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(54) Titre: ANTICORPS MONOCLONAL ANTI-SYNDROME ANTISYNTHETASE HUMANISE

(54) Title: HUMANIZED ANTI-Aβ MONOCLONAL ANTIBODY AND USE THEREOF

(57) Abrégé/Abstract:

Provided are a humanized anti-A β monoclonal antibody and application thereof. The humanized anti-A β monoclonal antibody provided can inhibit the polymerization of A β monomers, protect nerve cells from the toxicity of A β , and have a certain effect on improving the cognitive learning and memory ability of Alzheimer's dementia model mice, and can be used for the treatment and diagnosis of diseases and disorders related to amyloidosis, such as Alzheimer's disease.





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Abstract

Provided are a humanized anti-A β monoclonal antibody and use thereof. The humanized anti-A β monoclonal antibody provided can inhibit the polymerization of A β monomers, protect nerve cells from the toxicity of A β , and have a certain effect on improving the cognitive learning and memory ability of Alzheimer's dementia model mice, and can be used for the treatment and diagnosis of diseases and disorders related to amyloidosis, such as Alzheimer's disease.

Humanized anti-A \beta monoclonal antibody and use thereof

The present application claims the priority of the Chinese patent application filed with the Chinese Patent Office on February 1, 2019, with the application number 201910104326.1 and the invention title of "Humanized anti-A β monoclonal antibody and use thereof", the entire content of which is incorporated in the present application by reference.

Technical Field

The present disclosure relates to the technical field of antibody medicines, in particular to a humanized anti-Aβ monoclonal antibody and use thereof.

Background Art

<u>Αβ</u>

Amyloid β (A β) is encoded by the human chromosome 21 gene, contains 39-43 amino acids, has a β -sheet structure, is hydrophobic, and has a molecular weight of 4 KDa. A β is derived from the residue polypeptide produced by the fragmentation of amyloid precursor protein (APP) by proteolytic enzymes. APP can be decomposed by α -, β - and γ -proteases, and the products after decomposition have different biological functions. Among them, A β is produced by the continuous action of β -protease and γ -protease. The C-terminus of A β is produced by γ -protease, and a large number of residue subtypes with 39-43 amino acids are produced by cutting APP at the transmembrane region. The most common of all residue subtypes are A β 40 and A β 42. The former is typically formed by cutting APP at the endoplasmic reticulum, while the latter is formed in the trans-Golgi network.

Pathological mechanism of AB

It is currently known that all nerve cells, including neurons, astrocytes, microglia and endothelial cells, in the central nervous system (CNS) can express APP and produce $A\beta$. Under normal physiological conditions, APP is hydrolyzed by α -secretase to produce a soluble sAPP α fragment. This fragment contains the extracellular region of APP and the C-terminus with 83 amino acids located on the cell membrane. sAPP α can regulate the excitability of neurons, improve the plasticity, learning and memory of synapses, and enhance the resistance of neurons to oxidative and metabolic stress. Under neuropathological conditions, APP is first hydrolyzed by β -secretase 1 (BACE) to produce a sAPP β fragment and a peptide fragment with 99-amino acids (C99) connected to the cell membrane. Subsequently, the C99 peptide is subjected to the action of γ -secretase to produce $A\beta$. Different from sAPP α , $A\beta$ can cause the loss of nerve synapse function, reduce the plasticity of neurons, change cell energy metabolism, induce oxidative stress response

and mitochondrial dysfunction, thereby causing the imbalance of intracellular calcium ions. The formation, accumulation and deposition of $A\beta$, especially $A\beta_{42}$, may cause neurotoxicity and neurodegenerative diseases, and also play an important role in the pathogenesis of Alzheimer's disease (AD).

Aβ and Alzheimer's disease (AD)

As one of the main intracerebral pathological marker proteins of Alzmer's disease, the formation, deposition and degradation of $A\beta$ run through the whole pathological process of AD. $A\beta$ is divided into two types: soluble and insoluble. Soluble $A\beta$ itself has no neurotoxicity, but shows neurocytotoxicity after becoming insoluble precipitates upon formation of filamentous fiber aggregates by β -sheet. The primary structure of human $A\beta$ is a determinant factor of neurotoxicity. According to reports in the literature, the neurotoxicity of $A\beta$ is mainly reflected in the following four aspects: cholinergic neuron damage, nerve cell apoptosis, peroxidative damage and inflammatory response.

