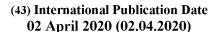
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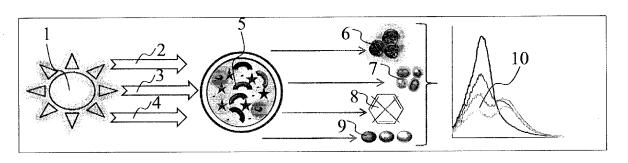
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(54) Title: A METHOD FOR IDENTIFYING THE PROPERTIES OF SUBSTANCES, IN PARTICULAR LIQUID SUBSTANCES

FIG. 1



(57) **Abstract:** The present invention relates to a method for identifying properties of substances, in particular liquid substances, wherein the method is based on subjecting a sample (5) of a substance containing on the one hand metal ions and on the other hand a thiol group or disulfide bonds first to the analysis in the original or untreated state, i.e. in the state not irradiated by UV radiation (2, 3, 4), and subsequently subjecting the sample to UV radiation (2, 3, 4), thereby causing spectral changes (10) which are characteristic for the given substance, and investigating the optical properties and possible differences between the original substance and the examined sample of substances based on the change in the spectral characteristics of the sample prior to its irradiation or information stored in the database about the non-radiated sample, and after its irradiation with UV radiation (2, 3, 4). Preferably, the UV radiation (2, 3, 4) in the sample (5) generates quantum nanocrystals (6), free radicals (7), degrades chemical bonds (8) or photo-bleaches (9).





A Method for Identifying the Properties of Substances, in Particular Liquid Substances

Technical Field

The invention relates to a method for identifying the properties of substances, in particular liquid substances, including liquefied substances, including biological samples, thus being included in the field of analytical chemistry. For identifying, the method uses natural spectral properties of substances, i.e. the absorption and emission of radiation, which varies depending on UV irradiation. Spectral analysis of these changes and chemical reactions yields valuable information on the physical, chemical or biological properties of the sample in a short time, without the need for any previously complicated preparation of the sample under investigation.

Background Art

The diverse composition of pharmacological products, especially drugs, drug mixtures, i.e. addictive and psychotropic substances, foods, beverages, and biological or clinical specimens, such as blood plasma or urine, is a fundamental problem that makes it difficult to analyze them within tens of minutes.

In most cases, an extraction, isolation and/or separation process must be performed prior to an analysis. The sample thus treated is subjected to an instrumental analysis performed by a highly qualified person using complicated and costly instrumentation. The whole process, though precise, is time-consuming and costly.

A rapid drug determination may be crucial in rescuing a patient overdosed with an unknown drug the transportation to a hospital. It is particularly important to detect a faulty or counterfeit drug batch before it is distributed. The ability to recognize important properties of the test substance within minutes appears to be crucial for the industry or food industry. In particular, this may involve checking the origin of cask wines or the quality control of the technological processing of fruit juices and vegetable extracts or beverages containing these juices and extracts.

In recent years, efforts have been exerted, particularly in the medical field, to develop methods which employ typical patterns of spectra, sometimes shapes or referred as so-called "fingerprints". These methods are based spectral analysis, in particular on for example Raman spectroscopy, Fourier transform infrared spectroscopy and clinical specimen mass spectrometry, wherein the shapes of the recorded spectra are evaluated. Although these methods not allow precise determination of the quantitative and qualitative representation individual of analytes in the sample, but these spectra can be assigned to a number of physiological or pathological conditions of the organism. In this way it is possible to create databases of typical spectra and compare them mutually. The proposed methods for identifying the properties of substances utilize and evaluate typical spectra shapes on the basis of which the sample can be accurately identified or studied.

Basic physicochemical methods used in the analysis of addictive drugs or food include chromatographic methods, i.e. a liquid or gas chromatography, electrophoresis or spectral methods, as a spectrophotometry, infrared or Raman spectroscopy. The extraction of analytes is one of the critical steps in the analytical determination substance properties. As extraction agents, both and non-polar solvents are used. The polar solvents include most often acetonitrile, ethanol and methanol. One of the most important conditions for achieving high extraction efficiency is the choice of a suitable pH value. The pH value of respective extracting agent affects the solubility and/or

ionization of analytes. The next step in the analysis may be to derivatize the sample. Derivatization reactions lead to the formation of detectable derivatives, to an increase in sensitivity and selectivity, an increase in the resolution, itself, respectively. allowing the separation process Generally, spectroscopic methods have less demands the sample preparation compared to chromatographic methods. For example, Raman spectroscopy is one of the methods of vibration molecular spectroscopy that is used to chemically identify analytes contained in a sample. The method uses the so-called Raman effect or Raman scattering, which occurs when the laser beam interacts with the electrons of the sample investigation. Since each substance a characteristic wavelength shift, this phenomenon can be used to non-destructively identify the chemical composition of a sample by creating a set of spectra typical for given analytes. The disadvantage of Raman scattering its relatively low intensity, especially when compared absorption or emission of radiation. Considerable purity of the analyzed sample is therefore necessary. Any fluorescence impurity or a possible photochemical reaction makes it impossible to carry out the analysis.

