Phe	Lys
			580						585						590
Glu	Asp	Gly	Asn	Ile	Leu	Gly	His	Lys	Leu	Glu	Tyr	Asn	Phe	Asn	Asn
		595					600						605		
Pro	Asn	Ala	Tyr	Gly	Gln	Ser	Ala	Met	Arg	Asp	Trp	Arg	Ser	Asn	Arg
	610					615									620
Ile	Val	Gly	Ser	Leu	Val	Ala	Gly	Ala	Val	Ala	Pro	Glu	Ser	Phe	Met
625					630						635				640
Ser	Gln	Phe	Gly	Thr	Val	Met	Glu	Ile	Phe	Leu	Gln	Thr	Arg	Asn	Pro
				645					650						655
Gln	Ala	Ala	Ala	Asn	Ala	Ala	Gln	Ala	Ile	Ala	Asp	Gln	Val	Gly	Leu
			660					665							670
Gly	Arg	Leu	Gly	Gln											
		675													

<210> SEQ ID NO 115
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 <212> TYPE: PRT
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 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic oligopeptide
 <220> FEATURE:
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 <223> OTHER INFORMATION: Any amino acid
 <400> SEQUENCE: 115

Xaa Pro Ser His Asn Val Tyr
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<210> SEQ ID NO 116
 <211> LENGTH: 7
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic oligopeptide

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<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (1)..(2)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 116

Xaa Xaa Ser His Asn Val Tyr
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<210> SEQ ID NO 117
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<212> TYPE: PRT
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<220> FEATURE:
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oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (1)..(2)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 117

Xaa Xaa Ser His Asn Val Phe
1 5

<210> SEQ ID NO 118
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<212> TYPE: PRT
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<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (2)..(2)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 118

Pro Xaa Ser His Asn Val Phe
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<212> TYPE: PRT
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<220> FEATURE:
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<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (2)..(2)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 119

Pro Xaa Ser Tyr Asn Val Phe
1 5

<210> SEQ ID NO 120
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<220> FEATURE:
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<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (1)..(2)
<223> OTHER INFORMATION: Any amino acid

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<400> SEQUENCE: 120

Xaa Xaa Ser Tyr Asn Val Phe
1 5

<210> SEQ ID NO 121

<211> LENGTH: 7

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
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<220> FEATURE:

<221> NAME/KEY: MOD_RES

<222> LOCATION: (2)..(2)

<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 121

Pro Xaa Ser Tyr Asn Val Phe
1 5

<210> SEQ ID NO 122

<211> LENGTH: 7

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
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<220> FEATURE:

<221> NAME/KEY: MOD_RES

<222> LOCATION: (1)..(2)

<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 122

Xaa Xaa Ser Tyr Asn Val Phe
1 5

<210> SEQ ID NO 123

<211> LENGTH: 7

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
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<220> FEATURE:

<221> NAME/KEY: MOD_RES

<222> LOCATION: (2)..(2)

<223> OTHER INFORMATION: Any amino acid

<220> FEATURE:

<221> NAME/KEY: MOD_RES

<222> LOCATION: (4)..(4)

<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 123

Pro Xaa Ser Xaa Asn Val Tyr
1 5

<210> SEQ ID NO 124

<211> LENGTH: 7

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
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<220> FEATURE:

<221> NAME/KEY: MOD_RES

<222> LOCATION: (2)..(2)

<223> OTHER INFORMATION: Any amino acid

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<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (5)..(5)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 124

Pro Xaa Ser His Xaa Val Tyr
1 5

<210> SEQ ID NO 125
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<220> FEATURE:
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<222> LOCATION: (2)..(2)
<223> OTHER INFORMATION: Any amino acid
<220> FEATURE:
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<222> LOCATION: (6)..(6)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 125

Pro Xaa Ser His Asn Xaa Tyr
1 5

<210> SEQ ID NO 126
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<212> TYPE: PRT
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<220> FEATURE:
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<221> NAME/KEY: MOD_RES
<222> LOCATION: (2)..(2)
<223> OTHER INFORMATION: Any amino acid
<220> FEATURE:
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<222> LOCATION: (7)..(7)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 126

Pro Xaa Ser His Asn Val Xaa
1 5

<210> SEQ ID NO 127
<211> LENGTH: 5
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (3)..(4)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 127

Phe Asn Xaa Xaa Tyr
1 5

<210> SEQ ID NO 128
<211> LENGTH: 4
<212> TYPE: PRT

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<213> ORGANISM: Artificial Sequence
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<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
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<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (3)..(3)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 128

Phe Asn Xaa Tyr

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<210> SEQ ID NO 129
<211> LENGTH: 3
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide

<400> SEQUENCE: 129

Phe Asn Tyr

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<210> SEQ ID NO 130
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<212> TYPE: PRT
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<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (2)..(2)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 130

Phe Xaa Tyr

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<210> SEQ ID NO 131
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<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
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<221> NAME/KEY: MOD_RES
<222> LOCATION: (1)..(2)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 131

Xaa Xaa Tyr

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<210> SEQ ID NO 132
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<212> TYPE: PRT
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<221> NAME/KEY: MOD_RES
<222> LOCATION: (2)..(2)
<223> OTHER INFORMATION: Any amino acid

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<400> SEQUENCE: 132

Trp Xaa Tyr
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<210> SEQ ID NO 133

<211> LENGTH: 3

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
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<220> FEATURE:

<221> NAME/KEY: MOD_RES

<222> LOCATION: (1)..(1)

<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 133

Xaa Lys Tyr
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<210> SEQ ID NO 134

<211> LENGTH: 5

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
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<220> FEATURE:

<221> NAME/KEY: MOD_RES

<222> LOCATION: (4)..(4)

<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 134

Phe Asn Pro Xaa Tyr
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<210> SEQ ID NO 135

<211> LENGTH: 5

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide

<220> FEATURE:

<221> NAME/KEY: MOD_RES

<222> LOCATION: (3)..(3)

<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 135

Phe Asn Xaa Pro Tyr
1 5

<210> SEQ ID NO 136

<211> LENGTH: 3

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide

<400> SEQUENCE: 136

His Asn Ser
1

<210> SEQ ID NO 137

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<211> LENGTH: 3
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide

<400> SEQUENCE: 137

Gly Gly Ser

1

<210> SEQ ID NO 138
<211> LENGTH: 3
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (1)..(2)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 138

Xaa Xaa Ser

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<210> SEQ ID NO 139
<211> LENGTH: 3
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (1)..(2)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 139

Xaa Xaa Lys

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<210> SEQ ID NO 140
<211> LENGTH: 3
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide

<400> SEQUENCE: 140

Gly Gly Lys

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<210> SEQ ID NO 141
<211> LENGTH: 3
<212> TYPE: PRT
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<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
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<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (2)..(2)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 141

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Pro Xaa Ser

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<210> SEQ ID NO 142
<211> LENGTH: 3
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
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<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (1)..(1)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 142

Xaa Pro Ser

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<210> SEQ ID NO 143
<211> LENGTH: 2
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (2)..(2)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 143

Pro Xaa

1

<210> SEQ ID NO 144
<211> LENGTH: 2
<212> TYPE: PRT
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<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (1)..(1)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 144

Xaa Pro

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<210> SEQ ID NO 145
<211> LENGTH: 4
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (2)..(3)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 145

Ile Xaa Xaa Ser

1

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<210> SEQ ID NO 146
<211> LENGTH: 4
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
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<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
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<220> FEATURE:
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<222> LOCATION: (2)..(2)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 146

Asn Xaa Pro Lys
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<210> SEQ ID NO 147
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<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide

<400> SEQUENCE: 147

Asn Pro Cys Lys
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<210> SEQ ID NO 148
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<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (3)..(3)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 148

Pro Pro Xaa Ser His
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<210> SEQ ID NO 149
<211> LENGTH: 6
<212> TYPE: PRT
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<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
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<222> LOCATION: (3)..(4)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 149

Pro Pro Xaa Xaa Ser His
1 5

<210> SEQ ID NO 150
<211> LENGTH: 6
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide

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<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (4)..(4)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 150

Pro Pro Pro Xaa Ser His
1 5

<210> SEQ ID NO 151
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<212> TYPE: PRT
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<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
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<222> LOCATION: (3)..(3)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 151

Pro Pro Xaa Pro Ser His
1 5

<210> SEQ ID NO 152
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<212> TYPE: PRT
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<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
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<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (1)..(2)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 152

Xaa Xaa Ser His
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<210> SEQ ID NO 153
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<212> TYPE: PRT
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<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (3)..(4)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 153

Pro Pro Xaa Xaa
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<210> SEQ ID NO 154
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<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (3)..(3)
<223> OTHER INFORMATION: Any amino acid

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<400> SEQUENCE: 154

Phe Asn Xaa Lys Asn
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<210> SEQ ID NO 155

<211> LENGTH: 6

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide

<220> FEATURE:

<221> NAME/KEY: MOD_RES

<222> LOCATION: (3)..(4)

<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 155

Phe Asn Xaa Xaa Lys Asn
1 5

<210> SEQ ID NO 156

<211> LENGTH: 6

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide

<220> FEATURE:

<221> NAME/KEY: MOD_RES

<222> LOCATION: (3)..(3)

<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 156

Phe Asn Xaa Pro Lys Asn
1 5

<210> SEQ ID NO 157

<211> LENGTH: 6

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
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<220> FEATURE:

<221> NAME/KEY: MOD_RES

<222> LOCATION: (4)..(4)

<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 157

Phe Asn Pro Xaa Lys Asn
1 5

<210> SEQ ID NO 158

<211> LENGTH: 4

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide

<220> FEATURE:

<221> NAME/KEY: MOD_RES

<222> LOCATION: (3)..(4)

<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 158

Phe Asn Xaa Xaa

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<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide

<400> SEQUENCE: 159

Asn Ala Asp Gly Ser Ser His
1 5

<210> SEQ ID NO 160
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<212> TYPE: PRT
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<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
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<222> LOCATION: (3)..(4)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 160

Ala Asp Xaa Xaa Ser His
1 5

<210> SEQ ID NO 161
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<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
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<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
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<222> LOCATION: (3)..(3)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 161

