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(54) **IMMUNOGENIC COMPOSITION
COMPRISING AN INACTIVATED
RECOMBINANT NON-PATHOGENIC
BACTERIUM**

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(57) **ABSTRACT**

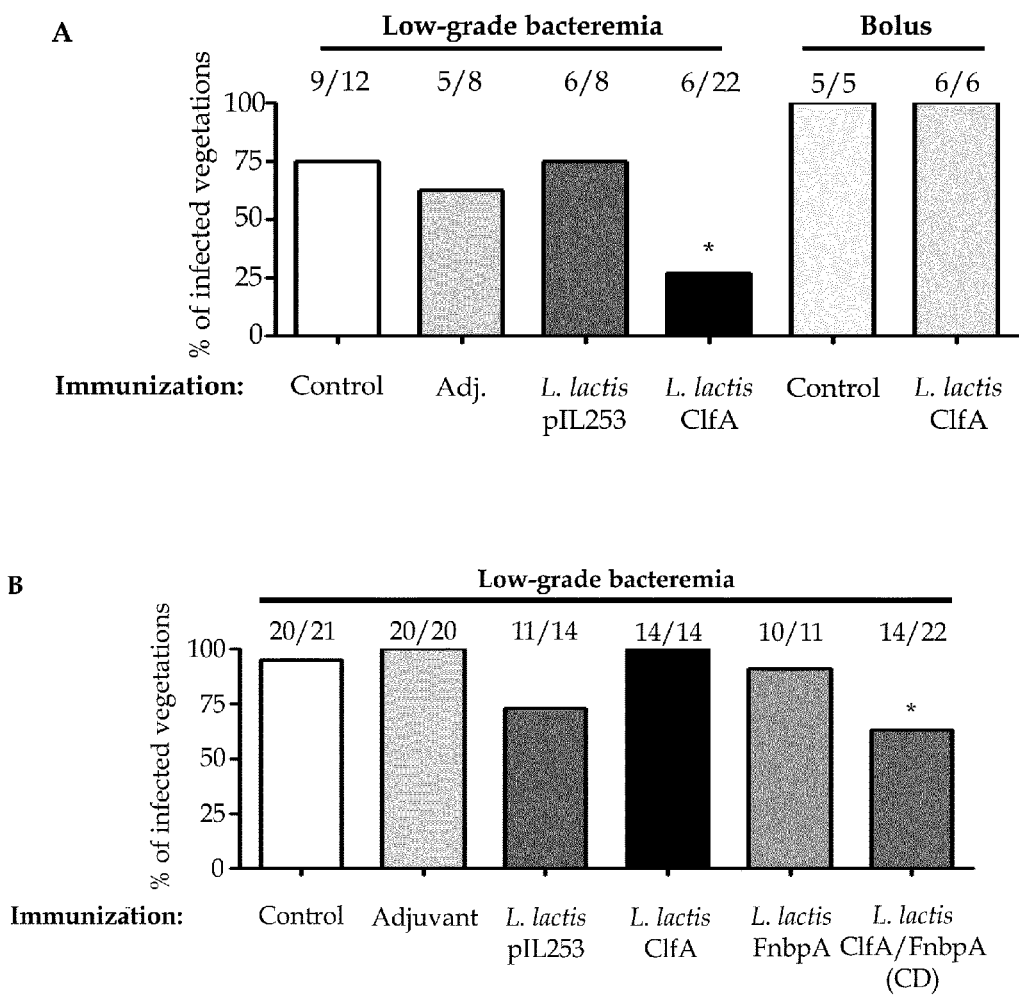
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§ 371 (c)(1),

(2) Date: **Mar. 5, 2015**

The present invention relates to immunogenic compositions, vaccines and antibodies for the treatment and/or prevention of infections and diseases caused by *S. aureus* in a subject in need thereof.

Fig. 1



**IMMUNOGENIC COMPOSITION
COMPRISING AN INACTIVATED
RECOMBINANT NON-PATHOGENIC
BACTERIUM**

FIELD OF THE INVENTION

[0001] The present invention relates to immunogenic compositions, vaccines and antibodies for the treatment and/or prevention of infections and diseases caused by *S. aureus* in a subject in need thereof.

BACKGROUND OF THE INVENTION

[0002] *Staphylococcus aureus* is a major pathogen responsible for a variety of diseases, from benign skin infections, such as folliculitis and furunculosis, to life-threatening conditions, including erysipelas, deep-seated abscesses, osteomyelitis, pneumonia, sepsis, and infective endocarditis (IE). In addition to infections in which the organism is physically present at the infected site, *S. aureus* is also capable of producing “distant” diseases, which are mediated by the secretion of toxins. This success is ensured by the coordinated expression of numerous surface adhesins, which mediate host-tissue colonization, and secreted proteins and toxins, which promote invasion as well as strategies to escape the host immune system (Que et al., 2009).

[0003] For instance, *S. aureus* adhesins—which are collectively referred to as MSCRAMMs for Microbial Surface Components Reacting with Adherence Matrix Molecules—encompass at least 21 surface-anchored proteins including fibrinogen-binding proteins A and B (clumping factors A and B, or ClfA and ClfB), fibronectin-binding proteins A and B (FnPBA and FnBPB), collagen-binding protein (Cna) and protein A (Spa) to mention just a few (Patti et al., 1994).

[0004] Previous studies have shown that ClfA is essential for the development of IE. Moreover, after individual expression of ClfA in the non-pathogenic bacteria *Lactococcus lactis*, it has been shown that fibrinogen-binding was pivotal in promoting IE. In addition ClfA also interacts with the GPIIb/III α receptor on the surface of platelets, a feature that plays an important indirect role in the ability of *S. aureus* to induce IE (Que et al., 2011).

[0005] Numerous attempts to develop vaccines against a variety of these structures, especially against the fibrinogen binding-domain of ClfA, have been attempted with various successes in animal models, but none of them have achieved sustainable efficacy in human clinical trials (Broughan et al., 2011).

[0006] In parallel, expressing antigens in non-pathogenic *L. lactis* has been attempted to trigger mucosal immunity. However, technical issues such as in vivo persistence of the bacterial strain as well as antigen release are as yet incompletely solved (Wells et al., 2008).

[0007] However, despite these studies and attempts, there is currently no efficacious immunogenic composition for treating and/or preventing *S. aureus* infections and diseases caused by *S. aureus*.

[0008] This object has been achieved by providing an immunogenic composition comprising an inactivated recombinant non-pathogenic bacterium, or a part thereof, expressing on its cell surface at least one folded sequence of a *S. aureus* adhesin, or a sequence having 80% or more sequence identity to said folded sequence of a *S. aureus* adhesin, wherein said at least one folded sequence of a *S. aureus*

adhesion, or sequence having 80% or more sequence identity to said folded sequence of a *S. aureus* adhesin, is entirely accessible to trypsin digestion.

SUMMARY OF THE INVENTION

[0009] The invention provides an immunogenic composition comprising an inactivated recombinant non-pathogenic bacterium, or a part thereof, expressing on its cell surface at least one folded sequence of a *S. aureus* adhesin, or a sequence having 80% or more sequence identity to said folded sequence of a *S. aureus* adhesin, wherein said at least one folded sequence of a *S. aureus* adhesin, or sequence having 80% or more sequence identity to said folded sequence of a *S. aureus* adhesin, is entirely accessible to trypsin digestion.

[0010] Furthermore, the invention also provides a vaccine comprising an immunogenic composition of the invention in an immunologically acceptable carrier or diluent.

[0011] The invention further provides the use of the vaccine of the invention for the treatment and/or prevention of infections and diseases caused by *S. aureus* in a subject in need thereof.

[0012] Also provided is an isolated and/or purified antibody, antibody fragment or derivative thereof able to bind to the at least one folded sequence of a *S. aureus* adhesin expressed on the cell surface of an inactivated recombinant non-pathogenic bacterium.

BRIEF DESCRIPTION OF THE FIGURE

[0013] FIG. 1. Prevention of *S. aureus* experimental endocarditis (i.e. infected vegetations in cardiac valves) in rats immunized with vaccine preparations. Rats were immunized with various vaccine preparations (see infra) for 6 weeks as described, before aortic vegetations were induced. 24 h later, they were challenged with identical inoculum sizes administered either by continuous infusion (0.0017 ml/min over 10 h), or by i.v. bolus (1 ml in 1 min). (A) Animals challenged with *S. aureus* Newman (10^4 CFU), and (B) with *S. aureus* P8 (10^6 CFU). * $P < 0.05$ compared to the control group by χ^2 test. Control: group immunized with PBS; Adj.: group immunized with Freund’s adjuvant group with the adjuvant emulsified at a 1:1 ratio in PBS; *L. lactis* pIL253, *L. lactis* ClfA and *L. lactis* ClfA/FnBPA(CD): groups immunized with *L. lactis* pIL253, ClfA or *L. lactis* ClfA/FnBPA (CD), respectively, emulsified at a 1:1 ratio in Freund’s adjuvant.

DESCRIPTION OF THE INVENTION

[0014] Although methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, suitable methods and materials are described below. All publications, patent applications, patents, and other references mentioned herein are incorporated by reference in their entirety. The publications and applications discussed herein are provided solely for their disclosure prior to the filing date of the present application. Nothing herein is to be construed as an admission that the present invention is not entitled to antedate such publication by virtue of prior invention. In addition, the materials, methods, and examples are illustrative only and are not intended to be limiting.

[0015] Unless defined otherwise, all technical and scientific terms used herein have the same meaning as is commonly

understood by one of skill in art to which the subject matter herein belongs. As used herein, the following definitions are supplied in order to facilitate the understanding of the present invention.

[0016] The term “comprise” or “comprising” is generally used in the sense of include/including, that is to say permitting the presence of one or more features or components.

[0017] As used in the specification and claims, the singular form “a”, “an” and “the” include plural references unless the context clearly dictates otherwise.

[0018] As used herein, “at least one” means “one or more.”

[0019] As used herein, the terms “protein”, “polypeptide”, “polypeptidic”, “peptide” and “peptidic” are used interchangeably herein to designate a series of amino acid residues connected to the other by peptide bonds between the alpha-amino and carboxy groups of adjacent residues.

[0020] As used herein the term “subject” is well-recognized in the art, and, is used herein to refer to a mammal, including dog, cat, rat, mouse, monkey, cow, horse, goat, sheep, pig, camel, and, most preferably, a human. The term does not denote a particular age or sex. Thus, adult and newborn subjects, whether male or female, are intended to be covered.

[0021] The present invention provides an immunogenic composition comprising an inactivated recombinant non-pathogenic bacterium, or a part thereof, expressing on its cell surface at least one folded sequence of a *S. aureus* adhesin, or a sequence having 80% or more sequence identity to said folded sequence of a *S. aureus* adhesin, wherein said at least one folded sequence of a *S. aureus* adhesin, or sequence having 80% or more sequence identity to said folded sequence of a *S. aureus* adhesin, is entirely accessible to trypsin digestion.

