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(54) **CELLULASE PREPARATIONS CONTAINING
REDUCING AGENT AND METHOD OF
PROCESSING FIBER**

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(57) **ABSTRACT**

A cellulase preparation comprising an endoglucanase derived from Zygomycetes, a cellulose-binding-domain-deleted endoglucanase, or a modified or homologous protein thereof, together with a reducing agent is disclosed. Further, a method of treating cellulose-containing fabric, comprising the step of treating the fabric with the cellulase preparation to improve a property of the fabric, a method of deinking waste paper, comprising the step of treating the waste paper with the cellulase preparation, together with a deinking agent, and a method of improving freeness of paper pulp, comprising the step of treating the paper pulp with the cellulase preparation, are disclosed.

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**CELLULASE PREPARATIONS CONTAINING
REDUCING AGENT AND METHOD OF
PROCESSING FIBER**

TECHNICAL FIELD

[0001] The present invention relates to a cellulase preparation containing a reducing agent by which an endoglucanase activity is improved, and a method of treating a fabric using the cellulose preparation.

[0002] 1. Background Art

[0003] Cellulase has three types of enzyme activities: a cellobiohydrolase activity which hydrolyzes solid crystal regions of cellulose from the nonreduced end in the exo manner to generate cellobiose; an endoglucanase activity which hydrolyzes amorphous regions of cellulose in the endo manner to transform cellulose molecules into low molecular weight molecules and to generate various types of cellooligosaccharides; and a β -glucosidase activity which decomposes cellobiose or cellooligosaccharides into glucose. Of these enzyme activities, it is known that cellulase having a high endoglucanase activity is advantageous when treating a fabric.

[0004] To impart desired properties to cellulose-containing fabric, the fabric has conventionally been treated with cellulase. For example, in the textile industry, treatment with cellulase is carried out to improve the touch and appearance of the cellulose-containing fabric, or to impart a "stone-wash" appearance to the colored cellulose-containing fabric, thereby providing the fabric with localized color variations (EP Patent No. 307,564).

[0005] In such textile processing, cellulase derived from wood-rotting fungi such as *Trichoderma* or *Humicola* is mainly used. Such cellulase was used as a mixture comprising multiple cellulase components obtained by processing a culture filtrate of microorganisms having a cellulolytic activity. However, in order to achieve a greater economy, among cellulase preparations obtained by isolation from cellulase components, only endoglucanase, which mainly acts in fabric treatment, and which is genetically enhanced, has recently been used.

[0006] Examples of such an endoglucanase with a high activity include: EGV [Unexamined International Publication (Kohyo) No. 5-509223] and NCE4 (WO98/03640) derived from *Humicola insolens*, which strongly act on cotton fabrics; RCE I, RCE II, and RCE III derived from *Rhizopus oryzae*, which strongly act on lyocell fabrics; MCE I and MCE II derived from *Mucor circinelloides*; and PCE I derived from *Phycomyces nitens* (WO00/24879).

[0007] To improve the effects of the cellulase, the combined use of additives has also been attempted. For example, Unexamined International Publication (Kohyo) No. 5-507615 describes that a water-soluble polymer such as polyvinylpyrrolidone, polyvinyl alcohol, and polyacrylamide enhances the effects of *Humicola insolens*-derived cellulase and improves its activity of removing fuzz from colored fabrics. Further, it is known that a CMCase activity in the culture solution of *Trichoderma viride* is improved by the addition of Tween 20 (Ooshima, H. et al., *Biotechnology and Bioengineering* 28: 1727-1734, 1986). Furthermore, it is shown that the fuzz-removing activity of RCE I derived

from *Rhizopus oryzae* is improved in the presence of a nonionic surfactant (WO02/38754).

DISCLOSURE OF THE INVENTION

[0008] The cellulases used for the above-described purposes are all expensive. Therefore, to achieve an industrial level application, the present inventors considered that a further improvement of the endoglucanase activity is desired, so that the above effects of cellulase can be more efficiently exerted. Further, where an attempt to obtain the effects of an improvement of the endoglucanase activity is carried out, the use of expensive additives increases costs in the textile processing treatment. Therefore, when selecting the additives, the present inventors considered that it is necessary to show the effects of the activity improvement by adding a low concentration of the additives, and that the additives should be readily available and inexpensive.

[0009] Therefore, the object of the present invention is to provide a cellulase preparation having the improved endoglucanase activity, which can be used in the fabric treatment for the purpose of improving cellulose-containing fabrics such that the removal of fuzz can be carried out efficiently and economically.

[0010] The present inventors have conducted intensive studies and, as a result, found that a reducing agent such as sodium thiosulfate or the like enhances the effects of Zygomycetes-derived endoglucanases such as RCE I, MCE I, PCE I, and the like at rates far higher than *Trichoderma*- and *Humicola*-derived known endoglucanases, and the present invention was completed.

[0011] The present invention relates to:

[0012] (1) a cellulase preparation comprising

[0013] an endoglucanase derived from Zygomycetes, a protein in which a cellulose binding domain is deleted in the endoglucanase, or a modified or homologous protein thereof; and a reducing agent;

[0014] (2) the cellulase preparation described in (1), wherein the Zygomycetes is a microorganism belonging to genus *Rhizopus*, *Mucor*, or *Phycomyces*;

[0015] (3) a cellulase preparation comprising at least one of:

[0016] (a) a protein consisting of any one of the amino acid sequences of SEQ ID NOS: 1 to 6;

[0017] (b) a protein consisting of an amino acid sequence in which a cellulose binding domain is deleted in any one of the amino acid sequences of SEQ ID NOS: 1 to 6; or

[0018] (c) a protein consisting of an amino acid sequence in which one or plural amino acids are deleted, substituted, inserted, or added, in any one of the amino acid sequences of SEQ ID NOS: 1 to 6 or in an amino acid sequence in which a cellulose binding domain is deleted therein, and exhibiting an endoglucanase activity; and

[0019] a reducing agent;

[0020] (4) a cellulase preparation comprising a protein encoded by at least one of:

[0021] (a) a polynucleotide which encodes any one of the amino acid sequences of SEQ ID NOS: 1 to 6;