Ala Asp Xaa Pro Ser His
1 5

<210> SEQ ID NO 162
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<212> TYPE: PRT
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<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
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<221> NAME/KEY: MOD_RES
<222> LOCATION: (4)..(4)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 162

Ala Asp Pro Xaa Ser His
1 5

<210> SEQ ID NO 163
<211> LENGTH: 4
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:

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<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (3)..(4)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 163

Ala Asp Xaa Xaa
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<210> SEQ ID NO 164
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<212> TYPE: PRT
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<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
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<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (3)..(4)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 164

Ala Asp Xaa Xaa Ser His
1 5

<210> SEQ ID NO 165
<211> LENGTH: 4
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide

<400> SEQUENCE: 165

Phe Asn Pro Gly
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<210> SEQ ID NO 166
<211> LENGTH: 6
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (3)..(4)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 166

Phe Asn Xaa Xaa Pro Gly
1 5

<210> SEQ ID NO 167
<211> LENGTH: 4
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (1)..(2)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 167

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Xaa Xaa Pro Gly

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<210> SEQ ID NO 168
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<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (3)..(4)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 168

Phe Asn Xaa Xaa

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<210> SEQ ID NO 169
<211> LENGTH: 4
<212> TYPE: PRT
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<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
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<221> NAME/KEY: MOD_RES
<222> LOCATION: (4)..(4)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 169

Phe Asn Pro Xaa

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<210> SEQ ID NO 170
<211> LENGTH: 6
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (3)..(4)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 170

Lys Tyr Xaa Xaa Ser His

1

5

<210> SEQ ID NO 171
<211> LENGTH: 6
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (4)..(4)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 171

Lys Tyr Pro Xaa Ser His

1

5

-continued

<210> SEQ ID NO 172
<211> LENGTH: 6
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (3)..(3)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 172

Lys Tyr Xaa Pro Ser His
1 5

<210> SEQ ID NO 173
<211> LENGTH: 4
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (2)..(3)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 173

Phe Xaa Xaa Pro
1

<210> SEQ ID NO 174
<211> LENGTH: 4
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (3)..(3)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 174

Phe Asn Xaa Pro
1

<210> SEQ ID NO 175
<211> LENGTH: 4
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (4)..(4)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 175

Phe Asn Pro Xaa
1

<210> SEQ ID NO 176
<211> LENGTH: 606
<212> TYPE: PRT

-continued

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic polypeptide

<400> SEQUENCE: 176

```

Met Glu Thr Asp Thr Leu Leu Leu Trp Val Leu Leu Leu Trp Val Pro
 1           5           10           15
Gly Ser Thr Gly Asp Arg Ser Ala Ala Gly Ser Thr Leu Asp Lys Ile
 20           25           30
Ala Lys Asn Gly Val Ile Val Val Gly His Arg Glu Ser Ser Val Pro
 35           40           45
Phe Ser Tyr Tyr Asp Asn Gln Gln Lys Val Val Gly Tyr Ser Gln Asp
 50           55           60
Tyr Ser Asn Ala Ile Val Glu Ala Val Lys Lys Lys Leu Asn Lys Pro
 65           70           75           80
Asp Leu Gln Val Lys Leu Ile Pro Ile Thr Ser Gln Asn Arg Ile Pro
 85           90           95
Leu Leu Gln Asn Gly Thr Phe Asp Phe Glu Cys Gly Ser Thr Thr Asn
 100          105          110
Asn Val Glu Arg Gln Lys Gln Ala Ala Phe Ser Asp Thr Ile Phe Val
 115          120          125
Val Gly Thr Arg Leu Leu Thr Lys Lys Gly Gly Asp Ile Lys Asp Phe
 130          135          140
Ala Asn Leu Lys Asp Lys Ala Val Val Val Thr Ser Gly Thr Thr Ser
 145          150          155          160
Glu Val Leu Leu Asn Lys Leu Asn Glu Glu Gln Lys Met Asn Met Arg
 165          170          175          175
Ile Ile Ser Ala Lys Asp His Gly Asp Ser Phe Arg Thr Leu Glu Ser
 180          185          190
Gly Arg Ala Val Ala Phe Met Met Asp Asp Val Leu Leu Ala Gly Glu
 195          200          205
Arg Ala Lys Ala Lys Lys Pro Asp Asn Trp Glu Ile Val Gly Lys Pro
 210          215          220
Gln Ser Gln Glu Ala Tyr Gly Cys Met Leu Arg Lys Asp Asp Pro Gln
 225          230          235          240
Phe Lys Lys Leu Met Asp Asp Thr Ile Ala Gln Val Gln Thr Ser Gly
 245          250          255
Glu Ala Glu Lys Trp Phe Asp Lys Trp Phe Lys Asn Pro Ile Leu Val
 260          265          270
Ser His Asn Val Tyr Ile Thr Ala Asp Lys Gln Lys Asn Gly Ile Lys
 275          280          285
Ala Asn Phe Lys Ile Arg His Asn Val Glu Asp Gly Ser Val Gln Leu
 290          295          300
Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu
 305          310          315          320
Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Val Leu Ser Lys Asp
 325          330          335
Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val Thr Ala
 340          345          350
Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Gly Gly Thr Gly
 355          360          365

```

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Gly Ser Met Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile
 370 375 380
 Leu Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Arg
 385 390 400
 Gly Glu Gly Glu Gly Asp Ala Thr Asn Gly Lys Leu Thr Leu Lys Phe
 405 410 415
 Ile Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr
 420 425 430
 Thr Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met
 435 440 445
 Lys Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln
 450 455 460
 Glu Arg Thr Ile Ser Phe Lys Asp Asp Gly Thr Tyr Lys Thr Arg Ala
 465 470 475 480
 Glu Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys
 485 490 495
 Gly Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu
 500 505 510
 Tyr Asn Phe Asn Asn Pro Leu Asn Met Asn Phe Glu Leu Ser Asp Glu
 515 520 525
 Met Lys Ala Leu Phe Lys Glu Pro Asn Asp Lys Ala Leu Lys Leu Gln
 530 535 540
 Val Asp Glu Gln Lys Leu Ile Ser Glu Glu Asp Leu Asn Ala Val Gly
 545 550 555 560
 Gln Asp Thr Gln Glu Val Ile Val Val Pro His Ser Leu Pro Phe Lys
 565 570 575
 Val Val Val Ile Ser Ala Ile Leu Ala Leu Val Val Leu Thr Ile Ile
 580 585 590
 Ser Leu Ile Ile Leu Ile Met Leu Trp Gln Lys Lys Pro Arg
 595 600 605

<210> SEQ ID NO 177

<211> LENGTH: 606

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic polypeptide

<400> SEQUENCE: 177

Met Glu Thr Asp Thr Leu Leu Leu Trp Val Leu Leu Leu Trp Val Pro
 1 5 10 15
 Gly Ser Thr Gly Asp Arg Ser Ala Ala Gly Ser Thr Leu Asp Lys Ile
 20 25 30
 Ala Lys Asn Gly Val Ile Val Val Gly His Arg Glu Ser Ser Val Pro
 35 40 45
 Phe Ser Tyr Tyr Asp Asn Gln Gln Lys Val Val Gly Tyr Ser Gln Asp
 50 55 60
 Tyr Ser Asn Ala Ile Val Glu Ala Val Lys Lys Lys Leu Asn Lys Pro
 65 70 75 80
 Asp Leu Gln Val Lys Leu Ile Pro Ile Thr Ser Gln Asn Arg Ile Pro
 85 90 95
 Leu Leu Gln Asn Gly Thr Phe Asp Phe Glu Cys Gly Ser Thr Thr Asn
 100 105 110

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Asn	Val	Glu	Arg	Gln	Lys	Gln	Ala	Ala	Phe	Ser	Asp	Thr	Ile	Phe	Val
	115						120					125			
Val	Gly	Thr	Arg	Leu	Leu	Thr	Lys	Lys	Gly	Gly	Asp	Ile	Lys	Asp	Phe
	130					135					140				
Ala	Asn	Leu	Lys	Asp	Lys	Ala	Val	Val	Val	Thr	Ser	Gly	Thr	Thr	Ser
145					150						155				160
Glu	Val	Leu	Leu	Asn	Lys	Leu	Asn	Glu	Glu	Gln	Lys	Met	Asn	Met	Arg
				165						170					175
Ile	Ile	Ser	Ala	Lys	Asp	His	Gly	Asp	Ser	Phe	Arg	Thr	Leu	Glu	Ser
			180					185						190	
Gly	Arg	Ala	Val	Ala	Phe	Met	Met	Asp	Asp	Ser	Leu	Leu	Ala	Gly	Glu
		195						200						205	
Arg	Ala	Lys	Ala	Lys	Lys	Pro	Asp	Asn	Trp	Glu	Ile	Val	Gly	Lys	Pro
	210					215						220			
Gln	Ser	Gln	Glu	Ala	Tyr	Gly	Cys	Met	Leu	Arg	Lys	Asp	Asp	Pro	Gln
225					230						235				240
Phe	Lys	Lys	Leu	Met	Asp	Asp	Thr	Ile	Ala	Gln	Val	Gln	Thr	Ser	Gly
				245					250						255
Glu	Ala	Glu	Lys	Trp	Phe	Asp	Lys	Trp	Phe	Lys	Asn	Pro	Ile	Leu	Val
			260					265						270	
Ser	His	Asn	Val	Tyr	Ile	Thr	Ala	Asp	Lys	Gln	Lys	Asn	Gly	Ile	Lys
		275						280					285		
Ala	Asn	Phe	Lys	Ile	Arg	His	Asn	Val	Glu	Asp	Gly	Ser	Val	Gln	Leu
	290					295					300				
Ala	Asp	His	Tyr	Gln	Gln	Asn	Thr	Pro	Ile	Gly	Asp	Gly	Pro	Val	Leu
305					310					315					320
Leu	Pro	Asp	Asn	His	Tyr	Leu	Ser	Thr	Gln	Ser	Val	Leu	Ser	Lys	Asp
				325					330						335
Pro	Asn	Glu	Lys	Arg	Asp	His	Met	Val	Leu	Leu	Glu	Phe	Val	Thr	Ala
			340					345						350	
Ala	Gly	Ile	Thr	Leu	Gly	Met	Asp	Glu	Leu	Tyr	Lys	Gly	Gly	Thr	Gly
		355					360					365			
Gly	Ser	Met	Ser	Lys	Gly	Glu	Glu	Leu	Phe	Thr	Gly	Val	Val	Pro	Ile
	370					375					380				
Leu	Val	Glu	Leu	Asp	Gly	Asp	Val	Asn	Gly	His	Lys	Phe	Ser	Val	Arg
385					390						395				400
Gly	Glu	Gly	Glu	Gly	Asp	Ala	Thr	Asn	Gly	Lys	Leu	Thr	Leu	Lys	Phe
				405					410						415
Ile	Cys	Thr	Thr	Gly	Lys	Leu	Pro	Val	Pro	Trp	Pro	Thr	Leu	Val	Thr
		420						425						430	
Thr	Leu	Thr	Tyr	Gly	Val	Gln	Cys	Phe	Ser	Arg	Tyr	Pro	Asp	His	Met
		435						440					445		
Lys	Gln	His	Asp	Phe	Phe	Lys	Ser	Ala	Met	Pro	Glu	Gly	Tyr	Val	Gln
		450				455						460			
Glu	Arg	Thr	Ile	Ser	Phe	Lys	Asp	Asp	Gly	Thr	Tyr	Lys	Thr	Arg	Ala
465					470					475					480
Glu	Val	Lys	Phe	Glu	Gly	Asp	Thr	Leu	Val	Asn	Arg	Ile	Glu	Leu	Lys
				485					490						495
Gly	Ile	Asp	Phe	Lys	Glu	Asp	Gly	Asn	Ile	Leu	Gly	His	Lys	Leu	Glu
			500					505							510

