



US 20240052391A1

(19) **United States**

(12) **Patent Application Publication** (10) **Pub. No.: US 2024/0052391 A1**

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(43) **Pub. Date: Feb. 15, 2024**

(54) **ENZYMATIC SYNTHESIS OF POLYNUCLEOTIDE PROBES**

(30) **Foreign Application Priority Data**

Oct. 29, 2020 (EP) 20306298

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Publication Classification

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(51) **Int. Cl.**
C12P 19/34 (2006.01)
C12N 9/16 (2006.01)

(21) Appl. No.: **18/034,487**

(52) **U.S. Cl.**
CPC **C12P 19/34** (2013.01); **C12N 9/16** (2013.01); **C12Y 301/21007** (2013.01)

(22) PCT Filed: **Oct. 27, 2021**

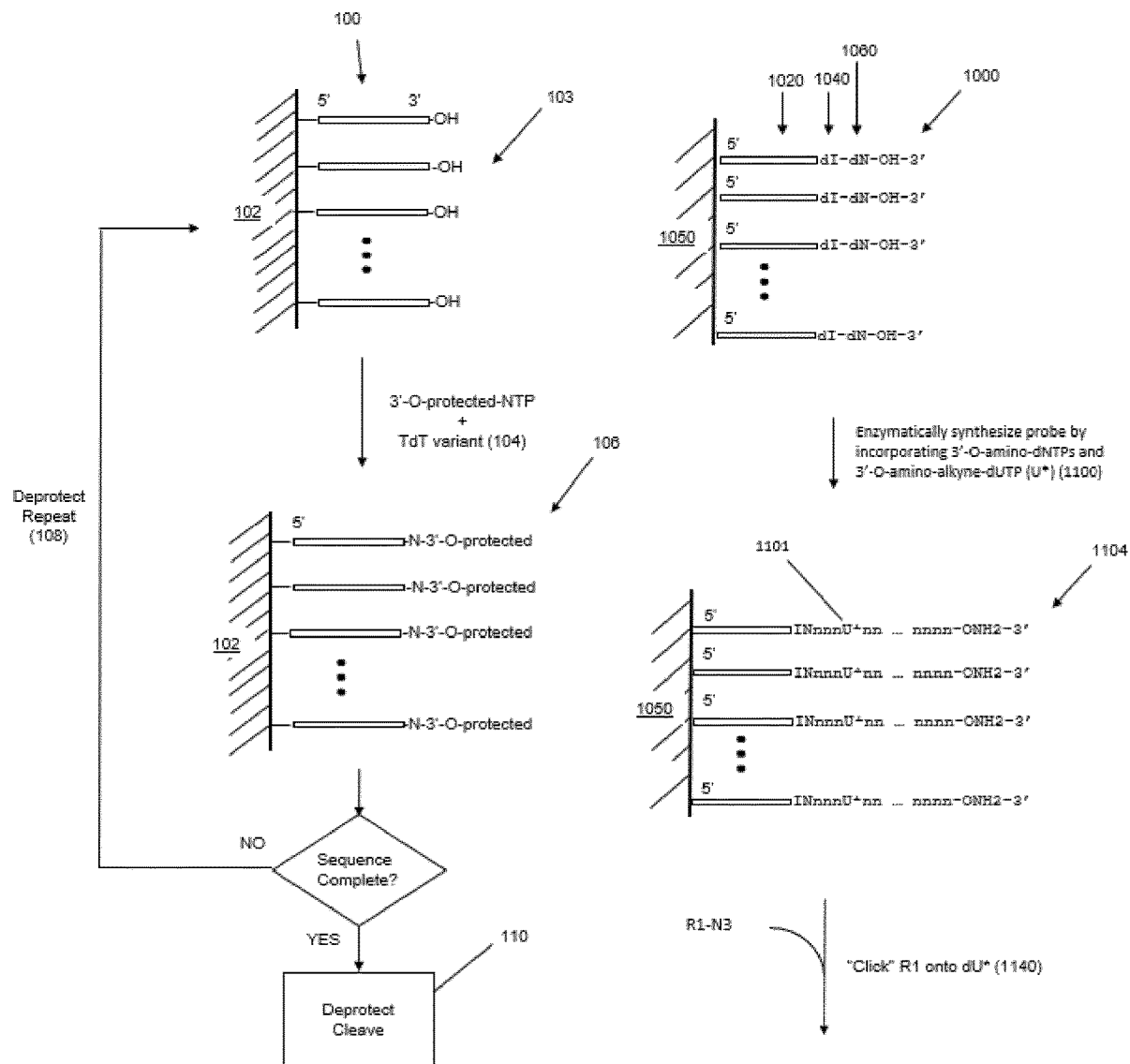
(57) **ABSTRACT**

(86) PCT No.: **PCT/EP2021/079847**

The present invention is directed to methods and kits for enzymatic synthesis of labeled polynucleotide probes using template-free DNA polymerases.

§ 371 (c)(1),
(2) Date: **Apr. 28, 2023**

Specification includes a Sequence Listing.



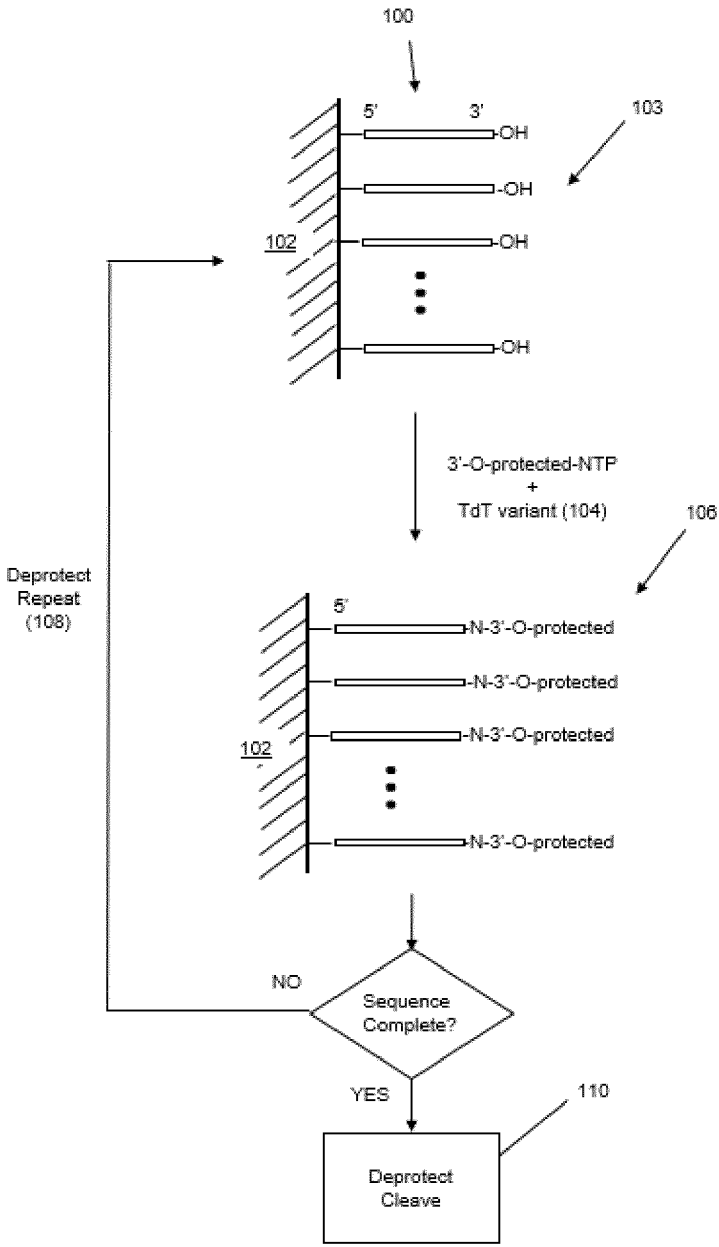


FIGURE 1A

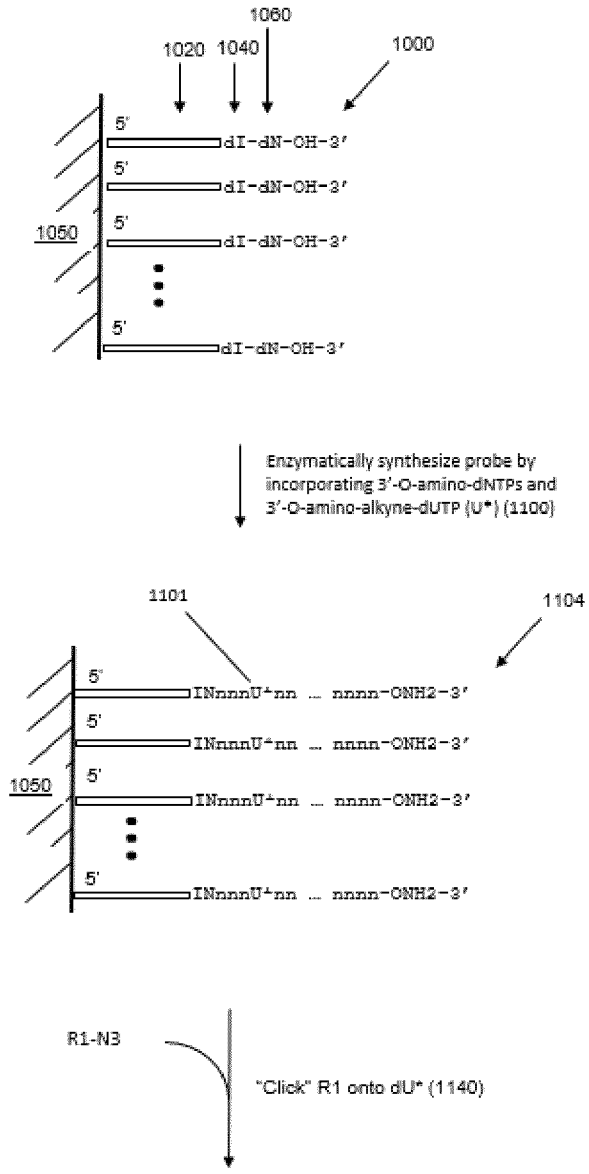


FIGURE 1B

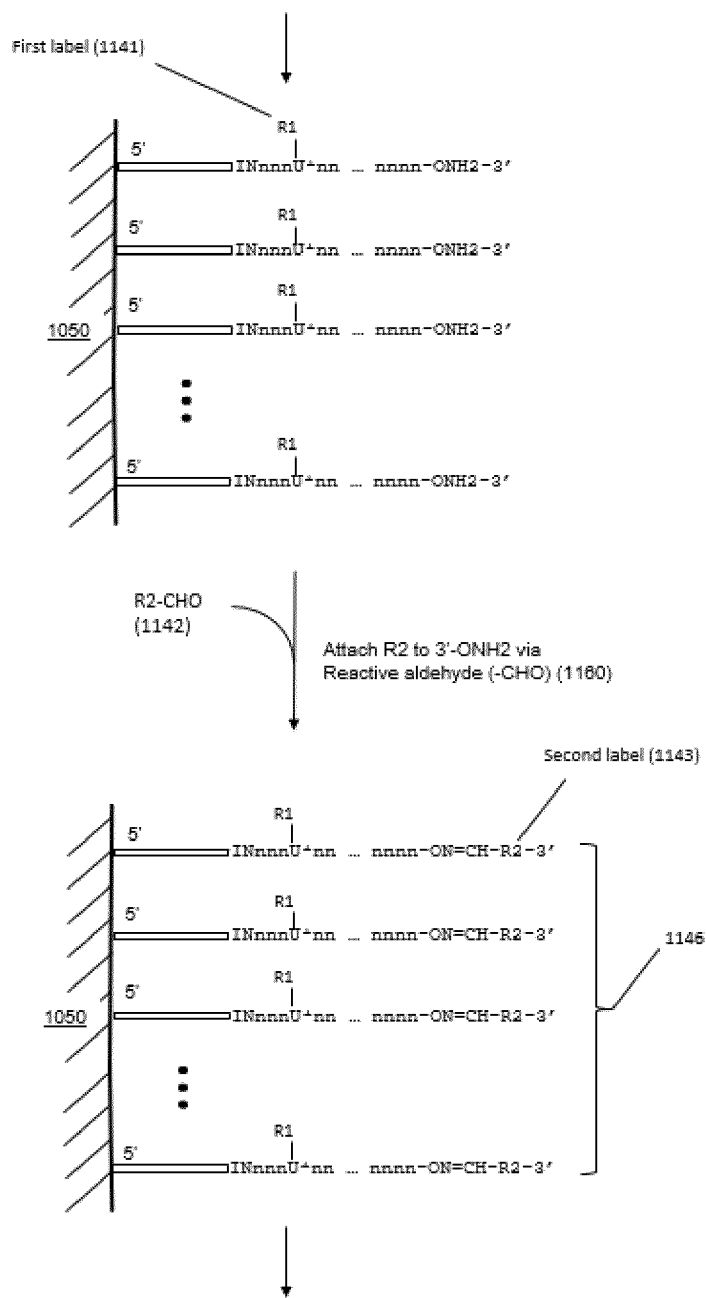


FIGURE 1C

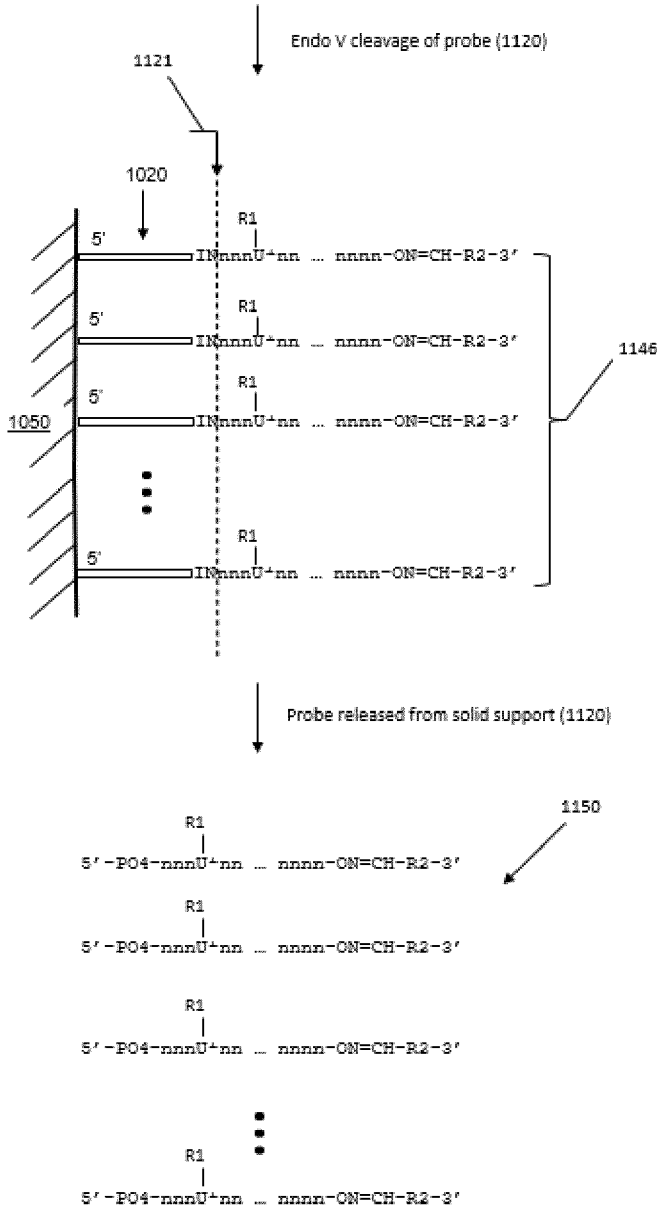


FIGURE 1D

ENZYMATIC SYNTHESIS OF POLYNUCLEOTIDE PROBES

BACKGROUND

[0001] Interest in enzymatic approaches to polynucleotide synthesis has recently increased both because of increased demand for synthetic polynucleotides in many areas, such as synthetic biology, CRISPR-Cas9 applications, high-throughput sequencing, and the like, and because of the limitations of chemical approaches to polynucleotide synthesis, such as the difficulty of performing multi-step synthesis reactions under inert atmospheres and moisture-free environments, the upper limits on product length, the use of, and needed disposal of, organic solvents, and so on, e.g., Jensen et al, *Biochemistry*, 57: 1821-1832 (2018). Currently, most enzymatic approaches employ a template-free polymerase to repeatedly add 3'-O-blocked nucleoside triphosphates to a single stranded initiator or an elongated strand attached to a support followed by deblocking until a polynucleotide of the desired sequence is obtained. In such enzymatic approaches to polynucleotide synthesis, enzyme specificity often presents a "double-edged sword" in that same specificity that was engineered for usual monomers may prevent or greatly reduce activity when it is desired to incorporate unusual monomers with labels or other adducts.

[0002] In view of the above, enzymatic synthesis of polynucleotides would be advanced if methods were available for high efficiency incorporation of various labels or other adducts into enzymatically synthesized nucleic acid probes.