[0022] (b) a polynucleotide which encodes a protein consisting of an amino acid sequence in which a cellulose binding domain is deleted in any one of the amino acid sequences of SEQ ID NOS: 1 to 6; or

[0023] (c) a polynucleotide which encodes a protein exhibiting an endoglucanase activity and is complementary to a polynucleotide which hybridizes under stringent conditions to a polynucleotide which encodes a protein consisting of any one of the amino acid sequences of SEQ ID NOS: 1 to 6 or a protein consisting of an amino acid sequence in which a cellulose binding domain is deleted therein; and

[0024] a reducing agent;

[0025] (5) the cellulase preparation described in any one of (1) to (4), containing 0.01 to 50% by weight of the reducing agent in the cellulase preparation;

[0026] (6) the cellulase preparation described in any one of (1) to (5), in which the reducing agent is sodium thiosulfate, sodium sulfite, or thiourea;

[0027] (7) the cellulase preparation described in any one of (1) to (6), which is a granule not having a dustability or stabilized liquid;

[0028] (8) a method of treating cellulose-containing fabric, comprising the step of:

[0029] treating the fabric with the cellulase preparation described in any one of (1) to (7) to improve a property of the fabric;

[0030] (9) the method described in (8), wherein the improvement of the property of the fabric is a color clarification;

[0031] (10) the method described in (8), wherein the improvement of the property of the fabric is a removal of fuzz;

[0032] (11) the method described in (8), wherein the improvement of the property of the fabric is an addition of a stonewash-like appearance and texture;

[0033] (12) the method described in (8), wherein the improvement of the property of the fabric is an improvement of touch and appearance;

[0034] (13) the method described in (8), wherein the improvement of the property of the fabric is a softening of the fabric;

[0035] (14) the method described in any one of (8) to (13), wherein the treatment of the fabric with the cellulase preparation is carried out by soaking or rinsing the fabric;

[0036] (15) a method of deinking waste paper, comprising the step of:

[0037] treating the waste paper with the cellulase preparation described in any one of (1) to (7) together with a deinking agent; and

[0038] (16) a method of improving a freeness of paper pulp, comprising the step of:

[0039] treating the paper pulp with the cellulase preparation described in any one of (1) to (7).

BEST MODE FOR CARRYING OUT THE INVENTION

[0040] The present invention will be explained in detail hereinafter.

[0041] [1] Cellulase Preparation

[0042] The cellulase preparation of the present invention comprises one or more reducing agents and at least one of:

[0043] (1a) an endoglucanase derived from *Zygomycetes*;

[0044] (1b) a protein in which a cellulose binding domain is deleted in the *Zygomycetes*-derived endoglucanase (1a) [hereinafter sometimes referred to as "CBD-deleted endoglucanase"]; or

[0045] (1c) a modified protein of the *Zygomycetes*-derived endoglucanase (1a) or the CBD-deleted endoglucanase (1b) [hereinafter sometimes simply referred to as "modified protein"]; or

[0046] (1d) a homologous protein of the *Zygomycetes*-derived endoglucanase (1a) or the CBD-deleted endoglucanase (1b) [hereinafter sometimes simply referred to as "homologous protein"].

[0047] The term "endoglucanase" as used herein means endo-1,4- β -glucanase (EC 3.2.1.4), which has an activity of hydrolyzing the β -1,4-glucopyranosyl bond of β -1,4-glucan.

[0048] The term "endoglucanase activity" as used herein means a CMCase activity. The term "CMCase activity" as used herein means an activity of hydrolyzing carboxymethylcellulose (CMC; Tokyo Kasei Kogyo Co., Ltd.). When a solution containing a protein (enzyme) to be assayed and CMC is incubated for a predetermined period and the amount of reducing sugar released is measured, the amount of the enzyme producing the reducing sugar corresponding to 1 μ mol of glucose per minute is defined as 1 unit of the CMCcase activity.

[0049] The endoglucanase activity can be measured, for example, by the following procedure. That is, 0.5 mL of a solution containing a protein to be assayed is added to 0.5 mL of 2% CMC solution dissolved in 50 mmol/L acetate-sodium acetate buffer (pH 6.0), and the mixture is incubated at 50° C. for 30 minutes. A concentration of reducing sugar generated in the reaction mixture is measured by the 3,5-dinitrosalicylic acid method (DNS method). More particularly, after the incubation for 30 minutes, 3.0 mL of a DNS reagent is added to 1.0 mL of the reaction mixture, the whole is incubated in a boiling water bath for 5 minutes and diluted with 8.0 mL of distilled water, and the absorbance at 540 nm is measured. A calibration curve is drawn using glucose solutions prepared by stepwise dilution, and an amount of reducing sugar generated in the enzyme reaction mixture is determined as that of converted glucose. The activity is calculated by defining the amount of the enzyme producing the reducing sugar corresponding to 1 μ mol of glucose per minute, as 1 unit.

[0050] The DNS reagent can be prepared in accordance with the disclosures in references such as Sakuzo Hukui, "Seikagaku Jikken-hou 1, Kangen-Tou no Teiryō-hou (Laboratory Manual for Biological Chemistry, Vol. 1, Assay of Reducing Sugar)", pp. 19-20, Japan Scientific Societies Press, or by the following procedure. To 300 mL of 4.5% aqueous solution of sodium hydrate, 880 mL of 1% 3,5-dinitrosalicylic acid solution and 255 g of Rochelle salt are added (Solution A). To 22 mL of 10% aqueous solution of sodium hydrate, 10 g of crystalline phenol is added, and then water is added so as to dissolve it and adjust the volume to 100 mL (Solution B). Then, 6.9 g of sodium hydrogencarbonate is dissolved in 69 mL of Solution B, and Solution A is poured thereinto. The whole is mixed with stirring so as to dissolve the Rochelle salt, allowed to stand for 2 days, and then filtrated.

[0051] As the endoglucanase derived from Zygomycetes which may be used in the present invention, there may be mentioned, for example, endoglucanases derived from *Rhizopus* sp., *Phycomyces* sp. or *Mucor* sp. More particularly, for example, RCE I (SEQ ID NO: 1), RCE II (SEQ ID NO: 2), RCE III (SEQ ID NO: 3), MCE I (SEQ ID NO: 4), MCE II (SEQ ID NO: 5), or PCE I (SEQ ID NO: 6) disclosed in WO00/24879 may be used. As the CBD-deleted endoglucanase, proteins in which the cellulose binding domain is deleted in the above-mentioned endoglucanases, such as CBD-deleted endoglucanases disclosed in WO02/42474, may be used. The CBD-deleted endoglucanases lack the cellulose binding domain, and exhibit the endoglucanase activity.