Another important method using a set of typical spectra represents also a mass spectrometry. This method can be used either alone or in combination with a separation technique.

Separate use of this method to determine the properties of substances without prior separation of samples can be problematic since the components of the laser ionized mixture and the degree of ionization are related to a number of factors. One of them is for example a type of matrix which is added to the sample as a part of the analysis. It absorbs most of the energy of the ionizing laser and transmits it to

the sample. Because the operator does not know the composition of the sample, an incorrectly selected matrix may result in poor or partial ionization of the sample, resulting in distorting or disabling the analysis.

Summary of the Invention

The subject matter of the present invention relates to a method for identifying the properties of substances, particular liquid substances, including liquefied substances, wherein the method is based on subjecting a sample of a substance containing on the one hand metal ions and on the other hand a thiol group or disulfide bonds first to the analysis in the original or untreated state, i.e. in the state not irradiated by UV radiation, and subsequently on subjecting the sample to UV radiation, thereby causing spectral changes are characteristic for the given substance, and evaluating the optical properties and possible differences between the original substance and the examined sample of substances based on the change in the spectral characteristics of the sample prior to its irradiation or information stored in the database about the non-radiated sample, and after its irradiation with UV radiation.

The sample of the substance contains metal ions, in particular some of $\mathrm{Zn^{2+}}$, $\mathrm{Cu^{2+}}$, $\mathrm{Fe^{2+}}$, $\mathrm{Mg^{2+}}$, $\mathrm{Se^{2+}}$ or $\mathrm{Cd^{2+}}$ or disulfide bonds or a thiol group, especially some of cysteine, homocysteine, glutathione oxidized, glutathione reduced, S-Adenosyl-L-homocysteine, S-Adenosyl-methionine, coenzyme A, cystamine, acetylcysteine, gamma-L-Glutamyl-L-cysteine.

Preferably, the sample is subjected to UV radiation at a wavelength of 100 to 400 nm.

The spectral changes induced by UV irradiation are manifested by a change in the absorption or emission or excitation spectrum or as an increase or decrease in the absorption, emission or excitation signal. These changes are mainly related to the formation of quantum nanocrystals, or to the change in the formation of radicals, or to the degradation of multiple chemical bonds, or to the change in photo-bleaching, or a combination of these changes.

The present invention provides a new, exceptional and unique fluorescence spectroscopic method which, based on typical spectra, can identify or distinguish individual samples or their properties.

Very advantageous are low demands on the sample preparation. The sample should only be at least partially dissolved in distilled water or another solvent, for example dimethylsulfoxide, or mixed with a suitable buffer solution. The method has no special demands on the quality of the samples, thus it is universal and can be used in the industry, agriculture, food, pharmacy and research workplaces.

The speed of sample analysis represents an extraordinary advantage. The entire procedure, including the sample preparation, analysis and evaluation, can be performed within minutes. For example, the drug analysis does not exceed 10 min.

The method does not require any special treatment of analytical instruments and is compatible with instruments using ultraviolet and visible radiation for its operation.

The simplicity of analysis allows the use of case or hand-held analyzers, allowing analysis to be performed wherever it is needed.

Brief Description of the Drawings

The results of exemplary embodiments and the method for identifying the properties of substances according to the invention are shown in the accompanying drawings, in which:

FIG. 1 schematically illustrates the nature of the method according to the invention, wherein the reference numeral 1 represents the ultraviolet radiation source, which is defined by the duration of irradiation as labeled by the reference numeral 2, by the wavelength as labeled by the reference numeral 3 and its intensity acting on the studied liquid sample as labeled by the reference numeral 4, and wherein the reference numeral 5 represents the sample being investigated where spectral changes are induced which are caused by the formation of fluorescent nanocrystals according to the reference numeral 6, the formation of free radicals shown by the reference numeral 7, degradation of chemical bonds shown by the reference numeral 8, and photo-bleaching illustrated by the reference numeral 9, wherein the changes can be recorded

as absorption, fluorescence and excitation spectra, as shown by the reference numeral 10.

FIG. 2 shows excitation spectra in the range $\lambda_{ex}=230\text{-}470$ nm and $\lambda_{em}=500$ nm of selected pharmacological agents, in this case antihistamines, dissolved in distilled water (20 mg / mL): the sample A is Dithiaden - manufactured by Zentiva, the sample B is Zenaro - manufactured by Zentiva, the sample C is Aerius - manufactured by SP Labo N. The sample D is Dasselta - manufactured by Krka d.d,. The curve 1 represents the excitation spectrum of the sample after 6 minutes of irradiation in the UV transilluminator ($\lambda_{em}=254$ nm), the curve 2 represents the excitation spectrum of the sample in the native state, i.e. without UV irradiation. The data were acquired using the fluorescence analyzer Infinite M200 pro. The samples were irradiated and analyzed in 100 µL batches in a UV transparent 96 well plate.