-continued

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Tyr Asn Phe Asn Asn Pro Leu Asn Met Asn Phe Glu Leu Ser Asp Glu
    515                               520                               525

Met Lys Ala Leu Phe Lys Glu Pro Asn Asp Lys Ala Leu Lys Leu Gln
    530                               535                               540

Val Asp Glu Gln Lys Leu Ile Ser Glu Glu Asp Leu Asn Ala Val Gly
    545                               550                               555                               560

Gln Asp Thr Gln Glu Val Ile Val Val Pro His Ser Leu Pro Phe Lys
    565                               570                               575

Val Val Val Ile Ser Ala Ile Leu Ala Leu Val Val Leu Thr Ile Ile
    580                               585                               590

Ser Leu Ile Ile Leu Ile Met Leu Trp Gln Lys Lys Pro Arg
    595                               600                               605

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<210> SEQ ID NO 178
<211> LENGTH: 606
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
                             polypeptide

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<400> SEQUENCE: 178

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Met Glu Thr Asp Thr Leu Leu Leu Trp Val Leu Leu Leu Trp Val Pro
 1      5      10      15

Gly Ser Thr Gly Asp Arg Ser Ala Ala Gly Ser Thr Leu Asp Lys Ile
 20     25     30

Ala Lys Asn Gly Val Ile Val Val Gly His Arg Glu Ser Ser Val Pro
 35     40     45

Phe Ser Tyr Tyr Asp Asn Gln Gln Lys Val Val Gly Tyr Ser Gln Asp
 50     55     60

Tyr Ser Asn Ala Ile Val Glu Ala Val Lys Lys Lys Leu Asn Lys Pro
 65     70     75     80

Asp Leu Gln Val Lys Leu Ile Pro Ile Thr Ala Gln Asn Arg Ile Pro
 85     90     95

Leu Leu Gln Asn Gly Thr Phe Asp Phe Glu Cys Gly Ser Thr Thr Asn
100    105    110

Asn Val Glu Arg Gln Lys Gln Ala Ala Phe Ser Asp Thr Ile Phe Val
115    120    125

Val Gly Thr Arg Leu Leu Thr Lys Lys Gly Gly Asp Ile Lys Asp Phe
130    135    140

Ala Asn Leu Lys Asp Lys Ala Val Val Val Thr Ser Gly Thr Thr Ser
145    150    155    160

Glu Val Leu Leu Asn Lys Leu Asn Glu Glu Gln Lys Met Asn Met Arg
165    170    175

Ile Ile Ser Ala Lys Asp His Gly Asp Ser Phe Arg Thr Leu Glu Ser
180    185    190

Gly Arg Ala Val Ala Phe Met Met Asp Asp Val Leu Leu Ala Gly Glu
195    200    205

Arg Ala Lys Ala Lys Lys Pro Asp Asn Trp Glu Ile Val Gly Lys Pro
210    215    220

Gln Ser Gln Glu Ala Tyr Gly Cys Met Leu Arg Lys Asp Asp Pro Gln
225    230    235    240