[0052] *Rhizopus oryzae* CP96001, from which the above-mentioned RCE I, RCE II, and RCE III are derived, was domestically deposited in the International Patent Organism Depository National Institute of Advanced Industrial Science and Technology (Address: AIST Tsukuba Central 6, 1-1, Higashi 1-chome Tukuba-shi, Ibaraki-ken 305-8566 Japan) on Apr. 21, 1997, and was transferred to an international deposit on Sep. 24, 1999. The international deposit number (a number in parenthesis [] following the international deposit number is a domestic deposit number) is FERM BP-6889 [FERM P-16201].

[0053] *Mucor circinelloides* CP99001, from which the above-mentioned MCE I and MCE II are derived, was domestically deposited in the International Patent Organism Depository National Institute of Advanced Industrial Science and Technology (Address: AIST Tsukuba Central 6, 1-1, Higashi 1-chome Tukuba-shi, Ibaraki-ken 305-8566 Japan) on Jul. 2, 1999, and was transferred to an international deposit on Sep. 24, 1999. The international deposit number (a number in parenthesis [] following the international deposit number is a domestic deposit number) is FERM BP-6890 [FERM P-17446].

[0054] *Phycomyces nitens* CP99002, from which the above-mentioned PCE I is derived, was domestically deposited in the International Patent Organism Depository National Institute of Advanced Industrial Science and Technology (Address: AIST Tsukuba Central 6, 1-1, Higashi 1-chome Tukuba-shi, Ibaraki-ken 305-8566 Japan) on Jul. 2, 1999, and was transferred to an international deposit on Sep. 24, 1999. The international deposit number (a number in parenthesis [] following the international deposit number is a domestic deposit number) is FERM BP-6891 [FERM P-17447].

[0055] The proteins which can be used in the cellulase preparation of the present invention include not only RCE I, RCE II, RCE III, MCE I, MCE II, and PCE I, and the CBD-deleted endoglucanases as described above, but also modified and/or homologous proteins thereof.

[0056] The term "modified protein" as used herein means a protein exhibiting the endoglucanase activity and consisting of an amino acid sequence in which one or plural (for example, one to several tens, more particularly, 1 to 50, preferably 1 to 30, more preferably 1 to 9) amino acids are deleted, substituted, inserted, or added in the amino acid sequence of RCE I, RCE II, RCE III, MCE I, MCE II, or PCE I, or any one of the CBD-deleted endoglucanases thereof.

[0057] The term "homologous protein" as used herein means a protein exhibiting the endoglucanase activity and having an amino acid sequence encoded by a polynucleotide (base sequence) complementary to a polynucleotide (base sequence) which hybridizes under stringent conditions to a polynucleotide (base sequence) which encodes the amino acid sequence of RCE I, RCE II, RCE III, MCE I, MCE II, or PCE I, or any one of the CBD-deleted endoglucanases thereof. The term "polynucleotide" as used herein includes DNA and RNA, and DNA is preferable.

[0058] The term "stringent conditions" as used herein means conditions in which a probe comprising a base sequence which encodes a partial or full-length sequence of

[0059] (a) the amino acid sequence of RCE I, RCE II, RCE III, MCE I, MCE II, or PCE I;

[0060] (b) the amino acid sequence of any one of the CBD-deleted endoglucanases thereof; or

[0061] (c) any one of the modified proteins thereof hybridizes to a polynucleotide which encodes a homologous protein, and the probe does not hybridize to the endoglucanase NCE 4 gene (SEQ ID NO: 7) described in WO98/03640 and the endoglucanase SCE 3 gene (SEQ ID NO: 8) described in WO98/54322. In this connection, it should be noted that the amount of each gene or polynucleotide used herein is equivalent to the amount of each of the NCE 4 gene, the SCE 3 gene, and the polynucleotide encoding a homologous protein.

[0062] More particularly, it means conditions in which, for example, using as a probe a labeled full-length DNA sequence encoding the amino acid sequence of RCE I, pre-hybridization is carried out at 42° C. for 1 hour according to the protocol attached to the ECL direct DNA/RNA labeling and detection system (Amersham), then the above probe is added thereto followed by hybridization at 42° C. for 15 hours, and thereafter, the resultant product is washed twice with 0.5×SSC (1×SSC; 15 mmol/L trisodium citrate, 150 mmol/L sodium chloride) containing 0.4% SDS and 6 mol/L urea at 42° C. for 20 minutes, and finally followed by washing the product twice with 5×SSC at room temperature for 10 minutes.

[0063] The above-mentioned "polynucleotide (base sequence) which encodes the amino acid sequence of RCE I, RCE II, RCE III, MCE I, MCE II, or PCE I" includes a polynucleotide in which codon usage and/or an intron recognition sequence are optimized in accordance with the type

of a host cell used for transformation, such as the codon-optimized endoglucanase RCE I gene (SEQ ID NO: 9) described in WO00/24879.

[0064] As the modified or homologous protein, there may be mentioned, for example, a protein having an amino acid sequence having preferably an 80% or more homology, more preferably a 90% or more homology, still further preferably a 95% or more homology, most preferably a 98% or more homology, with that of RCE I, RCE II, RCE III, MCE I, MCE II, or PCE I, or any one of the CBD-deleted endoglucanases thereof. In this connection, the above values of homology may be values calculated using a known program for homology search, preferably values calculated using FASTA3 [Science, 227, 1435-1441 (1985); Proc. Natl. Acad. Sci. USA, 85, 2444-2448 (1988); <http://www.ddbj.nig.ac.jp/E-mail/homology-j.html>] in accordance with default parameters.

[0065] The "reducing agent", which is contained in the cellulase preparation of the present invention, means a substance having an activity of reducing a molecule by accepting electrons from the molecule, thereby being itself oxidized. It is known that such reducing agents exhibit an activity of reducing and removing the remaining chlorine or the like in tap water. As the reducing agent used in the present invention, an inorganic reducing agent is preferable, and a substance which inhibits the enzyme activity cannot be used. Examples thereof include sulfurous acid, disulfurous acid, and thiosulfuric acid, and salts thereof, and thiourea. The reducing agents can be used alone or in a combination thereof.