FIG. 3 shows emission spectra ($\lambda_{\rm ex}$ = 250 nm and $\lambda_{\rm em}$ = 280 nm) of selected juices, where the sample labeled A represents orange juice from the company BASIC, denoted by the manufacturer as a 100% concentrate, the sample labeled B represents grapefruit juice from the same company and the sample labeled C represents apple juice, also from the same company. The curve 1 represents the result where 50 µL of the sample were mixed with 50 μL of 0.1 mol / liter phosphate buffer pH 7, the curve 2 represents the result where 50 µL of the sample diluted 1:1 with distilled water were mixed with 50 μL 0.1 mol / liter of phosphate buffer pH 7; the curve 3 represents the result where = 50 μ L of a sample diluted 1 : 2 with distilled water were mixed with 50 μL of 0.1 mol / liter phosphate buffer pH 7 and the curve 4 represents the result where 50 μL of a sample diluted 1 : 4 with distilled water were mixed with 50 μ L 0.1 mol / liter phosphate buffer pH 7.

The samples were irradiated with a UV transilluminator ($\lambda_{em}=254$ nm) in 100 μL batches in a UV transparent 96 well plate for 10 minutes. The data were acquired using the fluorescence analyzer Infinite M200 pro.

FIG. 4 shows the emission spectra ($\lambda_{\rm ex}$ = 250 nm and $\lambda_{\rm em}$ = 280 - 600 nm) of selected varietal wines, wherein the sample A represents Riesling, the sample B represents Riesling Vlašský, the sample C represents red Traminer wine and the sample D represents Sauvignon. The curve 1 shows the result where 50 µL of the sample were mixed with 50 μL of 0.1 mol / liter phosphate buffer pH 7, the curve 2 shows the result where 50 μL of the sample diluted 1: 1 with distilled water were mixed with 50 μ L 0.1 mol / liter phosphate buffer pH 7, the curve 3 shows the result where 50 μ L of the sample diluted 1 : 2 with distilled water were mixed with 50 μL 0.1 mol / liter phosphate buffer pH 7 and the curve 4 shows the result where 50 µL of the sample diluted 1:4 with distilled water were mixed with 50 μL 0.1 mol / liter phosphate buffer pH 7. The samples were irradiated with the UV transilluminator ($\lambda_{em} = 254$ nm) for 100 μL in the UV transparent 96-well plate for 15 minutes. The data were taken with the Infinite M200 pro fluorescence analyzer.

FIG 5 shows the excitation spectra in the range $\lambda_{\rm ex}$ 230 - 370 nm and $\lambda_{\rm em}$ = 400 nm of selected tomato variety extracts, where the sample A represents the variety Sassari, the sample B represents the variety Grammy and the sample C represents the variety Tirrenico. The curve 1 shows the excitation spectrum of the sample after 20 minutes of irradiation in the UV transilluinator ($\lambda_{\rm em}$ = 254 nm); the curve 2 shows the excitation spectrum of the sample in its native state, i.e. not irradiated with UV radiation. The data were acquired with the Infinite M200 pro fluorescence analyzer. Samples were

irradiated and analyzed in 100 μL batches in the UV transparent 96 well plate. The extracts, i.e. tomato pieces, were pressed and filtered through a 25 mm Syringe filter with a 0.2 μ m membrane.

FIG. 6 shows a comparison of a urine sample of a healthy human and a patient with a diagnosed prostate cancer. The urine sample was diluted each with 1:1, 1:2, 1:4 and 1:8 0.1 M phosphate buffer pH 7 having the concentration 0, mol / liter and pipetted into the 100 μL UV transparent 96 well plate. The fluorescence intensity (λ_{ex} - 400 nm and λ_{em} = 470 nm) of samples without UV irradiation and after 20 minutes irradiation with the UV transilluminator (λ_{em} = 254 nm) were then recorded. Graphs labeled A represent the fluorescence intensity of the urine control sample of the healthy human before irradiation (dotted bars) and after the UV exposure (cross-hatched bars). Graphs labeled В represent fluorescence intensity (diced bars) of the control sample after and before the UV exposure, showing a significantly increasing trend. Graphs labeled C represent the fluorescence intensity of the urine sample of a prostate cancer patient prior to the irradiation (dotted bars) and after the UV exposure (cross-hatched bars), and graphs labeled D represent the proportion of fluorescence intensity of the urine sample of the prostate cancer patient after and before the UV exposure, wherein a noticeable growing trend is apparent. The data were taken with the Infinite M200 pro fluorescence analyzer.

FIG. 7 shows a schematic performance of the intracellular analysis using a fluorescence correlation spectroscopy and a fluorescence microscope, wherein the reference numeral 11 represents a 10 μ L sample that is dropped onto a slide 12, the reference numeral 13 represents a sample that is overlaid by a

shielding sheath to remove the ambient illumination, the reference numeral 14 represents a fluorescent microscope objective focused on a confocal volume (typically 0.5 fL) into a selected portion of the sample under study 15, the reference numeral 16 represents an excitation laser ($\lambda_{em} = 385$ nm) which during the analysis induces fluorescent quantum dots in the sample, which are shown schematically under the reference numeral 17, and the reference numeral 18 represents a detector.