SUMMARY OF THE INVENTION

[0003] The present invention is directed to methods, kits and compositions for template-free enzymatic synthesis of polynucleotide probes having predetermined sequences and a plurality of attached fluorescent dyes and/or quencher molecules.

[0004] In one aspect, methods of the invention include a method of synthesizing a polynucleotide probe having a predetermined sequence and a plurality of labels with the following steps: (a) providing an initiator having a 3'-penultimate deoxyinosine and a 3'-terminal nucleotide having a free 3'-hydroxyl; (b) repeating elongation cycles comprising steps of: (i) contacting under elongation conditions the initiator or elongated fragments having free 3'-O-hydroxyls with a 3'-O-blocked nucleoside triphosphate and a template-independent DNA polymerase so that the initiator or elongated fragments are elongated by incorporation of a 3'-O-blocked nucleoside triphosphate to form 3'-O-blocked elongated fragments, and (ii) deblocking the elongated fragments to form elongated fragments having free 3'-hydroxyls, until the polynucleotide of the polynucleotide probe is formed, wherein (A) at least one of the 3'-O-blocked nucleoside triphosphate is a 3'-O-blocked-(alkyne/azide)-nucleoside triphosphate or a 3'-O-blocked-(protected alkyne/protected azide)-nucleoside triphosphate, (B) in a final elongation cycle a nucleoside triphosphate having a 3'-alternative blocking group or 3'-O-alternative blocking group is contacted with said elongated fragments having free 3'-hydroxyls, (C) the alternative blocking group has a reactive moiety and may be the same or different than the blocking group, and (D) said step of deblocking is omitted; (c) reacting a first label having an (alkyne/azide) group with the at least one incorporated 3'-O-blocked-(alkyne/azide)-

nucleoside triphosphate; (d) reacting the reactive moiety of the 3'-alternative blocking group or 3'-O-alternative blocking group with a second label having a complementary moiety to form a polynucleotide probe having a plurality of labels; and (e) enzymatically cleaving the polynucleotide probe from the initiator.

[0005] In another aspect, the invention includes kits and compositions of matter for carrying out methods of the invention. In particular, the invention includes a composition of matter comprising 3'-O—NH₂-alkyne-nucleoside triphosphates for use in the enzymatic synthesis of polynucleotide probes, wherein the alkyne moiety is attached to a base via a linker. The invention further includes a kit for enzymatically synthesizing a polynucleotide probe comprising at least one 3'-O-blocked-(alkyne/azide)-nucleoside triphosphate or 3'-O-blocked-(protected alkyne/protected azide)-nucleoside triphosphate and a nucleoside triphosphate having a 3'-alternative blocking group or 3'-O-alternative blocking group, wherein the alternative blocking group has a reactive moiety and may be the same or different than a 3'-O-blocking group of the 3'-O-blocked-(alkyne/azide)-nucleoside triphosphate or 3'-O-blocked-(protected alkyne/protected azide)-nucleoside triphosphate.

[0006] The invention is in part based on the recognition and appreciation that template-free DNA polymerases, especially terminal deoxynucleotidyl transferase (TdT) variants, and endonuclease V enzymes can accommodate substrates that have been drastically altered (e.g., labels or "click" groups added) for the purpose of making modified natural compounds, such as polynucleotide probes, which have useful applications in a wide variety of fields, especially in the medical and health related fields.

[0007] These above-characterized aspects, as well as other aspects, of the present invention are exemplified in a number of illustrated implementations and applications, some of which are shown in the figures and characterized in the claims section that follows. However, the above summary is not intended to describe each illustrated embodiment or every implementation of the present invention.

BRIEF DESCRIPTION OF THE DRAWINGS

[0008] FIG. 1A illustrates the basic steps of template-free enzymatic synthesis of polynucleotides.

[0009] FIGS. 1B-1D illustrate the steps of one embodiment of the method of the invention for synthesizing polynucleotide probes.

DETAILED DESCRIPTION OF THE INVENTION

[0010] The general principles of the invention are disclosed in more detail herein particularly by way of examples, such as those shown in the drawings and described in detail. It should be understood, however, that the intention is not to limit the invention to the particular embodiments described. The invention is amenable to various modifications and alternative forms, specifics of which are shown for several embodiments. The intention is to cover all modifications, equivalents, and alternatives falling within the principles and scope of the invention.

[0011] The practice of the present invention may employ, unless otherwise indicated, conventional techniques and descriptions of organic chemistry, molecular biology (including recombinant techniques), cell biology, and biochem-

istry, which are within the skill of the art. Such conventional techniques may include, but are not limited to, preparation and use of synthetic peptides, synthetic polynucleotides, monoclonal antibodies, nucleic acid cloning, amplification, sequencing and analysis, and related techniques. Protocols for such conventional techniques can be found in product literature from manufacturers and in standard laboratory manuals, such as *Genome Analysis: A Laboratory Manual Series (Vols. I-IV)*; *PCR Primer: A Laboratory Manual*; and *Molecular Cloning: A Laboratory Manual* (all from Cold Spring Harbor Laboratory Press); Lutz and Bornscheuer, Editors, *Protein Engineering Handbook* (Wiley-VCH, 2009); Hermanson, *Bioconjugate Techniques*, Second Edition (Academic Press, 2008); and like references.

[0012] The present invention is directed to methods, kits and compositions for enzymatic synthesis of labeled polynucleotide probes that may be used, for example, in “Taqman” assays. As used herein, the term “polynucleotide probe” refers to a polynucleotide that has a predetermined nucleotide sequence (typically the complement of a target polynucleotide) and that has one or more labels covalently attached. Such labels may include dyes, linkers, reactive groups, and the like, as described more fully below. In some embodiments, polynucleotide probes of the invention each have a plurality of labels attached. In particular, polynucleotide probes each may have two labels attached. In some embodiments, the two labels may be attached such that one label is at a 5' end of the polynucleotide probe and the other label at a 3' end of the polynucleotide probe. In some embodiments, one label is attached to a 3' terminal nucleotide and the second label is attached to a 5' terminal nucleotide. In a particular embodiment, such first label is a fluorescent dye and such second label is a fluorescent quencher that is capable of quenching the fluorescent emissions of the first label. As used herein, the term “labels” refers to any moiety that may be used directly or indirectly to detect the presence and/or quantity of a target nucleic acid to which the polynucleotide probe is capable of hybridizing. Typically, such detection is based on the appearance of a fluorescent signal and the intensity of a fluorescent signal, respectively. In some embodiments, labels are selected from the group consisting of fluorescent dyes and fluorescent quenchers, with the proviso that a polynucleotide probe comprises at least one fluorescent dye. Such fluorescent dyes and fluorescent quenchers are commercially available, e.g. *Molecular Probes* (ThermoFisher). In some embodiments, the length of the polynucleotide probe is selected so that when hybridized to a target nucleic acid the fluorescent dye and the fluorescent quencher are within a fluorescence resonant energy transfer (FRET) distance of one another so that no or minimal fluorescent emissions are emitted from the probe.

[0013] As mentioned above, methods of the invention for synthesizing a polynucleotide probe comprise the general steps of: (a) providing an initiator having a 3'-penultimate deoxyinosine and a 3'-terminal nucleotide having a free 3'-hydroxyl; (b) repeating elongation cycles comprising steps of: (i) contacting under elongation conditions the initiator or elongated fragments having free 3'-O-hydroxyls with a 3'-O-blocked nucleoside triphosphate and a template-independent DNA polymerase so that the initiator or elongated fragments are elongated by incorporation of a 3'-O-blocked nucleoside triphosphate to form 3'-O-blocked elongated fragments, and (ii) deblocking the elongated frag-

ments to form elongated fragments having free 3'-hydroxyls, until the polynucleotide of the polynucleotide probe is formed, wherein (A) at least one of the 3'-O-blocked nucleoside triphosphate is a 3'-O-blocked-(alkyne/azide)-nucleoside triphosphate or a 3'-O-blocked-(protected alkyne/protected azide)-nucleoside triphosphate, (B) in a final elongation cycle a nucleoside triphosphate having a 3'-alternative blocking group or 3'-O-alternative blocking group is contacted with said elongated fragments having free 3'-hydroxyls, (C) the alternative blocking group has a reactive moiety and may be the same or different than the blocking group, and (D) said step of deblocking is omitted; (c) reacting a first label having an (alkyne/azide) group with the at least one incorporated 3'-blocked-(alkyne/azide)-nucleoside triphosphate; (d) reacting the reactive moiety of the 3'-alternative blocking group or 3'-O-alternative blocking group with a second label having a complementary moiety to form a polynucleotide probe having a plurality of labels; and (e) enzymatically cleaving the polynucleotide probe from the initiator.

[0014] Regarding step (a), in some embodiments, the initiator is attached by its 5' end to a solid support which facilitates the addition and removal of reagents in the synthesis process. In some embodiments, the 3' terminal nucleotide is thymidine. In part the invention was a discovery that endonuclease V was unexpectedly able to cleave a polynucleotide probe from the initiator even when the first nucleotide of the polynucleotide probe contained a bulky label, such as a fluorescent dye. Likewise, the invention is in part a discovery that terminal deoxynucleotidyl transferases (TdT) were available that had the capability of incorporating 3'-O-blocked-(alkyne/azide)-nucleoside triphosphates.

[0015] Regarding step (b), part (A), as used herein, the expression “-(alkyne/azide)-” refers to the “click” pair consisting of the alkyne group and the azido group and circumstances in which the groups may be attached to either of two moieties intended to be ligated or coupled. That is, for example, the expression indicates that if it is desired to ligate chemical moieties R1 and R2, that it may be accomplished by either carrying out a “click” reaction between R1—N₃ and R2—C≡CH or carrying out a “click” reaction between R1—C≡CH and R2—N₃. Likewise, the expression “-(protected alkyne/protected azide)-” denotes the same relationship, except that deprotection must be carried out prior to performance of a “click” reaction. As used herein, the step of reacting or implementing a “click” reaction in reference to alkyne and azide groups means a (3+2) cycloaddition reaction between azide and alkyne groups forming a 1,2,3-triazole ring, e.g. Sharpless et al, U.S. Pat. No. 7,763,736; Carell et al, U.S. Pat. No. 8,129,315; and the like, which are incorporated herein by reference.

[0016] In further regard to step (b), part (A), in some embodiments, 3'-O-blocked-(alkyne/azide)-nucleoside triphosphates are 3'-O—NH₂-alkyne-nucleoside triphosphates. Such monomers may be synthesized from 3'-oxime-nucleoside starting materials using conventional chemistry, e.g. Winz et al, *Nucleic Acids Research*, 43(17): e110 (2015); Carell et al, U.S. Pat. No. 8,129,315; Sharpless et al, U.S. Pat. No. 7,375,234; and the like, which are incorporated herein by reference. For example, 3'-O—NH₂-5-alkyne-deoxyuridine triphosphates (5-octa(1,7)diynyl-3'-aminoxyl-dUTP) may be synthesized from 3'-oxime-5-iodo-deoxyuridine starting material. Typically, alkyne groups and azide

groups are attached to nucleobases, usually via a linker, at conventional positions for labeling nucleotides, e.g. 7-purine and 5-pyrimidine.

[0017] Regarding step (b), elongation cycles may also include one or more washing steps. In some embodiments, elongation cycles include a single washing step implemented after the deprotection step. In other embodiments, elongation cycles may include a washing step after each “contacting” step as well as after each “deblocking” step. In still further embodiments employing additional steps, such as capping steps, further washing steps may be implemented.

[0018] Regarding step (b), parts (B) and (C), in the final elongation cycle a monomer is added to which a second label is covalently coupled through an alternative blocking group. The alternative blocking group may be the same or different than the blocking group of the other monomers (i.e., non-final monomers). For example, when 3'-O—NH₂-dNTPs or 3'-O-azidomethyl-dNTPs are used, then the “alternative blocking” group may be the same as the “blocking” groups because both the —NH₂ and the -azidomethyl groups have a blocking function and are reactive to selected complementary groups. However, if the blocking groups are not reactive to a selected complementary group, then it is advantageous to employ an alternative blocking group in the monomer of the final cycle which has such reactivity, e.g. an azidomethyl or an amine. In some embodiments, the alternative blocking group of the final monomer may either be attached directly to the 3' carbon nucleoside triphosphate (e.g. Lui et al, *Nucleic Acids Resesarch*, 35(21): 7140-7149 (2007) disclosing 3'-azido-(2',3')-dideoxy-dNTPs) or may be attached indirectly to the 3' carbon via the 3' oxygen (e.g. Guo et al, *Acc. Chem. Res.*, 43(4): 551-563 (2010) disclosing 3'-O-azidomethyl-dNTPs). Thus, in some embodiments, the final monomer of the polynucleotide may be a 3'-azido-2',3'-dideoxynucleoside triphosphate. Or, alternatively, such monomer may be a 3'-O-azidomethyl-nucleoside triphosphate. Either case is advantageous since labels having an alkyne group may be readily coupled to the 3' end of the polynucleotide probe via a “click” reaction.

[0019] Regarding step (d), whenever the 3'-oxygen of monomers are protected (or blocked) with amine groups, the blocking groups and alternative blocking groups may be the same and the complementary moiety of the second label may be an aldehyde group. In some embodiments, there is a single first label which is a fluorescent dye and a second label which is a fluorescent quencher capable of quenching the fluorescent emissions of the selected fluorescent dye. Among such embodiments, a fluorescent quencher may be a Black Hole Quencher (BHQ) dye. Azide derivatives of BHQ dyes may be synthesized as disclosed in Chevalier et al, *Chem. Eur. J.*, 19: 1686-1699 (2013). The azide-BHQ dyes then may be converted into aldehyde-BHQ dyes by click reacting an alkyne-aldehyde linker (e.g. Biosearch Technologies) with the azide-BHQ dye. In other embodiments, the first label may be selected from the group of fluorescent dyes consisting of 6-carboxyfluorescein (FAM) and tetrachlorofluorescein (TET) and a second label may be tetramethylrhodamine (TAN/IRA).

[0020] Regarding step (e), enzymatic cleavage may be effected by a variety of enzymes and corresponding cleavable nucleotides or linkages. In some embodiments, enzymatic cleavage is carried out with an endonuclease V activity that cleaves a single stranded DNA containing a

deoxyinosine, as described in FIGS. 1B-1D. In other embodiments, cleavable nucleotides may be nucleotide analogs such as deoxyuridine or 8-oxo-deoxyguanosine that are recognized by specific glycosylases (e.g. uracil deoxyglycosylase followed by endonuclease VIII, and 8-oxoguanine DNA glycosylase, respectively). In some embodiments, cleavage by glycosylases and/or endonucleases may require a double stranded DNA substrate. In other embodiments, cleavable nucleotides include nucleotides comprising base analogs cleavable by endonuclease III which include, but are not limited to, urea, thymine glycol, methyl tartonyl urea, alloxan, uracil glycol, 6-hydroxy-5,6-dihydrocytosine, 5-hydroxyhydantoin, 5-hydroxycytocine, trans-1-carbamoyl-2-oxo-4,5-dihydroxyimidazolidine, 5,6-dihydrouracil, 5-hydroxycytosine, 5-hydroxyuracil, 5-hydroxy-6-hydrouracil, 5-hydroxy-6-hydrothymine, 5,6-dihydrothymine. In some embodiments, cleavable nucleotides include nucleotides comprising base analogs cleavable by formamidopyrimidine DNA glycosylase which include, but are not limited to, 7,8-dihydro-8-oxoguanine, 7,8-dihydro-8-oxoinosine, 7,8-dihydro-8-oxoadenine, 7,8-dihydro-8-oxonebularine, 4,6-diamino-5-formamidopyrimidine, 2,6-diamino-4-hydroxy-5-formamidopyrimidine, 2,6-diamino-4-hydroxy-5-N-methylformamidopyrimidine, 5-hydroxycytosine, 5-hydroxyuracil. In some embodiments, cleavable nucleotides include nucleotides comprising base analogs cleavable by hNeil 1 which include, but are not limited to, guanidinohydantoin, spiroiminodihydantoin, 5-hydroxyuracil, thymine glycol. In some embodiments, cleavable nucleotides include nucleotides comprising base analogs cleavable by thymine DNA glycosylase which include, but are not limited to, 5-formylcytosine and 5-carboxycytosine. In some embodiments, cleavable nucleotides include nucleotides comprising base analogs cleavable by human alkyladenine DNA glycosylase which include, but are not limited to, 3-methyladenine, 3-methylguanine, 7-methylguanine, 7-(2-chloroethyl)-guanine, 7-(2-hydroxyethyl)-guanine, 7-(2-ethoxyethyl)-guanine, 1,2-bis-(7-guanyl)ethane, 1,N6-ethenoadenine, 1,N2-ethenoguanine, N2,3-ethenoguanine, N2,3-ethanoguanine, 5-formyluracil, 5-hydroxymethyluracil, hypoxanthine. In some embodiments, cleavable nucleotides include 5-methylcytosine cleavable by 5-methylcytosine DNA glycosylase

[0021] FIGS. 1B-1D illustrate the steps of an exemplary embodiment of the method of template-free enzymatic synthesis of labeled polynucleotide probes which employs initiators with a penultimate deoxyinosine, 3'-O—NH₂-nucleoside triphosphate monomers, alkyne-derivatized monomers, and an aldehyde-derivatized label to couple to the 3'-O-amine of the final nucleotide of the probe. FIG. 1B shows initiators (1000) attached by their 5' ends to solid support (1050). Each initiator (1020) has a 3'-penultimate deoxyinosine (1040) next to 3'-terminal nucleotide (1060) that has a free 3' hydroxyl. Such initiators on solid supports (which is a starting material for the present invention) may be produced using conventional chemical nucleic acid synthesis. After a predetermined number of cycles of enzymatic incorporation and de-blocking (1100), except for the final cycle that omits the de-blocking step, a polynucleotide product (1104) having terminal 3'-O—NH₂ groups is formed. At least one of the nucleotides incorporated is a 3'-O—NH₂-(5-alkyne)-deoxyuridine triphosphate (designated as “U*”, 1101). At this point, in this embodiment, a first label is attached (1140) by “click” reacting an azide-

dervativized first label (R1—N3) to the alkyne of U* to give first label (1141). After first label (1141) is attached, aldehyde-derivative (1142) of second label (1143) is reacted (1160) with the 3'-O amines to form dual labeled probe (1146)

[0022] Dual labeled polynucleotide probe (1146) is then cleaved from initiators (1020) and support (1050) by treating the attached product with an endonuclease V activity which recognizes the presence of the deoxyinosine and cleaves the strand on the 3' side (1121 in FIG. 1D) of terminal nucleotide (1060) of the initiators. In some embodiments, the endonuclease V activity is provided by using a prokaryotic endonuclease V. In still other embodiments, the endonuclease V is an *E. coli* endonuclease V. As used herein, the term "endonuclease V activity" means an enzyme activity that catalyzes the following cleavage reaction in a single stranded DNA: 5' . . . NNINNNN . . . -3'→5' - . . . NNIN+5'-PO₄-NNNN . . . -3' where N is any nucleotide and I is deoxyinosine. Cleavage (1120) of polynucleotides (1146) by an endonuclease V activity releases polynucleotide probe (1150) which includes a 5'-monophosphate that optionally may be removed by a step of treating with a 5'-phosphatase.