Cholinergic neuron damage

The damage and loss of a large number of cholinergic system neurons and nerve synapses in the anterior basal projecting to the hippocampus and cortex are the main reasons for the decline of memory and cognitive ability of AD patients. A β activates protein kinase GSK-3/glycogen synthase kinase-3 β , which causes the phosphorylation of tau protein and mitochondrial pyruvate dehydrogenase, reduces the enzyme activity, and reduces the conversion of pyruvate into acetyl coenzyme A (actyl coenzyme A), thereby reducing the synthesis of acetylcholine (ACh), inhibiting succinate dehydrogenase, reducing energy supply, causing the damage, degeneration and transmitter transmission disorder of cholinergic neurons and synapses, and decreasing the activity of the cholinergic system. The reduction of ACh in turn leads to an increase in the production of A β , which in turn forms a vicious circle.

Nerve cell apoptosis

The main characteristic of AD is the decrease in the number of neurons in the cortex and hippocampus. When A β aggregates into a β -sheet folding structure, its neurotoxicity is significantly enhanced, and it can induce the apoptosis of nerve cells. This is an important reason for the lack of selective neurons and synapses in AD. The fibrous and aggregated A β and APP and other transmembrane receptors interact and cross-link through secretory pathways on the cell surface, leading to the inhibition and abnormal activation of signal transduction pathways, thereby starting the apoptosis of nerve cells. The A β -induced Ca²⁺ imbalance in the internal environment stimulates NMDA receptors or changes the membrane permeability through free radical damage effect, causing the Ca²⁺ influx to activate glutamate receptors, and causing the overexcitation and

death of glutamatergic neurons. In addition, $A\beta$ may also cause the increase of NO synthesis, thereby inducing the apoptosis of neuronal cells.

Peroxidative damage

Aβ may cause oxidative stress in many ways. Oxidative stress caused by the increase of free radicals induced by Aβ is an important reason. The toxicity of Aβ is mediated by H_2O_2 , and Aβ increases the accumulation of H_2O_2 in the body through the receptor of advanced glycation endoproduct (RAGE), causing oxidative damage and causing cell death. In addition, oxidative stress causes the microglia to proliferate and migrate along the Aβ concentration gradient, leading to aggregation of microglia around senile plaques, forming neuritic plaques, and generating more reactive oxygen free radicals. Aβ can also increase lipid peroxidation. H_2O_2 is not only a source of hydroxyl free radicals, but also increases the abnormal expression of nuclear factor κB (NF- κB) protein, which causes nerve cell membrane damage and leads to neuronal degeneration.

Inflammatory reaction

AD patients usually have inflammatory reaction in the brain. Glial cells proliferate around plaques and neurofibrillary tangles. A β may stimulate the release of a series of inflammatory proteins with strong neurotoxicity. For example, A β activates astrocytes and microglia to release inflammatory cytokines, such as NO, interleukin-1 (IL-1, IL-1 may cause abnormality in the production of cytoskeleton protein-neurofilament protein, thereby impairing the function of neurons), interleukin-6 (IL-6, IL-6 increases the overexpression of ADP and promotes the formation of A β , while A β may induce the expression of IL-6 in microglia, thereby forming a vicious circle in the immunopathological process of AD), tumor necrosis factor- α (TNF- α , TNF- α is involved in the pathological process of AD through the most important apolipoprotein ApoE of CNS), γ -interferon (γ -IFN), β -antitrypsin (ACT), complement C1, C3 and chemokines, adhesion factors, etc. Many inflammatory factors induce inflammation, promote the generation of free radicals, oxidative stress, resulting degeneration and necrosis of nerve cells.

Drugs and therapeutic mechanisms for AB target intervention

Based on the above, the toxicity of $A\beta$ to neurons is an important factor in the occurrence of AD. Therefore, by inhibiting the production of $A\beta$ and accelerating its clearance, the disease process of AD can be stopped, and the symptoms of the disease can be alleviated. The drugs currently being developed and used in clinic are also based on the production and clearance mechanism of $A\beta$, which include the following parts.

1. Inhibiting β - and γ -secretases

The hydrolysis of APP by β -secretase is the initial stage of amyloid production. Inhibiting the activity of β -secretase can inhibit the production of A β , but it may cause greater side effects. Because in addition to APP, β -secretase has numerous substrates, and the hydrolysis of these substrates plays an important role in the plasticity of neurons and synapses in the nervous system. Clinically, β -secretase inhibitors, such as E2609 (clinical trial ID# NCT01600859), MK-8931 (NCT01739348) and LY2886721 (NCT01807026, NCT01561430) all can reduce A β levels in human cerebrospinal fluid by 80-90%, but currently there is still no β -secretase inhibitor on the market.