FIG. 8 under the reference numeral 19 shows a real photo of a 2% agarose gel which comprised zinc and cysteine recorded by the fluorescent microscope, wherein bleaching or loss of autofluorescence took place, as shown by the reference numeral 20, and the formation of fluorescent nanocrystals was occurred, as shown by the reference numeral 21. The reference numeral 22 shows a real photo of a single cell taken with a fluorescence microscope where fluorescent nanocrystals were formed, which are indicated by the reference numerals 23 and 24, whose spectral analysis provides information about a given location in the cell. Fluorescent nanocrystals labeled by the reference numerals 21, 23 and 24 were created during the analysis by means of the excitation laser having $\lambda = 385$ nm.

Detailed Description of the Invention

The invention will now be described in more detail by way of examples, which are, however, not intended to limit the scope of the claims. The method for identifying the properties of substances according to the invention is schematically shown in Figure 1.

In the first step, a liquid sample is analyzed in its native state, i.e., not irradiated by UV radiation. The sample is then exposed to UV light at a precisely defined intensity for a few seconds or minutes. The irradiation time depends on the UV radiation intensity, wavelength and sample volume. UV radiation causes chemical changes in the sample, which transform as spectral changes specific to the relevant composition. UV photons have a high energy and can therefore break down chemical bonds. For example, chlorine does not react with alkanes under normal conditions. After the UV irradiation it begins to react because UV rays split the chemical bond in the Cl_2 molecule, which breaks down into extremely reactive radicals, which then react even with inert alkanes. The presence of metal ions, typically Zn^{2+} , Cu^{2+} , Fe^{2+} , Mq^{2+} , Se^{2+} , Cd^{2+} , and compounds having the disulfide bond (-SS-) and/or thiole group (-SH), especially cysteine, homocysteine, glutathione, S-Adenosyl-L-homocysteine, S-Adenosyl-methionine, coenzyme A, cystamine, acetylcysteine, gamma-L-Glutamyl-Lcysteine in the sample after interaction with UV causes the formation of a number of nano and micro particles, complexes clusters, which significantly influence the spectral properties of the studied matrix. The concentration of these precursors, i.e. metal ions and -SH, -S-S- containing compounds, required for cluster formation, is in the range of 1 nM to 3 mM. The analysis of the spectral signals of such created clusters provides valuable information on physical, chemical or biological properties of the sample. Therefore, changes in absorption and fluorescence, i.e. the excitation or emission signals, their shape, waveform, intensity, the lifetime of fluorescence, the fluorescent anisotropy, photobleaching, fluorophore diffusion coefficients before and after UV irradiation are evaluated. The content of

thiol groups or disulfide substances to be investigated within the scope of the invention is not added to the sample. Each substance, with only a few exceptions, contains them in trace amounts which are absolutely sufficient for the conventional analytical methods of the invention to achieve technical effects. These are substances that are naturally contained in each sample.

If a sample is changed by adding another component, by substitution for another component, or changing the type and concentration of thiol compounds (i.e. -SH, -S-S-containing compounds) or metal ions, the described method captures this change. Similarly, the method captures natural differences in samples, such as plant varieties or disease-related clinical changes.

The intended methodological procedures can be implemented into a conventional desktop spectrometer or even in the form of a handheld device, allowing on-site analyzes to be performed.

As a source of UV radiation, a table transilluminator $(\lambda_{em} = 254 \text{ nm})$ or a UV laser beam $(\lambda_{em} = 385 \text{ nm})$ can typically be used. Other light sources of suitable wavelengths such as electroluminescent diodes, LEDs and lamps can also be used. induced by the The changes UV emitter typical are of the chemical composition of the sample and can be monitored by conventional spectroscopic instrumentations, i.e. by means spectrophotometry, fluorescence spectroscopy, of the correlation fluorescence spectroscopy and the like.

Example 1

The test substance in this case is a chemical preparation, particularly a drug. This substance is first homogenized to a fine fraction by grinding and weighed to 10-20 mg doses. 1 distilled water or another solvent such dimethylsulfoxide is added to the drug. The sample shaken for vigorously stirred or one minute and after dissolution, at least partial, $50 \mu L$ of the resulting suspension is mixed with 50 μL of 0.1 mol / liter phosphate buffer pH 7 and / or distilled water in a UV transparent microtiter plate or spectroscopic cuvette. The sample is first analyzed in its native state using spectroscopic methods of excitation absorption and / or radiation emission. An example can be an absorption scan in the range of 230 to 800 nm or an excitation scan at $\lambda_{\rm ex}$ = 230 to 470 nm and $\lambda_{\rm em}$ = 500 nm (optionally 450 nm). Afterwards, the sample plate is exposed to UV radiation (λ_{em} = 254 nm) in a transilluminator or other radiation source for 6 minutes. Then the spectral analysis is repeated under the same parameters. The results of the analysis are unique spectra that are typical of a given analyte or sample composition. The spectra obtained are created based on the UV-induced degradation of the test i.e. by photobleaching or dissociation substance. functional groups, which leads to the formation of free radicals and subsequent typical reactions.