[0066] The cellulose preparation of the present invention may comprise components which are conventionally contained in cellulase preparations such as excipients and/or preservatives. The form of the cellulose preparation may be solid or liquid. Examples of the form include powder, particulate, granule, non-dusting granule and liquid formulation.

[0067] The non-dusting granule (preferably a granule not having a dustability) that is one form of cellulase preparation can be produced according to the common dry granulation method. That is to say, powder cellulase enzyme is mixed with one or plural substances selected from the group comprising inorganic salts such as sodium sulfate or sodium chloride which are neutral and do not have an effect on the endoglucanase activity; minerals such as bentonite or montmorillonite which do not have an effect on the endoglucanase activity; neutral organic substances such as starch or powder cellulose; and surfactants. Thereafter, the powders or the finely suspended suspension of one or plural reducing agents which improve the effects of endoglucanase are added to the mixture, and then the obtained product is fully mixed or kneaded.

[0068] Depending on the situation, a synthetic polymer such as polyethylene glycol or a natural polymer such as starch, which binds solids, is optionally added to the mixture and further kneaded. Thereafter, granulation is carried out by extrusion molding, using, for example, a disk pelleter, and the obtained molded material is then converted into a spherical form using a marumerizer followed by drying, so that non-dusting granules can be produced. Naturally, it is also possible to coat the surface of granules with a polymer or the like to control the permeation of oxygen or water. In

this case, one or plural reducing agents which improve the effect of endoglucanase can be added to the cellulase preparation at a ratio of 0.01 to 50% by weight, preferably 0.1 to 20% by weight, more preferably 0.1 to 10% by weight.

[0069] Further, the liquid preparation (preferably stabilized liquid) can be prepared by blending an endoglucanase stabilizer such as a synthetic or natural polymer with a cellulase enzyme solution and, if necessary, adding inorganic salts and/or a synthetic preservative. In this case, one or plural reducing agents which improve the effect of endoglucanase can be added. Similar to the case of the non-dusting granule, one or plural reducing agents which improve the effect of endoglucanase can be added to the cellulase preparation at a ratio of 0.01 to 50% by weight, preferably 0.1 to 20% by weight, more preferably 0.1 to 10% by weight.

[0070] [2] Method of Treating Fabric

[0071] The method of treating fabric according to the present invention comprising the step of: treating cellulose-containing fabric with the above-mentioned cellulase preparation.

[0072] The following properties of cellulose-containing fabric can be improved by the present treatment method:

[0073] (1) Color clarification of colored cellulose-containing fabric;

[0074] (2) Removal of fuzz (reduction of the rate of the formation of fuzz, and reduction of fuzz);

[0075] (3) Providing of localized color variation to colored cellulose-containing fabric, that is, providing a stonewash-like appearance and texture to colored cellulose-containing fabric, typically jeans;

[0076] (4) Improvement of the touch and appearance of fabric by reducing weight; and

[0077] (5) Softening of fabric (reduction of stiffness).

[0078] More particularly, the method of treating fabric according to the present invention can be carried out by adding the cellulase preparation of the present invention into water in which fabric is or will be soaked, for example, during a soaking or rinsing of fabric.

[0079] Conditions such as contact temperature or the amount of endoglucanase may be appropriately determined in accordance with various other conditions. For example, when reducing the rate of the formation of fuzz or reducing fuzz of the cellulose-containing fabric, the fabric can be treated at a temperature of approximately 30 to 60° C., using 0.2 $\mu\text{g/mL}$ or more of reducing agents and endoglucanases in a protein concentration of 0.001 to 20 mg/L. One or more reducing agents may be added, taking into consideration economical effects, so long as an amount thereof is 0.2 $\mu\text{g/mL}$ or more and the reducing agent does not inhibit the enzyme activity. Preferably 0.2 to 500 $\mu\text{g/mL}$, more preferably 0.3 to 150 $\mu\text{g/mL}$ thereof may be used.

[0080] When providing a localized color variation to colored cellulose-containing fabric, the fabric can be treated at a temperature of approximately 40 to 60° C., using 0.2 $\mu\text{g/mL}$ or more of reducing agents and endoglucanases in a protein concentration of 0.01 to 100 mg/L. One or more reducing agents may be added, taking into consideration

economical effects, so long as an amount thereof is 0.2 $\mu\text{g}/\text{mL}$ or more and the reducing agent does not inhibit the enzyme activity. Preferably 0.2 to 500 $\mu\text{g}/\text{mL}$, more preferably 0.3 to 150 $\mu\text{g}/\text{mL}$ thereof may be used.

[0081] In a processing of reducing weight to improve the touch and appearance of the cellulose-containing fabric, the fabric can be treated at a temperature of approximately 30 to 60° C., using 0.2 $\mu\text{g}/\text{mL}$ or more of reducing agents and endoglucanases in a protein concentration of 0.001 to 100 mg/L. One or more reducing agents may be added, taking into consideration economical effects, so long as an amount thereof is 0.2 $\mu\text{g}/\text{mL}$ or more and the reducing agent does not inhibit the enzyme activity. Preferably 0.2 to 500 $\mu\text{g}/\text{mL}$, more preferably 0.3 to 150 $\mu\text{g}/\text{mL}$ thereof may be used.

[0082] The protein concentration of each type of endoglucanase can be calculated, for example, by HPLC analysis using TSKgel TMS-250 column (4.6 mm I.D. \times 7.5 cm) (TOSOH Corporation). The HPLC analysis involves loading acetonitrile in 0.05% TFA (trifluoroacetic acid) with a linear concentration gradient of 0% to 80% at a flow rate of 1.0 mL/min so as to elute each type of endoglucanase, and calculating the protein concentration from the peak area at UV 280 nm. For example, a purified NCE4, the protein concentration of which is previously determined by a Protein Assay Kit (BioRad Laboratories), is subjected to the HPLC analysis in the same manner as above, so that it can be used as a standard. The purified NCE4 can be obtained, for example, by cultivating *Humicola insolens* MN200-1 and purifying it from the culture, in accordance with the method described in WO98/03640. As a standard for the determination of a protein concentration in the Protein Assay Kit, for example, Albumin Standard (Bovine serum albumin, fraction V; PIERCE) can be used.