Phe Lys Lys Leu Met Asp Asp Thr Ile Ala Gln Val Gln Thr Ser Gly
245    250    255

```

-continued

Glu	Ala	Glu	Lys	Trp	Phe	Asp	Lys	Trp	Phe	Lys	Asn	Pro	Ile	Leu	Val							
			260					265					270									
Ser	His	Asn	Val	Tyr	Ile	Thr	Ala	Asp	Lys	Gln	Lys	Asn	Gly	Ile	Lys							
		275					280					285										
Ala	Asn	Phe	Lys	Ile	Arg	His	Asn	Val	Glu	Asp	Gly	Ser	Val	Gln	Leu							
	290				295						300											
Ala	Asp	His	Tyr	Gln	Gln	Asn	Thr	Pro	Ile	Gly	Asp	Gly	Pro	Val	Leu							
305					310					315					320							
Leu	Pro	Asp	Asn	His	Tyr	Leu	Ser	Thr	Gln	Ser	Val	Leu	Ser	Lys	Asp							
				325					330					335								
Pro	Asn	Glu	Lys	Arg	Asp	His	Met	Val	Leu	Leu	Glu	Phe	Val	Thr	Ala							
			340					345					350									
Ala	Gly	Ile	Thr	Leu	Gly	Met	Asp	Glu	Leu	Tyr	Lys	Gly	Gly	Thr	Gly							
		355				360						365										
Gly	Ser	Met	Ser	Lys	Gly	Glu	Glu	Leu	Phe	Thr	Gly	Val	Val	Pro	Ile							
370					375						380											
Leu	Val	Glu	Leu	Asp	Gly	Asp	Val	Asn	Gly	His	Lys	Phe	Ser	Val	Arg							
385					390					395					400							
Gly	Glu	Gly	Glu	Gly	Asp	Ala	Thr	Asn	Gly	Lys	Leu	Thr	Leu	Lys	Phe							
				405					410					415								
Ile	Cys	Thr	Thr	Gly	Lys	Leu	Pro	Val	Pro	Trp	Pro	Thr	Leu	Val	Thr							
			420					425					430									
Thr	Leu	Thr	Tyr	Gly	Val	Gln	Cys	Phe	Ser	Arg	Tyr	Pro	Asp	His	Met							
	435					440						445										
Lys	Gln	His	Asp	Phe	Phe	Lys	Ser	Ala	Met	Pro	Glu	Gly	Tyr	Val	Gln							
	450					455					460											
Glu	Arg	Thr	Ile	Ser	Phe	Lys	Asp	Asp	Gly	Thr	Tyr	Lys	Thr	Arg	Ala							
465					470					475					480							
Glu	Val	Lys	Phe	Glu	Gly	Asp	Thr	Leu	Val	Asn	Arg	Ile	Glu	Leu	Lys							
			485					490						495								
Gly	Ile	Asp	Phe	Lys	Glu	Asp	Gly	Asn	Ile	Leu	Gly	His	Lys	Leu	Glu							
			500				505						510									
Tyr	Asn	Phe	Asn	Asn	Pro	Leu	Asn	Met	Asn	Phe	Glu	Leu	Ser	Asp	Glu							
	515					520						525										
Met	Lys	Ala	Leu	Phe	Lys	Glu	Pro	Asn	Asp	Lys	Ala	Leu	Lys	Leu	Gln							
	530					535					540											
Val	Asp	Glu	Gln	Lys	Leu	Ile	Ser	Glu	Glu	Asp	Leu	Asn	Ala	Val	Gly							
545					550					555					560							
Gln	Asp	Thr	Gln	Glu	Val	Ile	Val	Val	Pro	His	Ser	Leu	Pro	Phe	Lys							
			565					570						575								
Val	Val	Val	Ile	Ser	Ala	Ile	Leu	Ala	Leu	Val	Val	Leu	Thr	Ile	Ile							
			580				585						590									
Ser	Leu	Ile	Ile	Leu	Ile	Met	Leu	Trp	Gln	Lys	Lys	Pro	Arg									
	595					600						605										

<210> SEQ ID NO 179
 <211> LENGTH: 606
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic polypeptide

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<400> SEQUENCE: 179

Met Glu Thr Asp Thr Leu Leu Leu Trp Val Leu Leu Leu Trp Val Pro
1 5 10 15

Gly Ser Thr Gly Asp Arg Ser Ala Ala Gly Ser Thr Leu Asp Lys Ile
20 25 30

Ala Lys Asn Gly Val Ile Val Val Gly His Arg Glu Ser Ser Val Pro
35 40 45

Phe Ser Tyr Tyr Asp Asn Gln Gln Lys Val Val Gly Tyr Ser Gln Asp
50 55 60

Tyr Ser Asn Ala Ile Val Glu Ala Val Lys Lys Lys Leu Asn Lys Pro
65 70 75 80

Asp Leu Gln Val Lys Leu Ile Pro Ile Thr Ser Gln Asn Arg Ile Pro
85 90 95

Leu Leu Gln Asn Gly Thr Phe Asp Phe Glu Cys Gly Ser Thr Thr Asn
100 105 110

Asn Val Glu Arg Gln Lys Gln Ala Ala Phe Ser Asp Thr Ile Phe Val
115 120 125

Val Gly Thr Arg Leu Leu Thr Lys Lys Gly Gly Asp Ile Lys Asp Phe
130 135 140

Ala Asn Leu Lys Asp Lys Ala Val Val Val Thr Ser Gly Thr Thr Ser
145 150 155 160

Glu Val Leu Leu Asn Lys Leu Asn Glu Glu Gln Lys Met Asn Met Arg
165 170 175

Ile Ile Ser Ala Lys Asp His Gly Asp Ser Phe Arg Thr Leu Glu Ser
180 185 190

Gly Arg Ala Val Ala Phe Met Met Asp Asp Val Leu Leu Ala Gly Glu
195 200 205

Arg Ala Lys Ala Lys Lys Pro Asp Asn Trp Glu Ile Val Gly Lys Pro
210 215 220

Gln Ser Gln Glu Ala Tyr Gly Cys Met Leu Arg Lys Asp Asp Pro Gln
225 230 235 240

Phe Lys Lys Leu Met Asp Asp Thr Ile Ala Gln Val Gln Thr Ser Gly
245 250 255

Glu Ala Glu Lys Trp Phe Asp Lys Trp Phe Lys Asn Pro Ile Leu Val
260 265 270

Ser His Asn Val Tyr Ile Thr Ala Asp Lys Gln Lys Asn Gly Ile Lys
275 280 285

Ala Asn Phe Lys Ile Arg His Asn Val Glu Asp Gly Ser Val Gln Leu
290 295 300

Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu
305 310 315 320

Leu Pro Asp Asn His Tyr Leu Ser Tyr Gln Ser Val Leu Ser Lys Asp
325 330 335

Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val Thr Ala
340 345 350

Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Gly Gly Thr Gly
355 360 365

Gly Ser Met Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile
370 375 380

Leu Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Arg
385 390 395 400

-continued

Gly Glu Gly Glu Gly Asp Ala Thr Asn Gly Lys Leu Thr Leu Lys Leu
 405 410 415
 Ile Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr
 420 425 430
 Thr Leu Gly Tyr Gly Val Gln Cys Phe Ala Arg Tyr Pro Asp His Met
 435 440 445
 Lys Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln
 450 455 460
 Glu Arg Thr Ile Ser Phe Lys Asp Asp Gly Thr Tyr Lys Thr Arg Ala
 465 470 475 480
 Glu Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys
 485 490 495
 Gly Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu
 500 505 510
 Tyr Asn Phe Asn Asn Pro Leu Asn Met Asn Phe Glu Leu Ser Asp Glu
 515 520 525
 Met Lys Ala Leu Phe Lys Glu Pro Asn Asp Lys Ala Leu Lys Leu Gln
 530 535 540
 Val Asp Glu Gln Lys Leu Ile Ser Glu Glu Asp Leu Asn Ala Val Gly
 545 550 555 560
 Gln Asp Thr Gln Glu Val Ile Val Val Pro His Ser Leu Pro Phe Lys
 565 570 575
 Val Val Val Ile Ser Ala Ile Leu Ala Leu Val Val Leu Thr Ile Ile
 580 585 590
 Ser Leu Ile Ile Leu Ile Met Leu Trp Gln Lys Lys Pro Arg
 595 600 605

<210> SEQ ID NO 180

<211> LENGTH: 606

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic polypeptide

<400> SEQUENCE: 180

Met Glu Thr Asp Thr Leu Leu Leu Trp Val Leu Leu Leu Trp Val Pro
 1 5 10 15
 Gly Ser Thr Gly Asp Arg Ser Ala Ala Gly Ser Thr Leu Asp Lys Ile
 20 25 30
 Ala Lys Asn Gly Val Ile Val Val Gly His Arg Glu Ser Ser Val Pro
 35 40 45
 Phe Ser Tyr Tyr Asp Asn Gln Gln Lys Val Val Gly Tyr Ser Gln Asp
 50 55 60
 Tyr Ser Asn Ala Ile Val Glu Ala Val Lys Lys Lys Leu Asn Lys Pro
 65 70 75 80
 Asp Leu Gln Val Lys Leu Ile Pro Ile Thr Ala Gln Asn Arg Ile Pro
 85 90 95
 Leu Leu Gln Asn Gly Thr Phe Asp Phe Glu Cys Gly Ser Thr Thr Asn
 100 105 110
 Asn Val Glu Arg Gln Lys Gln Ala Ala Phe Ser Asp Thr Ile Phe Val
 115 120 125
 Val Gly Thr Arg Leu Leu Thr Lys Lys Gly Gly Asp Ile Lys Asp Phe

-continued

130			135			140									
Ala	Asn	Leu	Lys	Asp	Lys	Ala	Val	Val	Val	Thr	Ser	Gly	Thr	Thr	Ser
145					150						155				160
Glu	Val	Leu	Leu	Asn	Lys	Leu	Asn	Glu	Glu	Gln	Lys	Met	Asn	Met	Arg
				165						170					175
Ile	Ile	Ser	Ala	Lys	Asp	His	Gly	Asp	Ser	Phe	Arg	Thr	Leu	Glu	Ser
			180					185						190	
Gly	Arg	Ala	Val	Ala	Phe	Met	Met	Asp	Asp	Ser	Leu	Leu	Ala	Gly	Glu
		195						200						205	
Arg	Ala	Lys	Ala	Lys	Lys	Pro	Asp	Asn	Trp	Glu	Ile	Val	Gly	Lys	Pro
	210						215				220				
Gln	Ser	Gln	Glu	Ala	Tyr	Gly	Cys	Met	Leu	Arg	Lys	Asp	Asp	Pro	Gln
	225				230						235				240
Phe	Lys	Lys	Leu	Met	Asp	Asp	Thr	Ile	Ala	Gln	Val	Gln	Thr	Ser	Gly
				245						250					255
Glu	Ala	Glu	Lys	Trp	Phe	Asp	Lys	Trp	Phe	Lys	Asn	Pro	Ile	Leu	Val
			260					265						270	
Ser	His	Asn	Val	Tyr	Ile	Thr	Ala	Asp	Lys	Gln	Lys	Asn	Gly	Ile	Lys
		275						280						285	
Ala	Asn	Phe	Lys	Ile	Arg	His	Asn	Val	Glu	Asp	Gly	Ser	Val	Gln	Leu
	290						295				300				
Ala	Asp	His	Tyr	Gln	Gln	Asn	Thr	Pro	Ile	Gly	Asp	Gly	Pro	Val	Leu
	305				310						315				320
Leu	Pro	Asp	Asn	His	Tyr	Leu	Ser	Tyr	Gln	Ser	Val	Leu	Ser	Lys	Asp
				325						330					335
Pro	Asn	Glu	Lys	Arg	Asp	His	Met	Val	Leu	Leu	Glu	Phe	Val	Thr	Ala
			340					345						350	
Ala	Gly	Ile	Thr	Leu	Gly	Met	Asp	Glu	Leu	Tyr	Lys	Gly	Gly	Thr	Gly
		355						360						365	
Gly	Ser	Met	Ser	Lys	Gly	Glu	Glu	Leu	Phe	Thr	Gly	Val	Val	Pro	Ile
	370				375						380				
Leu	Val	Glu	Leu	Asp	Gly	Asp	Val	Asn	Gly	His	Lys	Phe	Ser	Val	Arg
	385				390						395				400
Gly	Glu	Gly	Glu	Gly	Asp	Ala	Thr	Asn	Gly	Lys	Leu	Thr	Leu	Lys	Leu
				405						410					415
Ile	Cys	Thr	Thr	Gly	Lys	Leu	Pro	Val	Pro	Trp	Pro	Thr	Leu	Val	Thr
			420					425						430	
Thr	Leu	Gly	Tyr	Gly	Val	Gln	Cys	Phe	Ala	Arg	Tyr	Pro	Asp	His	Met
		435						440						445	
Lys	Gln	His	Asp	Phe	Phe	Lys	Ser	Ala	Met	Pro	Glu	Gly	Tyr	Val	Gln
	450							455						460	