[0022] A “non-pathogenic bacterium” refers to a bacterium that does not cause a disease state when in contact with a subject. Examples of non-pathogenic bacteria are selected from the non-limiting group comprising the *Bacillus* genus, *Lactobacillus* genus, *Lactococcus* genus, *Sporolactobacillus* genus, *Bifidobacterium* genus, and the like bacteria. Preferably, the non-pathogenic bacterium is selected from the group comprising *L. lactis*, *L. acidophilus* and *Lactobacilli*. Particularly preferred is *L. lactis* and most particularly preferred is *L. lactis* subspecies *cremoris* 1363.

[0023] *Lactococcus lactis* is a nonpathogenic bacterium that is being developed, in its live form, as a vaccine delivery vehicle for immunization by mucosal routes.

[0024] However, the non-pathogenic bacterium of the invention is in an inactivated form. Inactivation is performed using methods and/or compounds known in the art, such as for example by H₂O₂, formaldehyde, heat or UV treatments.

[0025] The present invention also refers to a part of said inactivated recombinant non-pathogenic bacterium. Usually, this part consists in an isolated and/or purified cell wall of the inactivated recombinant non-pathogenic bacterium.

[0026] Generally, the non-pathogenic bacterium of the invention is a recombinant bacterium as it comprises at least one heterologous nucleic acid molecule encoding a folded sequence of a *S. aureus* adhesin.

[0027] Usually, the nucleic acid molecule encoding a folded sequence of a *S. aureus* adhesin is in the form of deoxyribonucleic acid (DNA). DNA which can be used herein is any polydeoxynucleotide sequence, including, e.g. double-stranded DNA, single-stranded DNA, double-stranded DNA wherein one or both strands are composed of

two or more fragments, double-stranded DNA wherein one or both strands have an uninterrupted phosphodiester backbone, DNA containing one or more single-stranded portion(s) and one or more double-stranded portion(s), double-stranded DNA wherein the DNA strands are fully complementary, double-stranded DNA wherein the DNA strands are only partially complementary, circular DNA, covalently-closed DNA, linear DNA, covalently cross-linked DNA, cDNA, chemically-synthesized DNA, semi-synthetic DNA, biosynthetic DNA, naturally-isolated DNA, enzyme-digested DNA, sheared DNA, labeled DNA, such as radiolabeled DNA and fluorochrome-labeled DNA, DNA containing one or more non-naturally occurring species of nucleic acid, genomic or complementary DNA. Preferably, the DNA is a genomic DNA or a complementary DNA (cDNA).

[0028] DNA sequences that encode a folded sequence of a *S. aureus* adhesin can be synthesized by standard chemical techniques, for example, the phosphotriester method or via automated synthesis methods and PCR methods.

[0029] The DNA sequence encoding a folded sequence of a *S. aureus* adhesin according to the invention may also be produced by enzymatic techniques. Thus, restriction enzymes, which cleave nucleic acid molecules at predefined recognition sequences can be used to isolate nucleic acid sequences from larger nucleic acid molecules containing the nucleic acid sequence, such as DNA (or RNA) that codes for a peptide consisting a folded sequence of a *S. aureus* adhesin.

[0030] Encompassed by the present invention is also a nucleic acid in the form of a polyribonucleotide (RNA), including, e.g., single-stranded RNA, cRNA, double-stranded RNA, double-stranded RNA wherein one or both strands are composed of two or more fragments, double-stranded RNA wherein one or both strands have an uninterrupted phosphodiester backbone, RNA containing one or more single-stranded portion(s) and one or more double-stranded portion(s), double-stranded RNA wherein the RNA strands are fully complementary, double-stranded RNA wherein the RNA strands are only partially complementary, covalently crosslinked RNA, enzyme-digested RNA, sheared RNA, mRNA, chemically-synthesized RNA, semi-synthetic RNA, biosynthetic RNA, naturally-isolated RNA, labeled RNA, such as radiolabeled RNA and fluorochrome-labeled RNA, RNA containing one or more non-naturally-occurring species of nucleic acid.

[0031] The present invention also includes variants of the aforementioned sequences that are nucleotide sequences that vary from the reference sequence by conservative nucleotide substitutions, whereby one or more nucleotides are substituted by another with same characteristics.

[0032] The invention also encompasses allelic and polymorphic variants of the aforementioned sequences; that is, naturally-occurring alternative forms of the folded sequence of a *S. aureus* adhesin that also encode peptides that are identical, homologous or related to that encoded by the sequence of a *S. aureus* adhesin. Alternatively, non-naturally occurring variants may be produced by mutagenesis techniques or by direct synthesis.

[0033] Also encompassed in the present invention is a sequence having 80% or more sequence identity to said folded sequence of a *S. aureus* adhesin. The percentage identity of a polynucleotide or polypeptide sequence is determined by aligning polynucleotide and polypeptide sequences; identifying the number of identical nucleic or amino acids over the aligned portions; dividing the number of

identical nucleic or amino acids by the total number of nucleic or amino acids of the polynucleotide or polypeptide of the present invention; and then multiplying by 100 to determine the percentage identity. Preferably, the sequence has at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or 100% sequence identity to said folded sequence of a *S. aureus* adhesin.

[0034] In case the sequence of a *S. aureus* adhesin is the clumping factor A (ClfA) then the acid nucleic molecule encoding the folded sequence of a *S. aureus* adhesin, namely the ClfA, is a genomic DNA as set forth in SEQ ID No 2.

TABLE 1

	SEQ ID No 2			
1	ggtaccataa	attacacatc	tgcttttgaa	aaaatatgat
	ttcaagctag	gattacatta		
61	ggtagagttc	atattaataa	taaaaaatgt	ttgcaatcaa
	atcgtacgtt	gtcgtttgta		
121	attcttaaaa	tagcaataaa	taaaatgttt	gttagtaaag
	tattattgtg	gataataaaa		
181	tatcgataca	aattaattgc	tataatgcaa	ttttagtgtg
	taattccatt	aacagagatt		
241	aatatatact	ttaaagggtg	tatagttaat	ataaaatgac
	tttttaaaaa	gaggggaataa		
301	aatgaatgat	aagaaaaaag	aaaaacacgc	aattcggaaa
	aaatcgattg	gcgtaggcttc		
361	agtgcttgta	ggtacgttaa	tcggttttgg	actactcagc
	agtaaagaag	cagatgcaag		
421	tgaaaatagt	gttacgcaat	ctgatagcgc	aagtaacgaa
	agcaaaagta	atgattcaag		
481	tagegttagt	gctgcacctg	aaacagacga	cacaaaactg
	agtgatacta	aaacatcgtc		
541	aaacactaat	aatggcgaaa	cgagtgtggc	gcaaaatcca
	gcacaacagg	aaacgacaca		
601	atcatcatca	acaaatgcaa	ctacggaaga	aacgccggtg
	actggtagaag	ctactactac		
661	gacaacgaat	caagctaata	caccggcaac	aactcaatca
	agcaatacaa	atgcggagga		
721	attagtgaat	caaaacagta	atgaaacgac	ttttaatgat
	actaatacag	tatcatctgt		
781	aaattcacct	caaaattcta	caaatgcgga	aaatgtttca
	acaacgcaag	atacttcaac		
841	tgaagcaaca	ccttcaaca	atgaaatcag	tccacagagt
	acagatgcaa	gtaataaaga		
901	tgtagttaat	caagcgggta	atacaagtgc	gcctagaatg
	agagcattta	gtttagcggc		
961	agtagctgca	gatgcaccgg	cagctggcac	agatattacg
	aatcagttga	cgaatgtgac		
1021	agttggatt	gactctggta	cgactgtgta	tccgcaccaa
	gcaggttatg	tcaaaactgaa		
1081	ttatggtttt	tcagtgccta	attctgctgt	taaaggtgac
	acattcaaaa	taactgtacc		

TABLE 1-continued

1141	taaagaatta	aacttaaagt	gtgtaacttc	aactgctaaa
	gtgccacca	ttatggctgg		
1201	agatcaagta	ttggcaaatg	gtgtaatcga	tagtgatggg
	aatgttattt	atacatttac		
1261	agactatgta	aataactaaag	atgatgtaaa	agcaactttg
	accatgcccc	cttatattga		
1321	ccctgaaaat	gttaaaaaga	caggtaatgt	gacattggct
	actggcatag	gtagtacaac		
1381	agcaaacaaa	acagtattag	tagattatga	aaaatatggg
	aagttttata	acttatctat		
1441	taaaggtaaa	attgacccaa	tcgataaaaac	aaataatcag
	tatcgtcaga	caatttatgt		
1501	caatccaagt	ggagataaac	ttattgcgcc	ggttttaaca
	ggtaatttaa	aaccaaatac		
1561	ggatagtaat	gcattaatag	atcagcaaaa	tacaagtatt
	aaagtatata	aagtagataa		
1621	tgcagctgat	ttatctgaaa	gttactttgt	gaatccagaa
	aactttgagg	atgtcactaa		
1681	tagtgatgat	attacattcc	caaatccaaa	tcaatataaa
	gtagagttaa	atcgcctgta		
1741	tgatcaaatt	acaacaccgt	atagtagtag	tgtaaatggg
	catattgatc	cgaatagcaa		
1801	aggtgattta	gctttacggt	caactttata	tgggtataac
	tcgaatataa	tttggcgctc		
1861	tatgtcatgg	gacaacgaag	tagcatttaa	taacggatca
	ggttctgggtg	acggatcaga		
1921	taaaccagtt	gttctggaac	aacctgatga	gcctgggtgaa
	attgaaccaa	ttccagagga		
1981	ttcagattct	gaccacgggt	cagattctgg	cagcgattct
	aattcagata	gcggttcaga		
2041	ttcgggtagt	gattctacat	cagatagtag	ttcagattca
	gcgagtgatt	cagattcagc		
2101	aagtgattca	gactcagcga	gtgattcaga	ttcagcaagc
	gattccgact	cagcagcaga		
2161	ttccgactca	gacaatgact	cggattcaga	tagcgattct
	gactcagaca	gtgactcaga		
2221	ttccgacagt	gactcagatt	cagatagcga	ttctgactca
	gacagtgact	cagattcaga		
2281	tagcgattca	gattcagata	gcgattcaga	ttccgacagt
	gattccgact	cagacagcga		
2341	ttctgactcc	gacagtgatt	ccgactcaga	cagcgattca
	gattccgaca	gtgattccga		
2401	ctcagatagc	gattccgact	cagatagcga	ctcagattca
	gacagcgatt	cagattcaga		
2461	cagcgattca	gattcagata	gcgattcaga	ttccgacagt
	gactcagatt	ccgacagtag		
2521	ctcggattca	gatagcgatt	cagattccga	cagtgactca
	gattccgaca	gtgactcaga		
2581	ctcagacagt	gattccgatt	cagcagtagta	ttcggattca
	gatagtgatt	ccgactccga		
2641	cagtgactcg	gattcagata	gcgactcaga	ctcggatagc
	gactccgatt	cagatagcga		