[0083] [3] Method of Deinking Waste Paper

[0084] The method of deinking waste paper according to the present invention comprises the step of:

[0085] treating the waste paper with the above-mentioned cellulase preparation together with a deinking agent.

[0086] More particularly, the present method can be carried out by treating waste paper with the cellulase preparation of the present invention together with a deinking agent, in a deinking step in a process of producing recycled paper from waste paper. The present method enables the deinking of waste paper, and thus the whiteness of waste paper can be improved. Waste paper which can be treated by the present method includes all types of common waste paper, for example, used newspaper, used magazine paper, and low to middle grade printed used paper which comprise mechanical pulp and chemical pulp; used wood-free paper comprising chemical pulp; and printed waste paper thereof such as coating paper. The deinking agent means an agent commonly used in the deinking of waste paper. Examples of the deinking agent include sodium chloride, alkalis such as sodium carbonate, sodium silicate, hydrogen peroxide, phosphates, anionic or nonionic surfactants, scavengers such as oleic acid, and assistant agents such as a pH stabilizer, a chelating agent, or a dispersing agent.

[0087] [4] Method of Improving Freeness of Paper Pulp

[0088] The method of improving the freeness of paper pulp according to the present invention comprises the step of:

[0089] treating the paper pulp with the above-mentioned cellulase preparation.

[0090] More particularly, the present method can be carried out by treating paper pulp with the cellulase preparation of the present invention. Examples of paper pulp which can be treated by the present method include waste paper pulp, recycled paperboard pulp, kraft pulp, sulfite pulp, thermo-mechanical treatment pulp, and other high-yield pulp.

EXAMPLES

[0091] The present invention now will be further illustrated by, but is by no means limited to, the following Examples.

[0092] All publications and patent applications mentioned in the present specification are herein incorporated by reference.

Example 1

Comparison Among Improvement Ratios of Fuzz-Removing Activities of Various Types of Cellulases by Addition of Reducing Agent

[0093] The cultivation of *Rhizopus oryzae*, *Mucor circinelloides*, and *Phycomyces nitens*, and the purification of RCE I, MCE I, and PCE I endoglucanases from the cultures were carried out by the method described in WO00/24879.

[0094] The cultivation of *Humicola insolens* MN200-1 and the purification of NCE4 endoglucanase from the culture were carried out by the method described in WO98/03640. The strain was domestically deposited in the International Patent Organism Depository National Institute of Advanced Industrial Science and Technology (Address: AIST Tsukuba Central 6, 1-1, Higashi 1-chome Tsukuba-shi, Ibaraki-ken 305-8566 Japan) on Jul. 15, 1996, and was transferred to an international deposit on Jun. 13, 1997. The international deposit number (a number in parenthesis [] following the international deposit number is a domestic deposit number) is FERM BP-5977 [FERM P-15736].

[0095] The cultivation of *Trichoderma viride* MC300-1 was carried out by the method described in WO98/54332. The strain was domestically deposited in the International Patent Organism Depository National Institute of Advanced Industrial Science and Technology (Address: AIST Tsukuba Central 6, 1-1, Higashi 1-chome Tsukuba-shi, Ibaraki-ken 305-8566 Japan) on Sep. 9, 1996, and was transferred to an international deposit on Aug. 11, 1997. The international deposit number (a number in parenthesis [] following the international deposit number is a domestic deposit number) is FERM BP-6047 [FERM P-15842].

[0096] Fuzz-removing treatment of a cotton knit fabric with fuzz formed in a large washer (a fabric of 6 cm \times 8 cm from Cotton Smooth Knit No. 3900, Nitto Boseki Co., Ltd. dyed brown by reactive dyeing in Tsuyatomo-Senko) was carried out using each of the obtained culture supernatants under the following conditions:

[0097] (Test Conditions)

[0098] Testing machine: Launder Meter L-20 (Daiei Kagaku Seiki MFG., Japan)

[0099] Temperature: 58° C. (only the *Trichoderma viride* culture supernatant); 40° C. (all other enzyme solutions)

[0100] Time: 120 minutes

[0101] Amount of reaction solution: 100 mL

[0102] Reaction pH: pH 4.5 (5 mmol/L acetate buffer) (only the *Trichoderma viride* culture supernatant pH 7.0 (5 mmol/L phosphate buffer) (all other enzyme solutions). All the buffers were prepared using tap water.

[0103] Type and amount of reducing agent: 1.2 μ g/mL sodium thiosulfate pentahydrate (Wako Pure Chemical Industries, Co., Ltd.)

[0104] To each of the treating solutions, four of about 16 g rubber balls were added together with each enzyme solution.

[0105] The amount of the enzyme solution required to remove approximately 50% of the formed fuzz on the basis of visual evaluation was determined in each of both cases of adding and not adding the reducing agent. Thereafter, a value was obtained by dividing the amount of the enzyme solution required to remove approximately 50% of the fuzz when not adding the reducing agent by the amount when adding the reducing agent, and the obtained value was defined as an improvement ratio of the fuzz-removing activity by the addition of the reducing agent. The results are shown in Table 1.

TABLE 1

Enzyme solutions	Improvement ratio of fuzz-removing activity by addition of reducing agent (fold)
<i>Humicola insolens</i> culture supernatant	1.2
<i>Trichoderma viride</i> culture supernatant	1.1
Purified NCE4	1.2
Purified RCEI	5.0
Purified MCEI	3.5
Purified PCEI	2.5

[0106] From the results of Table 1, it is found that the fuzz-removing activity of RCE I, MCE I, and PCE I, which are endoglucanases derived from Zygomycetes, is improved by the addition of the reducing agent at a level far higher than culture supernatants (i.e., cellulase) derived from *Humicola insolens* and *Trichoderma viride*.