[0023] Enzymes with endonuclease V activity are available from commercial enzyme suppliers, for example, New England Biolabs (Beverly, MA, USA), NzyTech (Lisbon, Portugal). Such enzymes may be used with the supplier's recommended cleavage buffers (e.g. 50 mM K—Ac, 20 mM Tris-Ac, 10 mM Mg—Ac, 1 mM DTT at pH 7.9). Typical cleavage conditions are as follows: 70U of Endo V in 500 of Nzytech buffer at 37° C. for 500 pmol synthesis scale on resin. Typical cleavage times are from 5 to 60 minutes, or from 10 to 30 minutes. Optionally, endonuclease activity of the above enzymes may be heat inactivated by incubation at 65° C. or higher for 20 minutes. Optionally, the Nzytech endonuclease V comprises a His tag that allows convenient removal of the enzyme from reaction mixtures in preparation of final products.

Template-Free Enzymatic Synthesis

[0024] Generally, methods of template-free (or equivalently, "template-independent") enzymatic DNA synthesis comprise repeated cycles of steps, such as are illustrated in FIG. 1A, in which a predetermined nucleotide is coupled to an initiator or growing chain in each cycle. The general elements of template-free enzymatic synthesis is described in the following references: Ybert et al, International patent publication WO/2015/159023; Ybert et al, International patent publication WO/2017/216472; Hyman, U.S. Pat. No. 5,436,143; Hiatt et al, U.S. Pat. No. 5,763,594; Jensen et al, *Biochemistry*, 57: 1821-1832 (2018); Mathews et al, *Organic & Biomolecular Chemistry*, DOI: 0.1039/c6ob01371f (2016); Schmitz et al, *Organic Lett.*, 1(11): 1729-1731 (1999).

[0025] Initiator polynucleotides (100) are provided, for example, attached to solid support (102), which have free 3'-hydroxyl groups (103). To the initiator polynucleotides (100) (or elongated initiator polynucleotides in subsequent cycles) are added a 3'-O-protected-dNTP and a template-free polymerase, such as a TdT or variant thereof (e.g. Ybert et al, WO/2017/216472; Champion et al, WO2019/135007) under conditions (104) effective for the enzymatic incorporation of the 3'-O-protected-dNTP onto the 3' end of the initiator polynucleotides (100) (or elongated initiator poly-

nucleotides). This reaction produces elongated initiator polynucleotides whose 3'-hydroxyls are protected (106). If the elongated sequence is not complete, then another cycle of addition is implemented (108). If the elongated initiator polynucleotide contains a competed sequence, then the 3'-O-protection group may be removed, or deprotected, and the desired sequence may be cleaved from the original initiator polynucleotide (110). Such cleavage may be carried out using any of a variety of single strand cleavage techniques, for example, by inserting a cleavable nucleotide at a predetermined location within the original initiator polynucleotide. An exemplary cleavable nucleotide may be a uracil nucleotide which is cleaved by uracil DNA glycosylase. If the elongated initiator polynucleotide does not contain a completed sequence, then the 3'-O-protection groups are removed to expose free 3'-hydroxyls (103) and the elongated initiator polynucleotides are subjected to another cycle of nucleotide addition and deprotection.

[0026] As used herein, the terms "protected" and "blocked" in reference to specified groups, such as, a 3'-hydroxyls of a nucleotide or a nucleoside, are used interchangeably and are intended to mean a moiety is attached covalently to the specified group that prevents a chemical change to the group during a chemical or enzymatic process. Whenever the specified group is a 3'-hydroxyl of a nucleoside triphosphate, or an extended fragment (or "extension intermediate") in which a 3'-protected (or blocked)-nucleoside triphosphate has been incorporated, the prevented chemical change is a further, or subsequent, extension of the extended fragment (or "extension intermediate") by an enzymatic coupling reaction.

[0027] As used herein, an "initiator" (or equivalent terms, such as, "initiating fragment," "initiator nucleic acid," "initiator oligonucleotide," or the like) usually refers to a short oligonucleotide sequence with a free 3'-hydroxyl at its end, which can be further elongated by a template-free polymerase, such as TdT. In one embodiment, the initiating fragment is a DNA initiating fragment. In an alternative embodiment, the initiating fragment is an RNA initiating fragment. In some embodiments, an initiating fragment possesses between 3 and 100 nucleotides, in particular between 3 and 20 nucleotides. In some embodiments, the initiating fragment is single-stranded. In alternative embodiments, the initiating fragment may be double-stranded. In some embodiments, an initiator oligonucleotide may be attached to a synthesis support by its 5'end; and in other embodiments, an initiator oligonucleotide may be attached indirectly to a synthesis support by forming a duplex with a complementary oligonucleotide that is directly attached to the synthesis support, e.g. through a covalent bond. In some embodiments a synthesis support is a solid support which may be a discrete region of a solid planar solid, or may be a bead.

[0028] In some embodiments, an initiator may comprise a non-nucleic acid compound having a free hydroxyl to which a TdT may couple a 3'-O-protected dNTP, e.g. Baiga, U.S. patent publications US2019/0078065 and US2019/0078126.

[0029] After synthesis is completed polynucleotides with the desired nucleotide sequence may be released from initiators and the solid supports by cleavage. A wide variety of cleavable linkages or cleavable nucleotides may be used for this purpose. In some embodiments, cleaving the desired polynucleotide leaves a natural free 5'-hydroxyl on a cleaved strand; however, in alternative embodiments, a cleaving step

may leave a moiety, e.g. a 5' -phosphate, that may be removed in a subsequent step, e.g. by phosphatase treatment.

[0030] Returning to FIG. 1A, in some embodiments, an ordered sequence of nucleotides are coupled to an initiator nucleic acid using a template-free polymerase, such as TdT, in the presence of 3'-O-protected dNTPs in each synthesis step. In some embodiments, the method of synthesizing an oligonucleotide comprises the steps of (a) providing an initiator having a free 3'-hydroxyl (100); (b) reacting (104) under extension conditions the initiator or an extension intermediate having a free 3'-hydroxyl with a template-free polymerase in the presence of a 3'-O-protected nucleoside triphosphate to produce a 3'-O-protected extension intermediate (106); (c) deprotecting the extension intermediate to produce an extension intermediate with a free 3'-hydroxyl (108); and (d) repeating steps (b) and (c) (110) until the polynucleotide is synthesized. (The terms "extension intermediate" and "elongation fragment" may be used interchangeably). In some embodiments, an initiator is provided as an oligonucleotide attached to a solid support, e.g. by its 5' end. The above method may also include washing step after each reaction, or extension, step, as well as after each de-protecting step. For example, the step of reacting may include a sub-step of removing unincorporated nucleoside triphosphates, e.g. by washing, after a predetermined incubation period, or reaction time. Such predetermined incubation periods or reaction times may be a few seconds, e.g. 30 sec, to several minutes, e.g. 30 min.

[0031] When the sequence of polynucleotides on a synthesis support includes reverse complementary subsequences, secondary intra-molecular or cross-molecular structures may be created by the formation of hydrogen bonds between the reverse complementary regions. In some embodiments, base protecting moieties for exocyclic amines are selected so that hydrogens of the protected nitrogen cannot participate in hydrogen bonding, thereby preventing the formation of such secondary structures. That is, base protecting moieties may be employed to prevent the formation of hydrogen bonds, such as are formed in normal base pairing, for example, between nucleosides A and T and between G and C. At the end of a synthesis, the base protecting moieties may be removed and the polynucleotide product may be cleaved from the solid support, for example, by cleaving it from its initiator.

[0032] In addition to providing 3'-O-blocked dNTP monomers with base protection groups, elongation reactions may be performed at higher temperatures using thermal stable template-free polymerases. For example, a thermal stable template-free polymerase having activity above 40° C. may be employed; or, in some embodiments, a thermal stable template-free polymerase having activity in the range of from 40-85° C. may be employed; or, in some embodiments, a thermal stable template-free polymerase having activity in the range of from 40-65° C. may be employed.

[0033] In some embodiments, elongation conditions may include adding solvents to an elongation reaction mixture that inhibit hydrogen bonding or base stacking. Such solvents include water miscible solvents with low dielectric constants, such as dimethyl sulfoxide (DMSO), methanol, and the like. Likewise, in some embodiments, elongation conditions may include the provision of chaotropic agents that include, but are not limited to, n-butanol, ethanol, guanidinium chloride, lithium perchlorate, lithium acetate, magnesium chloride, phenol, 2-propanol, sodium dodecyl

sulfate, thiourea, urea, and the like. In some embodiments, elongation conditions include the presence of a secondary-structure-suppressing amount of DMSO. In some embodiments, elongation conditions may include the provision of DNA binding proteins that inhibit the formation of secondary structures, wherein such proteins include, but are not limited to, single-stranded binding proteins, helicases, DNA glycolases, and the like.

[0034] 3'-O-blocked dNTPs without base protection may be purchased from commercial vendors or synthesized using published techniques, e.g. U.S. Pat. No. 7,057,026; Guo et al, Proc. Natl. Acad. Sci., 105(27): 9145-9150 (2008); Benner, U.S. Pat. Nos. 7,544,794 and 8,212,020; International patent publications WO2004/005667, WO91/06678; Canard et al, Gene (cited herein); Metzker et al, Nucleic Acids Research, 22: 4259-4267 (1994); Meng et al, J. Org. Chem., 14: 3248-3252 (3006); U.S. patent publication 2005/037991. 3'-O-blocked dNTPs with base protection may be synthesized as described below.

[0035] When base-protected dNTPs are employed the above method of FIG. 1A may further include a step (e) removing base protecting moieties, which in the case of acyl or amidine protection groups may (for example) include treating with concentrated ammonia. The above method may also include one or more capping steps in addition to washing steps after the reacting, or extending, step. A first capping step may cap, or render inert to further extensions, unreacted 3'-OH groups on partially synthesized polynucleotides. Such capping step is usually implemented after a coupling step, and whenever a capping compound is used, it is selected to be unreactive with protection groups of the monomer just coupled to the growing strands. In some embodiments, such capping steps may be implemented by coupling (for example, by a second enzymatic coupling step) a capping compound that renders the partially synthesized polynucleotide incapable of further couplings, e.g. with TdT. Such capping compounds may be a dideoxynucleoside triphosphate. In other embodiments, non-extended strands with free 3'-hydroxyls may be degraded by treating them with a 3' -exonuclease activity, e.g. Exo I. For example, see Hyman, U.S. Pat. No. 5,436,143. Likewise, in some embodiments, strands that fail to be deblocked may be treated to either remove the strand or render it inert to further extensions. A second capping step may be implemented after a deprotection step, to render the affected strands inert from any subsequent coupling or deprotection any 3'-O protection, or blocking groups. Capping compounds of such second capping step are selected so that they do not react with free 3'-hydroxyls that may be present. In some embodiments, such second capping compound may be a conjugate of an aldehyde group and a hydrophobic group. The latter group permits separation based on hydrophobicity, e.g. Andrus, U.S. Pat. No. 5,047,524.

[0036] In some embodiments, reaction conditions for an elongation step (also sometimes referred to as an extension step or a coupling step) may comprising the following: 2.0 μ M purified TdT; 125-600 μ M 3'-O-blocked dNTP (e.g. 3'-O—NH₂-blocked dNTP); about 10 to about 500 mM potassium cacodylate buffer (pH between 6.5 and 7.5) and from about 0.01 to about 10 mM of a divalent cation (e.g. CoCl₂ or MnCl₂), where the elongation reaction may be carried out in a 50 μ L reaction volume, at a temperature within the range RT to 45° C., for 3 minutes. In embodiments, in which the 3'-O-blocked dNTPs are 3'-O—NH₂-

blocked dNTPs, reaction conditions for a deblocking step may comprise the following: 700 mM NaNO₂; 1 M sodium acetate (adjusted with acetic acid to pH in the range of 4.8-6.5), where the deblocking reaction may be carried out in a 50 μL volume, at a temperature within the range of RT to 45° C. for 30 seconds to several minutes. Washes may be performed with the cacodylate buffer without the components of the coupling reaction (e.g. enzyme, monomer, divalent cations).

[0037] Depending on particular applications, the steps of deblocking and/or cleaving may include a variety of chemical or physical conditions, e.g. light, heat, pH, presence of specific reagents, such as enzymes, which are able to cleave a specified chemical bond. Guidance in selecting 3'-O-blocking groups and corresponding de-blocking conditions may be found in the following references, which are incorporated by reference: Benner, U.S. Pat. Nos. 7,544,794 and 8,212,020; 5,808,045; 8,808,988; International patent publication WO91/06678; and references cited below. In some embodiments, the cleaving agent (also sometimes referred to as a de-blocking reagent or agent) is a chemical cleaving agent, such as, for example, dithiothreitol (DTT). In alternative embodiments, a cleaving agent may be an enzymatic cleaving agent, such as, for example, a phosphatase, which may cleave a 3'-phosphate blocking group. It will be understood by the person skilled in the art that the selection of deblocking agent depends on the type of 3'-nucleotide blocking group used, whether one or multiple blocking groups are being used, whether initiators are attached to living cells or organisms or to solid supports, and the like, that necessitate mild treatment. For example, a phosphine, such as tris(2-carboxyethyl)phosphine (TCEP) can be used to cleave a 3'-O-azidomethyl groups, palladium complexes can be used to cleave a 3'-O-allyl groups, or sodium nitrite can be used to cleave a 3'-O-amino group. In particular embodiments, the cleaving reaction involves TCEP, a palladium complex or sodium nitrite.

[0038] As noted above, in some embodiments it is desirable to employ two or more blocking groups that may be removed using orthogonal de-blocking conditions. The following exemplary pairs of blocking groups may be used in parallel synthesis embodiments. It is understood that other blocking group pairs, or groups containing more than two, may be available for use in these embodiments of the invention.

3'-O-NH ₂	3'-O-azidomethyl
3'-O-NH ₂	3'-O-allyl, 3'-O-propargyl
3'-O-NH ₂	3'-O-phosphate
3'-O-azidomethyl	3'-O-allyl, 3'-O-propargyl
3'-O-azidomethyl	3'-O-phosphate
3'-O-allyl, 3'-O-propargyl	3'-O-phosphate

[0039] Synthesizing oligonucleotides on living cells requires mild deblocking, or deprotection, conditions, that is, conditions that do not disrupt cellular membranes, denature proteins, interfere with key cellular functions, or the like. In some embodiments, deprotection conditions are within a range of physiological conditions compatible with cell survival. In such embodiments, enzymatic deprotection is desirable because it may be carried out under physiological conditions. In some embodiments specific enzymatically removable blocking groups are associated with specific enzymes for their removal. For example, ester- or acyl-based

blocking groups may be removed with an esterase, such as acylesterase, or like enzyme, and a phosphate blocking group may be removed with a 3' phosphatase, such as T4 polynucleotide kinase. By way of example, 3'-O-phosphates may be removed by treatment with a solution of 100 mM Tris-HCl (pH 6.5) 10 mM MgCl₂, 5 mM 2-mercaptoethanol, and one Unit T4 polynucleotide kinase. The reaction proceeds for one minute at a temperature of 37° C.