TABLE 1-continued

2701	ttcggactca gatagcgatt cagaatcaga cagcgattca gaatcagaca gcgattcaga
2761	ttcagacagc gactcagaca gtgactcaga ttcagatagt gactcggatt cagcagagtga
2821	ttcagactca ggtagtgact cggattcatc aagtgattcc gactcagaaa gtgattcaaaa
2881	tagcgattcc gagtcagggt ctaacaataa tgtagttccg cctaattcac ctaaaaaatgg
2941	tactaatgct tctaataaaa atgaggctaa agatagtaaa gaaccattac cagatacaggy
3001	ttctgaagat gaagcaataa cgtcactaat ttggggatta ttagcatcaa tagggttcatt
3061	actacttttc agaagaaaaa aagaaaataa agataagaaa taagtaataa tgatattaaa
3121	ttaatcatat gattcatgaa gaagccacct taaaagggtgc ttcttttact tggattttcc
3181	aaatatattg tttgaatata attaataatt aattcatcaa cagttaatta ttttaaaaaag
3241	gtagatgta tataatttgg ctggcgaaa aaatagggtg taaggtaggt tgtaattag
3301	ggaaaattaa ggagaaaaa cagttgaaaa ataaattgct agttttatca ttggggagcat
3361	tatgtgatc acaaaatttgg gaaagtaatc gtgcgagtgc agtggtttct ggggagaaga
3421	atccatatgt atctgagtcg ttgaaactga ctaataataa aaataaatct agaacagtag
3481	aagagtataa gaaaagctt

[0035] In case the sequence of a *S. aureus* adhesin is the fibronectin-binding protein A (FnPBA) then the acid nucleic molecule encoding the folded sequence of a *S. aureus* adhesin, namely the FnPBA, is preferably a genomic DNA as set forth in SEQ ID No 3.

TABLE 2

	SEQ ID No 3
1	ttatgctttg tgattctttt tatttctcgg taataatgct aaacctagaa tgctgaataa
61	tccgccgaac aacatacctt tgtttggtga ttcttctoca cctgtttcag gtagttcaga
121	tttcttagat tgtggttttt tagttggtgc cactgcttta accttttcat tgatttcaat
181	aacagggtgt actactttac cttgttccac tggtttagaa ggctttttag gttcttcttt
241	ggcagggtgt actggtttac caggttcagc tggtaacctc ggtgttggcg gtgttggagt
301	ttctggctca ctccggacct ctgggtgcgg tgggtgtggg gtttccggct cacttggtag
361	ttctgggtgt ggtggcgttg gtgtttccgg ctcaactggg acttctgggt tcggtggcgt
421	tgggtggcac attggagggt ttgtatcttc ttcaatcgtt tgttgacctt cattttggcc

TABLE 2-continued

481	gcttactttt ggaagtgtat cttcttcaaa gtcaacacta ttgtgtccac cgaattgata
541	acttggttta tctttatttg tatcttcttc aataatttca gtgtgcttat tgaatccgtg
601	aatatgtggc acactgtcga agtcgatatac aatgatgta ccgccatggt cataacttagg
661	tttgtctttt tctgtatctt cctcgaatga ctgattaact ttattttgac catgaatttg
721	aggtacacta tcaaaatcga tatctacgat attgccacct tgttcatatt taggtttgtc
781	ttcttctgtg tcttctcga atgactgggt accgctattt tggccacctt cataacctaa
841	ttcactctta atatcaactg ggctattttc ttcgatttct tcaatcacgt cataattccc
901	gtgaccattt tcagttccta aaccagaatg agaaatatga tgattgtttt tagtaatttc
961	ctcgactggc ccttgtgctt gaccatgctc ttcaggtaat tcactcacta attcaatcag
1021	attactttca gttgtatatt ctttctgatac ttcaactggt gtatgatcgc tcaactgcgc
1081	agttacaata cctttttag actctctcgc aaattcaact aagttagact cagtagtaac
1141	ctgaccacca cctgggtttg tatcttcttc atattcaaca acatcagcgt gatgttttga
1201	attttcatgt gtagattctt caaagtcaat tggatttgat tcctcagagg actcagtgta
1261	tcctccaacg tgacctgctt cgctatccac agcagtagg taatcgatat caatagctga
1321	tgaatccggt tcttctattg tttcaatgta tccatcaaca tatccacctc caccatctat
1381	agctgtgtgg taatcaatgt caagagtga tgaatcatat tcctcttcaa cagtagttac
1441	taaattctta tcatattgac ctgtaagagt ttctttaatt gtatcttctt tatattcaaa
1501	tttattattt tgaataatcg gaccattttt ctcatctccg ttcgctttat tactgtataa
1561	aactaaacca ttatcccaag ttaaggata tcctctatca taataatact tataaagttg
1621	ctctggatgt cctaccattt gtgttctaaa atcaacttca tcagtacctt taaatactc
1681	tccatcatag tgaacaacat aagttttatc tagattttct atattcaatg aatagcttc
1741	attattttgt aaattcaaat tccactcat attactgtg acttctttaa atttagaagt
1801	atctgtcgtg tttgcatata cactcttcgc tatgtcttca ttattacca agtattcaaa
1861	tatcctaact tttggttgat ttccattctg attactaact ttcattaag ttccagtaac
1921	agtcacactt gtcgttttac cattattagg ttttaataat gcaacatcgc aaatctatt

TABLE 2-continued

1981	atcgcgttta ttaaagtgtc caatcgatcc atttaaattg gcataataat tccaataacc
2041	atctttatat ttaacatcta attcctttga agtttggtct tcatttagtg ttgaagttat
2101	agtttgattt ccattagttt gtacagtttt aggatcaata aataaattaa tttctagttc
2161	agccgttaca tcaaccttat cttcaatatic atttgaaat gtatatctaa tctttccacc
2221	ttctaaaact tcacctgtgc ccattacgac tgaaccattt ttaattttctg gtacttttct
2281	agcagttgat acgccatgcg tatttacatt atttgataaa gtaaaagcaaa agtagtcacc
2341	ttgatgtaaa ccattctcaa atttcaactt atattttagt accgctcggt gtccctgcatg
2401	aggttctact ttatttgat tgttatgccc ctcaatagaa ccaatttcta ctgtaacttt
2461	acttgttaca tctgtaccgc tttccacttt cgcgttacta gcttccttag ctcccgctac
2521	atctgctgat cttgtcacac gtggcttact ttctgatgcc gcttcctggt gtgccacttc
2581	aacttggttt tctgcgactt gattttgtgt agccttttta gggtgtaaat ctacttgctt
2641	ttgatctccg ctattgtctt gagattgtgt tgtttcctta acttgagggtt tegettcttc

TABLE 2-continued

2701	cttaactacc tcttctttaa ctgtttctat atttgctggt tgtgcagttt gtggtgcttg
2761	tactgctttt ggtgcttctt cagttgttac ttgtgttgccg tttgacgggt gttctgttac
2821	tgttgcgta tatgattgag tttcttctat atgattaacg ttagttgcag ttgtttgtgt
2881	ttcacttggt ttattatcag tagctgaatt cccattttct tctactgtag ttgtcttttg
2941	ttctgatget gcagcttctt tgtcttgtcc cat

[0036] Previously, it has been shown that *S. aureus* proteins expressed on the surface of Lactococci were entirely accessible to trypsin digestion for analysis by LC-MS (liquid chromatography coupled with mass spectrometry) (Ythier et al., 2012). In contrast, this was not the case in *S. aureus*, where trypsin had a limited access to the surface proteins, indicating that part of their structure was embedded in the wall and inaccessible to digestion, and thus to recognition by antibodies as well. The trypsin shaving protocol is a classical protocol described in Ythier et al. and in the Example parts.

[0037] The trypsin digestion of adhesins such as, for example, ClfA expressed on the surface of lactococci and/or FnPBA expressed on the surface of Lactococci, releases a digestion profile which is different from the digestion profile of the same adhesins expressed on the surface of *S. aureus*.

[0038] For example, the digestion profile of ClfA expressed on the surface of lactococci reveals 10 peptides versus 9 for ClfA expressed on the surface of *S. aureus*. Among these 10 peptides, 3 peptides (SEQ IDs 5, 6, and 12) were specific of Lactococci.

TABLE 3

Peptides released after trypsin digestion of ClfA expressed on		
SEQ ID/Position (aa)	<i>S. aureus</i>	<i>L. lactis</i>
SEQ ID No 5 40-55		SENSVTQSDSASNESK
SEQ ID No 6 56-67		SNDSSSVSAAPK
SEQ ID No 7 200-212	DVVNQAVNTSAPR	DVVNQAVNTSAPR
SEQ ID No 8 259-271	LNYGFSVPNSAVK	LNYGFSVPNSAVK
SEQ ID No 9 282-293	ELNLNGVTSTAK	
SEQ ID No 10 331-345	ATLTMPAYIDPENVK	ATLTMPAYIDPENVK
SEQ ID No 11 347-363	TGNVTLATGIGSTTANK	TGNVTLATGIGSTTANK
SEQ ID No 12 375-381		FYNLSIK
SEQ ID No 13 396-434	QTIYVNPsgDNVIAPVLTGnLkPNTD	QTIYVNPsgDNVIAPVLTGnLkPNTDSN ALIDQNTSISNALIDQNTSI

K

K

TABLE 3-continued

Peptides released after trypsin digestion of ClfA expressed on		
SEQ ID/Position (aa)	<i>S. aureus</i>	<i>L. lactis</i>
SEQ ID No 14 438-473	VDNAADLSESYFVNPENFEDVTNSVN ITFPNPQYK	
SEQ ID No 15 474-500	VEFNTPDDQITTPYIVVVNGHIDPNSK	VEFNTPDDQITTPYIVVVNGHIDPNSK

[0039] As another example, the digestion profile of FnbpA expressed on the surface of Lactococci reveals 10 peptides versus 22 for FnbpA expressed on the surface of *S. aureus*. Among these peptides, 2 peptides (SEQ IDs 38 and 39) were specific of lactococci.