Example 2

Improvement Effect of Fuzz-Removing Activity of RCE I Expressed in *Humicola* by Addition of Various Reducing Agents

[0107] RCE I endoglucanase was expressed in *Humicola insolens* in accordance with the method described in Examples D3 and D4 of WO00/24879. Fuzz-removing treatment of a cotton knit fabric with fuzz formed in a large washer (a fabric of 6 cm 8 cm from Cotton Smooth Knit No. 3900, Nitto Boseki Co., Ltd. dyed brown by reactive dyeing in Tsuyatomo-Senko) was carried out using the obtained culture supernatant under the following conditions:

[0108] (Test Conditions)

Testing machine:	Laundry Meter L-20 (Daiei Kagaku Seiki MFG., Japan)
Temperature:	40° C.
Time:	120 minutes
Amount of reaction solution:	100 mL
Reaction pH:	pH 7.0 (5 mmol/L phosphate buffer; prepared using tap water)
Amount of reducing agent:	1.2 μ g/mL
Type of reducing agent:	sodium thiosulfate pentahydrate (Wako Pure Chemical Industries, Co., Ltd.); sodium sulfite (anhydride) (Wako Pure Chemical Industries, Co., Ltd.); and thiourea (Kanto Kagaku, Co., Ltd.)

[0109] To each of the treating solutions, four of about 16 g rubber balls were added together with the enzyme solution.

[0110] The amount of the enzyme solution required to remove approximately 50% of the formed fuzz on the basis of visual evaluation was determined in each of both cases of adding and not adding the various reducing agents. Thereafter, a value was obtained by dividing the amount of the enzyme solution required to remove approximately 50% of the fuzz when not adding the reducing agent by the amount when adding each of the various reducing agents, and the obtained value was defined as an improvement ratio of the fuzz-removing activity by the addition of each of the various reducing agents. The results are shown in Table 2.

TABLE 2

Reducing agents	Improvement ratio of fuzz-removing activity by addition of reducing agent (fold)
Sodium thiosulfate pentahydrate	5.0
Sodium sulfite (anhydride)	5.0
Thiourea	5.0

[0111] From the results of Table 2, it is found that the fuzz-removing activity of the culture supernatant obtained by expressing and secreting RCE I in *Humicola insolens* was improved by any of the above reducing agents.

Example 3

Improvement Effect of Fuzz-Removing Activity of RCE I Expressed in *Humicola* by Addition of Reducing Agent with Various Concentrations

[0112] RCE I endoglucanase was expressed in *Humicola insolens* in accordance with the method described in Examples D3 and D4 of WO00/24879. Fuzz-removing treatment of a cotton knit fabric with fuzz formed in a large washer (a fabric of 6 cm×8 cm from Cotton Smooth Knit No. 3900, Nitto Boseki Co., Ltd. dyed brown by reactive dyeing in Tsuyatomo-Senko) was carried out using the obtained culture supernatant under the following conditions:

[0113] (Test Conditions)

Testing machine:	Launder Meter L-20 (Daiei Kagaku Seiki MFG., Japan)
Temperature:	40° C.
Time:	120 minutes
Amount of reaction solution:	100 mL
Reaction pH:	pH 7.0 (5 mmol/L phosphate buffer; prepared using tap water)
Type of reducing agent:	sodium thiosulfate pentahydrate (Wako Pure Chemical Industries, Co., Ltd.) Amount of reducing agent: 0.15 to 150 $\mu\text{g/mL}$

[0114] To each of the treating solutions, four of about 16 g rubber balls were added together with the enzyme solution.

[0115] The amount of the enzyme solution required to remove approximately 50% of the formed fuzz on the basis of visual evaluation was determined in each of cases of adding various concentrations of the reducing agent. Thereafter, a value was obtained by dividing the amount of the enzyme solution required to remove approximately 50% of the fuzz when not adding the reducing agent by the amount when adding each of various concentrations of the reducing agent, and the obtained value was defined as an improvement ratio of the fuzz-removing activity by the addition of each of various concentrations of the reducing agent. The results are shown in Table 3.

TABLE 3

Amount of reducing agent ($\mu\text{g/ml}$)	Improvement ratio of fuzz-removing activity by addition of reducing agent (fold)
0.15	1.0
0.3	2.5
0.6	4.5
0.9	5.0
1.2	5.0
1.8	4.5
3.0	4.5
6.0	4.5
15.0	4.5
30.0	4.5
60.0	4.0
150.0	4.0

[0116] From the results of Table 3, it is found that the fuzz-removing activity of the culture supernatant obtained (21) by expressing and secreting RCE I in *Humicola insolens* was improved by the addition of the reducing agent having a wide range of concentration from 0.3 $\mu\text{g/mL}$ to 150 $\mu\text{g/mL}$ or more.

Example 4: Production of RCE I Cellulase Preparation Comprising Reducing Agent

[0117] After mixing the following raw materials by the mixing ratios as described in Table 4, an appropriate amount of water was added thereto, and the mixture was kneaded. The obtained product was subjected to a disk pelletter for

molding, and the product obtained by injection molding was converted in a particle form using a marumerizer (Fuji Paudal Co., Ltd.) followed by drying and sieving the product so as to obtain a granulated product.

TABLE 4

Raw materials	Mixing ratio (%)
Sodium thiosulfate	1
S-220 (nonionic surfactant manufactured by NOF Corporation)	10
RCE I cellulase powder product	5
Magnesium chloride	0.5
Monopotassium phosphate (Wako Pure Chemical Industries, Co., Ltd.)	2
Dipotassium phosphate (Wako Pure Chemical Industries, Co., Ltd.)	1
Corn starch (Shikishima Starch Co.)	80.5

[0118] The RCE I cellulase powder product was prepared by concentrating the culture supernatant of RCE I expressed in *Humicola insolens* using ultrafiltration, according to the method described in Examples D3 and D4 of WO00/24879, followed by spray drying.

INDUSTRIAL APPLICABILITY

[0119] The present invention provides a cellulase preparation having a dramatically improved *Zygomycetes*-derived endoglucanase activity by adding a reducing agent into the preparation. When the cellulase preparation is used in the treatment of fabric (such as the reduction of fuzz of cellulose-containing fabric, the improvement of touch and appearance, the color clarification, localized color variation, or softening), the deinking of waste paper, or the processing of improving the freeness of paper pulp, each of the above treatments can be carried out with a less amount of enzyme, thereby significantly reducing cost.