[0040] A "3'-phosphate-blocked" or "3'-phosphate-protected" nucleotide refers to nucleotides in which the hydroxyl group at the 3'-position is blocked by the presence of a phosphate containing moiety. Examples of 3'-phosphate-blocked nucleotides in accordance with the invention are nucleotidyl-3'-phosphate monoester/nucleotidyl-2',3'-cyclic phosphate, nucleotidyl-2'-phosphate monoester and nucleotidyl-2' or 3'-alkylphosphate diester, and nucleotidyl-2' or 3'-pyrophosphate. Thiophosphate or other analogs of such compounds can also be used, provided that the substitution does not prevent dephosphorylation resulting in a free 3'-OH by a phosphatase.

[0041] Further examples of synthesis and enzymatic deprotection of 3'-O-ester-protected dNTPs or 3'-O-phosphate-protected dNTPs are described in the following references: Canard et al, Proc. Natl. Acad. Sci., 92:10859-10863 (1995); Canard et al, Gene, 148: 1-6 (1994); Cameron et al, Biochemistry, 16(23): 5120-5126 (1977); Rasolonjatovo et al, Nucleosides & Nucleotides, 18(4&5): 1021-1022 (1999); Ferrero et al, Monatshefte fur Chemie, 131: 585-616 (2000); Taunton-Rigby et al, J. Org. Chem., 38(5): 977-985 (1973); Uemura et al, Tetrahedron Lett., 30(29): 3819-3820 (1989); Becker et al, J. Biol. Chem., 242(5): 936-950 (1967); Tsien, International patent publication W01991/006678.

In some embodiments, the modified nucleotides comprise a modified nucleotide or nucleoside molecule comprising a purine or pyrimidine base and a ribose or deoxyribose sugar moiety having a removable 3'-OH blocking group covalently attached thereto, such that the 3' carbon atom has attached a group of the structure:

$$-O-Z$$

wherein -Z is any of $-C(R')_2-O-R''$, $-C(R')_2-N(R'')_2$, $-C(R')_2-N(H)R''$, $-C(R')_2-S-R''$ and $-C(R')_2-F$, wherein each R'' is or is part of a removable protecting group; each R' is independently a hydrogen atom, an alkyl, substituted alkyl, arylalkyl, alkenyl, alkynyl, aryl, heteroaryl, heterocyclic, acyl, cyano, alkoxy, aryloxy, heteroaryloxy or amido group, or a detectable label attached through a linking group; with the proviso that in some embodiments such substituents have up to 10 carbon atoms and/or up to 5 oxygen or nitrogen heteroatoms; or (R')₂ represents a group of formula $=C(R''')_2$ wherein each R''' may be the same or different and is selected from the group comprising hydrogen and halogen atoms and alkyl groups, with the proviso that in some embodiments the alkyl of each R'' has from 1 to 3 carbon atoms; and wherein the molecule may be reacted to yield an intermediate in which each R'' is exchanged for H or, where Z is $-(R')_2-F$, the F is exchanged for OH, SH or NH₂, preferably OH, which intermediate dissociates under aqueous conditions to afford a molecule with a free 3'-OH; with the proviso that where Z is $-C(R')_2-S-R''$, both R' groups are not H. In certain embodiments, R' of the modified nucleotide or nucleoside is an alkyl or substituted alkyl, with the proviso that such alkyl or substituted alkyl has from 1 to 10 carbon atoms and from

0 to 4 oxygen or nitrogen heteroatoms. In certain embodiments, -Z of the modified nucleotide or nucleoside is of formula $-C(R')_2-N_3$. In certain embodiments, Z is an azidomethyl group.

[0042] In some embodiments, Z is a cleavable organic moiety with or without heteroatoms having a molecular weight of 200 or less. In other embodiments, Z is a cleavable organic moiety with or without heteroatoms having a molecular weight of 100 or less. In other embodiments, Z is a cleavable organic moiety with or without heteroatoms having a molecular weight of 50 or less. In some embodiments, Z is an enzymatically cleavable organic moiety with or without heteroatoms having a molecular weight of 200 or less. In other embodiments, Z is an enzymatically cleavable organic moiety with or without heteroatoms having a molecular weight of 100 or less. In other embodiments, Z is an enzymatically cleavable organic moiety with or without heteroatoms having a molecular weight of 50 or less. In other embodiments, Z is an enzymatically cleavable ester group having a molecular weight of 200 or less. In other embodiments, Z is a phosphate group removable by a 3'-phosphatase. In some embodiments, one or more of the following 3'-phosphatases may be used with the manufacturer's recommended protocols: T4 polynucleotide kinase, calf intestinal alkaline phosphatase, recombinant shrimp alkaline phosphatase (e.g. available from New England Biolabs, Beverly, MA).

[0043] In a further embodiment, the 3'-blocked nucleotide triphosphate is blocked by either a 3'-O-azidomethyl, 3'-O-NH₂ or 3'-O-allyl group. In other embodiments, 3'-O-blocking groups of the invention include 3'-O-methyl, 3'-O-(2-nitrobenzyl), 3'-O-allyl, 3'-O-amine, 3'-O-azidomethyl, 3'-O-tert-butoxy ethoxy, 3'-O-(2-cyanoethyl), 3'-O-nitro, and 3'-O-propargyl. In other embodiments, the 3'-blocked nucleotide triphosphate is blocked by either a 3'-O-azidomethyl or a 3'-O-NH₂. Synthesis and use of such 3'-blocked nucleoside triphosphates are disclosed in the following references: U.S. Pat. Nos. 9,410,197; 8,808,988; 6,664,097; 5,744,595; 7,544,794; 8,034,923; 8,212,020; 10,472,383; Guo et al, Proc. Natl. Acad. Sci., 105(27): 9145-9150 (2008); and like references.

[0044] In some embodiments, 3'-O- protection groups are electrochemically labile groups. That is, deprotection or cleavage of the protection group is accomplished by changing the electrochemical conditions in the vicinity of the protection group which result in cleavage. Such changes in electrochemical conditions may be brought about by changing or applying a physical quantity, such as a voltage difference or light to activate auxiliary species which, in turn, cause changes in the electrochemical conditions at the site of the protection group, such as an increase or decrease in pH. In some embodiments, electrochemically labile groups include, for example, pH-sensitive protection groups that are cleaved whenever the pH is changed to a predetermined value. In other embodiments, electrochemically labile groups include protecting groups which are cleaved directly whenever reducing or oxidizing conditions are changed, for example, by increasing or decreasing a voltage difference at the site of the protection group.

[0045] In some embodiments, enzymatic synthesis methods employ TdT variants that display increased incorporation activity with respect to 3'-O-modified nucleoside triphosphates. For example, such TdT variants may be produced using techniques described in Champion et al, U.S. Pat. No. 10,435,676, which is incorporated herein by reference. In some embodiments, a TdT variant is employed having an amino acid sequence at least 60 percent identical to SEQ ID NO: 2 and a substitution at a first arginine at position 207 and a substitution at a second arginine at position 325, or functionally equivalent residues thereof. In some embodiments, a terminal deoxynucleotidyl transferase (TdT) variant is employed that has an amino acid sequence at least sixty percent identical to an amino acid sequence selected from SEQ ID NO: 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14 or 15 with a substitution of arginine ("first arginine") at position 207 with respect to SEQ ID NOs 2, 3, 4, 6, 7, 9, 12 and 13, at position 206 with respect to SEQ ID NO 5, at position 208 with respect to SEQ ID NOs 8 and 10, at position 205 with respect to SEQ ID NO 11, at position 216 with respect to SEQ ID NO 14 and at position 210 with respect to SEQ ID NO 15; and a substitution of arginine ("second arginine") at position 325 with respect to SEQ ID NOs 2, 9 and 13, at position 324 with respect to SEQ ID NOs 3 and 4, at position 320 with respect to SEQ ID NO 320, at position 331 with respect to SEQ ID NOs 6 and 8, at position 323 with respect to SEQ ID NO 11, at position 328 with respect to SEQ ID NOs 12 and 15, and at position 338 with respect to SEQ ID NO 14; or functionally equivalent residues thereof; wherein the TdT variant (i) is capable of synthesizing a nucleic acid fragment without a template and (ii) is capable of incorporating a 3'-O-modified nucleotide onto a free 3'-hydroxyl of a nucleic acid fragment. In some embodiments, the above percent identity value is at least 80 percent identity with the indicated SEQ ID NOs; in some embodiments, the above percent identity value is at least 90 percent identity with the indicated SEQ ID NOs; in some embodiments, the above percent identity value is at least 95 percent identity with the indicated SEQ ID NOs; in some embodiments, the above percent identity value is at least 97 percent identity; in some embodiments, the above percent identity value is at least 98 percent identity; in some embodiments, the above percent identity value is at least 99 percent identity. As used herein, the percent identity values used to compare a reference sequence to a variant sequence do not include the expressly specified amino acid positions containing substitutions of the variant sequence; that is, the percent identity relationship is between sequences of a reference protein and sequences of a variant protein outside of the expressly specified positions containing substitutions in the variant. Thus, for example, if the reference sequence and the variant sequence each comprised 100 amino acids and the variant sequence had mutations at positions 25 and 81, then the percent homology would be in regard to sequences 1-24, 26-80 and 82-100.

[0046] In regard to (ii), such 3'-O-modified nucleotide may comprise a 3'-O-NH₂-nucleoside triphosphate, a 3'-O-azidomethyl-nucleoside triphosphate, a 3'-O-allyl-nucleoside triphosphate, a 3'-O-(2-nitrobenzyl)-nucleoside triphosphate, or a 3'-O-propargyl-nucleoside triphosphate.

[0047] In some embodiments, the above TdT variants have substitutions at the first and second arginines as shown in Table 1.

TABLE 1

SEQ ID NO	Substitutions				
1	M192R/Q	C302G/R	R336L/N	R454P/N/A/V	E457N/L/T/S/K
2	M63R/Q	C173G/R	R207L/N	R325P/N/A/V	E328N/L/T/S/K
3	M63R/Q	C173G/R	R207L/N	R324P/N/A/V	E327N/L/T/S/K
4	M63R/Q	C173G/R	R207L/N	R324P/N/A/V	E327N/L/T/S/K
5	—	C172G/R	R206L/N	R320P/N/A/V	—
6	M63R/Q	C173G/R	R207L/N	R331P/N/A/V	E334N/L/T/S/K
7	M63R/Q	C173G/R	R207L/N	—	E328N/L/T/S/K
8	—	C174G/R	R208L/N	R331P/N/A/V	E334N/L/T/S/K
9	M73R/Q	C173G/R	R207L/N	R325P/N/A/V	E328N/L/T/S/K
10	M64R/Q	C174G/R	R208L/N	—	E329N/L/T/S/K
11	M61R/Q	C171G/R	R205L/N	R323P/N/A/V	E326N/L/T/S/K
12	M63R/Q	C173G/R	R207L/N	R328P/N/A/V	E331N/L/T/S/K
13	—	C173G/R	R207L/N	R325P/N/A/V	E328N/L/T/S/K
14	M63R/Q	C182G/R	R216L/N	R338P/N/A/V	E341N/L/T/S/K
15	M66R/Q	C176G/R	R210L/N	R328P/N/A/V	E331N/L/T/S/K

[0048] In some embodiments, further TdT variants for use with methods of the invention include one or more of the further substitutions of methionine, cysteine or glutamic acid, as shown in Table 1.

[0049] Further specific TdT variants that may be used in methods of the invention are set forth in Table 2. Each of the TdT variants DS1001 through DS1018 of Table 2 comprises an amino acid sequence at least 60 percent identical to SEQ ID NO 2 and comprises the substitutions at the indicated positions. In some embodiments, TdT variants DS1001 through DS1018 comprises an amino acid sequence at least 80 percent identical to SEQ ID NO 2 and comprises the substitutions at the indicated positions; in some embodiments, TdT variants DS1001 through DS1018 comprises an amino acid sequence at least 90 percent identical to SEQ ID

NO 2 and comprises the substitutions at the indicated positions; in some embodiments, TdT variants DS1001 through DS1018 comprises an amino acid sequence at least 95 percent identical to SEQ ID NO 2 and comprises the substitutions at the indicated positions; in some embodiments, TdT variants DS1001 through DS1018 comprises an amino acid sequence at least 97 percent identical to SEQ ID NO 2 and comprises the substitutions at the indicated positions; in some embodiments, TdT variants DS1001 through DS1018 comprises an amino acid sequence at least 98 percent identical to SEQ ID NO 2 and comprises the substitutions at the indicated positions; in some embodiments, TdT variants DS1001 through DS1018 comprises an amino acid sequence at least 99 percent identical to SEQ ID NO 2 and comprises the substitutions at the indicated positions.

TABLE 2

Specific TdT Variants for Use with Methods of the Invention	
DS1001 (TH M27)	A17V + L52F + M63R + A108V + C173G + R207L + K265T + G284P + E289V + R325P + E328N + R351K
DS1002 (M44)	A17V + Q37E + D41R + L52F + G57E + M63R + S94R + G98E + A108V + S119A + L131R + S146E + Q149R + C173G + R207L + K265T + G284P + E289V + R325P + Q326F + E328N + H337D + R351K + W377R
DS1003	A17V + Q37E + D41R + L52F + G57E + M63R + S94R + G98E + A108V + S146E + Q149R + C173G + F193Y + V199M + M201V + R207L + K265T + G284P + E289V + Q326F + E328N + R351K
DS1004 (M45)	A17V + Q37E + D41R + L52F + G57E + M63R + S94R + G98E + A108V + S146E + Q149R + C173G + F193Y + V199M + M201V + R207L + K265T + G284P + E289V + R325A + Q326F + E328N + R351K
DS1005	A17V + Q37E + D41R + L52F + G57E + M63R + S94R + G98E + A108V + S146E + Q149R + C173G + F193Y + V199M + M201V + R207L + K265T + G284P + E289V + Q326F + E328N + R351K
DS1006 (M46)	L52F + A108V + R351K + A17V + Q37E + D41R + G57E + C59R + L60D + M63R + S94R + G98E + S119A + L131R + S146E + Q149R + C173G + R207L + K265T + G284P + E289V + R325A + Q326F + E328N
DS1007 (M47)	L52F + A108V + R351K + A17V + Q37E + D41R + G57E + C59R + L60D + M63R + S94R + G98E + K118Q + S119A + L131R + S146E + Q149R + C173G + R207L + K265T + G284P + E289V + R325A + Q326F + E328N + W377R
DS1008	A17V + Q37E + D41R + L52F + G57E + C59R + L60D + M63R + S94R + G98E + A108V + S119A + L131R + S146E + Q149R + C173G + R207L + F259S + Q261L + G284P + E289V + R325A + Q326F + E328N + R351K + W377R
DS1009 (MS 13-34)	A17V + D41R + L53F + G57E + C59R + L60D + M63R + S94R + G98E + K118Q + S119A + L131R + S146E + Q149R + C173G + R207L + K265T + G284P + E289V + R325A + Q326F + R351K + W377R

TABLE 2-continued

Specific TdT Variants for Use with Methods of the Invention	
DS1010 (MS 34-1)	A17V + D41R + L52F + G57E + C59R + L60D + M63R + S94R + G98E + A108V + S119A + L131R + S146E + Q149R + R207L + K265T + G284P + E289V + R325A + Q326F + R351K
DS1011	A17V + D41R + L53F + G57E + C59R + L60D + M63R + S94R + G98E + K118Q + S119A + L131R + S146E + Q149R + C173G + R207L + K265T + G284P + E289V + Q326F + R351K + W377R
DS1012 (M48)	A17V + Q37E + D41R + L52F + G57E + C59R + L60D + M63R + S94R + G98E + A108V + S119A + L131R + S146E + Q149R + C173G + R207L + F259S + Q261L, G284P + E289V + R325A + Q326F + E328N + R351K + W377R
DS1013	A17V + Q37E + D41R + L52F + G57E + M63R + S94R + G98E + A108V + S146E + Q149R + C173G + R207L + K265T + G284P + E289V + R325A + Q326F + E328N + R351K
DS1014 (M49)	A17V + Q37E + D41R + L52F + G57E + C59R + L60D + M63R + S94R + G98E + A108V + S119A + L131R + S146E + Q149R + C173G + R207L + E257D + F259S + K260R + Q261L + G284P + E289V + R325A + Q326F + E328N + R351K + W377R
DS1015	A17V + Q37E + D41R + L52F + G57E + C59R + L60D + M63R + S94R + G98E + A108V + S119A + L131R + S146E + Q149R + C173G + F193Y + V199M + M201V + R207L + E257D + F259S + K260R + Q261L + G284P + E289V + R325A + Q326F + E328N + R351K + W377R
DS1016 TH e2_5	A17V + D41R + L52F + G57E + M63R + S94R + G98E + A108V + S146E + Q149R + C173G + M184T + R207L + K209H + G284L + E289A + R325V + E328K + R351K
DS1017 (M27)	A17V + L52F + G57E + M63R + A108V + C173G + R207L + K265T + G284P + E289V + R325P + E328N + R351K
DS1018 (M60)	A17V + L32T + Q37R + D41R + L52F + G57E + C59R + L60D + M63R + S67A + S94R + G98E + A108V + S119A + L131R + S146E + Q149R + V171A + S172E + C173R + V182I + S183E + R207L + K209H + M210K + T211I + E223G + A224P + E228D + Q261L + G284P + E289V + R325A + Q326F + E328N + R351K + D372E

[0050] TdT variants of the invention as described above each comprise an amino acid sequence having a percent sequence identity with a specified SEQ ID NO, subject to the presence of indicated substitutions. In some embodiments, the number and type of sequence differences between a TdT variant of the invention described in this manner and the specified SEQ ID NO may be due to substitutions, deletion and/or insertions, and the amino acids substituted, deleted and/or inserted may comprise any amino acid. In some embodiments, such deletions, substitutions and/or insertions comprise only naturally occurring amino acids. In some embodiments, substitutions comprise only conservative, or synonymous, amino acid changes, as described in Grantham, Science, 185: 862-864 (1974). That is, a substitution of an amino acid can occur only among members of its set of synonymous amino acids. In some embodiments, sets of synonymous amino acids that may be employed are set forth in Table 3A.