Glu	Arg	Thr	Ile	Ser	Phe	Lys	Asp	Asp	Gly	Thr	Tyr	Lys	Thr	Arg	Ala
	465				470						475				480
Glu	Val	Lys	Phe	Glu	Gly	Asp	Thr	Leu	Val	Asn	Arg	Ile	Glu	Leu	Lys
				485						490					495
Gly	Ile	Asp	Phe	Lys	Glu	Asp	Gly	Asn	Ile	Leu	Gly	His	Lys	Leu	Glu
				500				505						510	
Tyr	Asn	Phe	Asn	Asn	Pro	Leu	Asn	Met	Asn	Phe	Glu	Leu	Ser	Asp	Glu
		515						520						525	
Met	Lys	Ala	Leu	Phe	Lys	Glu	Pro	Asn	Asp	Lys	Ala	Leu	Lys	Leu	Gln
	530							535							540

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Val Asp Glu Gln Lys Leu Ile Ser Glu Glu Asp Leu Asn Ala Val Gly
545 550 555 560

Gln Asp Thr Gln Glu Val Ile Val Val Pro His Ser Leu Pro Phe Lys
565 570 575

Val Val Val Ile Ser Ala Ile Leu Ala Leu Val Val Leu Thr Ile Ile
580 585 590

Ser Leu Ile Ile Leu Ile Met Leu Trp Gln Lys Lys Pro Arg
595 600 605

<210> SEQ ID NO 181
 <211> LENGTH: 606
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
 polypeptide

<400> SEQUENCE: 181

Met Glu Thr Asp Thr Leu Leu Leu Trp Val Leu Leu Leu Trp Val Pro
1 5 10 15

Gly Ser Thr Gly Asp Arg Ser Ala Ala Gly Ser Thr Leu Asp Lys Ile
20 25 30

Ala Lys Asn Gly Val Ile Val Val Gly His Arg Glu Ser Ser Val Pro
35 40 45

Phe Ser Tyr Tyr Asp Asn Gln Gln Lys Val Val Gly Tyr Ser Gln Asp
50 55 60

Tyr Ser Asn Ala Ile Val Glu Ala Val Lys Lys Lys Leu Asn Lys Pro
65 70 75 80

Asp Leu Gln Val Lys Leu Ile Pro Ile Thr Ser Gln Asn Arg Ile Pro
85 90 95

Leu Leu Gln Asn Gly Thr Phe Asp Phe Glu Cys Gly Ser Thr Thr Asn
100 105 110

Asn Val Glu Arg Gln Lys Gln Ala Ala Phe Ser Asp Thr Ile Phe Val
115 120 125

Val Gly Thr Arg Leu Leu Thr Lys Lys Gly Gly Asp Ile Lys Asp Phe
130 135 140

Ala Asn Leu Lys Asp Lys Ala Val Val Val Thr Ser Gly Thr Thr Ser
145 150 155 160

Glu Val Leu Leu Asn Lys Leu Asn Glu Glu Gln Lys Met Asn Met Arg
165 170 175

Ile Ile Ser Ala Lys Asp His Gly Asp Ser Phe Arg Thr Leu Glu Ser
180 185 190

Gly Arg Ala Val Ala Phe Met Met Asp Asp Val Leu Leu Ala Gly Glu
195 200 205

Arg Ala Lys Ala Lys Lys Pro Asp Asn Trp Glu Ile Val Gly Lys Pro
210 215 220

Gln Ser Gln Glu Ala Tyr Gly Cys Met Leu Arg Lys Asp Asp Pro Gln
225 230 235 240

Phe Lys Lys Leu Met Asp Asp Thr Ile Ala Gln Val Gln Thr Ser Gly
245 250 255

Glu Ala Glu Lys Trp Phe Asp Lys Trp Phe Lys Asn Pro Ile Leu Val
260 265 270

Ser His Asn Val Tyr Ile Thr Ala Asp Lys Gln Lys Asn Gly Ile Lys

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275	280	285																		
Ala Asn Phe Lys Ile Arg His Asn Val Glu Asp Gly Ser Val Gln Leu																				
290					295					300										
Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu																				
305				310						315										320
Leu Pro Asp Asn His Tyr Leu Ser Tyr Gln Ser Val Leu Ser Lys Asp																				
			325							330										335
Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val Thr Ala																				
		340								345										350
Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Gly Gly Thr Gly																				
		355								360										365
Gly Ser Met Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile																				
		370								375										380
Leu Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Arg																				
		385								390										395
Gly Glu Gly Glu Gly Asp Ala Thr Asn Gly Lys Leu Thr Leu Lys Leu																				
				405						410										415
Ile Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr																				
				420						425										430
Thr Leu Gly Tyr Gly Val Gln Cys Phe Ala Arg Tyr Pro Asp His Met																				
		435								440										445
Lys Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln																				
		450								455										460
Glu Arg Thr Ile Ser Phe Lys Asp Asp Gly Thr Tyr Lys Thr Arg Ala																				
		465								470										475
Glu Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys																				
				485						490										495
Gly Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu																				
				500						505										510
Tyr Asn Phe Asn Asn Pro Leu Asn Met Asn Phe Glu Leu Ser Asp Glu																				
		515								520										525
Met Lys Ala Leu Phe Lys Glu Pro Asn Asp Lys Ala Leu Lys Leu Gln																				
		530								535										540
Val Asp Glu Gln Lys Leu Ile Ser Glu Glu Asp Leu Asn Ala Val Gly																				
		545								550										555
Gln Asp Thr Gln Glu Val Ile Val Val Pro His Ser Leu Pro Phe Lys																				
				565						570										575
Val Val Val Ile Ser Ala Ile Leu Ala Leu Val Val Leu Thr Ile Ile																				
				580						585										590
Ser Leu Ile Ile Leu Ile Met Leu Trp Gln Lys Lys Pro Arg																				
		595								600										605

<210> SEQ ID NO 182
 <211> LENGTH: 606
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic polypeptide

<400> SEQUENCE: 182
 Met Glu Thr Asp Thr Leu Leu Leu Trp Val Leu Leu Leu Trp Val Pro
 1 5 10 15

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Gly	Ser	Thr	Gly	Asp	Arg	Ser	Ala	Ala	Gly	Ser	Thr	Leu	Asp	Lys	Ile
			20					25					30		
Ala	Lys	Asn	Gly	Val	Ile	Val	Val	Gly	His	Arg	Glu	Ser	Ser	Val	Pro
		35					40					45			
Phe	Ser	Tyr	Tyr	Asp	Asn	Gln	Gln	Lys	Val	Val	Gly	Tyr	Ser	Gln	Asp
	50					55					60				
Tyr	Ser	Asn	Ala	Ile	Val	Glu	Ala	Val	Lys	Lys	Lys	Leu	Asn	Lys	Pro
	65				70					75					80
Asp	Leu	Gln	Val	Lys	Leu	Ile	Pro	Ile	Thr	Ser	Gln	Asn	Arg	Ile	Pro
				85					90					95	
Leu	Leu	Gln	Asn	Gly	Thr	Phe	Asp	Phe	Glu	Cys	Gly	Ser	Thr	Thr	Asn
			100					105					110		
Asn	Val	Glu	Arg	Gln	Lys	Gln	Ala	Ala	Phe	Ser	Asp	Thr	Ile	Phe	Val
		115					120					125			
Val	Gly	Thr	Arg	Leu	Leu	Thr	Lys	Lys	Gly	Gly	Asp	Ile	Lys	Asp	Phe
	130					135					140				
Ala	Asn	Leu	Lys	Asp	Lys	Ala	Val	Val	Val	Thr	Ser	Gly	Thr	Thr	Ser
	145				150					155					160
Glu	Val	Leu	Leu	Asn	Lys	Leu	Asn	Glu	Glu	Gln	Lys	Met	Asn	Met	Arg
				165					170					175	
Ile	Ile	Ser	Ala	Lys	Asp	His	Gly	Asp	Ser	Phe	Arg	Thr	Leu	Glu	Ser
			180					185					190		
Gly	Arg	Ala	Val	Ala	Phe	Met	Met	Asp	Asp	Val	Leu	Leu	Ala	Gly	Glu
		195					200					205			
Arg	Ala	Lys	Ala	Lys	Lys	Pro	Asp	Asn	Trp	Glu	Ile	Val	Gly	Lys	Pro
	210					215					220				
Gln	Ser	Gln	Glu	Ala	Tyr	Gly	Cys	Met	Leu	Arg	Lys	Asp	Asp	Pro	Gln
	225				230					235					240
Phe	Lys	Lys	Leu	Met	Asp	Asp	Thr	Ile	Ala	Gln	Val	Gln	Thr	Ser	Gly
				245					250					255	
Glu	Ala	Glu	Lys	Trp	Phe	Asp	Lys	Trp	Phe	Lys	Asn	Pro	Ile	Leu	Gly
			260					265					270		
Tyr	His	Asn	Ile	Tyr	Ile	Thr	Ala	Asp	Lys	Gln	Lys	Asn	Gly	Ile	Lys
		275					280					285			
Ala	Asn	Phe	Lys	Ile	Arg	His	Asn	Val	Glu	Asp	Gly	Ser	Val	Gln	Leu
	290					295					300				
Ala	Asp	His	Tyr	Gln	Gln	Asn	Thr	Pro	Ile	Gly	Asp	Gly	Pro	Val	Leu
	305				310					315					320
Leu	Pro	Asp	Asn	His	Tyr	Leu	Ser	Thr	Gln	Ser	Val	Leu	Ser	Lys	Asp
				325					330					335	
Pro	Asn	Glu	Lys	Arg	Asp	His	Met	Val	Leu	Leu	Glu	Phe	Arg	Thr	Ala
			340					345					350		
Ala	Gly	Ile	Thr	Leu	Gly	Met	Asp	Glu	Leu	Tyr	Lys	Gly	Gly	Thr	Gly
		355					360					365			
Gly	Ser	Met	Ser	Lys	Gly	Glu	Glu	Leu	Phe	Thr	Gly	Val	Val	Pro	Ile
	370					375					380				
Leu	Val	Glu	Leu	Asp	Gly	Asp	Val	Asn	Gly	His	Lys	Phe	Ser	Val	Arg
	385				390					395					400
Gly	Glu	Gly	Glu	Gly	Asp	Ala	Thr	Asn	Gly	Lys	Leu	Thr	Leu	Lys	Phe
				405					410					415	
Ile	Cys	Thr	Thr	Gly	Lys	Leu	Pro	Val	Pro	Trp	Pro	Thr	Leu	Val	Thr

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420					425					430					
Thr	Leu	Ser	His	Gly	Val	Gln	Cys	Phe	Ser	Arg	Tyr	Pro	Asp	His	Met
	435						440					445			
Lys	Gln	His	Asp	Phe	Phe	Lys	Ser	Ala	Met	Pro	Glu	Gly	Tyr	Val	Gln
	450					455					460				
Glu	Arg	Thr	Ile	Ser	Phe	Lys	Asp	Asp	Gly	Thr	Tyr	Lys	Thr	Arg	Ala
	465					470					475				480
Glu	Val	Lys	Phe	Glu	Gly	Asp	Thr	Leu	Val	Asn	Arg	Ile	Glu	Leu	Lys
			485					490						495	
Gly	Ile	Asp	Phe	Lys	Glu	Asp	Gly	Asn	Ile	Leu	Gly	His	Lys	Leu	Glu
			500					505					510		
Tyr	Asn	Phe	Asn	Glu	Gln	Leu	Asn	Met	Asn	Phe	Glu	Leu	Ser	Asp	Glu
		515					520					525			
Met	Lys	Ala	Leu	Phe	Lys	Glu	Pro	Asn	Asp	Lys	Ala	Leu	Lys	Leu	Gln
	530					535					540				
Val	Asp	Glu	Gln	Lys	Leu	Ile	Ser	Glu	Glu	Asp	Leu	Asn	Ala	Val	Gly
	545					550					555				560
Gln	Asp	Thr	Gln	Glu	Val	Ile	Val	Val	Pro	His	Ser	Leu	Pro	Phe	Lys
				565					570					575	
Val	Val	Val	Ile	Ser	Ala	Ile	Leu	Ala	Leu	Val	Val	Leu	Thr	Ile	Ile
			580					585					590		