[0120] Free Text in Sequence Listing

[0121] Features of "Artificial Sequence" are described in the numeric identifier <223> in the Sequence Listing. More particularly, the base sequence of SEQ ID NO: 9 is a codon-optimized sequence corresponding to RCE I protein (SEQ ID NO: 1).

[0122] Although the present invention has been described with reference to specific embodiments, various changes and modifications obvious to those skilled in the art are possible without departing from the scope of the appended claims.

SEQUENCE LISTING

<160> NUMBER OF SEQ ID NOS: 9

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 <222> LOCATION: (-23)..(-1)
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Thr Cys Lys Val Ser Asn Asp Tyr Tyr Ser Gln Cys Leu Pro Ser Gly
      30                35                40

Ser Ser Gly Asn Lys Ser Ser Glu Ser Ala His Lys Lys Thr Thr Thr
      45                50                55

Ala Ala His Lys Lys Thr Thr Thr Ala Ala His Lys Lys Thr Thr Thr
      60                65                70

Ala Pro Ala Lys Lys Thr Thr Thr Val Ala Lys Ala Ser Thr Pro Ser
      75                80                85

Asn Ser Ser Ser Ser Ser Ser Gly Lys Tyr Ser Ala Val Ser Gly Gly
  90                95                100                105

Ala Ser Gly Asn Gly Val Thr Thr Arg Tyr Trp Asp Cys Cys Lys Ala
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Ser Cys Ser Trp Pro Gly Lys Ala Asn Val Ser Ser Pro Val Lys Ser
      125                130                135

Cys Asn Lys Asp Gly Val Thr Ala Leu Ser Asp Ser Asn Ala Gln Ser
      140                145                150

Gly Cys Asn Gly Gly Asn Ser Tyr Met Cys Asn Asp Asn Gln Pro Trp
      155                160                165

Ala Val Asn Asp Asn Leu Ala Tyr Gly Phe Ala Ala Ala Ala Ile Ser
      170                175                180                185

Gly Gly Gly Glu Ser Arg Trp Cys Cys Ser Cys Phe Glu Leu Thr Phe
      190                195                200

Thr Ser Thr Ser Val Ala Gly Lys Lys Met Val Val Gln Val Thr Asn
      205                210                215

Thr Gly Gly Asp Leu Gly Ser Ser Thr Gly Ala His Phe Asp Leu Gln
      220                225                230

Met Pro Gly Gly Gly Val Gly Ile Phe Asn Gly Cys Ser Ser Gln Trp
      235                240                245

Gly Ala Pro Asn Asp Gly Trp Gly Ser Arg Tyr Gly Gly Ile Ser Ser
      250                255                260                265

Ala Ser Asp Cys Ser Ser Leu Pro Ser Ala Leu Gln Ala Gly Cys Lys
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Trp Arg Phe Asn Trp Phe Lys Asn Ala Asp Asn Pro Ser Met Thr Tyr

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Thr Cys Lys Val Ser Asn Asp Tyr Tyr Ser Gln Cys Leu Ala Pro Glu	30	35	40		
Ser Asn Gly Asn Lys Ser Ser Glu Cys Ser Lys Leu Tyr Gly Gln Cys	45	50	55		
Gly Gly Lys Asp Trp Asn Gly Pro Thr Cys Cys Glu Ser Gly Ser Thr	60	65	70		
Cys Lys Val Ser Asn Asp Tyr Tyr Ser Gln Cys Leu Ala Pro Glu Ser	75	80	85		
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Ala Pro Ala Lys Glu Ile Thr Thr Thr Ala Lys Ala Ser Asn Ser Ser	110	115	120		
Asn Ser Ser Gly Lys Tyr Ser Ile Val Ser Gly Gly Ala Ser Gly Asn	125	130	135		
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Pro Gly Lys Ala Asn Val Ser Ser Pro Val Lys Ser Cys Asn Lys Asp	155	160	165		
Gly Val Thr Ala Leu Ser Asp Ser Asn Val Gln Ser Gly Cys Asn Gly	170	175	180	185	
Gly Asn Ser Tyr Met Cys Asn Asp Asn Gln Pro Trp Ala Val Asn Asp	190	195	200		
Asn Leu Ala Tyr Gly Phe Ala Ala Ala Ala Ile Ser Gly Gly Gly Glu	205	210	215		
Ser Arg Trp Cys Cys Ser Cys Phe Glu Leu Thr Phe Thr Ser Thr Ser	220	225	230		
Val Ala Gly Lys Lys Met Val Ile Gln Val Thr Asn Thr Gly Gly Asp	235	240	245		
Leu Gly Ser Ser Thr Gly Ala His Phe Asp Leu Gln Met Pro Gly Gly	250	255	260	265	
Gly Val Gly Ile Phe Asn Gly Cys Ser Lys Gln Trp Gly Ala Pro Asn					

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  140                145                150
Val Thr Ser Pro Val Gly Ser Cys Asn Lys Asp Gly Lys Thr Leu Ala
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Asp Asn Asn Thr Gln Asn Gly Cys Val Gly Gly Ser Ser Tyr Thr Cys
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Cys Phe Glu Leu Thr Phe Thr Ser Thr Ala Val Lys Gly Lys Lys Met
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 270 275 280