TABLE 3A

Synonymous Sets of Amino Acids I	
Amino Acid	Synonymous Set
Ser	Ser, Thr, Gly, Asn
Arg	Arg, Gln, Lys, Glu, His
Leu	Ile, Phe, Tyr, Met, Val, Leu
Pro	Gly, Ala, Thr, Pro
Thr	Pro, Ser, Ala, Gly, His, Gln, Thr
Ala	Gly, Thr, Pro, Ala
Val	Met, Tyr, Phe, Ile, Leu, Val
Gly	Gly, Ala, Thr, Pro, Ser
Ile	Met, Tyr, Phe, Val, Leu, Ile
Phe	Trp, Met, Tyr, Ile, Val, Leu, Phe
Tyr	Trp, Met, Phe, Ile, Val, Leu, Tyr

TABLE 3A-continued

Synonymous Sets of Amino Acids I	
Amino Acid	Synonymous Set
Cys	Cys, Ser, Thr
His	His, Glu, Lys, Gln, Thr, Arg
Gln	Gln, Glu, Lys, Asn, His, Thr, Arg
Asn	Asn, Gln, Asp, Ser
Lys	Lys, Glu, Gln, His, Arg
Asp	Asp, Glu, Asn
Glu	Glu, Asp, Lys, Asn, Gln, His, Arg
Met	Met, Phe, Ile, Val, Leu
Trp	Trp

[0051] In some embodiments, sets of synonymous amino acids that may be employed are set forth in Table 3B.

TABLE 3B

Synonymous Sets of Amino Acids II	
Amino Acid	Synonymous Set
Ser	Ser
Arg	Arg, Lys, His
Leu	Ile, Phe, Met, Leu
Pro	Ala, Pro
Thr	Thr
Ala	Pro, Ala
Val	Met, Ile Val
Gly	Gly
Ile	Met, Phe, Val, Leu, Ile
Phe	Met, Tyr, Ile, Leu, Phe
Tyr	Trp, Met
Cys	Cys, Ser
His	His, Gln, Arg
Gln	Gln, Glu, His

TABLE 3B-continued

Synonymous Sets of Amino Acids II	
Amino Acid	Synonymous Set
Asn	Asn, Asp
Lys	Lys, Arg
Asp	Asp, Asn
Glu	Glu, Gln
Met	Met, Phe, Ile, Val, Leu
Trp	Trp

Kits

[0052] The invention includes kits for carrying out methods of the invention. In one aspect, a kit for synthesizing a polynucleotide probe comprises at least one 3'-O—NH₂-alkyne-nucleoside triphosphate. In a particular embodiment, such 3'-O—NH₂-alkyne-nucleoside triphosphate includes a 5-octa(1,7)diynyl-3'-aminoxy-dUTP.

[0053] In another aspect, a kit for enzymatically synthesizing a polynucleotide probe comprising at least one 3'-O-blocked-(alkyne/azide)-nucleoside triphosphate or 3'-O-blocked-(protected alkyne/protected azide)-nucleoside triphosphate and a nucleoside triphosphate having a 3'-alternative blocking group or 3'-O-alternative blocking group, wherein the alternative blocking group has a reactive moiety and may be the same or different than a 3'-O-blocking group of the 3'-O-blocked-(alkyne/azide)-nucleoside triphosphate or 3'-O-blocked-(protected alkyne/protected azide)-nucleoside triphosphate. In some embodiments, such at least one 3'-O-blocked-(alkyne/azide)-nucleoside triphosphate or 3'-O-blocked-(protected alkyne/protected azide)-nucleoside triphosphate is at least one 3'-O—NH₂-alkyne-nucleoside triphosphate, and such nucleoside triphosphate having a 3'-alternative blocking group or 3'-O-alternative blocking group is a 3'-O—NH₂-nucleoside triphosphate. In some embodiments, such kit further comprises a second label with an aldehyde group capable of reacting with the 3'-O-amine of the polynucleotide probe. In some embodiments, such kit further includes an azide-derivatized first label capable of click reacting with the alkyne group incorporated into the polynucleotide probe.

[0054] In some embodiments, the above kits of the invention may further comprise an initiator attached to a support by a 5' end and having a deoxyinosine penultimate to a 3' end and free 3'-hydroxyl. In some embodiments, a kit of the invention further includes an endonuclease V capable of cleaving an initiator-polynucleotide conjugate 3' of a terminal nucleotide of the initiator. In some such kits, the endonuclease V has a capture moiety to permit removal from a reaction mixture. In some kits, such capture moiety is a His tag. In some embodiments, initiators of a kit have a 3'-terminal sequence of 5'-dI-dT-3'. In some embodiments, initiators of a kit have a 3'-terminal sequence of 5'-dI-dG-3'. In some embodiments, initiators of a kit have a 3'-terminal sequence of 5'-dI-dA-3'. In some embodiments, initiators of a kit have a 3'-terminal sequence of 5'-dI-dT-3', 5'-dI-dG-3', or 5'-dI-dA-3'. In some embodiments, such support is a solid support. Such solid support may comprise beads, such as magnetic bead, a planar solid, such as a glass slide, or a membrane, or the like. In some embodiments, a kit of the invention may further include a template-free polymerase and 3'-O-blocked nucleoside triphosphates of one or more of

deoxyadenosine, deoxyguanosine, thymidine, deoxyuridine and deoxycytidine. In some kits, such template-free polymerase may be a TdT. In some embodiments, such TdT may be a TdT variant described herein. In some embodiments, a kit of the invention may further include a de-blocking agent which is capable of removing 3' blocking groups from incorporated 3'-O-blocked nucleotides.

Real Time PCR with Taqman Probes

[0055] Polynucleotide probes of the invention may be used in Taqman real-time PCR assays, for example, using commercially available kits. In some embodiments, a Taqman real-time PCR assay is the same as a PCR assay with the addition of a concentration of polynucleotide probe which is within about a factor of two of the primer concentrations. Preferably the amplified fragment is in the range of from about 50 to about 150 base pairs. The length of the polynucleotide probe may be in the range of from 10 to 30 nucleotides, or in the range of from 15 to 25 nucleotides, or in the range of from 20 to 22 nucleotides. Preferably the assay uses Taq DNA polymerase, or an equivalent polymerase, that has 5'->3' exonuclease activity.

Definitions

[0056] Unless otherwise specifically defined herein, terms and symbols of nucleic acid chemistry, biochemistry, genetics, and molecular biology used herein follow those of standard treatises and texts in the field, e.g. Kornberg and Baker, DNA Replication, Second Edition (W. H. Freeman, New York, 1992); Lehninger, Biochemistry, Second Edition (Worth Publishers, New York, 1975); Strachan and Read, Human Molecular Genetics, Second Edition (Wiley-Liss, New York, 1999).

[0057] "Functionally equivalent" in reference to amino acid positions in two or more different TdTs means (i) the amino acids at the respective positions play the same functional role in an activity of the TdTs, and (ii) the amino acids occur at homologous amino acid positions in the amino acid sequences of the respective TdTs. It is possible to identify positionally equivalent or homologous amino acid residues in the amino acid sequences of two or more different TdTs on the basis of sequence alignment and/or molecular modelling. In some embodiments, functionally equivalent amino acid positions belong to sequence motifs that are conserved among the amino acid sequences of TdTs of evolutionarily related species, e.g. genus, families, or the like. Examples of such conserved sequence motifs are described in Motea et al, Biochim. Biophys. Acta. 1804(5): 1151-1166 (2010); Delarue et al, EMBO J., 21: 427-439 (2002); and like references.

[0058] "Kit" refers to any delivery system for delivering materials or reagents for carrying out a method of the invention. In the context of reaction assays, such delivery systems include systems and/or compounds (such as dilutants, surfactants, carriers, or the like) that allow for the storage, transport, or delivery of reaction reagents (e.g., fluorescent labels, such as mutually quenching fluorescent labels, fluorescent label linking agents, enzymes, quenching agents, etc. in the appropriate containers) and/or supporting materials (e.g., buffers, written instructions for performing the assay etc.) from one location to another. For example, kits include one or more enclosures (e.g., boxes) containing the relevant reaction reagents and/or supporting materials. Such contents may be delivered to the intended recipient together or separately. For example, a first container may

contain an enzyme for use in an assay, while a second or more containers contain mutually quenching fluorescent labels and/or quenching agents.

[0059] “Mutant” or “variant,” which are used interchangeably, refer to polypeptides derived from a natural or reference TdT polypeptide described herein, and comprising a modification or an alteration, i.e., a substitution, insertion, and/or deletion, at one or more positions. Variants may be obtained by various techniques well known in the art. In particular, examples of techniques for altering the DNA sequence encoding the wild-type protein, include, but are not limited to, site-directed mutagenesis, random mutagenesis, sequence shuffling and synthetic oligonucleotide construction. Mutagenesis activities consist in deleting, inserting or substituting one or several amino-acids in the sequence of a protein or in the case of the invention of a polymerase. The following terminology is used to designate a substitution: L238A denotes that amino acid residue (Leucine, L) at position 238 of a reference, or wild type, sequence is changed to an Alanine (A). A132V/I/M denotes that amino acid residue (Alanine, A) at position 132 of the parent sequence is substituted by one of the following amino acids: Valine (V), Isoleucine (I), or Methionine (M). The substitution can be a conservative or non-conservative substitution. Examples of conservative substitutions are within the groups of basic amino acids (arginine, lysine and histidine), acidic amino acids (glutamic acid and aspartic acid), polar amino acids (glutamine, asparagine and threonine), hydrophobic amino acids (methionine, leucine, isoleucine, cysteine and valine), aromatic amino acids (phenylalanine, tryptophan and tyrosine), and small amino acids (glycine, alanine and serine).

[0060] “Polymerase chain reaction,” or “PCR,” means a reaction for the in vitro amplification of specific DNA sequences by the simultaneous primer extension of complementary strands of DNA. In other words, PCR is a reaction for making multiple copies or replicates of a target nucleic acid flanked by primer binding sites, such reaction comprising one or more repetitions of the following steps: (i) denaturing the target nucleic acid, (ii) annealing primers to the primer binding sites, and (iii) extending the primers by a nucleic acid polymerase in the presence of nucleoside triphosphates. Usually, the reaction is cycled through different temperatures optimized for each step in a thermal cycler instrument. Particular temperatures, durations at each step, and rates of change between steps depend on many factors well-known to those of ordinary skill in the art, e.g. exemplified by the references: McPherson et al, editors, PCR: A Practical Approach and PCR2: A Practical Approach (IRL Press, Oxford, 1991 and 1995, respectively). For example, in a conventional PCR using Taq DNA polymerase, a double stranded target nucleic acid may be denatured at a temperature $>90^{\circ}\text{C}$., primers annealed at a temperature in the range $50\text{--}75^{\circ}\text{C}$., and primers extended at a temperature in the range $72\text{--}78^{\circ}\text{C}$. The term “PCR” encompasses derivative forms of the reaction, including but not limited to, RT-PCR, real-time PCR, nested PCR, quantitative PCR, multiplexed PCR, and the like. Reaction volumes range from a few hundred nanoliters, e.g. 200 nL, to a few hundred μL , e.g. 200 μL . “Reverse transcription PCR,” or “RT-PCR,” means a PCR that is preceded by a reverse transcription reaction that converts a target RNA to a complementary single stranded DNA, which is then amplified, e.g. Tecott et al, U.S. Pat. No. 5,168,038, which patent

is incorporated herein by reference. “Real-time PCR” means a PCR for which the amount of reaction product, i.e. amplicon, is monitored as the reaction proceeds. There are many forms of real-time PCR that differ mainly in the detection chemistries used for monitoring the reaction product, e.g. Gelfand et al, U.S. Pat. No. 5,210,015 (“taqman”); Wittwer et al, U.S. Pat. Nos. 6,174,670 and 6,569,627 (intercalating dyes); Tyagi et al U.S. Pat. No. 5,925,517 (molecular beacons); which patents are incorporated herein by reference. Detection chemistries for real-time PCR are reviewed in Mackay et al, Nucleic Acids Research, 30: 1292-1305 (2002), which is also incorporated herein by reference.

[0061] “Polynucleotide” or “oligonucleotide” are used interchangeably and each mean a linear polymer of nucleotide monomers or analogs thereof. Monomers making up polynucleotides and oligonucleotides are capable of specifically binding to a natural polynucleotide by way of a regular pattern of monomer-to-monomer interactions, such as Watson-Crick type of base pairing, base stacking, Hoogsteen or reverse Hoogsteen types of base pairing, or the like. Such monomers and their internucleosidic linkages may be naturally occurring or may be analogs thereof, e.g. naturally occurring or non-naturally occurring analogs. Non-naturally occurring analogs may include PNAs, phosphorothioate internucleosidic linkages, bases containing linking groups permitting the attachment of labels, such as fluorophores, or haptens, and the like. Whenever the use of an oligonucleotide or polynucleotide requires enzymatic processing, such as extension by a polymerase, ligation by a ligase, or the like, one of ordinary skill would understand that oligonucleotides or polynucleotides in those instances would not contain certain analogs of internucleosidic linkages, sugar moieties, or bases at any or some positions. Polynucleotides typically range in size from a few monomeric units, e.g. 5-40, when they are usually referred to as “oligonucleotides,” to several thousand monomeric units. Whenever a polynucleotide or oligonucleotide is represented by a sequence of letters (upper or lower case), such as “ATGCCTG,” it will be understood that the nucleotides are in 5'→3' order from left to right and that “A” denotes deoxyadenosine, “C” denotes deoxycytidine, “G” denotes deoxyguanosine, and “T” denotes thymidine, “I” denotes deoxyinosine, “U” denotes uridine, unless otherwise indicated or obvious from context. Unless otherwise noted the terminology and atom numbering conventions will follow those disclosed in Strachan and Read, Human Molecular Genetics 2 (Wiley-Liss, New York, 1999). Usually polynucleotides comprise the four natural nucleosides (e.g. deoxyadenosine, deoxycytidine, deoxyguanosine, deoxythymidine for DNA or their ribose counterparts for RNA) linked by phosphodiester linkages; however, they may also comprise non-natural nucleotide analogs, e.g. including modified bases, sugars, or internucleosidic linkages. It is clear to those skilled in the art that where an enzyme has specific oligonucleotide or polynucleotide substrate requirements for activity, e.g. single stranded DNA, RNA/DNA duplex, or the like, then selection of appropriate composition for the oligonucleotide or polynucleotide substrates is well within the knowledge of one of ordinary skill, especially with guidance from treatises, such as Sambrook et al, Molecular Cloning, Second Edition (Cold Spring Harbor Laboratory, New York, 1989), and like references. Likewise, the oligonucleotide and polynucleotide may refer to either a

single stranded form or a double stranded form (i.e. duplexes of an oligonucleotide or polynucleotide and its respective complement). It will be clear to one of ordinary skill which form or whether both forms are intended from the context of the terms usage.