This procedure and its results correspond to FIG. 2. The analysis of other liquid, dissolved or suspension samples is carried out in an analogous way. In this way, the profiling of chemical mixtures can be performed to identify them.

Example 2

The test substance in this case is a fruit drink or wine. The beverage is diluted to four concentrations by the half-dilution with distilled water. The beverage is then mixed 1: 1 with 0.1 mol / liter phosphate buffer pH 7 in a microtiter plate or spectroscopic cuvette. The plate is exposed for 10 to 15 minutes to UV radiation ($\lambda_{em}=254$ nm) in a transilluminator or by means of other radiation source. Emission or excitation spectra are then recorded. An example can be an emission scan at $\lambda_{ex}=250$ nm and $\lambda_{em}=280$ to 800 nm. The emission or excitation spectra recorded are typical of the beverage. The spectra obtained are the result of UV radiation caused by the photooxidation of thiol groups (SH) and subsequent interaction with metal ions. Furthermore, free radicals are generated under the influence of UV radiation, which interact with other molecules.

This procedure and its results correspond to FIGs. 3 and 4. In this way, it is possible to determine precisely what fruit the juice originates from, or whether the necessary technology has been observed in processing, storage, peeling prior to pressing, or similar beverage preparation operations.

Example 3

The subject under examination corresponds to vegetable or fruit extracts. The extract or juice obtained by pressing or leaching from fruit or vegetables is filtered to remove the pulp or other impurities and solid fractions. Then the sample is mixed 1: 1 with 0.1 mol / liter phosphate buffer pH 7 in a microtiter plate or spectroscopic cuvette. The sample is first analyzed in its native state, i.e. without UV

irradiation by spectroscopic methods, i.e. the absorption, excitation and emission of radiation. An example can be an excitation scan at $\lambda_{\rm ex}=230$ to 370 nm and $\lambda_{\rm em}=400$ nm. Afterwards, the sample plate is exposed to UV radiation ($\lambda_{\rm m}=254$ nm) for 20 minutes in a transilluminator or other UV device. Then the spectral analysis is repeated under the same parameters. The results of the analysis are unique spectra, which are typical of a given variety of fruits or vegetables. The spectra obtained are the result of UV radiation caused by the photooxidation of thiol groups (SH) and subsequent interaction with metal ions. Furthermore, free radicals are generated under the influence of UV radiation, which interact with other molecules.

This procedure and its results correspond to FIG. 5. In this way, different varieties of vegetables and fruits can be distinguished and identified.

Example 4

The subject under investigation corresponds to clinical specimens. As in the previous examples, biological or clinical specimens such as blood plasma or urine may be used. These samples can be used to study the physiological or pathological conditions of an organism associated with metal ions and -S-S-, -SH molecules. A change in the concentration of metal ions and / or -SS-, -SH compounds in the study sample results in a change in the emission or excitation spectrum of the shape or as an increase or decrease in the emission or excitation signal at a particular wavelength (for example $\lambda_{\rm ex}$ = 400 nm and $\lambda_{\rm em}$ = 470 nm) after UV irradiation.

This procedure and its results correspond to FIG. 6. In this way, control tests can be performed to detect serious diseases at an early stage.

Example 5

In this case, the investigation focuses on the localization and identification of cellular processes. A cell line or histological section is placed or dropped in the volume 1-50 μ L on a glass slide and placed in a fluorescent microscope. Using the microscope and appropriate software, cell locations to be analyzed are selected. Next, a laser with a suitable wavelength (typically $\lambda_{em}=385$ nm) is selected, which for a few seconds or minutes, most often in the range of 1 to 90 seconds, induces cluster formation at the analysis site. These clusters are evaluated in real time or after the analysis. The resulting clusters are typical of a given site in a cell, for its composition and for the biological or chemical processes taking place therein. An example of the analysis performance is shown in FIG. 7.

Industrial applicability

The use of the method according to the invention can be expected in areas where it is necessary to monitor the composition of pharmacological agents, the quality and composition of beverages or foods, or to diagnose serious diseases from body fluids or at the level of a histological section or cell samples.

The method makes it possible, for example, to detect counterfeiting or to determine illegal additives in mixtures such as medicines, food, and many other types of substances. Similarly, the method can be used to detect a number of misdemeanors associated with food counterfeiting, or failure to adhere to a given technology or recipe in beverage and food production.

Furthermore, the procedure may also serve to identify a drug by the rescue service during the transfer of a patient to the hospital. The procedure provides a cheap and fast screening tool to hospitals or research institutes by the study of clinical specimens. Using advanced spectroscopic techniques, such as fluorescence correlation spectroscopy, the method can localize or monitor cellular processes within a single cell. The intended method is compatible with all spectroscopic methods using electromagnetic waves in the classical range.

Legend for Figures of Drawings

Fig. 1

- 1) The source of ultraviolet radiation that can be characterized according to:
 - 2) duration time,
 - 3) wavelength, and
 - 4) intensity.

Exposure of a sample

- 5) by ultraviolet radiation causes spectral changes based on:
 - 6) formation of fluorescence nanoparticles/complexes,
 - 7) formation of free radicals,
 - 8) degradation of chemical bonds, and/or
 - 9) photobleaching.