The hydrolysis of APP by γ-secretase is the last step in the production of amyloid, which directly produces A β 40 and A β 42 fragments. Therefore, it is also considered that the inhibition of γ -secretase may effectively inhibit the production of A β , so as to achieve the purpose of treating AD. However, in addition to hydrolyzing APP, γ -secretase also hydrolyzes other substrate proteins, including Notch protein. Notch protein is important for cell proliferation, differentiation and intercellular signal transduction. Semagacestat (LY450139) as a γ-secretase inhibitor has been clinically tested in 3000 patients (NCT00762411, NCT01035138, NCT00762411). The results of the test showed that the subjects' cognition did not improve, but deteriorated, and was accompanied by side effects such as weight loss, increased skin cancer probability, and high risk of infection. Other γ-secretase inhibitors, such as Avagacestat, have also failed in clinical trials (NCT00810147, NCT00890890, NCT00810147, NCT01079819). The selective γ-secretase modulator (SGSM) can theoretically avoid the side effects caused by the total inhibition of γ secretase, and only inhibit the hydrolysis pathway of APP without interfering with other signal channels, such as the hydrolysis of Notch protein. Some non-steroidal anti-inflammatory drugs, such as ibuprofen, sulindac, indomethacin, and flurbiprofen, can regulate the level of γ -secretase, and can reduce the level of Aβ₄₂ in in vivo and in vitro activity experiments. Although such drugs have been shown to relieve mild cognitive impairment and reduce the level of inflammatory factors in the cerebrospinal fluid, long-term use of non-steroidal anti-inflammatory drugs for the treatment of AD still needs to be clinically verified.

2. Inhibiting Aβ aggregation

Inhibition of senile plaque can be achieved by interfering with or antagonizing the accumulation of A β . For example, 3-Amino-1-propane sulfonic acid (3-APS, Alzhemed, tramiprosate) interferes with the interaction between dissoluble A β and endogenous aminodextran, in which the later can promote the formation and precipitation of A β amyloid fibers, thereby inhibiting the accumulation of A β . However, the results of the Phase III clinical trial of 3-APS were not satisfactory, which led to the suspension of the trial. Other anti-A β aggregation drugs have also failed in phase III and phase III clinical trials, including Colostrinin, which could inhibit

A β aggregation and neutralize the neurotoxicity of A β in an in vitro test, and could also improve the cognition ability of mice in an in vivo test, but it did not achieve satisfactory results in the clinical phase II trial. Scyllo-inositol (ELND005) is an oral anti-A β aggregation drug, and the mouse experiments have shown that Scyllo-inositol could reduce the toxicity of A β , but did not achieve the expected results in the 18-month phase II clinical trial for patients with mild to moderate AD.

3. Promoting the clearance of A β deposition and polymer

There are three main ways to remove $A\beta$ deposition and polymer: activating the activity of amyloid plaque degrading enzymes; regulating the transport of $A\beta$ in the brain and peripheral circulation; and anti- $A\beta$ immunotherapy.

The deposition and polymer of $A\beta$ can be degraded by a variety of proteolytic enzymes, including plasmin, endothelin-converting enzyme, angiotensin-converting enzyme, metalloproteinase, etc. The levels of these enzymes in the brains of AD patients are relatively low, but due to the lack of specificity for these enzymes, no such drugs have entered the clinic at present.

The transport of $A\beta$ between the central nervous system and the peripheral circulatory system is regulated by apolipoprotein. Low-density lipoprotein receptor-related protein (LRP-1) can promote the flow of $A\beta$ from the brain into the blood. The receptor for advanced glycation end products (RAGE) can assist $A\beta$ to pass through the blood-brain barrier. This treatment mechanism is to reduce the load of amyloid in the brain by restricting $A\beta$ from entering the peripheral circulation. So far, only RAGE inhibitors/modulators have entered clinical trials, including PF-0449470052 and TTP4000. The former failed in phase II clinical trials, while the latter did not have reliable data to show that the expected results were achieved in phase I clinical trials.

Anti-A β antibodies can neutralize the toxicity of A β and improve the cognition of transgenic animals. Anti-A β antibodies have become a hot topic in AD treatment. Anti-A β antibodies mainly aim at the early treatment of AD, as well as the treatment of mild to moderate AD. This is also related to the pathogenic mechanism of A β , that is, once neurons are injured, it is difficult to reverse and repair them, therefore, the early removal of A β may more effectively treat and alleviate AD.