[0062] “Primer” means an oligonucleotide, either natural or synthetic that is capable, upon forming a duplex with a polynucleotide template, of acting as a point of initiation of nucleic acid synthesis and being extended from its 3' end along the template so that an extended duplex is formed. Extension of a primer is usually carried out with a nucleic acid polymerase, such as a DNA or RNA polymerase. The sequence of nucleotides added in the extension process is determined by the sequence of the template polynucleotide. Usually primers are extended by a DNA polymerase. Primers usually have a length in the range of from 14 to 40 nucleotides, or in the range of from 18 to 36 nucleotides. Primers are employed in a variety of nucleic acid amplification reactions, for example, linear amplification reactions using a single primer, or polymerase chain reactions, employing two or more primers. Guidance for selecting the lengths and sequences of primers for particular applications is well known to those of ordinary skill in the art, as evidenced by the following references that are incorporated by reference: Dieffenbach, editor, PCR Primer: A Laboratory Manual, 2nd Edition (Cold Spring Harbor Press, New York, 2003).

[0063] “Sequence identity” refers to the number (or fraction, usually expressed as a percentage) of matches (e.g., identical amino acid residues) between two sequences, such as two polypeptide sequences or two polynucleotide sequences. The sequence identity is determined by comparing the sequences when aligned so as to maximize overlap and identity while minimizing sequence gaps. In particular, sequence identity may be determined using any of a number of mathematical global or local alignment algorithms, depending on the length of the two sequences. Sequences of similar lengths are preferably aligned using a global alignment algorithm (e.g. Needleman and Wunsch algorithm; Needleman and Wunsch, 1970) which aligns the sequences optimally over the entire length, while sequences of substantially different lengths are preferably aligned using a local alignment algorithm (e.g. Smith and Waterman algorithm (Smith and Waterman, 1981) or Altschul algorithm (Altschul et al., 1997; Altschul et al., 2005)). Alignment for purposes of determining percent amino acid sequence identity can be achieved in various ways that are within the skill in the art, for instance, using publicly available computer software available on internet web sites such as <http://blast.ncbi.nlm.nih.gov/> or <http://www.ebi.ac.uk/Tools/emboss/>. Those skilled in the art can determine appropriate parameters for measuring alignment, including any algorithm needed to achieve maximal alignment over the full length of the sequences being compared. For purposes herein, % amino acid sequence identity values refer to values gener-

ated using the pair wise sequence alignment program EMBOSS Needle, that creates an optimal global alignment of two sequences using the Needleman-Wunsch algorithm, wherein all search parameters are set to default values, i.e. Scoring matrix=BLOSUM62, Gap open=10, Gap extend=0.5, End gap penalty=false, End gap open=10 and End gap extend=0.5.

[0064] A “substitution” means that an amino acid residue is replaced by another amino acid residue. Preferably, the term “substitution” refers to the replacement of an amino acid residue by another selected from the naturally-occurring standard 20 amino acid residues, rare naturally-occurring amino acid residues (e.g. hydroxyproline, hydroxylysine, allohydroxylysine, 6-N-methyllysine, N-ethylglycine, N-methylglycine, N-ethylasparagine, allo-isoleucine, N-methylisoleucine, N-methylvaline, pyroglutamine, aminobutyric acid, ornithine, norleucine, norvaline), and non-naturally occurring amino acid residue, often made synthetically, (e.g. cyclohexyl-alanine). Preferably, the term “substitution” refers to the replacement of an amino acid residue by another selected from the naturally-occurring standard 20 amino acid residues. The sign “+” indicates a combination of substitutions. The amino acids are herein represented by their one-letter or three-letters code according to the following nomenclature: A: alanine (Ala); C: cysteine (Cys); D: aspartic acid (Asp); E: glutamic acid (Glu); F: phenylalanine (Phe); G: glycine (Gly); H: histidine (His); I: isoleucine (Ile); K: lysine (Lys); L: leucine (Leu); M: methionine (Met); N: asparagine (Asn); P: proline (Pro); Q: glutamine (Gln); R: arginine (Arg); S: serine (Ser); T: threonine (Thr); V: valine (Val); W: tryptophan (Trp) and Y: tyrosine (Tyr). In the present document, the following terminology is used to designate a substitution: L238A denotes that amino acid residue (Leucine, L) at position 238 of the parent sequence is changed to an Alanine (A). A132V/I/M denotes that amino acid residue (Alanine, A) at position 132 of the parent sequence is substituted by one of the following amino acids: Valine (V), Isoleucine (I), or Methionine (M). The substitution can be a conservative or non-conservative substitution. Examples of conservative substitutions are within the groups of basic amino acids (arginine, lysine and histidine), acidic amino acids (glutamic acid and aspartic acid), polar amino acids (glutamine, asparagine and threonine), hydrophobic amino acids (methionine, leucine, isoleucine, cysteine and valine), aromatic amino acids (phenylalanine, tryptophan and tyrosine), and small amino acids (glycine, alanine and serine).

[0065] This disclosure is not intended to be limited to the scope of the particular forms set forth, but is intended to cover alternatives, modifications, and equivalents of the variations described herein. Further, the scope of the disclosure fully encompasses other variations that may become obvious to those skilled in the art in view of this disclosure. The scope of the present invention is limited only by the appended claims.

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Arg Asp Leu Val Leu Phe Ile Leu Glu Lys Lys Met Gly Thr Thr Arg
          35          40          45
Arg Ala Phe Leu Met Glu Leu Ala Arg Arg Lys Gly Phe Arg Val Glu
          50          55          60
Asn Glu Leu Ser Asp Ser Val Thr His Ile Val Ala Glu Asn Asn Ser
          65          70          75          80
Gly Ser Asp Val Leu Glu Trp Leu Gln Leu Gln Asn Ile Lys Ala Ser
          85          90          95
Ser Glu Leu Glu Leu Leu Asp Ile Ser Trp Leu Ile Glu Cys Met Gly
          100          105          110
Ala Gly Lys Pro Val Glu Met Met Gly Arg His Gln Leu Val Val Asn
          115          120          125
Arg Asn Ser Ser Pro Ser Pro Val Pro Gly Ser Gln Asn Val Pro Ala
          130          135          140
Pro Ala Val Lys Lys Ile Ser Gln Tyr Ala Cys Gln Arg Arg Thr Thr
          145          150          155          160
Leu Asn Asn Tyr Asn Gln Leu Phe Thr Asp Ala Leu Asp Ile Leu Ala
          165          170          175
Glu Asn Asp Glu Leu Arg Glu Asn Glu Gly Ser Cys Leu Ala Phe Met
          180          185          190
Arg Ala Ser Ser Val Leu Lys Ser Leu Pro Phe Pro Ile Thr Ser Met
          195          200          205
Lys Asp Thr Glu Gly Ile Pro Cys Leu Gly Asp Lys Val Lys Ser Ile
          210          215          220
Ile Glu Gly Ile Ile Glu Asp Gly Glu Ser Ser Glu Ala Lys Ala Val
          225          230          235          240
Leu Asn Asp Glu Arg Tyr Lys Ser Phe Lys Leu Phe Thr Ser Val Phe
          245          250          255
Gly Val Gly Leu Lys Thr Ala Glu Lys Trp Phe Arg Met Gly Phe Arg
          260          265          270
Thr Leu Ser Lys Ile Gln Ser Asp Lys Ser Leu Arg Phe Thr Gln Met
          275          280          285
Gln Lys Ala Gly Phe Leu Tyr Tyr Glu Asp Leu Val Ser Cys Val Asn
          290          295          300
Arg Pro Glu Ala Glu Ala Val Ser Met Leu Val Lys Glu Ala Val Val
          305          310          315          320
Thr Phe Leu Pro Asp Ala Leu Val Thr Met Thr Gly Gly Phe Arg Arg
          325          330          335
Gly Lys Met Thr Gly His Asp Val Asp Phe Leu Ile Thr Ser Pro Glu
          340          345          350
Ala Thr Glu Asp Glu Glu Gln Gln Leu Leu His Lys Val Thr Asp Phe
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Trp Lys Gln Gln Gly Leu Leu Leu Tyr Cys Asp Ile Leu Glu Ser Thr
          370          375          380

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 Ser Glu Lys Ser Gly Gln Gln Glu Gly Lys Gly Trp Lys Ala Ile Arg
 420 425 430
 Val Asp Leu Val Met Cys Pro Tyr Asp Arg Arg Ala Phe Ala Leu Leu
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 Gly Trp Thr Gly Ser Arg Gln Phe Glu Arg Asp Leu Arg Arg Tyr Ala
 450 455 460
 Thr His Glu Arg Lys Met Met Leu Asp Asn His Ala Leu Tyr Asp Arg
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 <220> FEATURE:
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 35 40 45
 Asn Asp Glu Leu Arg Glu Asn Glu Gly Ser Cys Leu Ala Phe Met Arg
 50 55 60
 Ala Ser Ser Val Leu Lys Ser Leu Pro Phe Pro Ile Thr Ser Met Lys
 65 70 75 80
 Asp Thr Glu Gly Ile Pro Cys Leu Gly Asp Lys Val Lys Ser Ile Ile
 85 90 95
 Glu Gly Ile Ile Glu Asp Gly Glu Ser Ser Glu Ala Lys Ala Val Leu
 100 105 110
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 115 120 125
 Val Gly Leu Lys Thr Ala Glu Lys Trp Phe Arg Met Gly Phe Arg Thr
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 145 150 155 160
 Lys Ala Gly Phe Leu Tyr Tyr Glu Asp Leu Val Ser Cys Val Asn Arg
 165 170 175
 Pro Glu Ala Glu Ala Val Ser Met Leu Val Lys Glu Ala Val Val Thr
 180 185 190
 Phe Leu Pro Asp Ala Leu Val Thr Met Thr Gly Gly Phe Arg Arg Gly
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 Lys Met Thr Gly His Asp Val Asp Phe Leu Ile Thr Ser Pro Glu Ala
 210 215 220

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Thr Glu Asp Glu Glu Gln Gln Leu Leu His Lys Val Thr Asp Phe Trp
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 Lys Gln Gln Gly Leu Leu Leu Tyr Cys Asp Ile Leu Glu Ser Thr Phe
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 Glu Lys Phe Lys Gln Pro Ser Arg Lys Val Asp Ala Leu Asp His Phe
 260 265 270
 Gln Lys Cys Phe Leu Ile Leu Lys Leu Asp His Gly Arg Val His Ser
 275 280 285
 Glu Lys Ser Gly Gln Gln Glu Gly Lys Gly Trp Lys Ala Ile Arg Val
 290 295 300
 Asp Leu Val Met Cys Pro Tyr Asp Arg Arg Ala Phe Ala Leu Leu Gly
 305 310 315 320
 Trp Thr Gly Ser Arg Gln Phe Glu Arg Asp Leu Arg Arg Tyr Ala Thr
 325 330 335
 His Glu Arg Lys Met Met Leu Asp Asn His Ala Leu Tyr Asp Arg Thr
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 Lys Arg Val Phe Leu Glu Ala Glu Ser Glu Glu Glu Ile Phe Ala His
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 35 40 45
 Asn Ser Glu Phe Lys Glu Asn Glu Val Ser Tyr Val Thr Phe Met Arg
 50 55 60
 Ala Ala Ser Val Leu Lys Ser Leu Pro Phe Thr Ile Ile Ser Met Lys
 65 70 75 80
 Asp Thr Glu Gly Ile Pro Cys Leu Gly Asp Lys Val Lys Cys Ile Ile
 85 90 95
 Glu Glu Ile Ile Glu Asp Gly Glu Ser Ser Glu Val Lys Ala Val Leu
 100 105 110
 Asn Asp Glu Arg Tyr Gln Ser Phe Lys Leu Phe Thr Ser Val Phe Gly
 115 120 125
 Val Gly Leu Lys Thr Ser Glu Lys Trp Phe Arg Met Gly Phe Arg Ser
 130 135 140
 Leu Ser Lys Ile Met Ser Asp Lys Thr Leu Lys Phe Thr Lys Met Gln
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 Lys Ala Gly Phe Leu Tyr Tyr Glu Asp Leu Val Ser Cys Val Thr Arg
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 Ala Glu Ala Glu Ala Val Gly Val Leu Val Lys Glu Ala Val Trp Ala
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  210                                215                                220

Ala Glu Asp Glu Glu Gln Leu Leu Pro Lys Val Ile Asn Leu Trp Glu
  225                                230                                235                                240

Lys Lys Gly Leu Leu Leu Tyr Tyr Asp Leu Val Glu Ser Thr Phe Glu
                                245                                250                                255

Lys Phe Lys Leu Pro Ser Arg Gln Val Asp Thr Leu Asp His Phe Gln
                                260                                265                                270

Lys Cys Phe Leu Ile Leu Lys Leu His His Gln Arg Val Asp Ser Ser
                                275                                280                                285

Lys Ser Asn Gln Gln Glu Gly Lys Thr Trp Lys Ala Ile Arg Val Asp
  290                                295                                300

Leu Val Met Cys Pro Tyr Glu Asn Arg Ala Phe Ala Leu Leu Gly Trp
  305                                310                                315                                320

Thr Gly Ser Arg Gln Phe Glu Arg Asp Ile Arg Arg Tyr Ala Thr His
                                325                                330                                335

Glu Arg Lys Met Met Leu Asp Asn His Ala Leu Tyr Asp Lys Thr Lys
                                340                                345                                350

Arg Val Phe Leu Lys Ala Glu Ser Glu Glu Glu Ile Phe Ala His Leu
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Asn Cys Glu Phe Arg Glu Asn Glu Asp Ser Cys Val Thr Phe Met Arg
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Ala Ala Ser Val Leu Lys Ser Leu Pro Phe Thr Ile Ile Ser Met Lys
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Asp Thr Glu Gly Ile Pro Cys Leu Gly Ser Lys Val Lys Gly Ile Ile
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Glu Glu Ile Ile Glu Asp Gly Glu Ser Ser Glu Val Lys Ala Val Leu
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Asn Asp Glu Arg Tyr Gln Ser Phe Lys Leu Phe Thr Ser Val Phe Gly
  115         120         125

Val Gly Leu Lys Thr Ser Glu Lys Trp Phe Arg Met Gly Phe Arg Thr
  130         135         140

Leu Ser Lys Val Arg Ser Asp Lys Ser Leu Lys Phe Thr Arg Met Gln
  145         150         155         160

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Lys Ala Gly Phe Leu Tyr Tyr Glu Asp Leu Val Ser Cys Val Thr Arg
 165 170 175
 Ala Glu Ala Glu Ala Val Ser Val Leu Val Lys Glu Ala Val Trp Ala
 180 185 190
 Phe Leu Pro Asp Ala Phe Val Thr Met Thr Gly Gly Phe Arg Arg Gly
 195 200 205
 Lys Lys Met Gly His Asp Val Asp Phe Leu Ile Thr Ser Pro Gly Ser
 210 215 220
 Thr Glu Asp Glu Glu Gln Leu Leu Gln Lys Val Met Asn Leu Trp Glu
 225 230 235 240
 Lys Lys Gly Leu Leu Leu Tyr Tyr Asp Leu Val Glu Ser Thr Phe Glu
 245 250 255
 Lys Leu Arg Leu Pro Ser Arg Lys Val Asp Ala Leu Asp His Phe Gln
 260 265 270
 Lys Cys Phe Leu Ile Phe Lys Leu Pro Arg Gln Arg Val Asp Ser Asp
 275 280 285
 Gln Ser Ser Trp Gln Glu Gly Lys Thr Trp Lys Ala Ile Arg Val Asp
 290 295 300
 Leu Val Leu Cys Pro Tyr Glu Arg Arg Ala Phe Ala Leu Leu Gly Trp
 305 310 315 320
 Thr Gly Ser Arg Gln Phe Glu Arg Asp Leu Arg Arg Tyr Ala Thr His
 325 330 335
 Glu Arg Lys Met Ile Leu Asp Asn His Ala Leu Tyr Asp Lys Thr Lys
 340 345 350
 Arg Ile Phe Leu Lys Ala Glu Ser Glu Glu Glu Ile Phe Ala His Leu
 355 360 365
 Gly Leu Asp Tyr Ile Glu Pro Trp Glu Arg Asn Ala
 370 375 380

<210> SEQ ID NO 5

<211> LENGTH: 376

<212> TYPE: PRT

<213> ORGANISM: ARTIFICIAL SEQUENCE

<220> FEATURE:

<223> OTHER INFORMATION: Chicken 1 truncated

<400> SEQUENCE: 5

Gln Tyr Pro Thr Leu Lys Thr Pro Glu Ser Glu Val Ser Ser Phe Thr
 1 5 10 15
 Ala Ser Lys Val Ser Gln Tyr Ser Cys Gln Arg Lys Thr Thr Leu Asn
 20 25 30
 Asn Cys Asn Lys Lys Phe Thr Asp Ala Phe Glu Ile Met Ala Glu Asn
 35 40 45
 Tyr Glu Phe Lys Glu Asn Glu Ile Phe Cys Leu Glu Phe Leu Arg Ala
 50 55 60
 Ala Ser Val Leu Lys Ser Leu Pro Phe Pro Val Thr Arg Met Lys Asp
 65 70 75 80
 Ile Gln Gly Leu Pro Cys Met Gly Asp Arg Val Arg Asp Val Ile Glu
 85 90 95
 Glu Ile Ile Glu Glu Gly Glu Ser Ser Arg Ala Lys Asp Val Leu Asn
 100 105 110
 Asp Glu Arg Tyr Lys Ser Phe Lys Glu Phe Thr Ser Val Phe Gly Val
 115 120 125