TABLE 4

Peptides released after trypsin digestion of FnbpA expressed on		
Position (aa)	<i>S. aureus</i>	<i>L. lactis</i>
SEQ ID No 16 57-97	TSETQTTATNVNHIETQSYNATVTEQPSNATQ VTTEEAPK	TSETQTTATNVNHIETQSYNAT VTEQPSNATQVTTEEAPK
SEQ ID No 17 98-114	AVQAPQTAQPANIETVK	AVQAPQTAQPANIETVK
SEQ ID No 18 98-119	AVQAPQTAQPANIETVKEEVVK	
SEQ ID No 19 115-127	EEVVKEEAKPQVK	
SEQ ID No 20 148-165	KATQNQVAETQVEVAQPR	KATQNQVAETQVEVAQPR
SEQ ID No 21 149-165	ATQNQVAETQVEVAQPR	ATQNQVAETQVEVAQPR
SEQ ID No 22 201-216	VTVEIGSIEGHNTNK	VTVEIGSIEGHNTNK
SEQ ID No 23 233-260	FENGLHQGDYDFDFTLSNNVNTHGVSSTAR	
SEQ ID No 24 267-282	NGSWMATGEVLEGGK	
SEQ ID No 25 285-294	YTFTNDIEDK	
SEQ ID No 26 311-331	TVQTNGNQITITSLNNEEQTSK	TVQTNGNQITITSLNNEEQTSK
SEQ ID No 27 337-357	YKDGIGNYYANLNGSIETFNK	
SEQ ID No 28 339-357	DGIGNYYANLNGSIETFNK	
SEQ ID No 29 362-374	FSHVAFIKPNNGK	
SEQ ID No 30 375-386	TTSVTVTGTLMK	
SEQ ID No 31 399-411	IFEYLGNNEDIAK	IFEYLGNNEDIAK

TABLE 5-continued

DSDSASDSDSGSDSDSSSDSESDSNDSSESGSNMNVPPNSPKNGTN
 ASNKNEAKDSKEPLPDTGSEDEANTSLIWGLLASIGSLLLFRKKNKD
 KK

[0046] More preferably also, the sequence of the invention consists in a whole functional amino acid sequence of FnPBA (amino acid 1 to 990, table 6).

TABLE 6

	SEQ ID No 4
MGQDKAAASEQKTTTVEENGNSATDNKTSETQTATNVNHIETQSY	
NATVTEQPSNATQVTTEEAPKAVQAPQTAQPANIETVKEEVVKEEAKP	
QVKETTQSQDNSGDQRQVDLTPKKATQNVQVAETQVEVAQPRTASESKP	
RVTRSADVAEAKESNAKVEGTGTDVISKVTVIEIGSIEGHNNNTKVEPH	
AGQRAVLKYKLFENGLHQGDYDFDTLSNNVNTHGVSSTARVKVPEIKNG	
SVVMATGEVLEGGKIRYFTTNDIEDKVDVTAELEINLFDPKTVQTNG	
NQTIITSTLNEEQTSKELDVKYKDIGNYANLNGSIETFNKANRFSH	
VAFIKPNNKGTTSVTVTGTLMKGSNQNQPKVRIFEYLGNNEDIAKS	
VYANTTDTSKFKEVTSNMSGNLNLQNGNSYSLNIENLDKTYVVHYDGE	
YLNGTDEVDPRQMGVHPEQLYKYYDRGYTLTWDNGLVLYSNKANGN	
EKNGP I IQNNKFEYKEDTIKETLTGOYDKNLVTTVEEYDSSTLIDY	
HTAIDGGGGYVDGYIETIETDSSAIDIDYHTAVDSEAGHVGGYTES	
EESNPIDFEESTHENSKHHADVVEEEDTNPGGGQVTTESNLVEFDDE	
STKGIVTGAVDHHTVEDTKEYTTESNLIELVDELPEEHGQAQGPVEE	
ITKNHHISHSGLGTENGHGNVDVIEEIEENSHVDIKSELGYEGGQNS	
GNQSFEEDEEDKPKYEQGGNIVDIDFDSVPQIHGQNKGNQSFEEDE	
KDKPKYEHGGNIIDIDFDSVPHIHGFNKHTEIEEDTNKDKPSYQFGG	
HNSVDFEEDTLFPKVSQNEGQQTIEEDTTPPIVPPPTPEVPSEPET	
PTPPTPEVPSEPETPTPPTPEVPSEPETPTPPTPEVPAEPGKVPVPAK	
EPPKPKSPVEQGVVTPVIEINEKVKAVPTKPKQSKKSELPEVGGE	
ESTNKGMLFGGLFSILGLALLRRNKKNHKA	

[0047] Surprisingly, the Applicants of the present invention have successfully prevented *S. aureus* experimental endocarditis in rats vaccinated with UV-killed *L. lactis* expressing heterologously the staphylococcal adhesin ClfA, or ClfA/FnBPA (CD). The success of vaccination is due to the method of antigen delivery, i.e. a whole functional *S. aureus* surface adhesin on a genuine bacterial surface, rather than only restricted peptides of the protein. This mode of delivery largely increases the repertoire of anti-staphylococcal antibodies generated by the host, and thus increases the efficacy of protection.

[0048] The cell wall anchored fibrinogen-binding protein ClfA has been the major target of vaccine candidates, due to its ability to binds to the γ -chain of the fibrinogen. Indeed, previous studies have shown that ClfA is essential for the

development of IE due to the pivotal role of fibrinogen-binding (Moreillon et al., 1995; Yok-ai Que et al., 2005). However, numerous attempts to develop vaccines against a variety of *S. aureus* virulence factors have been attempted with various successes in animal models, but have as yet not achieved sustainable efficacy in human clinical trials.

[0049] The reasons for these failures are not entirely clarified, but there are at least two parameters of the *S. aureus* camouflage system that might have been underestimated until now. First, the fact that the bacterium may vary the way it exposes its antigenic structure on the surface, and second the fact that different strains may undergo antigenic variations. As a result, vaccination against one particular structure that is valid against one strain may not be efficacious against another strain. Therefore, a blocking or opsonizing antibody that is active in certain circumstances may become inactive in other settings.

[0050] Staphylococcal adhesins of the LPXTG-protein family are equipped with a spacer domain between the cell wall anchor and the outermost binding domain. This spacer is important to expose the binding domain on top of the plethora of other constituents of the staphylococcal envelope, in order to bind to the target host tissue. Thus, depending on the length of this spacer (which may vary between strains) and the size of other surface components (for instance polysaccharides and protein A), the binding domain of the adhesin may become embedded in other surface structures and hidden to the immune system (Scarpa et al., 2010). Indeed, it was shown that *S. aureus* exposed differently the fibrinogen-binding protein domain “A” of ClfA (Mcdevitt et al., 1994), as well as the surface capsule, at various stages of in vivo infection (Risley et al., 2007).

[0051] Another example comes from the length of anti-phagocytic protein A, which binds antibodies by their Fc fragment. It was recently shown that a longer spacer sequence allowed protein A to better prevent binding of antibodies to various antigenic structures presented on the staphylococcal surface (Scarpa et al., 2010). Moreover, the polymorphism of the protein A gene may affect the efficacy of vaccines in a strain-dependent manner. Indeed, the length of the spacer is determined by series of repeats that are known to vary between different staphylococcal strains, and thus are currently used as phylogenetic markers (Kuhn et al., 2007).

[0052] Eventually, *S. aureus* produces a plethora of toxins and superantigens that may interfere with the immune response, and thus help the organism to circumvent existing host immune strategies. Vaccination against such structures were also recently attempted (Broughan et al., 2011).

[0053] The importance of antigenic variation in *S. aureus* is less clear. Indeed, only two major capsular types (types 5 and 8) are implicated in infection in human, and surface proteins and toxins are relatively well conserved (Shinefield et al., 2002). However, many gram-positive pathogens, including group A streptococci and *Streptococcus pneumoniae*, undergo wide genetic variability at the level of the anti-phagocytic M protein and the polysaccharidic capsule, respectively. Taken together, the current consensus for anti-*S. aureus* vaccines is that they should comprise several antigens in order to be effective, and that single antigen-based vaccines are bound to fail (Broughan et al., 2011). However, the ideal antigen mixture to be used has yet to be elucidated.

[0054] Examples showed hereafter that *S. aureus* adhesins could be expressed functionally in *L. Lactis* in vitro, and could promote experimental endocarditis by the recombinant

Lactococci in vivo. When conjugated with the proteomic dissection of the surface proteome of *S. aureus*, it appeared that the *S. aureus* proteins expressed on the surface of Lactococci were entirely accessible to trypsin digestion for analysis by LC-MS (liquid chromatography coupled with mass spectrometry) (Ythier et al., 2012). In contrast, this was not the case in *S. aureus*, where trypsin had a limited access to the surface proteins, indicating that part of their structure was embedded in the wall and inaccessible to digestion, and thus to recognition by antibodies as well. Since all these domains were variously exposed on the *S. aureus* surface, it was conceivable that vaccination against only one binding-domain, which might become hidden in certain circumstances, might be less effective than vaccination against the whole protein presented in a functional conformation on the surface of lactococci.

[0055] As shown in the examples, immunizing series of rats with UV-killed lactococci expressing *S. aureus* ClfA, or ClfA/FnBPA (CD) protected animals from subsequent experimental endocarditis due to *S. aureus* induced by low-grade bacteremia ($P < 0.05$ when compared to the several control groups). Remarkably, when the same vaccination schedule was tested in animals where the experimental endocarditis due to *S. aureus* was induced by high-grade bacteremia (traditional bolus infection) the infection rates increased (2/15 vs 6/6) and the protective effect was lost.

[0056] Attempts to vaccinate against severe infections in animal models are usually biased by the fact that most protocols administer very large bacterial inocula, in order to ensure that all untreated control animals will become infected. However, such large inocula—often in the range of 10 s of million bacteria injected intravenously—are incommensurably larger than inoculum sized expected during “natural” infection in human. Thus, it is possible that such large inocula may overwhelm the immune system, and falsely underestimate the efficacy of preventive or therapeutic strategies that would otherwise be efficacious. The Applicants recently showed that this was realistic in the model of experimental endocarditis, and that challenging animals with continuous low-grade bacteremia was as infectious that transient high-grade inoculation, but represented much more the reality of the disease in human (Veloso et al., 2011). Importantly, the good results of the vaccination strategy reported above were achieved with this very realistic model. Indeed, vaccination was less effective against the standard high-grade bacteremia model, an observation that could also explain failures with other types of potential anti-*S. aureus* vaccines tested in animals in the past, and abandoned.

[0057] The present invention also concerns a vaccine. Preferably, the vaccine comprises an immunogenic composition of the invention in an immunologically acceptable carrier or diluent. Preferably said vaccine is for treating and/or preventing infections and diseases caused by *S. aureus*. Examples of diseases caused by *S. aureus* are folliculitis, furunculosis, erysipelas, deep-seated abscesses, osteomyelitis, pneumonia, sepsis, and infective endocarditis (IE).

[0058] The vaccine of the invention may also contain several antibodies or antibody fragments, e.g. two or three antibodies or antibody fragments that recognize(s) at least one folded sequence of a *S. aureus* adhesin expressed on the cell surface of an inactivated recombinant non-pathogenic bacterium of the invention.