Ser	Leu	Ile	Ile	Leu	Ile	Met	Leu	Trp	Gln	Lys	Lys	Pro	Arg		
		595					600					605			

<210> SEQ ID NO 183

<211> LENGTH: 728

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic polypeptide

<400> SEQUENCE: 183

Met	Glu	Thr	Asp	Thr	Leu	Leu	Leu	Trp	Val	Leu	Leu	Leu	Trp	Val	Pro
1				5					10					15	
Gly	Ser	Thr	Gly	Asp	Arg	Ser	Lys	Leu	Glu	Ile	Phe	Ser	Trp	Trp	Ala
			20				25						30		
Gly	Asp	Glu	Gly	Pro	Ala	Leu	Glu	Ala	Leu	Ile	Arg	Leu	Tyr	Lys	Gln
		35				40						45			
Lys	Tyr	Pro	Gly	Val	Glu	Val	Ile	Asn	Ala	Thr	Val	Thr	Gly	Gly	Ala
	50					55						60			
Gly	Val	Asn	Ala	Arg	Ala	Val	Leu	Lys	Thr	Arg	Met	Leu	Gly	Gly	Asp
	65			70							75				80
Pro	Pro	Asp	Thr	Phe	Gln	Val	Ala	Ala	Gly	Met	Glu	Leu	Ile	Gly	Thr
				85					90					95	
Trp	Val	Val	Ala	Asn	Arg	Met	Glu	Asp	Leu	Ser	Ala	Leu	Phe	Arg	Gln
			100					105					110		
Glu	Gly	Trp	Leu	Gln	Ala	Phe	Pro	Lys	Gly	Leu	Ile	Asp	Leu	Ile	Ser
			115					120					125		
Tyr	Lys	Gly	Gly	Ile	Trp	Ser	Val	Pro	Val	Asn	Ile	His	Arg	Ser	Asn
	130					135						140			
Val	Met	Trp	Tyr	Leu	Pro	Ala	Lys	Leu	Lys	Glu	Trp	Gly	Val	Asn	Pro
	145				150						155				160

-continued

Pro	Arg	Thr	Trp	Asp	Glu	Phe	Leu	Ala	Thr	Cys	Gln	Thr	Leu	Lys	Gln
				165					170					175	
Lys	Gly	Leu	Glu	Ala	Pro	Leu	Ala	Leu	Gly	Glu	Asn	Trp	Thr	Gln	Gln
		180						185					190		
His	Leu	Trp	Glu	Ser	Val	Ala	Leu	Ala	Val	Leu	Gly	Pro	Asp	Asp	Trp
		195					200					205			
Asn	Asn	Leu	Trp	Asn	Gly	Lys	Leu	Lys	Phe	Thr	Asp	Pro	Lys	Ala	Val
	210					215					220				
Arg	Ala	Trp	Glu	Val	Phe	Gly	Arg	Val	Leu	Asp	Cys	Ala	Asn	Lys	Asp
	225				230					235					240
Ala	Ala	Gly	Leu	Ser	Trp	Gln	Gln	Ala	Val	Asp	Arg	Val	Val	Gln	Gly
				245					250					255	
Lys	Ala	Ala	Phe	Asn	Val	Met	Gly	Asp	Trp	Ala	Ala	Gly	Tyr	Met	Thr
			260					265					270		
Thr	Thr	Leu	Lys	Leu	Lys	Pro	Gly	Thr	Asp	Phe	Ala	Trp	Ala	Pro	Ser
		275					280					285			
Pro	Gly	Thr	Gln	Gly	Val	Phe	Met	Met	Leu	Ser	Asp	Ser	Phe	Gly	Leu
	290					295					300				
Pro	Lys	Gly	Ala	Lys	Asn	Arg	Gln	Asn	Ala	Ile	Asn	Trp	Leu	Arg	Leu
	305				310					315					320
Val	Gly	Ser	Lys	Glu	Gly	Gln	Asp	Thr	Phe	Asn	Pro	Leu	Lys	Gly	Ser
				325					330					335	
Ile	Ala	Ala	Arg	Leu	Asp	Ser	Asp	Pro	Ser	Lys	Tyr	Pro	Ala	Ser	His
			340					345					350		
Asn	Val	Tyr	Ile	Thr	Ala	Asp	Lys	Gln	Lys	Asn	Gly	Ile	Lys	Ala	Asn
		355					360					365			
Phe	Lys	Ile	Arg	His	Asn	Val	Glu	Asp	Gly	Ser	Val	Gln	Leu	Ala	Asp
	370					375					380				
His	Tyr	Gln	Gln	Asn	Thr	Pro	Ile	Gly	Asp	Gly	Pro	Val	Leu	Leu	Pro
	385				390					395					400
Asp	Asn	His	Tyr	Leu	Ser	Thr	Gln	Ser	Val	Leu	Ser	Lys	Asp	Pro	Asn
				405					410					415	
Glu	Lys	Arg	Asp	His	Met	Val	Leu	Leu	Glu	Phe	Val	Thr	Ala	Ala	Gly
			420						425				430		
Ile	Thr	Leu	Gly	Met	Asp	Glu	Leu	Tyr	Lys	Gly	Gly	Thr	Gly	Gly	Ser
		435					440					445			
Met	Ser	Lys	Gly	Glu	Glu	Leu	Phe	Thr	Gly	Val	Val	Pro	Ile	Leu	Val
	450					455					460				
Glu	Leu	Asp	Gly	Asp	Val	Asn	Gly	His	Lys	Phe	Ser	Val	Arg	Gly	Glu
	465				470					475					480
Gly	Glu	Gly	Asp	Ala	Thr	Asn	Gly	Lys	Leu	Thr	Leu	Lys	Phe	Ile	Cys
				485					490					495	
Thr	Thr	Gly	Lys	Leu	Pro	Val	Pro	Trp	Pro	Thr	Leu	Val	Thr	Thr	Leu
			500					505					510		
Thr	Tyr	Gly	Val	Gln	Cys	Phe	Ser	Arg	Tyr	Pro	Asp	His	Met	Lys	Gln
		515					520					525			
His	Asp	Phe	Phe	Lys	Ser	Ala	Met	Pro	Glu	Gly	Tyr	Val	Gln	Glu	Arg
	530					535					540				
Thr	Ile	Ser	Phe	Lys	Asp	Asp	Gly	Thr	Tyr	Lys	Thr	Arg	Ala	Glu	Val
	545				550					555					560
Lys	Phe	Glu	Gly	Asp	Thr	Leu	Val	Asn	Arg	Ile	Glu	Leu	Lys	Gly	Ile

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	565		570		575														
Asp	Phe	Lys	Glu	Asp	Gly	Asn	Ile	Leu	Gly	His	Lys	Leu	Glu	Tyr	Asn				
			580					585					590						
Phe	Asn	Asn	Pro	Asn	Ala	Tyr	Gly	Gln	Ser	Ala	Met	Arg	Asp	Trp	Arg				
			595				600					605							
Ser	Asn	Arg	Ile	Val	Gly	Ser	Leu	Val	Ala	Gly	Ala	Val	Ala	Pro	Glu				
	610					615					620								
Ser	Phe	Met	Ser	Gln	Phe	Gly	Thr	Val	Met	Glu	Ile	Phe	Leu	Gln	Thr				
	625				630					635					640				
Arg	Asn	Pro	Gln	Ala	Ala	Ala	Asn	Ala	Ala	Gln	Ala	Ile	Ala	Asp	Gln				
				645					650					655					
Val	Gly	Leu	Gly	Arg	Leu	Gly	Gln	Leu	Gln	Val	Asp	Glu	Gln	Lys	Leu				
			660					665					670						
Ile	Ser	Glu	Glu	Asp	Leu	Asn	Ala	Val	Gly	Gln	Asp	Thr	Gln	Glu	Val				
		675					680						685						
Ile	Val	Val	Pro	His	Ser	Leu	Pro	Phe	Lys	Val	Val	Val	Ile	Ser	Ala				
	690					695					700								
Ile	Leu	Ala	Leu	Val	Val	Leu	Thr	Ile	Ile	Ser	Leu	Ile	Ile	Leu	Ile				
	705				710					715					720				
Met	Leu	Trp	Gln	Lys	Lys	Pro	Arg												
					725														

<210> SEQ ID NO 184
 <211> LENGTH: 650
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic polypeptide

<400> SEQUENCE: 184

Met	Glu	Thr	Asp	Thr	Leu	Leu	Leu	Trp	Val	Leu	Leu	Leu	Trp	Val	Pro				
1				5					10					15					
Gly	Ser	Thr	Gly	Asp	Arg	Ser	Glu	Ser	Ile	Asn	Phe	Val	Ser	Trp	Gly				
			20					25					30						
Gly	Ser	Thr	Gln	Asp	Ala	Gln	Lys	Gln	Ala	Trp	Ala	Asp	Pro	Phe	Ser				
			35				40					45							
Lys	Ala	Ser	Gly	Ile	Thr	Val	Val	Gln	Asp	Gly	Pro	Thr	Asp	Tyr	Gly				
	50					55					60								
Lys	Leu	Lys	Ala	Met	Val	Glu	Ser	Gly	Asn	Val	Gln	Trp	Asp	Val	Val				
	65				70				75					80					
Asp	Val	Glu	Ala	Asp	Phe	Ala	Leu	Arg	Ala	Ala	Ala	Glu	Gly	Leu	Leu				
			85						90					95					
Glu	Pro	Leu	Asp	Phe	Ser	Val	Ile	Gln	Arg	Asp	Lys	Ile	Asp	Pro	Arg				
			100					105					110						
Phe	Val	Ser	Asp	His	Gly	Val	Gly	Ser	Phe	Leu	Phe	Ser	Phe	Val	Leu				
			115					120					125						
Gly	Tyr	Asn	Glu	Gly	Lys	Leu	Gly	Ala	Ser	Lys	Pro	Gln	Asp	Trp	Thr				
	130					135					140								
Ala	Leu	Phe	Asp	Thr	Lys	Thr	Tyr	Pro	Gly	Lys	Arg	Ala	Leu	Tyr	Lys				
	145				150					155					160				
Trp	Pro	Ser	Pro	Gly	Val	Leu	Glu	Leu	Ala	Leu	Leu	Ala	Asp	Gly	Val				
				165					170					175					

-continued

Pro	Ala	Asp	Lys	Leu	Tyr	Pro	Leu	Asp	Leu	Asp	Arg	Ala	Phe	Lys	Lys
			180					185					190		
Leu	Asp	Thr	Ile	Lys	Lys	Asp	Ile	Val	Trp	Trp	Gly	Gly	Gly	Ala	Gln
		195					200					205			
Ser	Gln	Gln	Leu	Leu	Ala	Ser	Gly	Glu	Val	Ser	Met	Gly	Gln	Phe	Trp
	210					215					220				
Asn	Gly	Arg	Ile	His	Ala	Leu	Gln	Glu	Asp	Gly	Ala	Pro	Val	Gly	Val
225					230					235					240
Ser	Trp	Lys	Gln	Asn	Leu	Val	Met	Ala	Asp	Ile	Leu	Val	Val	Pro	Lys
				245					250					255	
Gly	Thr	Lys	Asn	Lys	Ala	Ala	Ala	Met	Lys	Phe	Leu	Ala	Ser	Ala	Ser
			260					265					270		
Ser	Ala	Lys	Gly	Gln	Asp	Asp	Phe	Ser	Ala	Leu	Thr	Ala	Tyr	Ala	Pro
		275					280					285			
Val	Asn	Ile	Asp	Ser	Val	Gln	Arg	Leu	Asp	Leu	Ala	Gln	Val	Arg	Ile
	290					295					300				
Thr	Ala	Asp	Lys	Gln	Lys	Asn	Gly	Ile	Met	Ala	Asn	Phe	Lys	Ile	Arg
305					310					315					320
His	Asn	Val	Glu	Asp	Gly	Ser	Val	Gln	Leu	Ala	Asp	His	Tyr	Gln	Gln
				325					330					335	
Asn	Thr	Pro	Ile	Gly	Asp	Gly	Pro	Val	Leu	Leu	Pro	Asp	Asn	His	Tyr
			340					345					350		
Leu	Ser	Thr	Gln	Ser	Val	Leu	Ser	Lys	Asp	Pro	Asn	Glu	Lys	Arg	Asp
		355					360					365			
His	Met	Val	Leu	Leu	Glu	Phe	Val	Thr	Ala	Ala	Gly	Ile	Thr	Leu	Gly
	370					375					380				
Met	Asp	Glu	Leu	Tyr	Lys	Gly	Gly	Thr	Gly	Gly	Ser	Met	Ser	Lys	Gly
385					390					395					400
Glu	Glu	Leu	Phe	Thr	Gly	Val	Val	Pro	Ile	Leu	Val	Glu	Leu	Asp	Gly
				405					410					415	
Asp	Val	Asn	Gly	His	Lys	Phe	Ser	Val	Arg	Gly	Glu	Gly	Glu	Gly	Asp
			420					425					430		
Ala	Thr	Asn	Gly	Lys	Leu	Thr	Leu	Lys	Phe	Ile	Cys	Thr	Thr	Gly	Lys
		435					440					445			
Leu	Pro	Val	Pro	Trp	Pro	Thr	Leu	Val	Thr	Thr	Leu	Thr	Tyr	Gly	Val
	450					455					460				
Gln	Cys	Phe	Ser	Arg	Tyr	Pro	Asp	His	Met	Lys	Gln	His	Asp	Phe	Phe
465					470					475					480
Lys	Ser	Ala	Met	Pro	Glu	Gly	Tyr	Val	Gln	Glu	Arg	Thr	Ile	Ser	Phe
				485					490					495	
Lys	Asp	Asp	Gly	Thr	Tyr	Lys	Thr	Arg	Ala	Glu	Val	Lys	Phe	Glu	Gly
			500					505					510		
Asp	Thr	Leu	Val	Asn	Arg	Ile	Glu	Leu	Lys	Gly	Ile	Asp	Phe	Lys	Glu
		515					520					525			
Asp	Gly	Asn	Ile	Leu	Gly	His	Lys	Leu	Glu	Tyr	Asn	Trp	Asn	Ala	Asn
	530					535					540				
Leu	Ala	Pro	Asn	Leu	Pro	Thr	Ala	Tyr	Val	Lys	Asp	Gln	Ile	Thr	Leu
545					550					555					560
Asp	Phe	Ala	Tyr	Trp	Ala	Lys	Asn	Gly	Pro	Ala	Ile	Ala	Thr	Arg	Trp
				565					570					575	
Asn	Glu	Trp	Leu	Val	Lys	Leu	Gln	Val	Asp	Leu	Gln	Val	Asp	Glu	Gln

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580	585	590
Lys Leu Ile Ser Glu Glu Asp 595	Leu Asn Ala Val 600	Gly Gln Asp Thr Gln 605
Glu Val Ile Val Val Pro His Ser Leu Pro Phe 610	615	Lys Val Val Val Ile 620