Tyr Gly Gly Val Ser Ser Ala Ser Asp Cys Ser Asn Leu Pro Ser Ala
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Leu Gln Ala Gly Cys Lys Trp Arg Phe Gly Trp Phe Lys Asn Ala Asp
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Ser His Ser Asn Asn Ala Gly Asn Ala Ser Ser Thr Lys Lys Thr Ser
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Thr Lys Thr Ser Thr Thr Thr Ala Lys Ala Thr Ala Thr Val Thr Thr
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Lys Thr Val Thr Lys Thr Thr Thr Lys Thr Thr Thr Lys Thr Ser Thr
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Ile Ser Gly Gly Lys Ser Gly Ser Gly Ser Thr Thr Arg Tyr Trp Asp
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Cys Cys Lys Ala Ser Cys Ser Trp Pro Gly Lys Ala Ser Val Thr Gly
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 Val Ser Asp Cys Ala Ser Leu Pro Ser Ala Leu Gln Ala Gly Cys Lys
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 225 230 235
 Ser Thr Asn His Phe Asp Leu Gln Met Pro Gly Gly Gly Val Gly Tyr
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<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
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<222> LOCATION: (16)..(1032)
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Ala Leu Ala Leu Gly Thr Glu Met Ala Ser Ala Ala Glu Cys Ser Lys
-10 -5 1 5
ctc tac gga cag tgc ggc gga aag aac tgg aac ggc ccc acc tgc tgc 147
Leu Tyr Gly Gln Cys Gly Gly Lys Asn Trp Asn Gly Pro Thr Cys Cys
10 15 20
gag agc ggc tcg acc tgc aag gtc tcg aat gac tac tac agc cag tgc 195
Glu Ser Gly Ser Thr Cys Lys Val Ser Asn Asp Tyr Tyr Ser Gln Cys
25 30 35
ctg ccg agc ggc tcc tcg gga aac aag tcg agc gag tcg gcc cac aag 243
Leu Pro Ser Gly Ser Ser Gly Asn Lys Ser Ser Glu Ser Ala His Lys
40 45 50
aag acc acg acc gct gcc cac aag aag acc acg acc gcc gct cac aag 291
Lys Thr Thr Thr Ala Ala His Lys Lys Thr Thr Thr Ala Ala His Lys
55 60 65
aag act acg acc gct ccc gcc aag aag acc acg acc gtc gcc aag gct 339
Lys Thr Thr Thr Ala Pro Ala Lys Lys Thr Thr Thr Val Ala Lys Ala
70 75 80 85
tcg act ccg tcc aac tcg agc agc tcg tct tcg gga aag tac agc gct 387
Ser Thr Pro Ser Asn Ser Ser Ser Ser Ser Ser Gly Lys Tyr Ser Ala
90 95 100
gtc agc ggt ggc gct agc ggc aac ggc gtc act acc cgc tac tgg gac 435
Val Ser Gly Gly Ala Ser Gly Asn Gly Val Thr Thr Arg Tyr Trp Asp
105 110 115
tgc tgc aag gct tcg tgc tcg tgg ccc gcc aag gct aac gtc agc tcg 483
Cys Cys Lys Ala Ser Cys Ser Trp Pro Gly Lys Ala Asn Val Ser Ser
120 125 130
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Pro Val Lys Ser Cys Asn Lys Asp Gly Val Thr Ala Leu Ser Asp Ser
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Asn Gln Pro Trp Ala Val Asn Asp Asn Leu Ala Tyr Gly Phe Ala Ala	
170	175 180
gct gcc att agc ggc ggt ggc gag agc cgc tgg tgc tgc tcc tgc ttc	675
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Gln Val Thr Asn Thr Gly Gly Asp Leu Gly Ser Ser Thr Gly Ala His	
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Gly Ile Ser Ser Ala Ser Asp Cys Ser Ser Leu Pro Ser Ala Leu Gln	
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Ala Gly Cys Lys Trp Arg Phe Asn Trp Phe Lys Asn Ala Asp Asn Pro	
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295	300 305
acc gga tgc tcg cgc aag taa acgcaggatc c	1043
Thr Gly Cys Ser Arg Lys	
310	315

1. A cellulase preparation comprising an endoglucanase derived from Zygomycetes, a protein in which a cellulose binding domain is deleted in the endoglucanase, or a modified or homologous protein thereof; and a reducing agent.

2. The cellulase preparation according to claim 1, wherein the Zygomycetes is a microorganism belonging to genus *Rhizopus*, *Mucor*, or *Phycomyces*.

3. A cellulase preparation comprising at least one of:

- (a) a protein consisting of any one of the amino acid sequences of SEQ ID NOS: 1 to 6;
- (b) a protein consisting of an amino acid sequence in which a cellulose binding domain is deleted in any one of the amino acid sequences of SEQ ID NOS: 1 to 6; or
- (c) a protein consisting of an amino acid sequence in which one or plural amino acids are deleted, substituted, inserted, or added, in any one of the amino acid sequences of SEQ ID NOS: 1 to 6 or in an amino acid sequence in which a cellulose binding domain is deleted therein, and exhibiting an endoglucanase activity; and

a reducing agent.

4. A cellulase preparation comprising a protein encoded by at least one of:

- (a) a polynucleotide which encodes any one of the amino acid sequences of SEQ ID NOS: 1 to 6;
- (b) a polynucleotide which encodes a protein consisting of an amino acid sequence in which a cellulose binding domain is deleted in any one of the amino acid sequences of SEQ ID NOS: 1 to 6; or
- (c) a polynucleotide which encodes a protein exhibiting an endoglucanase activity and is complementary to a polynucleotide which hybridizes under stringent conditions to a polynucleotide which encodes a protein consisting of any one of the amino acid sequences of SEQ ID NOS: 1 to 6 or a protein consisting of an amino acid sequence in which a cellulose binding domain is deleted therein; and

a reducing agent.

5. The cellulase preparation according to claim 1, containing 0.01 to 50% by weight of the reducing agent in the cellulase preparation.

6. The cellulase preparation according to claim 1, in which the reducing agent is sodium thiosulfate, sodium sulfite, or thiourea.

7. The cellulase preparation according to claim 1, which is a granule not having a dustability or stabilized liquid.

8. A method of treating cellulose-containing fabric, comprising the step of:

treating the fabric with the cellulase preparation according to claim 1 to improve a property of the fabric.

9. The method according to claim 8, wherein the improvement of the property of the fabric is a color clarification.

10. The method according to claim 8, wherein the improvement of the property of the fabric is a removal of fuzz.

11. The method according to claim 8, wherein the improvement of the property of the fabric is an addition of a stonewash-like appearance and texture.

12. The method according to claim 8, wherein the improvement of the property of the fabric is an improvement of touch and appearance.

13. The method according to claim 8, wherein the improvement of the property of the fabric is a softening of the fabric.

14. The method according to claim 8, wherein the treatment of the fabric with the cellulase preparation is carried out by soaking or rinsing the fabric.

15. A method of deinking waste paper, comprising the step of: treating the waste paper with the cellulase preparation according to claim 1 together with a deinking agent.

16. A method of improving a freeness of paper pulp, comprising the step of:

treating the paper pulp with the cellulase preparation according to claim 1.

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