These changes are typical for sample and can by monitored as a 10) absorbance, fluorescence and/or excitation spectra.

Fig. 7

11) The drop of sample containing prokaryotic and/or eukaryotic cells is placed on

- 12) plain glass microscope slide and
- 13) covered with a shield box to remove a ambient light.
- 14) The lens is focused on selected part of cell (sample). Into the
- 15) confocal volume is concentrated the
- 16) excitation laser and causing the
- 17) formation of fluorescent nanocrystals/complexes, which are detected by the
 - 18) detector.

Fig. 8

- 19) Real image from fluorescence microscopy of 2% agarose gel containing zinc and cysteine. The point of analysis is characterized by
- 20) photobleaching (loss of autofluorescence) and by formation of new
 - 21) fluorescent nanocrystals.
- 22) Real photo of single cell from a fluorescence microscope. The site of analysis in the cell is characterized by the formation of
 - 23) and
- 24) fluorescent nanocrystals. Spectral analysis of these fluorescent nanocrystals provides information about a given cell site.

CLAIMS

- A method for identifying the properties of substances, in 1. particular liquid substances, characterized a sample of a substance containing on the one hand metal ions and on the other hand a thiol group or disulphide bonds is first subjected to the analysis in the original or initial untreated state, i.e. not irradiated by UV radiation, then it is subjected to UV radiation at a wavelength of 250 to 400 nm, thereby causing spectral changes that are characteristic of the given substance, and based on the change in the spectral characteristics the sample before irradiation, or information contained in the non-irradiated sample information database, and after its irradiation with UV radiation, the inherent optical properties and relevant differences between the original substance and the sample under investigation are investigated.
- 2. The method for identifying the properties of substances, in particular liquid substances, according to claim 1, characterized in that the sample of the substance contains metal ions, in particular some of Zn^{2+} , Cu^{2+} , Fe^{2+} , Mg^{2+} , Se^{2+} or Cd^{2+} .

3. The method for identifying the properties of substances, in particular liquid substances, according to claim 1, characterized in that the sample of the substance contains a thiol group, in particular some of cysteine, homocysteine, glutathione oxidized, glutathione reduced, S-Adenosyl-L-homocysteine, S-Adenosyl-methionine, coenzyme A, cystamine, acetylcysteine, gamma-L-glutamyl-L-cysteine.

- 4. The method for identifying the properties of substances, in particular liquid substances, according to claim 1, characterized in that UV radiation in the sample generates quantum nanocrystals, free radicals, degrades chemical bonds and bleaches.
- 5. The method for identifying the properties of substances, in particular liquid substances, according to claim 1, characterized in that the concentration change of metal ions concentration, in particular Zn²+, Cu²+, Fe²+, Mg²+, Se²+ or Cd²+ and thiol compounds, in particular cysteine, homocysteine, glutathione oxidized, glutathione reduced, S-Adenosyl-L-homocysteine, S-Adenosyl-methionine, coenzyme A, cystamine, acetylcysteine, gamma-L-glutamyl-L-cysteine in the study sample results in a change in the absorption or emission or excitation spectrum or as an increase or decrease of the absorption, emission or excitation signal.
- 6. The method for identifying the properties of substances, in particular liquid substances, according to claim 1, characterized in that the concentration change of the metal ions and the thiol group results in a change in the formation of quantum nanocrystals.

7. The method for identifying the properties of substances, in particular liquid substances, according to claim 1, characterized in that the concentration change of the metal ions and the thiol group results in a change in the formation of radicals.

- 8. The method for identifying the properties of substances, in particular liquid substances, according to claim 1, characterized in that the concentration change of the metal ions and the thiol group results in a change in the degradation of multiple chemical bonds.
- 9. The method for identifying the properties of substances, in particular liquid substances, according to claim 1, characterized in that the concentration change of the metal ions and the thiol group results in a change in photo-bleaching.

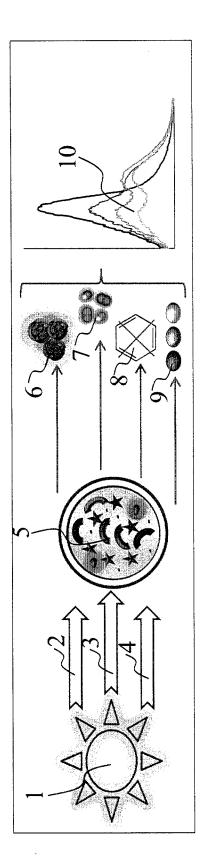
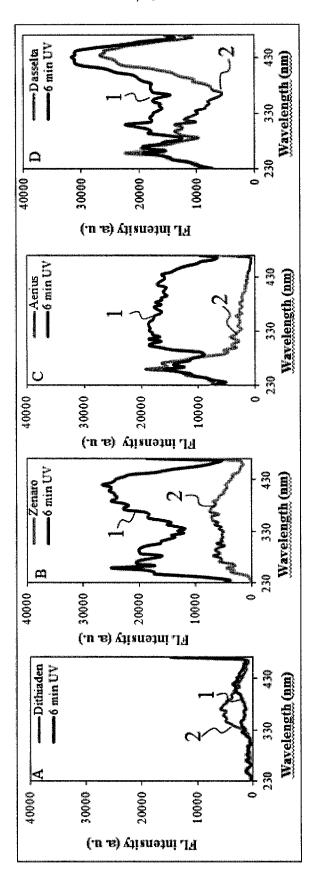


FIG.