[0059] The immunologically acceptable carrier is usually selected from the group comprising polysaccharide materials

forming hydrogels, bacterial ghosts and vesicular carriers. Preferably, the vesicular carriers are selected from the group comprising liposomes, niosomes, transfersomes, and ethosomes, and others known in the art.

[0060] Hydrogels envisioned as immunologically acceptable carrier as known in the art and are particularly adapted for mucosal, topical, oral or injectable delivery.

[0061] The vaccines of the invention may further comprise one or more adjuvant. Adjuvants can include, but are not limited to, MPL+TDM+CWS (SIGMA), MF59 (an oil-in-water emulsion that includes 5% squalene, 0.5% sorbitan monoleate and 0.5% sorbitan trioleate Chiron), Heat-labile toxin (HLT), CRMig (nontoxic genetic mutant of diphtheria toxin), Squalene (IDEC PHARMACEUTICALS CORP.), Ovalbumin (SIGMA), Quil A (SARGEANT, INC.), Aluminum phosphate gel (SUPERFOS BIOSECTOR), Cholera holotoxin (CT LIST BIOLOGICAL LAB.), Cholera toxin B subunit (CTB), Cholera toxin A subunit-Protein A D-fragment fusion protein, Muramyl dipeptide (MDP), Adjuvera (polyphosphazene, VIRUS RESEARCH INSTITUTE), Montanide ISA 720, SPT (an emulsion of 5% squalene, 0.2% Tween 80, 1.25% Pluronic L121 with phosphate-buffered saline pH 7.4), Avridine (M6 PHARMACEUTICALS), Bay R1005 (BAYER), Calcitrol (SIGMA), Calcium phosphate gel (SARGEANT INC.), CRL 1005 (Block co-polymer P1205, VAXCEL CORP.), DHEA (MERCK), DMPC (GENZYME PHARMACEUTICALS and FINE CHEMICALS), DMPG (GENZYME PHARMACEUTICALS and FINE CHEMICALS), Gamma Inulin, Gerbu Adjuvant (CC BIOTECH CORP.), GM-CSF, (IMMUNE CORP.), GMDP (PEPTECH LIMITED), Imiquimod (3M PHARMACEUTICALS), ImmTher (ENDOREX CORPORATION), ISCOMTM (ISCOTEC AB), Iscoprep 7.0.3 TM (ISCOTEC AB), Loxoribine, LT-Oral Adjuvant (*E. coli* labile enterotoxin, prototoxin, BERNA PRODUCTS CORP.), MTP-PE (CIBA-GEIGY LTD), Murametide, (VACSYN S. A.), Murapalmitine (VACSYN S. A.), Pluronic L121 (IDEC PHARMACEUTICALS CORP.), PMMA (INSTITUT FÜR PHARMAZEUTISCHE TECHNOLOGIE), SAF-1 (SYNTEX ADJUVANT FORMULATION CHIRON), Stearyl tyrosine (BIOCHEM THERAPEUTIC INC.), Theramidea (IMMUNO THERAPEUTICS INC.), Threonyl-MDP (CHIRON), FREUNDS adjuvant (complete or incomplete), aluminum hydroxide, dimethyldioctadecyl-ammonium bromide, Adjuvax (ALPHA-BETA TECHNOLOGY), Inject Alum (PIERCE), Monophosphoryl Lipid A (RIBI IMMUNOCHEM RESEARCH), MPL+TDM (RIBI IMMUNOCHEM RESEARCH), Titermax (CYTRX), QS21, t Ribi Adjuvant System, TiterMaxGold, QS21, Adjuver, Calcitrol, CTB, LT (*E. coli* toxin), LPS (lipopolysaccharide), Avridine, the CpG sequences (Singh et al., 1999 Singh, M. and Hagem, D., Nature Biotechnology 1999 17: 1075-81) toxins, toxoids, glycoproteins, lipids, glycolipids, bacterial cell walls, subunits (bacterial or viral), carbohydrate moieties (mono-, di-, tri-, tetra-, oligo- and polysaccharide), or saponins. Combinations of various adjuvants may be used with the antigen to prepare the immunogen formulations. Adjuvants administered parentally or for the induction of mucosal immunity may also be used.

[0062] The present invention further contemplates isolated and/or purified antibody, antibody fragment or derivative thereof able to bind to the at least one folded sequence of a *S. aureus* adhesion of the invention, or to the sequence having 80% or more sequence identity to said folded sequence of a *S.*

aureus adhesion of the invention, expressed on the cell surface of an inactivated recombinant non-pathogenic bacterium.

[0063] As used herein, an “antibody” is a protein molecule that reacts with a specific antigenic determinant or epitope and belongs to one or five distinct classes based on structural properties: IgA, IgD, IgE, IgG and IgM. The antibody may be a polyclonal (e.g. a polyclonal serum) or a monoclonal antibody, including but not limited to fully assembled antibody, single chain antibody, antibody fragment, and chimeric antibody, humanized antibody as long as these molecules are still biologically active and still bind to one folded sequence of a *S. aureus* adhesin of the invention. Preferably the antibody is a monoclonal antibody. Preferably also the monoclonal antibody will be selected from the group comprising the IgG1, IgG2, IgG2a, IgG2b, IgG3 and IgG4 or a combination thereof. Most preferably, the monoclonal antibody is selected from the group comprising the IgG1, IgG2, IgG2a, and IgG2b, or a combination thereof.

[0064] A typical antibody is composed of two immunoglobulin (Ig) heavy chains and two Ig light chains. Several different types of heavy chain exist that define the class or isotype of an antibody. These heavy chain types vary between different animals. All heavy chains contain a series of immunoglobulin domains, usually with one variable (VH) domain that is important for binding antigen and several constant (CH) domains. Each light chain is composed of two tandem immunoglobulin domains: one constant (CL) domain and one variable domain (VL) that is important for antigen binding.

[0065] The term “isolated”, when used as a modifier of an antibody of the invention means that the antibody is made by the hand of man or is separated, completely or at least in part, from their naturally occurring in vivo environment. Generally, isolated antibodies are substantially free of one or more materials with which they normally associate with in nature, for example, one or more protein. The term “isolated” does not exclude alternative physical forms of the antibodies, such as multimers/oligomers, modifications (e.g., phosphorylation, glycosylation, lipidation) or derivatized forms, or forms expressed in host cells produced by the hand of man

[0066] An “isolated” antibody can also be “substantially pure” or “purified” when free of most or all of the materials with which it typically associates with in nature. Thus, an isolated antibody that also is substantially pure or purified does not include polypeptides or polynucleotides present among millions of other sequences, such as antibodies of an antibody library or nucleic acids in a genomic or cDNA library.

[0067] Antibodies used in the present invention are not limited to whole antibody molecules and may be antibody fragments or derivatives as long as they are able to bind to the at least one folded sequence of a *S. aureus* adhesin expressed on the cell surface of an inactivated recombinant non-pathogenic bacterium and that they specifically recognize said folded sequence of a *S. aureus* adhesin.

[0068] Examples of isolated and/or purified antibody fragment or derivative thereof are selected amongst the group comprising a Fab-fragment, a F(ab2)^γ-fragment, a single-chain antibody, a chimeric antibody, a CDR-grafted antibody, a bivalent antibody-construct, a humanized antibody, a synthetic antibody, a chemically modified derivative thereof, a multispecific antibody, a diabody, a scFv-fragment; a dsFv-fragment, a labeled antibody, or another type of recombinant antibody. Specifically, an antibody fragment is synthesized

by treating the antibody with an enzyme such as papain or pepsin, or genes encoding these antibody fragments are constructed, and expressed by appropriate host cells as known to the skilled artisan.

[0069] Yet another concern of the present invention is to provide an expression vector comprising at least one isolated and/or purified nucleic acid sequence encoding for at least one folded sequence of a *S. aureus* adhesin, or a sequence having 80% or more sequence identity to said folded sequence of a *S. aureus* adhesin. Preferably the nucleic acid molecule sequence encoding a peptide of the invention is DNA.

[0070] As used herein, “vector”, “plasmid” and “expression vector” are used interchangeably, as the plasmid is the most commonly used vector form.

[0071] The vector may further comprise a promoter operably linked to the sequence encoding a folded sequence of a *S. aureus* adhesin. This means that the linked isolated and purified DNA sequence encoding the peptide of the present invention is under control of a suitable regulatory sequence which allows expression, i.e. transcription and translation of the inserted isolated and purified DNA sequence.

[0072] As used herein, the term “promoter” designates any additional regulatory sequences as known in the art e.g. a promoter and/or an enhancer, polyadenylation sites and splice junctions usually employed for the expression of the polypeptide or may include additionally one or more separate targeting sequences and may optionally encode a selectable marker. Promoters which can be used provided that such promoters are compatible with the host cell are e.g. promoters obtained from the genomes of viruses such as polyoma virus, adenovirus (such as Adenovirus 2), papilloma virus (such as bovine papilloma virus), avian sarcoma virus, cytomegalovirus (such as murine or human cytomegalovirus immediate early promoter), a retrovirus, hepatitis-B virus, and Simian Virus 40 (such as SV 40 early and late promoters) or promoters obtained from heterologous mammalian promoters, such as the actin promoter or an immunoglobulin promoter or heat shock promoters.

[0073] Enhancers, which can be used, are e.g. enhancer sequences known from mammalian genes (globin, elastase, albumin, alpha-fetoprotein, and insulin) or enhancer from a eukaryotic cell virus. e.g. the SV40 enhancer, the cytomegalovirus early promoter enhancer, the polyoma, and adenovirus enhancers.

[0074] Useful expression vectors, for example, may consist of segments of chromosomal, non-chromosomal and synthetic DNA sequences. Suitable vectors include derivatives of SV40 and known bacterial plasmids, phage DNAs, yeast plasmids such as the 2μ plasmid or derivatives thereof; vectors useful in eukaryotic cells, such as vectors useful in insect or mammalian cells; vectors derived from combinations of plasmids and phage DNAs, such as plasmids that have been modified to employ phage DNA or other expression control sequences; and the like. Most preferably the expression vector is a lactococcal plasmid. More preferably, the lactococcal plasmid is pOri23.

[0075] Also provided is a method for treating and/or preventing infections and diseases caused by *S. aureus*, in a subject in need thereof, comprising administering a pharmaceutically effective amount of an immunogenic composition according to the invention.

[0076] Usually, infections and diseases caused by *S. aureus* are selected among the non limiting group comprising IE,

intravascular and intravascular device infections, blood-stream infections, deep-seated abscesses, osteomyelitis, infection of prosthetic materials, and skin and soft tissue infections. Examples of diseases caused by *S. aureus* are folliculitis, furunculosis, erysipelas, deep-seated abscesses, osteomyelitis, pneumonia, sepsis, and infective endocarditis (IE).