Ser Ala Ile Leu Ala Leu Val Val Leu Thr Ile Ile Ser Leu Ile Ile 625	630	635 640
Leu Ile Met Leu Trp Gln Lys Lys Pro Arg 645	650	

<210> SEQ ID NO 185
 <211> LENGTH: 560
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic polypeptide

<400> SEQUENCE: 185

Met His His His His His His Gly Tyr Pro Tyr Asp Val Pro Asp Tyr 1 5 10 15
Ala Gly Ala Gln Pro Ala Arg Ser Ala Asn Asp Thr Val Val Val Gly 20 25 30
Ser Ile Ile Phe Thr Glu Gly Ile Ile Val Ala Asn Met Val Ala Glu 35 40 45
Met Ile Glu Ala His Thr Asp Leu Lys Val Val Arg Lys Leu Asn Leu 50 55 60
Gly Gly Val Asn Val Asn Phe Glu Ala Ile Lys Arg Gly Gly Ala Asn 65 70 75 80
Asn Gly Ile Asp Ile Tyr Val Glu Tyr Thr Gly His Gly Leu Val Asp 85 90 95
Ile Leu Gly Phe Pro Glu Pro Asn Val Tyr Ile Thr Ala Asp Lys Gln 100 105 110
Lys Asn Gly Ile Lys Ala Asn Phe Lys Ile Arg His Asn Val Glu Asp 115 120 125
Gly Ser Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly 130 135 140
Asp Gly Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser 145 150 155 160
Val Leu Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu 165 170 175
Glu Phe Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr 180 185 190
Lys Gly Gly Thr Gly Gly Ser Met Ser Lys Gly Glu Glu Leu Phe Thr 195 200 205
Gly Val Val Pro Ile Leu Val Glu Leu Asp Gly Asp Val Asn Gly His 210 215 220
Lys Phe Ser Val Arg Gly Glu Gly Glu Gly Asp Ala Thr Asn Gly Lys 225 230 235 240
Leu Thr Leu Lys Phe Ile Cys Thr Thr Gly Lys Leu Pro Val Pro Trp 245 250 255
Pro Thr Leu Val Thr Thr Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg 260 265 270

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Tyr Pro Asp His Met Lys Gln His Asp Phe Phe Lys Ser Ala Met Pro
 275 280 285

Glu Gly Tyr Val Gln Glu Arg Thr Ile Ser Phe Lys Asp Asp Gly Thr
 290 295 300

Tyr Lys Thr Arg Ala Glu Val Lys Phe Glu Gly Asp Thr Leu Val Asn
 305 310 315

Arg Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu
 325 330 335

Gly His Lys Leu Glu Tyr Asn Phe Pro Pro Pro Ala Thr Thr Asp Pro
 340 345 350

Glu Gly Ala Tyr Glu Thr Val Lys Lys Glu Tyr Lys Arg Lys Trp Asn
 355 360 365

Ile Val Trp Leu Lys Pro Leu Gly Phe Asn Asn Thr Tyr Thr Leu Thr
 370 375 380

Val Lys Asp Glu Leu Ala Lys Gln Tyr Asn Leu Lys Thr Phe Ser Asp
 385 390 395 400

Leu Ala Lys Ile Ser Asp Lys Leu Ile Leu Gly Ala Thr Met Phe Phe
 405 410 415

Leu Glu Gly Pro Asp Gly Tyr Pro Gly Leu Gln Lys Leu Tyr Asn Phe
 420 425 430

Lys Phe Lys His Thr Lys Ser Met Asp Met Gly Ile Arg Tyr Thr Ala
 435 440 445

Ile Asp Asn Asn Glu Val Gln Val Ile Asp Ala Trp Ala Thr Asp Gly
 450 455 460

Leu Leu Val Ser His Lys Leu Lys Ile Leu Glu Asp Asp Lys Ala Phe
 465 470 475 480

Phe Pro Pro Tyr Tyr Ala Ala Pro Ile Ile Arg Gln Asp Val Leu Asp
 485 490 495

Lys His Pro Glu Leu Lys Asp Val Leu Asn Lys Leu Ala Asn Gln Ile
 500 505 510

Ser Leu Glu Glu Met Gln Lys Leu Asn Tyr Lys Val Asp Gly Glu Gly
 515 520 525

Gln Asp Pro Ala Lys Val Ala Lys Glu Phe Leu Lys Glu Lys Gly Leu
 530 535 540

Ile Leu Gln Val Asp Glu Gln Lys Leu Ile Ser Glu Glu Asp Leu Asn
 545 550 555 560

<210> SEQ ID NO 186
 <211> LENGTH: 608
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
 polypeptide

<400> SEQUENCE: 186

Met Glu Thr Asp Thr Leu Leu Leu Trp Val Leu Leu Leu Trp Val Pro
 1 5 10 15

Gly Ser Thr Gly Asp Arg Ser Ala Asn Asp Thr Val Val Val Gly Ser
 20 25 30

Ile Ile Phe Thr Glu Gly Ile Ile Val Ala Asn Met Val Ala Glu Met
 35 40 45

Ile Glu Ala His Thr Asp Leu Lys Val Val Arg Lys Leu Asn Leu Gly
 50 55 60

-continued

Gly Val Asn Val Asn Phe Glu Ala Ile Lys Arg Gly Gly Ala Asn Asn
 65 70 75 80
 Gly Ile Asp Ile Tyr Val Glu Tyr Thr Gly His Gly Leu Val Asp Ile
 85 90 95
 Leu Gly Phe Pro Glu Pro Asn Val Tyr Ile Thr Ala Asp Lys Gln Lys
 100 105 110
 Asn Gly Ile Lys Ala Asn Phe Lys Ile Arg His Asn Val Glu Asp Gly
 115 120 125
 Ser Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp
 130 135 140
 Gly Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Val
 145 150 155 160
 Leu Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu
 165 170 175
 Phe Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys
 180 185 190
 Gly Gly Thr Gly Gly Ser Met Ser Lys Gly Glu Glu Leu Phe Thr Gly
 195 200 205
 Val Val Pro Ile Leu Val Glu Leu Asp Gly Asp Val Asn Gly His Lys
 210 215 220
 Phe Ser Val Arg Gly Glu Gly Glu Gly Asp Ala Thr Asn Gly Lys Leu
 225 230 235 240
 Thr Leu Lys Phe Ile Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro
 245 250 255
 Thr Leu Val Thr Thr Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr
 260 265 270
 Pro Asp His Met Lys Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu
 275 280 285
 Gly Tyr Val Gln Glu Arg Thr Ile Ser Phe Lys Asp Asp Gly Thr Tyr
 290 295 300
 Lys Thr Arg Ala Glu Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg
 305 310 315 320
 Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly
 325 330 335
 His Lys Leu Glu Tyr Asn Phe Pro Pro Pro Ala Thr Thr Asp Pro Glu
 340 345 350
 Gly Ala Tyr Glu Thr Val Lys Lys Glu Tyr Lys Arg Lys Trp Asn Ile
 355 360 365
 Val Trp Leu Lys Pro Leu Gly Phe Asn Asn Thr Tyr Thr Leu Thr Val
 370 375 380
 Lys Asp Glu Leu Ala Lys Gln Tyr Asn Leu Lys Thr Phe Ser Asp Leu
 385 390 395 400
 Ala Lys Ile Ser Asp Lys Leu Ile Leu Gly Ala Thr Met Phe Phe Leu
 405 410 415
 Glu Gly Pro Asp Gly Tyr Pro Gly Leu Gln Lys Leu Tyr Asn Phe Lys
 420 425 430
 Phe Lys His Thr Lys Ser Met Asp Met Gly Ile Arg Tyr Thr Ala Ile
 435 440 445
 Asp Asn Asn Glu Val Gln Val Ile Asp Ala Trp Ala Thr Asp Gly Leu
 450 455 460

-continued

Leu Val Ser His Lys Leu Lys Ile Leu Glu Asp Asp Lys Ala Phe Phe
 465 470 475 480

Pro Pro Tyr Tyr Ala Ala Pro Ile Ile Arg Gln Asp Val Leu Asp Lys
 485 490 495

His Pro Glu Leu Lys Asp Val Leu Asn Lys Leu Ala Asn Gln Ile Ser
 500 505 510

Leu Glu Glu Met Gln Lys Leu Asn Tyr Lys Val Asp Gly Glu Gly Gln
 515 520 525

Asp Pro Ala Lys Val Ala Lys Glu Phe Leu Lys Glu Lys Gly Leu Ile
 530 535 540

Leu Gln Val Asp Glu Gln Lys Leu Ile Ser Glu Glu Asp Leu Asn Ala
 545 550 555 560

Val Gly Gln Asp Thr Gln Glu Val Ile Val Val Pro His Ser Leu Pro
 565 570 575

Phe Lys Val Val Val Ile Ser Ala Ile Leu Ala Leu Val Val Leu Thr
 580 585 590

Ile Ile Ser Leu Ile Ile Leu Ile Met Leu Trp Gln Lys Lys Pro Arg
 595 600 605

<210> SEQ ID NO 187
 <211> LENGTH: 8
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
 peptide

<400> SEQUENCE: 187

Pro Ile Leu Val Ser His Asn Val
 1 5

<210> SEQ ID NO 188
 <211> LENGTH: 8
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
 peptide

<400> SEQUENCE: 188

Pro Ile Leu Gly Tyr His Asn Val
 1 5

<210> SEQ ID NO 189
 <211> LENGTH: 8
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
 peptide

<400> SEQUENCE: 189

Tyr Asn Phe Asn Asn Pro Leu Asn
 1 5

<210> SEQ ID NO 190
 <211> LENGTH: 8
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic

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peptide

<400> SEQUENCE: 190

Tyr Asn Phe Asn Glu Gln Leu Asn
1 5

<210> SEQ ID NO 191

<211> LENGTH: 5

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
peptide

<400> SEQUENCE: 191

Ser His Asn Val Tyr
1 5

<210> SEQ ID NO 192

<211> LENGTH: 5

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
peptide

<400> SEQUENCE: 192

Leu Ala Gln Val Arg
1 5

<210> SEQ ID NO 193

<211> LENGTH: 5

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
peptide

<400> SEQUENCE: 193

Ser Phe Gly Phe Pro
1 5

<210> SEQ ID NO 194

<211> LENGTH: 5

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
peptide

<400> SEQUENCE: 194

Ser Val Leu Ala Pro
1 5

<210> SEQ ID NO 195

<211> LENGTH: 5

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
peptide

<400> SEQUENCE: 195

Ala Asn Leu Ala Pro
1 5

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<210> SEQ ID NO 196
<211> LENGTH: 7
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
      oligopeptide
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (2)..(2)
<223> OTHER INFORMATION: Any amino acid

<400> SEQUENCE: 196

Pro Xaa Ser His Asn Val Tyr
1             5

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<210> SEQ ID NO 197
<211> LENGTH: 6
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
      6xHis tag

<400> SEQUENCE: 197

His His His His His His
1             5