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Gly Val Lys Thr Ser Glu Lys Trp Phe Arg Met Gly Leu Arg Thr Val
 130 135 140
 Glu Glu Val Lys Ala Asp Lys Thr Leu Lys Leu Ser Lys Met Gln Arg
 145 150 155 160
 Ala Gly Phe Leu Tyr Tyr Glu Asp Leu Val Ser Cys Val Ser Lys Ala
 165 170 175
 Glu Ala Asp Ala Val Ser Ser Ile Val Lys Asn Thr Val Cys Thr Phe
 180 185 190
 Leu Pro Asp Ala Leu Val Thr Ile Thr Gly Gly Phe Arg Arg Gly Lys
 195 200 205
 Lys Ile Gly His Asp Ile Asp Phe Leu Ile Thr Ser Pro Gly Gln Arg
 210 215 220
 Glu Asp Asp Glu Leu Leu His Lys Gly Leu Leu Leu Tyr Cys Asp Ile
 225 230 235 240
 Ile Glu Ser Thr Phe Val Lys Glu Gln Ile Pro Ser Arg His Val Asp
 245 250 255
 Ala Met Asp His Phe Gln Lys Cys Phe Ala Ile Leu Lys Leu Tyr Gln
 260 265 270
 Pro Arg Val Asp Asn Ser Ser Tyr Asn Met Ser Lys Lys Cys Asp Met
 275 280 285
 Ala Glu Val Lys Asp Trp Lys Ala Ile Arg Val Asp Leu Val Ile Thr
 290 295 300
 Pro Phe Glu Gln Tyr Ala Tyr Ala Leu Leu Gly Trp Thr Gly Ser Arg
 305 310 315 320
 Gln Phe Gly Arg Asp Leu Arg Arg Tyr Ala Thr His Glu Arg Lys Met
 325 330 335
 Met Leu Asp Asn His Ala Leu Tyr Asp Lys Arg Lys Arg Val Phe Leu
 340 345 350
 Lys Ala Gly Ser Glu Glu Glu Ile Phe Ala His Leu Gly Leu Asp Tyr
 355 360 365
 Val Glu Pro Trp Glu Arg Asn Ala
 370 375

<210> SEQ ID NO 6
 <211> LENGTH: 387
 <212> TYPE: PRT
 <213> ORGANISM: ARTIFICIAL SEQUENCE
 <220> FEATURE:
 <223> OTHER INFORMATION: Possum truncated

<400> SEQUENCE: 6

Ser Ala Asn Pro Asp Pro Thr Ala Gly Thr Leu Asn Ile Leu Pro Pro
 1 5 10 15
 Thr Thr Lys Thr Ile Ser Gln Tyr Ala Cys Gln Arg Arg Thr Thr Ile
 20 25 30
 Asn Asn His Asn Gln Arg Phe Thr Asp Ala Phe Glu Ile Leu Ala Lys
 35 40 45
 Asn Tyr Glu Phe Lys Glu Asn Asp Asp Thr Cys Leu Thr Phe Met Arg
 50 55 60
 Ala Ile Ser Val Leu Lys Cys Leu Pro Phe Glu Val Val Ser Leu Lys
 65 70 75 80
 Asp Thr Glu Gly Leu Pro Trp Ile Gly Asp Glu Val Lys Gly Ile Met
 85 90 95

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Glu Glu Ile Ile Glu Asp Gly Glu Ser Leu Glu Val Gln Ala Val Leu
 100 105 110

Asn Asp Glu Arg Tyr Gln Ser Phe Lys Leu Phe Thr Ser Val Phe Gly
 115 120 125

Val Gly Leu Lys Thr Ala Asp Lys Trp Tyr Arg Met Gly Phe Arg Thr
 130 135 140

Leu Asn Lys Ile Arg Ser Asp Lys Thr Leu Lys Leu Thr Lys Met Gln
 145 150 155 160

Lys Ala Gly Leu Cys Tyr Tyr Glu Asp Leu Ile Asp Cys Val Ser Lys
 165 170 175

Ala Glu Ala Asp Ala Val Ser Leu Leu Val Gln Asp Ala Val Trp Thr
 180 185 190

Phe Leu Pro Asp Ala Leu Val Thr Ile Thr Gly Gly Phe Arg Arg Gly
 195 200 205

Lys Glu Phe Gly His Asp Val Asp Phe Leu Ile Thr Ser Pro Gly Ala
 210 215 220

Glu Lys Glu Gln Glu Asp Gln Leu Leu Gln Lys Val Thr Asn Leu Trp
 225 230 235 240

Lys Lys Gln Gly Leu Leu Leu Tyr Cys Asp Leu Ile Glu Ser Thr Phe
 245 250 255

Glu Asp Leu Lys Leu Pro Ser Arg Lys Ile Asp Ala Leu Asp His Phe
 260 265 270

Gln Lys Cys Phe Leu Ile Leu Lys Leu Tyr His His Lys Glu Asp Lys
 275 280 285

Arg Lys Trp Glu Met Pro Thr Gly Ser Asn Glu Ser Glu Ala Lys Ser
 290 295 300

Trp Lys Ala Ile Arg Val Asp Leu Val Val Cys Pro Tyr Asp Arg Tyr
 305 310 315 320

Ala Phe Ala Leu Leu Gly Trp Ser Gly Ser Arg Gln Phe Glu Arg Asp
 325 330 335

Leu Arg Arg Tyr Ala Thr His Glu Lys Lys Met Met Leu Asp Asn His
 340 345 350

Ala Leu Tyr Asp Lys Thr Lys Lys Ile Phe Leu Lys Ala Lys Ser Glu
 355 360 365

Glu Glu Ile Phe Ala His Leu Gly Leu Glu Tyr Ile Gln Pro Ser Glu
 370 375 380

Arg Asn Ala
 385

<210> SEQ ID NO 7
 <211> LENGTH: 381
 <212> TYPE: PRT
 <213> ORGANISM: ARTIFICIAL SEQUENCE
 <220> FEATURE:
 <223> OTHER INFORMATION: New truncated shrew

<400> SEQUENCE: 7

Asp Cys Pro Ala Ser His Asp Ser Ser Pro Gln Lys Thr Glu Ser Ala
 1 5 10 15

Ala Val Gln Lys Ile Ser Gln Tyr Ala Cys Gln Arg Arg Thr Thr Leu
 20 25 30

Asn Asn His Asn His Ile Phe Thr Asp Ala Phe Glu Ile Leu Ala Glu
 35 40 45

-continued

Leu Asp Arg Asp Ser Ile Ser Glu Tyr Ala Cys Gln Arg Arg Thr Thr
 20 25 30
 Leu Lys Asn Tyr Asn Gln Lys Phe Thr Asp Ala Phe Glu Ile Leu Ala
 35 40 45
 Glu Asn Tyr Glu Phe Asn Glu Asn Lys Gly Phe Cys Thr Ala Phe Arg
 50 55 60
 Arg Ala Ala Ser Val Leu Lys Cys Leu Pro Phe Thr Ile Val Gln Val
 65 70 75 80
 His Asp Ile Glu Gly Val Pro Trp Met Gly Lys Gln Val Lys Gly Ile
 85 90 95
 Ile Glu Asp Ile Ile Glu Glu Gly Glu Ser Ser Lys Val Lys Ala Val
 100 105 110
 Leu Asp Asn Glu Asn Tyr Arg Ser Val Lys Leu Phe Thr Ser Val Phe
 115 120 125
 Gly Val Gly Leu Lys Thr Ser Asp Lys Trp Tyr Arg Met Gly Leu Arg
 130 135 140
 Thr Leu Glu Glu Val Lys Arg Asp Lys Asn Leu Lys Leu Thr Arg Met
 145 150 155 160
 Gln Lys Ala Gly Phe Leu His Tyr Asp Asp Leu Thr Ser Cys Val Ser
 165 170 175
 Lys Ala Glu Ala Asp Ala Ala Ser Leu Ile Val Gln Asp Val Val Trp
 180 185 190
 Lys Ile Val Pro Asn Ala Ile Val Thr Ile Ala Gly Gly Phe Arg Arg
 195 200 205
 Gly Lys Gln Thr Gly His Asp Val Asp Phe Leu Ile Thr Val Pro Gly
 210 215 220
 Ser Lys Gln Glu Glu Glu Glu Leu Leu His Thr Val Ile Asp Ile Trp
 225 230 235 240
 Lys Lys Gln Glu Leu Leu Leu Tyr Tyr Asp Leu Ile Glu Ser Thr Phe
 245 250 255
 Glu Asp Thr Lys Leu Pro Ser Arg Lys Val Asp Ala Leu Asp His Phe
 260 265 270
 Gln Lys Cys Phe Ala Ile Leu Lys Val His Lys Glu Arg Glu Asp Lys
 275 280 285
 Gly Asn Ser Ile Arg Ser Lys Ala Phe Ser Glu Glu Glu Ile Lys Asp
 290 295 300
 Trp Lys Ala Ile Arg Val Asp Leu Val Val Val Pro Phe Glu Gln Tyr
 305 310 315 320
 Ala Phe Ala Leu Leu Gly Trp Thr Gly Ser Thr Gln Phe Glu Arg Asp
 325 330 335
 Leu Arg Arg Tyr Ala Thr His Glu Lys Lys Met Met Leu Asp Asn His
 340 345 350
 Ala Leu Tyr Asp Lys Thr Lys Lys Ile Phe Leu Asn Ala Ala Ser Glu
 355 360 365
 Glu Glu Ile Phe Ala His Leu Gly Leu Asp Tyr Leu Glu Pro Trp Glu
 370 375 380
 Arg Asn Ala
 385

<210> SEQ ID NO 9

<211> LENGTH: 381

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<212> TYPE: PRT
<213> ORGANISM: ARTIFICIAL SEQUENCE
<220> FEATURE:
<223> OTHER INFORMATION: truncated dog

<400> SEQUENCE: 9

Asp Tyr Thr Ala Ser Pro Asn Pro Glu Leu Gln Lys Thr Leu Pro Val
1          5          10          15
Ala Val Lys Lys Ile Ser Gln Tyr Ala Cys Gln Arg Arg Thr Thr Leu
20          25          30
Asn Asn Tyr Asn Asn Val Phe Thr Asp Ala Phe Glu Val Leu Ala Glu
35          40          45
Asn Tyr Glu Phe Arg Glu Asn Glu Val Phe Ser Leu Thr Phe Met Arg
50          55          60
Ala Ala Ser Val Leu Lys Ser Leu Pro Phe Thr Ile Ile Ser Met Lys
65          70          75          80
Asp Thr Glu Gly Ile Pro Cys Leu Gly Asp Gln Val Lys Cys Ile Ile
85          90          95
Glu Glu Ile Ile Glu Asp Gly Glu Ser Ser Glu Val Lys Ala Val Leu
100         105         110
Asn Asp Glu Arg Tyr Gln Ser Phe Lys Leu Phe Thr Ser Val Phe Gly
115         120         125
Val Gly Leu Lys Thr Ser Glu Lys Trp Phe Arg Met Gly Phe Arg Thr
130         135         140
Leu Ser Lys Ile Lys Ser Asp Lys Ser Leu Lys Phe Thr Pro Met Gln
145         150         155         160
Lys Ala Gly Phe Leu Tyr Tyr Glu Asp Leu Val Ser Cys Val Thr Arg
165         170         175
Ala Glu Ala Glu Ala Val Gly Val Leu Val Lys Glu Ala Val Gly Ala
180         185         190
Phe Leu Pro Asp Ala Phe Val Thr Met Thr Gly Gly Phe Arg Arg Gly
195         200         205
Lys Lys Met Gly His Asp Val Asp Phe Leu Ile Thr Ser Pro Gly Ser
210         215         220
Thr Asp Glu Asp Glu Glu Gln Leu Leu Pro Lys Val Ile Asn Leu Trp
225         230         235         240
Glu Arg Lys Gly Leu Leu Leu Tyr Cys Asp Leu Val Glu Ser Thr Phe
245         250         255
Glu Lys Leu Lys Leu Pro Ser Arg Lys Val Asp Ala Leu Asp His Phe
260         265         270
Gln Lys Cys Phe Leu Ile Leu Lys Leu His His Gln Arg Val Asp Gly
275         280         285
Gly Lys Cys Ser Gln Gln Glu Gly Lys Thr Trp Lys Ala Ile Arg Val
290         295         300
Asp Leu Val Met Cys Pro Tyr Glu Arg Arg Ala Phe Ala Leu Leu Gly
305         310         315         320
Trp Thr Gly Ser Arg Gln Phe Glu Arg Asp Leu Arg Arg Tyr Ala Ser
325         330         335
His Glu Arg Lys Met Ile Leu Asp Asn His Ala Leu Tyr Asp Lys Thr
340         345         350
Lys Lys Ile Phe Leu Lys Ala Glu Ser Glu Glu Glu Ile Phe Ala His
355         360         365

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Leu Gly Leu Asp Tyr Ile Glu Pro Trp Glu Arg Asn Ala
 370 375 380

<210> SEQ ID NO 10
 <211> LENGTH: 382
 <212> TYPE: PRT
 <213> ORGANISM: ARTIFICIAL SEQUENCE
 <220> FEATURE:
 <223> OTHER INFORMATION: TRUNC MOLE

<400> SEQUENCE: 10

Gly Asp Cys Pro Ala Ser His Asp Ser Ser Pro Gln Lys Thr Glu Ser
 1 5 10 15
 Ala Ala Val Gln Lys Ile Ser Gln Tyr Ala Cys Gln Arg Arg Thr Thr
 20 25 30
 Leu Asn Asn His Asn His Ile Phe Thr Asp Ala Phe Glu Ile Leu Ala
 35 40 45
 Glu Asn Cys Glu Phe Arg Glu Asn Glu Gly Ser Tyr Val Thr Tyr Met
 50 55 60
 Arg Ala Ala Ser Val Leu Lys Ser Leu Pro Phe Ser Ile Ile Ser Met
 65 70 75 80
 Lys Asp Thr Glu Gly Ile Pro Cys Leu Ala Asp Lys Val Lys Cys Val
 85 90 95
 Ile Glu Glu Ile Ile Glu Asp Gly Glu Ser Ser Glu Val Lys Ala Val
 100 105 110
 Leu Asn Asp Glu Arg Tyr Lys Ser Phe Lys Leu Phe Thr Ser Val Phe
 115 120 125
 Gly Val Gly Leu Lys Thr Ala Glu Lys Trp Phe Arg Leu Gly Phe Arg
 130 135 140
 Thr Leu Ser Gly Ile Met Asn Asp Lys Thr Leu Lys Leu Thr His Met
 145 150 155 160
 Gln Lys Ala Gly Phe Leu Tyr Tyr Glu Asp Leu Val Ser Cys Val Thr
 165 170 175
 Arg Ala Glu Ala Glu Ala Val Gly Val Leu Val Lys Glu Ala Val Trp
 180 185 190
 Ala Phe Leu Pro Asp Ala Ile Val Thr Met Thr Gly Gly Phe Arg Arg
 195 200 205
 Gly Lys Lys Val Gly His Asp Val Asp Phe Leu Ile Thr Ser Pro Glu
 210 215 220
 Ala Thr Glu Glu Gln Glu Gln Gln Leu Leu His Lys Val Ile Thr Phe
 225 230 235 240
 Trp Glu Lys Glu Gly Leu Leu Leu Tyr Cys Asp Leu Tyr Glu Ser Thr
 245 250 255
 Phe Glu Lys Leu Lys Met Pro Ser Arg Lys Val Asp Ala Leu Asp His
 260 265 270
 Phe Gln Lys Cys Phe Leu Ile Leu Lys Leu His Arg Glu Cys Val Asp
 275 280 285
 Asp Gly Thr Ser Ser Gln Leu Gln Gly Lys Thr Trp Lys Ala Ile Arg
 290 295 300
 Val Asp Leu Val Val Cys Pro Tyr Glu Cys Arg Ala Phe Ala Leu Leu
 305 310 315 320
 Gly Trp Thr Gly Ser Pro Gln Phe Glu Arg Asp Leu Arg Arg Tyr Ala
 325 330 335

-continued

Thr His Glu Arg Lys Met Met Leu Asp Asn His Ala Leu Tyr Asp Lys
 340 345 350

Thr Lys Arg Lys Phe Leu Ser Ala Asp Ser Glu Glu Asp Ile Phe Ala
 355 360 365

His Leu Gly Leu Asp Tyr Ile Glu Pro Trp Glu Arg Asn Ala
 370 375 380

<210> SEQ ID NO 11
 <211> LENGTH: 379
 <212> TYPE: PRT
 <213> ORGANISM: ARTIFICIAL SEQUENCE
 <220> FEATURE:
 <223> OTHER INFORMATION: Pika trunk