[0077] Also envisioned is a method of inducing active immunity against a *S. aureus* infection in a subject in need thereof, comprising administering to said subject in need thereof i) an immunogenic composition comprising an inactivated recombinant non-pathogenic bacterium, or a part thereof, expressing on its cell surface, at least one folded sequence of a *S. aureus* adhesin or ii) a vaccine of the invention.

[0078] Also encompassed in the present invention is a method of inducing passive immunity against a *S. aureus* infection in a subject in need thereof, comprising administering to said subject in need thereof an isolated and/or purified antibody, antibody fragment or derivative thereof of the invention.

EXAMPLES

Example 1

Materials and Methods

1.1 Plasmid Constructs and Bacterial Strains

[0079] The immunization protocol in this study was done using the recombinant strain of the non-pathogenic *L. lactis* subsp. *cremoris* 1363 expressing individual *S. aureus* ClfA, described elsewhere (Piroth et al., 2008; Que et al., 2000; Que et al., 2001) or *S. aureus* ClfA/FnBPA(CD). The *S. aureus* Newman ClfA gene was inserted in the lactococcal plasmid pOri23 together with an erythromycin resistance determinant as described by Que et al (Que et al., 2000). In the strain *L. lactis* ClfA/FnBPA(CD) *S. aureus* Newman ClfA gene was expressed as described above, and the CD domain of the *S. aureus* 8325 FnBPA gene was inserted in lactococcal plasmid pOri23 together with an kanamycin resistance determinant (Elonora Widmer MD/PhD thesis). The *L. lactis* pIL253, containing the lactococcal plasmid pOri23 expressing only the erythromycin resistance determinant, and expressing no pathogenic factors, was used as the control mutant strain. All lactococci were grown at 30° C. without shaking in M17 medium (Oxoid) or on M17 agar plates supplemented with 0.5% glucose and 5 µg/ml erythromycin (plus kanamycin when appropriate).

[0080] The well-described *S. aureus* strain Newman (i.e. methicillin-susceptible *S. aureus*) *S. aureus* strain P8 (i.e. methicillin-resistant *S. aureus*) (Entenza et al., 2001) was used in the animal model of endocarditis. The *S. aureus* bacterial isolates was grown at 37° C. in tryptic soy broth (Difco).

[0081] All the bacterial stocks were kept at -80° C. in liquid medium supplemented with 20% (vol/vol) of glycerol.

1.2 Immunization Protocol

[0082] (i) Animals. Four to six-week females (100 g of weight) Wistar Han rats were purchased from Charles River, France. The rats were supplied with water and food ad libitum, and randomly allocated to 6 treatment groups as follows: (a) Control, (b) Freund's adjuvant (Sigma), (c) *L. lactis*

pIL253 and (d) *L. lactis* ClfA, *L. lactis* FnBPA or *L. lactis* ClfA/FnBPA(CD). All animal experiments were carried out according to Swiss regulations (authorization 879.8).

(ii) Preparation of bacterial vaccine. The inactivated *L. lactis* vaccine was prepared as follows. *L. lactis* strains carrying either the empty plasmid pOri23, or the plasmid expressing ClfA or ClfA/FnBPA(CD) were cultured overnight at 30° C. without shaking in M17 medium (Oxoid), harvested by centrifugation, resuspended in sterilized PBS, and adjusted to 1×10⁸ CFU/ml. The bacteria were then inactivated during 60 min under U.V. Then, the bacteria were emulsified at a 1:1 ratio in Freund's adjuvant. The first immunization was done with Freund's complete adjuvant, and the subsequent with Freund's incomplete adjuvant. The control group was immunized with PBS, and the Freund's adjuvant group with the adjuvant emulsified at a 1:1 ratio in PBS.

(iii) Vaccination schedule. The rats were immunized at 2-week intervals (days 0, 14 and 28). Three hundred microlitres of the preparations were injected intra-peritoneal to the respective groups. Blood samples were collected on days 7, 21 and 35; and the sera were harvested and stored at -80° C. to posterior in vitro analysis (Gong et al., 2010).

1.3. Animal Model of Endocarditis

[0083] Catheter-induced aortic vegetations were produced according to the method of Heraïef et al (Héraïef et al., 1982) Insertion of an intravenous (i.v.) line in the jugular vein and connection to a programmable infusion pump (Pump 44; Harvard Apparatus, Inc., South Natick, Mass.) to deliver the inocula was performed as described (Fluckiger et al., 1994; Pea et al., 2011) on the day 40 of the vaccination schedule. Bacterial inocula were prepared from overnight cultures. Microorganisms were recovered by centrifugation, washed and adjusted to the desired inoculum size in saline. The inoculum size was confirmed by colony counts on blood agar plates. Animals were inoculated 24 h after catheterization, via the infusion pump, with 1 ml of 10⁴ CFU (*S. aureus* Newman) or 10⁶ CFU (*S. aureus* P8) progressively delivered at a pace of 0.0017 ml/min over 10 h in order to produce a low-grade of bacteremia (Veloso et al. 2011).

[0084] The traditional i.v. bolus inoculation (10⁴ CFU/ml) provoking transient high-grade bacteremia was also performed. Rats were sacrificed 24 h after the end of inoculation. Quantitative valve cultures were performed as previously described (Fluckiger et al., 1994) This method permitted the detection of 2 log 10 CFU/g of vegetation.

1.4. Statistical Analysis

[0085] Statistical analyses were performed using GraphPad software (GraphPad Software, Inc., USA). The rates of valve infections of the various groups were compared by the x2 test. P<0.05 was considered to be statistically significant.

1.5. Trypsin Shaving Protocol (Ythier et al. 2012)

[0086] In brief, bacteria were grown in 300 ml liquid cultures in the different media described above. At various times of the logarithmic or stationary growth phases, samples (between 10 and 100 ml depending on the cell density) were removed, immediately chilled at 4° C., and harvested by centrifugation. Pellets were washed three times with ice-cold phosphate-buffered saline (PBS) and finally re-suspended in 1 ml of the same buffer. To allow semi-quantitative comparisons between the proteomes of different samples, cell con-

centrations were adjusted to 1×10^9 bacteria/ml in all samples. Cell counts were validated by optical microscopy (Neubauer cell) and viable colony counts on nutrient agar. There were $<0.5 \log_{10}$ differences between the Neubauer cell and viable counts, indicating that the large majority of cells were alive. Samples were then shaved for 1 h with $1 \mu\text{g/ml}$ (final concentration) of trypsin (Promega, Madison, Wis.) at 37°C ., after which they were chilled at 4°C . and bacterial cells removed by centrifugation for 10 min at 4000 rpm and 4°C . Supernatants containing trypsin-shaved peptides were filtered ($0.22 \mu\text{m}$) and freeze-dried until further use.

Example 2

Results

[0087] The efficacy of the immunization with *L. lactis* ClfA or *L. lactis* ClfA/FnBPA(CD) against the *S. aureus* induced experimental endocarditis due to low-grade bacteremia was compared to the different control groups. The results of the infectivity rate are shown in FIG. 1.

[0088] In FIG. 1A the proportion of infection in the group immunized with *L. lactis* ClfA (6/22; 27.2%) was significantly lower than in the control groups PBS (9/12; 75%), Adj. (5/8; 62.5%) and *L. lactis* pIL253 (6/8; 75%) (χ^2 test; $P < 0.05$), in the case of low-grade bacteremia experimental endocarditis induced by *S. aureus* Newman. These results confirm the protective effect of the immunization using the non-pathogenic *L. lactis* expressing heterologously the staphylococcal adhesin ClfA. In contrast, when the same immunization schedule was used in animals exposed to high-grade bacteremia experimental endocarditis, the protective effect was not observed.

[0089] In FIG. 1B the proportion of infection in the group immunized with *L. lactis* ClfA/FnBPA(CD) (14/22; 63.6%) was significantly lower ($P < 0.05$) than in the control groups PBS (20/21; 95.2%), Adj. (20/20; 100%) but not for *L. lactis* pIL253 (11/15; 73.3%; χ^2 test; $P = 0.53$). These results demonstrate a diminution in the infection after the effect of the immunization using the non-pathogenic *L. lactis* expressing heterologously the staphylococcal adhesins ClfA/FnBPA(CD).

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 Ser Val Val Met Ala Thr Gly Glu Val Leu Glu Gly Gly Lys Ile Arg
 245 250 255
 Tyr Thr Phe Thr Asn Asp Ile Glu Asp Lys Val Asp Val Thr Ala Glu
 260 265 270
 Leu Glu Ile Asn Leu Phe Ile Asp Pro Lys Thr Val Gln Thr Asn Gly
 275 280 285
 Asn Gln Thr Ile Thr Ser Thr Leu Asn Glu Glu Gln Thr Ser Lys Glu
 290 295 300
 Leu Asp Val Lys Tyr Lys Asp Gly Ile Gly Asn Tyr Tyr Ala Asn Leu
 305 310 315 320
 Asn Gly Ser Ile Glu Thr Phe Asn Lys Ala Asn Asn Arg Phe Ser His
 325 330 335
 Val Ala Phe Ile Lys Pro Asn Asn Gly Lys Thr Thr Ser Val Thr Val
 340 345 350
 Thr Gly Thr Leu Met Lys Gly Ser Asn Gln Asn Gly Asn Gln Pro Lys
 355 360 365
 Val Arg Ile Phe Glu Tyr Leu Gly Asn Asn Glu Asp Ile Ala Lys Ser
 370 375 380
 Val Tyr Ala Asn Thr Thr Asp Thr Ser Lys Phe Lys Glu Val Thr Ser
 385 390 395 400
 Asn Met Ser Gly Asn Leu Asn Leu Gln Asn Asn Gly Ser Tyr Ser Leu
 405 410 415
 Asn Ile Glu Asn Leu Asp Lys Thr Tyr Val Val His Tyr Asp Gly Glu
 420 425 430
 Tyr Leu Asn Gly Thr Asp Glu Val Asp Phe Arg Thr Gln Met Val Gly
 435 440 445
 His Pro Glu Gln Leu Tyr Lys Tyr Tyr Tyr Asp Arg Gly Tyr Thr Leu
 450 455 460