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FIG.

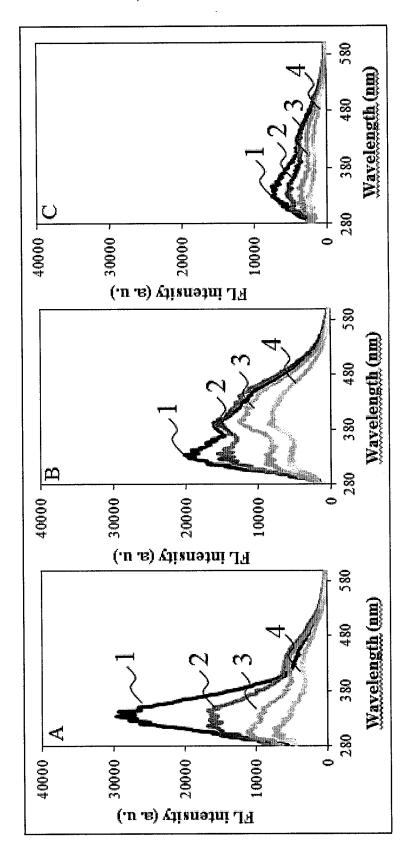


FIG. 3

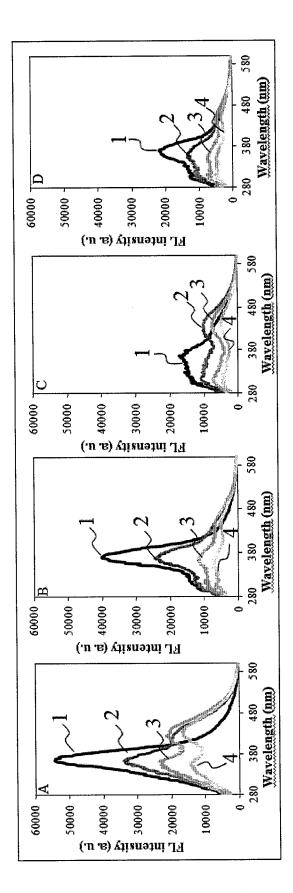


FIG. 4

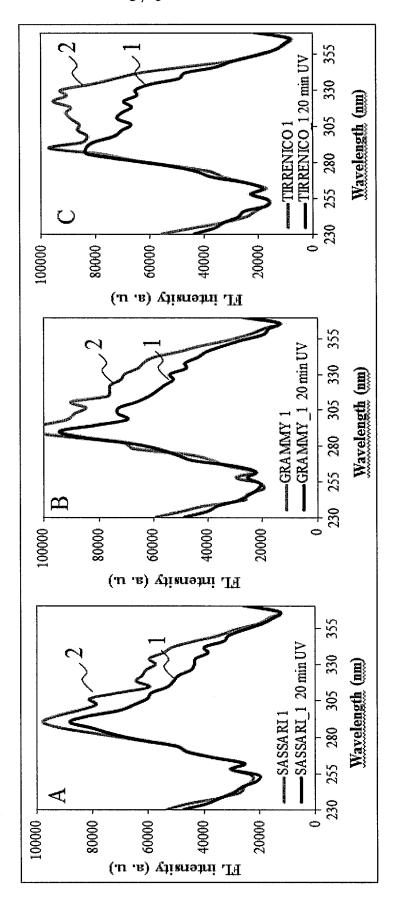


FIG. 5

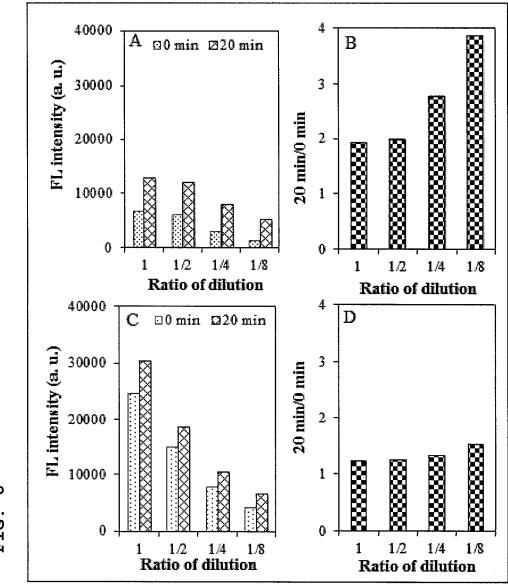
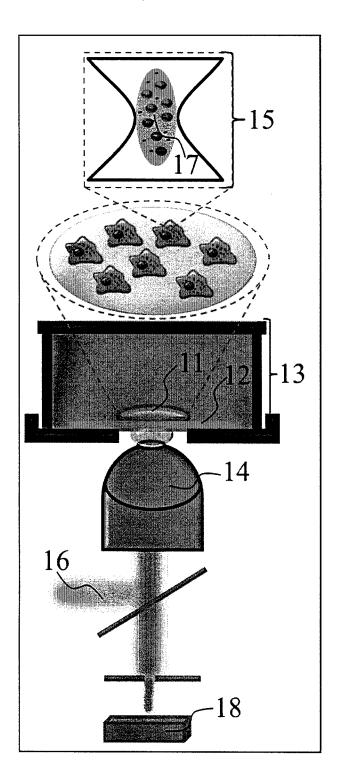