```

What is claimed is:

1. A recombinant peptide biosensor comprising an analyte-binding framework portion and a signaling portion, wherein the signaling portion is present within the framework portion at a site or amino acid position that undergoes a conformational change upon interaction of the framework portion with a defined, specific, or selected analyte, wherein the recombinant peptide biosensor comprises a amino acid sequence having at least 85% sequence identity to SEQ ID NO: 184.

2. The recombinant peptide biosensor of claim 1, wherein the signaling portion is allosterically regulated by the framework portion such that signaling from the signaling portion is altered upon interaction of the framework portion with the analyte.

3. The recombinant peptide biosensor of claim 1, wherein signaling by the signaling portion detectably increases upon interaction of the framework portion with the analyte.

4. The recombinant peptide biosensor of claim 1, wherein signaling by the signaling portion detectably decreases upon interaction of the framework portion with the analyte.

5. The recombinant peptide biosensor of claim 1, wherein signaling by the signaling portion is proportional to the level of interaction between the framework portion and the analyte.

6. The recombinant peptide biosensor of claim 1, wherein the framework portion has a first structure in the absence of an analyte and a second structure, that is detectably distinct from the first structure, in the presence of the analyte.

7. The recombinant peptide biosensor of claim 6, wherein the conformational change between the first structure and the second structure allosterically regulates the signaling portion.

8. The recombinant peptide biosensor of claim 1, wherein the framework portion is a periplasmic binding protein (PBP) or a variant of a PBP.

9. The recombinant peptide biosensor of claim 1, wherein the signaling portion is a circularly permuted super fluorescent (SF) protein.

10. The recombinant peptide biosensor of claim 9, wherein the SF protein is selected from the group consisting of a green fluorescent protein, a yellow fluorescent protein, a red fluorescent protein, and a blue fluorescent protein.

11. The recombinant peptide biosensor of claim 1, wherein the analyte-binding framework portion binds specifically to gamma aminobutyric acid (GABA).

12. A recombinant peptide biosensor comprising an amino acid sequence having at least 90% sequence identity to SEQ ID NO:184, wherein the recombinant peptide biosensor binds specifically to GABA.

13. The recombinant peptide biosensor of claim 12 having SEQ ID NO:184 and comprising 10 or fewer conservative amino acid substitutions, wherein the recombinant peptide biosensor binds specifically to GABA.

14. The recombinant peptide biosensor of claim 12, comprising a recombinant peptide biosensor having SEQ ID NO:184.

15. A nucleic acid encoding the recombinant peptide biosensor of claim 1.

16. A vector comprising the nucleic acid of claim 15.

17. A cell comprising the nucleic acid of claim 15.

18. A cell comprising the vector of claim 16.

19. A kit comprising the recombinant peptide biosensor of claim 1, the nucleic acid of claim 15, the vector of claim 16, the cell of claim 17, and/or the cell of claim 18.

20. A method for detecting GABA, the method comprising detecting a level of fluorescence emitted by a recombi-

nant peptide biosensor, the peptide biosensor having the amino acid sequence shown in SEQ ID NO:184, and correlating the level of fluorescence with the presence of GABA.

21. The method of claim **20**, wherein the recombinant peptide biosensor is expressed from a nucleic acid.

22. The method of claim **20**, comprising contacting the recombinant peptide biosensor with a sample comprising GABA.

23. The method of claim **22**, comprising correlating the level of fluorescence with a concentration of GABA.

24. The method of claim **23**, comprising comparing the level of fluorescence with a level of fluorescence emitted by the recombinant peptide biosensor in the presence of a sample comprising a known concentration or range of concentrations of GABA.

25. The method of claim **24**, wherein the method is performed in vitro.

26. A method for detecting a defined, selected, or specific analyte, the method comprising detecting a level of fluorescence emitted by a recombinant peptide biosensor of claim

1 in the presence of said analyte; and correlating the level of fluorescence with the presence of a defined, selected, or specific analyte.

27. The method of claim **26**, wherein the recombinant peptide biosensor is expressed from a nucleic acid.

28. The method of claim **26**, comprising contacting the recombinant peptide biosensor with a sample comprising the analyte.

29. The method of claim **28**, comprising correlating the level of fluorescence with a concentration of the analyte.

30. The method of claim **29**, comprising comparing the level of fluorescence with a level of fluorescence emitted by the recombinant peptide biosensor in the presence of a sample comprising a known concentration or range of concentrations of the analyte.

31. The method of claim **30**, wherein the method is performed in vitro.

32. The method of claim **30**, wherein the analyte is GABA.

* * * * *