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Thr Trp Asp Asn Gly Leu Val Leu Tyr Ser Asn Lys Ala Asn Gly Asn
 465 470 475 480
 Glu Lys Asn Gly Pro Ile Ile Gln Asn Asn Lys Phe Glu Tyr Lys Glu
 485 490 495
 Asp Thr Ile Lys Glu Thr Leu Thr Gly Gln Tyr Asp Lys Asn Leu Val
 500 505 510
 Thr Thr Val Glu Glu Glu Tyr Asp Ser Ser Thr Leu Asp Ile Asp Tyr
 515 520 525
 His Thr Ala Ile Asp Gly Gly Gly Tyr Val Asp Gly Tyr Ile Glu
 530 535 540
 Thr Ile Glu Glu Thr Asp Ser Ser Ala Ile Asp Ile Asp Tyr His Thr
 545 550 555 560
 Ala Val Asp Ser Glu Ala Gly His Val Gly Gly Tyr Thr Glu Ser Ser
 565 570 575
 Glu Glu Ser Asn Pro Ile Asp Phe Glu Glu Ser Thr His Glu Asn Ser
 580 585 590
 Lys His His Ala Asp Val Val Glu Tyr Glu Glu Asp Thr Asn Pro Gly
 595 600 605
 Gly Gly Gln Val Thr Thr Glu Ser Asn Leu Val Glu Phe Asp Glu Glu
 610 615 620
 Ser Thr Lys Gly Ile Val Thr Gly Ala Val Ser Asp His Thr Thr Val
 625 630 635 640
 Glu Asp Thr Lys Glu Tyr Thr Thr Glu Ser Asn Leu Ile Glu Leu Val
 645 650 655
 Asp Glu Leu Pro Glu Glu His Gly Gln Ala Gln Gly Pro Val Glu Glu
 660 665 670
 Ile Thr Lys Asn Asn His His Ile Ser His Ser Gly Leu Gly Thr Glu
 675 680 685
 Asn Gly His Gly Asn Tyr Asp Val Ile Glu Glu Ile Glu Glu Asn Ser
 690 695 700
 His Val Asp Ile Lys Ser Glu Leu Gly Tyr Glu Gly Gly Gln Asn Ser
 705 710 715 720
 Gly Asn Gln Ser Phe Glu Glu Asp Thr Glu Glu Asp Lys Pro Lys Tyr
 725 730 735
 Glu Gln Gly Gly Asn Ile Val Asp Ile Asp Phe Asp Ser Val Pro Gln
 740 745 750
 Ile His Gly Gln Asn Lys Gly Asn Gln Ser Phe Glu Glu Asp Thr Glu
 755 760 765
 Lys Asp Lys Pro Lys Tyr Glu His Gly Gly Asn Ile Ile Asp Ile Asp
 770 775 780
 Phe Asp Ser Val Pro His Ile His Gly Phe Asn Lys His Thr Glu Ile
 785 790 795 800
 Ile Glu Glu Asp Thr Asn Lys Asp Lys Pro Ser Tyr Gln Phe Gly Gly
 805 810 815
 His Asn Ser Val Asp Phe Glu Glu Asp Thr Leu Pro Lys Val Ser Gly
 820 825 830
 Gln Asn Glu Gly Gln Gln Thr Ile Glu Glu Asp Thr Thr Pro Pro Ile
 835 840 845
 Val Pro Pro Thr Pro Pro Thr Pro Glu Val Pro Ser Glu Pro Glu Thr
 850 855 860
 Pro Thr Pro Pro Thr Pro Glu Val Pro Ser Glu Pro Glu Thr Pro Thr

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865		870		875		880									
Pro	Pro	Thr	Pro	Glu	Val	Pro	Ser	Glu	Pro	Glu	Thr	Pro	Thr	Pro	Pro
				885						890					895
Thr	Pro	Glu	Val	Pro	Ala	Glu	Pro	Gly	Lys	Pro	Val	Pro	Pro	Ala	Lys
			900						905					910	
Glu	Glu	Pro	Lys	Lys	Pro	Ser	Lys	Pro	Val	Glu	Gln	Gly	Lys	Val	Val
		915					920						925		
Thr	Pro	Val	Ile	Glu	Ile	Asn	Glu	Lys	Val	Lys	Ala	Val	Ala	Pro	Thr
	930					935					940				
Lys	Lys	Pro	Gln	Ser	Lys	Lys	Ser	Glu	Leu	Pro	Glu	Thr	Gly	Gly	Glu
945					950					955					960
Glu	Ser	Thr	Asn	Lys	Gly	Met	Leu	Phe	Gly	Gly	Leu	Phe	Ser	Ile	Leu
			965						970					975	
Gly	Leu	Ala	Leu	Leu	Arg	Arg	Asn	Lys	Lys	Asn	His	Lys	Ala		
			980					985					990		

<210> SEQ ID NO 5
 <211> LENGTH: 16
 <212> TYPE: PRT
 <213> ORGANISM: Lactobacillus lactis

<400> SEQUENCE: 5

Ser	Glu	Asn	Ser	Val	Thr	Gln	Ser	Asp	Ser	Ala	Ser	Asn	Glu	Ser	Lys
1			5						10					15	

<210> SEQ ID NO 6
 <211> LENGTH: 12
 <212> TYPE: PRT
 <213> ORGANISM: Lactobacillus lactis

<400> SEQUENCE: 6

Ser	Asn	Asp	Ser	Ser	Ser	Val	Ser	Ala	Ala	Pro	Lys
1			5						10		

<210> SEQ ID NO 7
 <211> LENGTH: 13
 <212> TYPE: PRT
 <213> ORGANISM: Lactobacillus lactis

<400> SEQUENCE: 7

Asp	Val	Val	Asn	Gln	Ala	Val	Asn	Thr	Ser	Ala	Pro	Arg
1			5						10			

<210> SEQ ID NO 8
 <211> LENGTH: 13
 <212> TYPE: PRT
 <213> ORGANISM: Lactobacillus lactis

<400> SEQUENCE: 8

Leu	Asn	Tyr	Gly	Phe	Ser	Val	Pro	Asn	Ser	Ala	Val	Lys
1			5						10			

<210> SEQ ID NO 9
 <211> LENGTH: 12
 <212> TYPE: PRT
 <213> ORGANISM: Staphylococcus aureus

<400> SEQUENCE: 9

Glu	Leu	Asn	Leu	Asn	Gly	Val	Thr	Ser	Thr	Ala	Lys
-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----

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1 5 10

<210> SEQ ID NO 10
 <211> LENGTH: 15
 <212> TYPE: PRT
 <213> ORGANISM: *Lactobacillus lactis*

<400> SEQUENCE: 10

Ala Thr Leu Thr Met Pro Ala Tyr Ile Asp Pro Glu Asn Val Lys
 1 5 10 15

<210> SEQ ID NO 11
 <211> LENGTH: 17
 <212> TYPE: PRT
 <213> ORGANISM: *Lactobacillus lactis*

<400> SEQUENCE: 11

Thr Gly Asn Val Thr Leu Ala Thr Gly Ile Gly Ser Thr Thr Ala Asn
 1 5 10 15

Lys

<210> SEQ ID NO 12
 <211> LENGTH: 7
 <212> TYPE: PRT
 <213> ORGANISM: *Lactobacillus lactis*

<400> SEQUENCE: 12

Phe Tyr Asn Leu Ser Ile Lys
 1 5

<210> SEQ ID NO 13
 <211> LENGTH: 38
 <212> TYPE: PRT
 <213> ORGANISM: *Lactobacillus lactis*

<400> SEQUENCE: 13

Gln Thr Ile Tyr Val Asn Pro Ser Gly Asp Asn Val Ile Ala Pro Val
 1 5 10 15

Leu Thr Gly Asn Leu Lys Pro Asn Thr Asp Ser Asn Ala Leu Ile Asp
 20 25 30

Gln Gln Asn Thr Ser Ile
 35

<210> SEQ ID NO 14
 <211> LENGTH: 36
 <212> TYPE: PRT
 <213> ORGANISM: *Staphylococcus aureus*

<400> SEQUENCE: 14

Val Asp Asn Ala Ala Asp Leu Ser Glu Ser Tyr Phe Val Asn Pro Glu
 1 5 10 15

Asn Phe Glu Asp Val Thr Asn Ser Val Asn Ile Thr Phe Pro Asn Pro
 20 25 30

Asn Gln Tyr Lys
 35

<210> SEQ ID NO 15
 <211> LENGTH: 27
 <212> TYPE: PRT
 <213> ORGANISM: *Lactobacillus lactis*

-continued

<400> SEQUENCE: 15

Val Glu Phe Asn Thr Pro Asp Asp Gln Ile Thr Thr Pro Tyr Ile Val
 1 5 10 15

Val Val Asn Gly His Ile Asp Pro Asn Ser Lys
 20 25

<210> SEQ ID NO 16

<211> LENGTH: 41

<212> TYPE: PRT

<213> ORGANISM: Lactobacillus lactis

<400> SEQUENCE: 16

Thr Ser Glu Thr Gln Thr Thr Ala Thr Asn Val Asn His Ile Glu Glu
 1 5 10 15

Thr Gln Ser Tyr Asn Ala Thr Val Thr Glu Gln Pro Ser Asn Ala Thr
 20 25 30

Gln Val Thr Thr Glu Glu Ala Pro Lys
 35 40

<210> SEQ ID NO 17

<211> LENGTH: 17

<212> TYPE: PRT

<213> ORGANISM: Lactobacillus lactis

<400> SEQUENCE: 17

Ala Val Gln Ala Pro Gln Thr Ala Gln Pro Ala Asn Ile Glu Thr Val
 1 5 10 15

Lys

<210> SEQ ID NO 18

<211> LENGTH: 22

<212> TYPE: PRT

<213> ORGANISM: Staphylococcus aureus

<400> SEQUENCE: 18

Ala Val Gln Ala Pro Gln Thr Ala Gln Pro Ala Asn Ile Glu Thr Val
 1 5 10 15

Lys Glu Glu Val Val Lys
 20

<210> SEQ ID NO 19

<211> LENGTH: 13

<212> TYPE: PRT

<213> ORGANISM: Staphylococcus aureus

<400> SEQUENCE: 19

Glu Glu Val Val Lys Glu Glu Ala Lys Pro Gln Val Lys
 1 5 10

<210> SEQ ID NO 20

<211> LENGTH: 18

<212> TYPE: PRT

<213> ORGANISM: Lactobacillus lactis

<400> SEQUENCE: 20

Lys Ala Thr Gln Asn Gln Val Ala Glu Thr Gln Val Glu Val Ala Gln
 1 5 10 15

Pro Arg

-continued

<210> SEQ ID NO 21
 <211> LENGTH: 17
 <212> TYPE: PRT
 <213> ORGANISM: Lactobacillus lactis

<400> SEQUENCE: 21

Ala Thr Gln Asn Gln Val Ala Glu Thr Gln Val Glu Val Ala Gln Pro
 1 5 10 15

Arg

<210> SEQ ID NO 22
 <211> LENGTH: 16
 <212> TYPE: PRT
 <213> ORGANISM: Lactobacillus lactis

<400> SEQUENCE: 22

Val Thr Val Glu Ile Gly Ser Ile Glu Gly His Asn Asn Thr Asn Lys
 1 5 10 15

<210> SEQ ID NO 23
 <211> LENGTH: 28
 <212> TYPE: PRT
 <213> ORGANISM: Staphylococcus aureus