4. Aβ target antibodies currently undergoing clinical trials

There are currently 15 anti-A β antibody drugs undergoing clinical trials. In comparison, Aducanumab, Gantenerumab and Solanezumab have advanced rapidly and have entered phase III clinical trials. As mentioned in the mechanism, various companies have targeted mild Alzheimer's disease as their indications.

Although there is a certain theoretical knowledge in the field of treatment and prevention of Alzheimer's disease, there is still a need to improve the composition and method for the treatment and/or prevention of the disease, and there is a need for antibodies and treatments that can target $A\beta$. Although some humanized monoclonal antibodies with great therapeutic advantages have been obtained, it is not an easy task to screen out humanized monoclonal antibodies with the required properties and functions. In reality, there is still an urgent need for such humanized monoclonal antibodies.

Contents of the present disclosure

In view of this, the technical problem to be solved by the present disclosure is to provide a humanized anti-A β monoclonal antibody and use thereof, and also provide a carrier and host cell for a nucleotide encoding the monoclonal antibody and use thereof. From the sequence of the variable region of the antibody gene involved in the present disclosure, a full-length antibody molecule can be constructed, which can be used as a drug for treatment and diagnosis of an amyloidosis-related disease and disorder (such as Alzheimer's disease) in clinic.

In order to achieve the above-mentioned purpose of the present disclosure, the present disclosure provides the following technical solutions.

The present disclosure provides an anti-Aß humanized monoclonal antibody, in which

(I) the amino acid sequences of three heavy chain CDR regions of the monoclonal antibody are the amino acid sequences set forth in SEQ ID NOs: 1, 2 and 3, respectively; and (II) the amino acid sequences of three light chain CDR regions of the monoclonal antibody are the amino acid sequences set forth in SEQ ID NOs: 4, 5 and 6, respectively;

or

(III) the amino acid sequences are the amino acid sequences obtained from the amino acids of (I) or (II) via substitution, deletion or addition of one or more amino acids, and are the amino acid sequences having the same function as the amino acid sequence of (I) or (II);

further, the function comprises two or three functions selected from the group consisting of inhibition of $A\beta$ polymerization, improvement of cognitive learning and memory ability in an Alzheimer's dementia model, and cytotoxic protective activity;

or

(IV) the amino acid sequences are the amino acid sequences having at least 97% homology with the amino acid sequences of (I), (II) or (III).

Further, the monoclonal antibody of the present disclosure has an antigen binding epitope of $A\beta_{30\sim42}$.

In some specific embodiments of the present disclosure, the present disclosure provides a humanized anti-A β monoclonal antibody, wherein:

its heavy chain comprises three CDR regions, in which at least one of the CDR regions has an amino acid sequence that is the amino acid sequence as shown in SEQ ID NO: 1, 2 or 3, or an amino acid sequence that has at least 97% homology with the amino acid sequence;

its light chain comprises three CDR regions, in which at least one of the CDR regions has an amino acid sequence that is the amino acid sequence as shown in SEQ ID NO: 4, 5 or 6, or an amino acid sequence that has at least 97% homology with the amino acid sequence.

In some specific embodiments of the present disclosure, the three heavy chain CDR regions of the monoclonal antibody have amino acid sequences that are the amino acid sequences as shown in SEQ ID NOs: 1, 2 and 3, respectively;

the three light chain CDR regions of the monoclonal antibody have amino acid sequences that are the amino acid sequences as shown in SEQ ID NOs: 4, 5 and 6, respectively.

Therein, the sequence shown in SEQ ID NO:1 is SYAMS;

the sequence shown in SEQ ID NO: 2 is SISTTSNTYYPDSVKG;

the sequence shown in SEQ ID NO: 3 is GVITNQAWFAY;

the sequence shown in SEQ ID NO: 4 is RASQSISNNLH;

the sequence shown in SEQ ID NO: 5 is YASQSIS;

the sequence shown in SEQ ID NO: 6 is QQSNSWPLT.

In some specific embodiments of the present disclosure, the present disclosure provides a monoclonal antibody, in which

(V) the amino acid sequences of 4 heavy chain FR regions of the monoclonal antibody are the amino acid sequences set forth in SEQ ID NOs: 7, 8, 9 and 10, respectively; and (VI) the amino acid sequences of 4 light chain FR regions of the monoclonal antibody are the amino acid sequences set forth in SEQ ID NOs: 11, 12, 13 and 14, respectively;

or

(VII) the amino acid sequences are the amino acid sequences obtained from the amino acids of (V) or (VI) via substitution, deletion or addition of one or more amino acids, and are the amino acid sequences having the same function as the amino acid sequences of (V) or (VI);

further, the function comprises two or three functions selected from the group consisting of inhibition of $A\beta$ polymerization, improvement of cognitive learning and memory ability in an Alzheimer's dementia model, and cytotoxic protective activity;

or

(VIII) the amino acid sequences are the amino acid sequences having at least 97% homology with the amino acid sequences of (V), (VI) or (VII).