FIG. 6

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FIG

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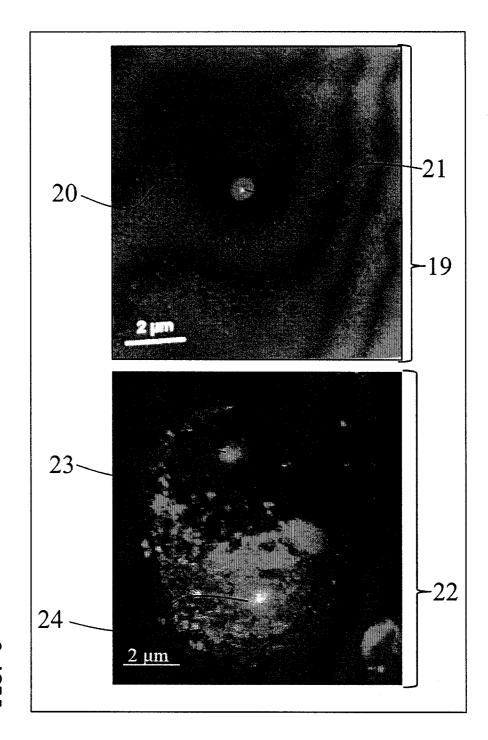


FIG. 8

INTERNATIONAL SEARCH REPORT

International application No PCT/CZ2019/000046

a. classification of subject matter INV. G01N21/31 G01N2 INV. ADD. G01N21/17

G01N21/64 G01N33/02

G01N33/14 G01N21/75

G01N33/52

C09K11/00

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

G01N H05B C09K A61B

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

EPO-Internal, WPI Data

C. DOCUMENTS CONSIDERED TO BE RELEVANT					
0	0.00				

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US 2013/165330 A1 (NATARAJAN ARUNKUMAR [US] ET AL) 27 June 2013 (2013-06-27) paragraphs [0001], [0005] - [0051], [0098] - [0100], [0110] - [0113], [0121], [0138], [0155] - [0156], [0179] - [0203] paragraphs [0234] - [0237]	1-5,9
X	US 2007/141726 A1 (YING JACKIE Y [SG] ET AL) 21 June 2007 (2007-06-21) paragraphs [0001], [0004], [0005], [0020], [0023], [0029] - [0032], [0037]; figure 4; examples 1,3,5,6	1-6,8,9

Χ See patent family annex.

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Date of mailing of the international search report

Date of the actual completion of the international search

18 November 2019 03/12/2019

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Authorized officer

Duijs, Eric

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INTERNATIONAL SEARCH REPORT

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PCT/CZ2019/000046

	ation). DOCUMENTS CONSIDERED TO BE RELEVANT	
ategory*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
ategory*	US 6 313 274 B1 (SYKES THOMAS R [CA] ET AL) 6 November 2001 (2001-11-06) column 1, lines 10-13 column 4, lines 5-13 column 5, line 21 - column 13, line 1 column 15, lines 15-32; figures 2,5; examples 6,102	1-5,7

INTERNATIONAL SEARCH REPORT

Information on patent family members

International application No
PCT/CZ2019/000046

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
US 2013165330 A1	27-06-2013	AU 2012355736 A1 BR 112014015603 A2 CA 2860097 A1 CN 104114713 A EP 2794908 A1 JP 6148682 B2 JP 2015506469 A KR 20140103350 A NZ 626334 A RU 2014124974 A SG 11201403460S A US 2013165330 A1 US 2013345089 A1 WO 2013095896 A1	17-07-2014 13-06-2017 27-06-2013 22-10-2014 29-10-2014 14-06-2017 02-03-2015 26-08-2014 29-04-2016 20-02-2016 30-07-2014 27-06-2013 26-12-2013
US 2007141726 A1	21-06-2007	CN 101365938 A EP 1969349 A2 JP 2009520207 A US 2007141726 A1 WO 2007075495 A2	11-02-2009 17-09-2008 21-05-2009 21-06-2007 05-07-2007
US 6313274 B1	06-11-2001	AT 160222 T AU 675235 B2 CA 2137561 A1 DE 69315213 D1 DE 69315213 T2 EP 0649532 A1 FI 950062 A JP H08505119 A US 6313274 B1 WO 9401773 A1	15-11-1997 30-01-1997 20-01-1994 18-12-1997 19-03-1998 26-04-1995 05-01-1995 04-06-1996 06-11-2001 20-01-1994