<400> SEQUENCE: 23

Phe Glu Asn Gly Leu His Gln Gly Asp Tyr Phe Asp Phe Thr Leu Ser
 1 5 10 15

Asn Asn Val Asn Thr His Gly Val Ser Thr Ala Arg
 20 25

<210> SEQ ID NO 24
 <211> LENGTH: 16
 <212> TYPE: PRT
 <213> ORGANISM: Staphylococcus aureus

<400> SEQUENCE: 24

Asn Gly Ser Val Val Met Ala Thr Gly Glu Val Leu Glu Gly Gly Lys
 1 5 10 15

<210> SEQ ID NO 25
 <211> LENGTH: 10
 <212> TYPE: PRT
 <213> ORGANISM: Staphylococcus aureus

<400> SEQUENCE: 25

Tyr Thr Phe Thr Asn Asp Ile Glu Asp Lys
 1 5 10

<210> SEQ ID NO 26
 <211> LENGTH: 21
 <212> TYPE: PRT
 <213> ORGANISM: Lactobacillus lactis

<400> SEQUENCE: 26

Thr Val Gln Thr Asn Gly Asn Gln Thr Ile Thr Ser Thr Leu Asn Glu
 1 5 10 15

Glu Gln Thr Ser Lys
 20

-continued

<210> SEQ ID NO 27
 <211> LENGTH: 21
 <212> TYPE: PRT
 <213> ORGANISM: Staphylococcus aureus

<400> SEQUENCE: 27

Tyr Lys Asp Gly Ile Gly Asn Tyr Tyr Ala Asn Leu Asn Gly Ser Ile
 1 5 10 15

Glu Thr Phe Asn Lys
 20

<210> SEQ ID NO 28
 <211> LENGTH: 19
 <212> TYPE: PRT
 <213> ORGANISM: Staphylococcus aureus

<400> SEQUENCE: 28

Asp Gly Ile Gly Asn Tyr Tyr Ala Asn Leu Asn Gly Ser Ile Glu Thr
 1 5 10 15

Phe Asn Lys

<210> SEQ ID NO 29
 <211> LENGTH: 13
 <212> TYPE: PRT
 <213> ORGANISM: Staphylococcus aureus

<400> SEQUENCE: 29

Phe Ser His Val Ala Phe Ile Lys Pro Asn Asn Gly Lys
 1 5 10

<210> SEQ ID NO 30
 <211> LENGTH: 12
 <212> TYPE: PRT
 <213> ORGANISM: Staphylococcus aureus

<400> SEQUENCE: 30

Thr Thr Ser Val Thr Val Thr Gly Thr Leu Met Lys
 1 5 10

<210> SEQ ID NO 31
 <211> LENGTH: 13
 <212> TYPE: PRT
 <213> ORGANISM: Lactobacillus lactis

<400> SEQUENCE: 31

Ile Phe Glu Tyr Leu Gly Asn Asn Glu Asp Ile Ala Lys
 1 5 10

<210> SEQ ID NO 32
 <211> LENGTH: 29
 <212> TYPE: PRT
 <213> ORGANISM: Staphylococcus aureus

<400> SEQUENCE: 32

Phe Lys Glu Val Thr Ser Asn Met Ser Gly Asn Leu Asn Leu Gln Asn
 1 5 10 15

Asn Gly Ser Tyr Ser Leu Asn Ile Glu Asn Leu Asp Lys
 20 25

<210> SEQ ID NO 33
 <211> LENGTH: 20

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<212> TYPE: PRT
 <213> ORGANISM: Staphylococcus aureus

<400> SEQUENCE: 33

Thr Tyr Val Val His Tyr Asp Gly Glu Tyr Leu Asn Gly Thr Asp Glu
 1 5 10 15

Val Asp Phe Arg
 20

<210> SEQ ID NO 34
 <211> LENGTH: 12
 <212> TYPE: PRT
 <213> ORGANISM: Staphylococcus aureus

<400> SEQUENCE: 34

Thr Gln Met Val Gly His Pro Glu Gln Leu Tyr Lys
 1 5 10

<210> SEQ ID NO 35
 <211> LENGTH: 14
 <212> TYPE: PRT
 <213> ORGANISM: Staphylococcus aureus

<400> SEQUENCE: 35

Glu Asp Thr Ile Lys Glu Thr Leu Thr Gly Gln Tyr Asp Lys
 1 5 10

<210> SEQ ID NO 36
 <211> LENGTH: 34
 <212> TYPE: PRT
 <213> ORGANISM: Staphylococcus aureus

<400> SEQUENCE: 36

His His Ala Asp Val Val Glu Tyr Glu Glu Asp Thr Asn Pro Gly Gly
 1 5 10 15

Gly Gln Val Thr Thr Glu Ser Asn Leu Val Glu Phe Asp Glu Glu Ser
 20 25 30

Thr Lys

<210> SEQ ID NO 37
 <211> LENGTH: 17
 <212> TYPE: PRT
 <213> ORGANISM: Lactobacillus lactis

<400> SEQUENCE: 37

Gly Ile Val Thr Gly Ala Val Ser Asp His Thr Thr Val Glu Asp Thr
 1 5 10 15

Lys

<210> SEQ ID NO 38
 <211> LENGTH: 31
 <212> TYPE: PRT
 <213> ORGANISM: Lactobacillus lactis

<400> SEQUENCE: 38

Glu Tyr Thr Thr Glu Ser Asn Leu Ile Glu Leu Val Asp Glu Leu Pro
 1 5 10 15

Glu Glu His Gly Gln Ala Gln Gly Pro Val Glu Glu Ile Thr Lys
 20 25 30

-continued

<210> SEQ ID NO 39
 <211> LENGTH: 23
 <212> TYPE: PRT
 <213> ORGANISM: Lactobacillus lactis

<400> SEQUENCE: 39

Tyr Glu Gln Gly Gly Asn Ile Val Asp Ile Asp Phe Asp Ser Val Pro
 1 5 10 15
 Gln Ile His Gly Gln Asn Lys
 20

1. An immunogenic composition comprising an inactivated recombinant non-pathogenic bacterium, or a part thereof, expressing on its cell surface at least one folded sequence of a *S. aureus* adhesin, or a sequence having 80% or more sequence identity to said folded sequence of a *S. aureus* adhesin, wherein said at least one folded sequence of a *S. aureus* adhesin, or sequence having 80% or more sequence identity to said folded sequence of a *S. aureus* adhesin, is entirely accessible to trypsin digestion.

2. The immunogenic composition of claim 1, wherein the trypsin digestion of said at least one folded sequence of a *S. aureus* adhesin, or of the sequence having 80% or more sequence identity to said folded sequence of a *S. aureus* adhesin, releases a digestion profile upon trypsin digestion which is different from a digestion profile upon trypsin digestion of said adhesin expressed on the surface of *S. aureus*.

3. The immunogenic composition of claim 1, wherein the part of the inactivated recombinant non-pathogenic bacterium consists of an isolated and/or purified cell wall.

4. The immunogenic composition of claim 1, wherein the at least one folded sequence of a *S. aureus* adhesin is selected from the group consisting of fibrinogen-binding protein A (clumping factor A ClfA), fibrinogen-binding protein B (ClfB), fibronectin-binding protein A (FnPBA) and fibronectin-binding protein B (FnBPB), collagen-binding protein (Cna) and protein A (Spa), Serine-aspartate repeat protein C, D and E (SdrC-E), Plasmin-sensitive protein (Pls), Factor affecting methicillin resistance in the presence of Triton X-100 (FmtB), and surface protein A-K (SasA-K), or a combination thereof.

5. The immunogenic composition of claim 4, wherein the at least one folded sequence of a *S. aureus* adhesin consists of the fibrinogen-binding protein A (clumping factor A (ClfA)) and/or fibronectin-binding protein A (FnPBA).

6. The immunogenic composition of claim 1, wherein the inactivated recombinant non-pathogenic bacterium is the *L. lactis* subspecies *cremoris* 1363.

7. The immunogenic composition of claim 4, wherein the sequence of *S. aureus* fibrinogen-binding protein A (clumping factor A (ClfA)) is as set forth in SEQ ID No.1.

8. The immunogenic composition of claim 1, wherein the *S. aureus* fibrinogen-binding protein A (clumping factor A (ClfA)) releases a digestion profile of 9 peptides upon trypsin digestion.

9. The immunogenic composition of claim 1, wherein the *S. aureus* fibronectin-binding protein A (FnPBA) releases a digestion profile of 23 peptides upon trypsin digestion.

10. The immunogenic composition of claim 1, wherein the inactivated recombinant non-pathogenic bacterium is UV inactivated.

11. A vaccine comprising an immunogenic composition of claim 1 in an immunologically acceptable carrier and/or diluent.

12. The vaccine of claim 11, wherein the immunologically acceptable carrier is selected from the group consisting of polysaccharide materials forming hydrogels and vesicular carriers.

13. The vaccine of claim 12, wherein the vesicular carriers are selected from the group consisting of bacterial ghosts, liposomes, niosomes, transfersomes, and ethosomes.

14. The vaccine of claim 11, further comprising an adjuvant.

15. (canceled)

16. The method of claim 19, wherein the infection or disease caused by *S. aureus* is selected from the group consisting of IE, intravascular and intravascular device infections, bloodstream infections, deep-seated abscesses, osteomyelitis, infection of prosthetic materials, and skin and soft tissue infections.

17. An isolated and/or purified antibody, antibody fragment or derivative thereof able to bind to the at least one folded sequence of a *S. aureus* adhesin, or to a sequence having 80% or more sequence identity to said folded sequence of a *S. aureus* adhesin, expressed on the cell surface of an inactivated recombinant non-pathogenic bacterium.

18. An expression vector comprising an isolated and/or purified nucleic acid sequence encoding for at least one folded sequence of a *S. aureus* adhesin, or a sequence having 80% or more sequence identity to said folded sequence of a *S. aureus* adhesin.

19. A method for treating and/or preventing an infection or disease caused by *S. aureus*, in a subject in need thereof, comprising administering a pharmaceutically effective amount of an immunogenic composition of claim 1.

20. A method for inducing active immunity against an infection or disease caused by *S. aureus* in a subject in need thereof, comprising administering to said subject in need thereof i) an immunogenic composition of claim 1 or ii) a vaccine of claim 11.

21. A method for inducing passive immunity against an infection or disease caused by *S. aureus* in a subject in need thereof, comprising administering to said subject in need thereof an isolated and/or purified antibody, antibody fragment or derivative thereof of claim 17.

22. The method of claim 20, wherein the infection or disease caused by *S. aureus* is selected from the group consisting

of IE, intravascular and intravascular device infections, bloodstream infections, deep-seated abscesses, osteomyelitis, infection of prosthetic materials, and skin and soft tissue infections.

23. The method of claim **21**, wherein the infection or disease caused by *S. aureus* is selected from the group consisting of IE, intravascular and intravascular device infections, bloodstream infections, deep-seated abscesses, osteomyelitis, infection of prosthetic materials, and skin and soft tissue infections.

* * * * *