<400> SEQUENCE: 11

Glu Tyr Ser Ala Asn Pro Ser Pro Gly Pro Gln Ala Thr Pro Ala Val
 1 5 10 15

Tyr Lys Ile Ser Gln Tyr Ala Cys Gln Arg Arg Thr Thr Leu Asn Asn
 20 25 30

His Asn His Ile Phe Thr Asp Ala Phe Glu Ile Leu Ala Glu Asn Tyr
 35 40 45

Glu Phe Lys Glu Asn Glu Gly Cys Tyr Val Thr Tyr Met Arg Ala Ala
 50 55 60

Ser Val Leu Lys Ser Leu Pro Phe Thr Ile Val Ser Met Lys Asp Thr
 65 70 75 80

Glu Gly Ile Pro Cys Leu Glu Asp Lys Val Lys Ser Ile Met Glu Glu
 85 90 95

Ile Ile Glu Glu Gly Glu Ser Ser Glu Val Lys Ala Val Leu Ser Asp
 100 105 110

Glu Arg Tyr Gln Cys Phe Lys Leu Phe Thr Ser Val Phe Gly Val Gly
 115 120 125

Leu Lys Thr Ser Glu Lys Trp Phe Arg Met Gly Phe Arg Ser Leu Ser
 130 135 140

Asn Ile Arg Leu Asp Lys Ser Leu Lys Phe Thr Gln Met Gln Lys Ala
 145 150 155 160

Gly Phe Arg Tyr Tyr Glu Asp Ile Val Ser Cys Val Thr Arg Ala Glu
 165 170 175

Ala Glu Ala Val Asp Val Leu Val Asn Glu Ala Val Arg Ala Phe Leu
 180 185 190

Pro Asp Ala Phe Ile Thr Met Thr Gly Gly Phe Arg Arg Gly Lys Lys
 195 200 205

Ile Gly His Asp Val Asp Phe Leu Ile Thr Ser Pro Glu Leu Thr Glu
 210 215 220

Glu Asp Glu Gln Gln Leu Leu His Lys Val Met Asn Leu Trp Glu Lys
 225 230 235 240

Lys Gly Leu Leu Leu Tyr His Asp Leu Val Glu Ser Thr Phe Glu Lys
 245 250 255

Leu Lys Gln Pro Ser Arg Lys Val Asp Ala Leu Asp His Phe Gln Lys
 260 265 270

Cys Phe Leu Ile Phe Lys Leu Tyr His Glu Arg Val Gly Gly Asp Arg
 275 280 285

Cys Arg Gln Pro Glu Gly Lys Asp Trp Lys Ala Ile Arg Val Asp Leu
 290 295 300

-continued

Gln Lys Cys Phe Leu Ile Leu Lys Leu His Leu Gln His Val Asn Gly
 275 280 285

Val Gly Asn Ser Lys Thr Gly Gln Gln Glu Gly Lys Asn Trp Lys Ala
 290 295 300

Ile Arg Val Asp Leu Val Met Cys Pro Tyr Glu Arg Arg Ala Phe Ala
 305 310 315 320

Leu Leu Gly Trp Thr Gly Ser Arg Gln Phe Glu Arg Asp Leu Arg Arg
 325 330 335

Phe Ala Thr His Glu Arg Lys Met Met Leu Asp Asn His Ala Leu Tyr
 340 345 350

Asp Lys Thr Lys Arg Ile Phe Leu Lys Ala Glu Ser Glu Glu Glu Ile
 355 360 365

Phe Ala His Leu Gly Leu Asp Tyr Ile Asp Pro Trp Glu Arg Asn Ala
 370 375 380

<210> SEQ ID NO 13
 <211> LENGTH: 381
 <212> TYPE: PRT
 <213> ORGANISM: ARTIFICIAL SEQUENCE
 <220> FEATURE:
 <223> OTHER INFORMATION: truncated tree shrew

<400> SEQUENCE: 13

Asp His Ser Thr Ser Pro Ser Pro Gly Pro Gln Lys Thr Pro Ala Leu
 1 5 10 15

Ala Val Gln Lys Ile Ser Gln Tyr Ala Cys Gln Arg Arg Thr Thr Leu
 20 25 30

Asn Asn Cys Asn Arg Val Phe Thr Asp Ala Phe Glu Thr Leu Ala Glu
 35 40 45

Asn Tyr Glu Phe Arg Glu Asn Glu Asp Ser Ser Val Ile Phe Leu Arg
 50 55 60

Ala Ala Ser Val Leu Arg Ser Leu Pro Phe Thr Ile Thr Ser Met Arg
 65 70 75 80

Asp Thr Glu Gly Leu Pro Cys Leu Gly Asp Lys Val Lys Cys Val Ile
 85 90 95

Glu Glu Ile Ile Glu Asp Gly Glu Ser Ser Glu Val Asn Ala Val Leu
 100 105 110

Asn Asp Glu Arg Tyr Lys Ser Phe Lys Leu Phe Thr Ser Val Phe Gly
 115 120 125

Val Gly Leu Lys Thr Ser Glu Lys Trp Phe Arg Met Gly Phe Arg Thr
 130 135 140

Leu Ser Arg Val Arg Ser Asp Lys Ser Leu His Leu Thr Arg Met Gln
 145 150 155 160

Gln Ala Gly Phe Leu Tyr Tyr Glu Asp Leu Ala Ser Cys Val Thr Arg
 165 170 175

Ala Glu Ala Glu Ala Val Gly Val Leu Val Lys Glu Ala Val Gly Ala
 180 185 190

Phe Leu Pro Asp Ala Leu Val Thr Ile Thr Gly Gly Phe Arg Arg Gly
 195 200 205

Lys Lys Thr Gly His Asp Val Asp Phe Leu Ile Thr Ser Pro Gly Ser
 210 215 220

Thr Glu Glu Lys Glu Glu Glu Leu Leu Gln Lys Val Leu Asn Leu Trp
 225 230 235 240

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Thr Leu Thr Gly Gly Phe Arg Arg Gly Lys Lys Ile Gly His Asp Val
 210 215 220

Asp Phe Leu Ile Ser Asp Pro Glu Ser Gly Gln Asp Glu Gln Leu Leu
 225 230 235 240

Pro Asn Ile Ile Lys Leu Trp Glu Lys Gln Glu Leu Leu Leu Tyr Tyr
 245 250 255

Asp Leu Val Glu Ser Thr Phe Glu Lys Thr Lys Ile Pro Ser Arg Lys
 260 265 270

Val Asp Ala Met Asp His Phe Gln Lys Cys Phe Leu Ile Leu Lys Leu
 275 280 285

His His Gln Lys Val Asp Ser Gly Arg Tyr Lys Pro Pro Pro Glu Ser
 290 295 300

Lys Asn His Glu Ala Lys Asn Trp Lys Ala Ile Arg Val Asp Leu Val
 305 310 315 320

Met Cys Pro Phe Glu Gln Tyr Ala Tyr Ala Leu Leu Gly Trp Thr Gly
 325 330 335

Ser Arg Gln Phe Glu Arg Asp Leu Arg Arg Tyr Ala Thr His Glu Lys
 340 345 350

Lys Met Met Leu Asp Asn His Ala Leu Tyr Asp Lys Thr Lys Lys Ile
 355 360 365

Phe Leu Lys Ala Glu Ser Glu Glu Asp Ile Phe Thr His Leu Gly Leu
 370 375 380

Asp Tyr Ile Glu Pro Trp Glu Arg Asn Ala
 385 390

<210> SEQ ID NO 15
 <211> LENGTH: 384
 <212> TYPE: PRT
 <213> ORGANISM: ARTIFICIAL SEQUENCE
 <220> FEATURE:
 <223> OTHER INFORMATION: TRUNCATED JERBOA

<400> SEQUENCE: 15

Ser Ser Glu Leu Glu Leu Leu Asp Val Ser Trp Leu Ile Glu Cys Met
 1 5 10 15

Gly Ala Gly Lys Pro Val Glu Met Thr Gly Arg His Gln Leu Val Lys
 20 25 30

Gln Thr Phe Cys Leu Pro Gly Phe Ile Leu Gln Asp Ala Phe Asp Ile
 35 40 45

Leu Ala Glu Asn Cys Glu Phe Arg Glu Asn Glu Ala Ser Cys Val Glu
 50 55 60

Phe Met Arg Ala Ala Ser Val Leu Lys Ser Leu Pro Phe Pro Ile Ile
 65 70 75 80

Ser Val Lys Asp Thr Glu Gly Ile Pro Trp Leu Gly Gly Lys Val Lys
 85 90 95

Cys Val Ile Glu Glu Ile Ile Glu Asp Gly Glu Ser Ser Glu Val Lys
 100 105 110

Ala Leu Leu Asn Asp Glu Arg Tyr Lys Ser Phe Lys Leu Phe Thr Ser
 115 120 125

Val Phe Gly Val Gly Leu Lys Thr Ala Glu Arg Trp Phe Arg Met Gly
 130 135 140

Phe Arg Thr Leu Ser Thr Val Lys Leu Asp Lys Ser Leu Thr Phe Thr
 145 150 155 160

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Arg	Met	Gln	Lys	Ala	Gly	Phe	Leu	His	Tyr	Glu	Asp	Leu	Val	Ser	Cys
				165					170					175	
Val	Thr	Arg	Ala	Glu	Ala	Glu	Ala	Val	Ser	Val	Leu	Val	Gln	Gln	Ala
			180					185					190		
Val	Val	Ala	Phe	Leu	Pro	Asp	Ala	Leu	Val	Ser	Met	Thr	Gly	Gly	Phe
		195					200					205			
Arg	Arg	Gly	Lys	Lys	Ile	Gly	His	Asp	Val	Asp	Phe	Leu	Ile	Thr	Ser
	210					215					220				
Pro	Glu	Ala	Thr	Glu	Glu	Glu	Gln	Gln	Leu	Leu	His	Lys	Val	Thr	
225				230					235					240	
Asn	Phe	Trp	Glu	Gln	Lys	Gly	Leu	Leu	Leu	Tyr	Cys	Asp	His	Val	Glu
			245						250					255	
Ser	Thr	Phe	Glu	Lys	Cys	Lys	Leu	Pro	Ser	Arg	Lys	Val	Asp	Ala	Leu
		260						265					270		
Asp	His	Phe	Gln	Lys	Cys	Phe	Leu	Ile	Leu	Lys	Leu	Tyr	Arg	Glu	Arg
		275					280						285		
Val	Asp	Ser	Val	Lys	Ser	Ser	Gln	Gln	Glu	Gly	Lys	Gly	Trp	Lys	Ala
	290					295					300				
Ile	Arg	Val	Asp	Leu	Val	Met	Cys	Pro	Tyr	Glu	Cys	Arg	Ala	Phe	Ala
305					310					315					320
Leu	Leu	Gly	Trp	Thr	Gly	Ser	Arg	Gln	Phe	Glu	Arg	Asp	Leu	Arg	Arg
				325					330					335	
Tyr	Ala	Thr	His	Glu	Arg	Lys	Met	Arg	Leu	Asp	Asn	His	Ala	Leu	Tyr
			340					345					350		
Asp	Lys	Thr	Lys	Arg	Val	Phe	Leu	Lys	Ala	Glu	Ser	Glu	Glu	Glu	Ile
		355					360					365			
Phe	Ala	His	Leu	Gly	Leu	Glu	Tyr	Ile	Glu	Pro	Leu	Glu	Arg	Asn	Ala
	370					375					380				

1-18. (canceled)

19. A method of synthesizing a polynucleotide probe having a predetermined sequence and a plurality of labels, the method comprising the steps of:

- providing an initiator having a 3'-penultimate deoxyinosine and a 3'-terminal nucleotide having a free 3' -hydroxyl;
- repeating elongation cycles comprising steps of: (i) contacting under elongation conditions the initiator or elongated fragments having free 3'-O-hydroxyls with a 3'-O-blocked nucleoside triphosphate and a template-independent DNA polymerase so that the initiator or elongated fragments are elongated by incorporation of a 3'-O-blocked nucleoside triphosphate to form 3'-O-blocked elongated fragments, and (ii) deblocking the elongated fragments to form elongated fragments having free 3' -hydroxyls, until the polynucleotide probe is formed, wherein (A) at least one of the 3'-O-blocked nucleoside triphosphate is a 3'-O-blocked-(alkyne/azide)-nucleoside triphosphate or a 3'-O-blocked-(protected alkyne/protected azide)-nucleoside triphosphate, (B) in a final elongation cycle a nucleoside triphosphate having a 3' -alternative blocking group or 3'-O-alternative blocking group is contacted with said elongated fragments having free 3'-hydroxyls, (C) the alternative blocking group has a reactive moiety and

may be the same or different than the blocking group, and (D) said step of deblocking is omitted;

- reacting a first label having an (alkyne/azide) group with the at least one incorporated 3' -blocked-(alkyne/azide)-nucleoside triphosphate;
- reacting the reactive moiety of the 3'-alternative blocking group or 3'-O-alternative blocking group with a second label having a complementary moiety to form a polynucleotide probe having a plurality of labels; and
- enzymatically cleaving the polynucleotide probe from the initiator.

20. The method of claim **19**, wherein said step of enzymatically cleaving comprises treating said polynucleotide probe with an endonuclease V activity.

21. The method of claim **20**, wherein (i) said endonuclease V activity is provided by a prokaryotic endonuclease V, (ii) said template-independent DNA polymerase is a terminal deoxynucleotidyl transferase (TdT), and (iii) said initiator is attached to a solid support by a 5' end.

22. The method of claim **21**, wherein said prokaryotic endonuclease V is an *E. coli* endonuclease V.

23. The method of claim **21**, further including a step of removing said prokaryotic endonuclease V from said cleaved polynucleotide probe.

24. The method of claim **21**, wherein said initiator has a 3'-terminal sequence of 5'-dI-dT-3'.

25. The method of claim 21, wherein said TdT is a TdT variant.

26. The method of claim 19, wherein each of said first labels is selected from the group consisting of fluorescent dyes and fluorescent quenchers such that at least one of said first labels is a fluorescent dye.

27. The method of claim 19, wherein said 3'-O-blocked-(alkyne/azide)-nucleoside triphosphate or said 3'-O-blocked-(protected alkyne/protected azide)-nucleoside triphosphate is a 3'-O-blocked-alkyne-nucleoside triphosphate or a 3'-O-blocked-protected alkyne-nucleoside triphosphate and wherein each of said first labels has an azide group.

28. The method of claim 19, wherein said plurality of labels comprises two labels and said first label is a fluorescent dye and said second label is a fluorescent quencher, and wherein said 3'-O-blocked-alkyne-nucleoside triphosphate or 3'-O-blocked-protected alkyne-nucleoside triphosphate is a 3'-O-blocked-alkyne-nucleoside triphosphate.

29. The method of claim 28, wherein said 3'-O-blocked-alkyne-nucleoside triphosphate is incorporated immediately after said terminal nucleotide of said initiator.

30. The method of claim 19, wherein said 3'-O-blocked-nucleoside triphosphates are 3'-O—NH₂-nucleoside triphosphates.

31. The method of claim 19, wherein said 3' -alternative blocking group or 3'-O-alternative blocking group is 3' -azido or 3'-O-azido and said second label is a fluorescent quencher whose said complementary moiety is an alkyne.

32. The method of claim 19, wherein said 3' -alternative blocking group or 3'-O-alternative blocking group is 3'-O—

NH₂ and wherein said second label is a fluorescent quencher whose said complementary moiety is an aldehyde.

33. A kit for enzymatically synthesizing a polynucleotide probe comprising at least one 3'-O-blocked-(alkyne/azide)-nucleoside triphosphate or 3'-O-blocked-(protected alkyne/protected azide)-nucleoside triphosphate and a nucleoside triphosphate having a 3'-alternative blocking group or 3'-O-alternative blocking group, wherein the alternative blocking group has a reactive moiety and may be the same or different than a 3'-O-blocking group of the 3'-O-blocked-(alkyne/azide)-nucleoside triphosphate or 3'-O-blocked-(protected alkyne/protected azide)-nucleoside triphosphate.

34. The kit of claim 33, further comprising at least one first label having an (alkyne/azide) group and a second label having a complementary moiety to said reactive moiety.

35. The kit of claim 33, wherein (i) said at least one 3'-O-blocked-(alkyne/azide)-nucleoside triphosphate or 3'-O-blocked-(protected alkyne/protected azide)-nucleoside triphosphate is at least one 3'-O—NH₂-alkyne-nucleoside triphosphate, (ii) each of said at least one first labels has an azide group, (iii) said 3'-alternative blocking group or 3'-O-alternative blocking group is 3'-O—NH₂, and (iv) said complementary moiety of said second label is an aldehyde group.

36. The kit of claim 33, wherein at least one of said first labels is a fluorescent dye and said second label is a fluorescent quencher capable of quenching fluorescent emissions of the fluorescent dye.

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