In some specific embodiments of the present disclosure, its heavy chain comprises 4 FR regions, in which at least one of the FR regions has an amino acid sequence that is the amino acid sequence as shown in SEQ ID NO: 7, 8, 9 or 10, or an amino acid sequence that has at least 97% homology with the amino acid sequence;

its light chain comprises 4 FR regions, in which at least one of the FR regions has an amino acid sequence that is the amino acid sequence as shown in SEQ ID NO: 11, 12, 13 or 14, or an amino acid sequence that has at least 97% homology with the amino acid sequence.

In some specific embodiments of the present disclosure, the 4 heavy chain FR regions of the monoclonal antibody have amino acid sequences that are the amino acid sequence as shown in SEQ ID NOs: 7, 8, 9 and 10, respectively, or amino acid sequences that have at least 97% homology with the amino acid sequences;

the 4 light chain FR regions of the monoclonal antibody have amino acid sequences that are the amino acid sequence as shown in SEQ ID NOs: 11, 12, 13 and 14, respectively, or amino acid sequences that have at least 97% sequence homology with the amino acid sequences.

Therein, the sequence shown in SEQ ID NO: 7 is EVQLVESGGGLVQPGGSLRLSCVASGFTFR;

the sequence shown in SEQ ID NO: 8 is WVRQAPGKGLEWVA;

the sequence shown in SEQ ID NO: 9 is RFTTSRDNSKNTVYLQMSSLRAEDTAVYYCGR;

the sequence shown in SEQ ID NO: 10 is WGQGTLVTVSS;

the sequence shown in SEQ ID NO: 11 is DIVLTQSPATLSVSPGERATLSC;

the sequence shown in SEQ ID NO: 12 is WYQQKPGQAPRLLIK;

the sequence shown in SEQ ID NO: 13 GIPARFSGSGSGTDFTLTISSLQSEDFAVYFC;

the sequence shown in SEQ ID NO: 14 is FGGGTKVEIK.

In some specific embodiments of the present disclosure, for the monoclonal antibody,

(IX) its heavy chain variable region has the amino acid sequence as shown in any one of SEQ ID NOs: 15 to 19; and (X) its light chain variable region has the amino acid sequence as shown in any one of SEQ ID NOs: 20 to 24;

or

(XI) its heavy chain variable region or its light chain variable region has the amino acid sequence is obtained from the amino acids of (IX) or (X) via substitution, deletion or addition of one or more amino acids, and are the amino acid sequence having the same function as the amino acid sequence of (IX) or (X);

further, the function comprises two or three functions selected from the group consisting of inhibition of $A\beta$ polymerization, improvement of cognitive learning and memory ability in an Alzheimer's dementia model, and cytotoxic protective activity;

or

(XII) its heavy chain variable region or its light chain variable region has the amino acid sequence having at least 97% homology with the amino acid sequence of (IX), (X) or (XI).

In some specific embodiments of the present disclosure, its heavy chain variable region has the amino acid sequence as shown in any one of SEQ ID NOs: 15 to 19; and its light chain variable region has the amino acid sequence as shown in any one of SEQ ID NOs: 20 to 24.

In some specific embodiments of the present disclosure, the humanized anti-A β monoclonal antibody comprises:

a heavy chain variable region that has the amino acid sequence as shown in SEQ ID NO: 15, 16, 17, 18 or 19, and a light chain variable region that has the amino acid sequence as shown in SEQ ID NO: 20; or

a heavy chain variable region that has the amino acid sequence as shown in SEQ ID NO: 15, 16, 17, 18 or 19, and a light chain variable region that has the amino acid sequence as shown in SEQ ID NO: 21; or

a heavy chain variable region that has the amino acid sequence as shown in SEQ ID NO: 15, 16, 17, 18 or 19, and a light chain variable region that has the amino acid sequence as shown in SEQ ID NO: 22; or

a heavy chain variable region that has the amino acid sequence as shown in SEQ ID NO: 15, 16, 17, 18 or 19, and a light chain variable region that has the amino acid sequence as shown in SEQ ID NO: 23; or

a heavy chain variable region that has the amino acid sequence as shown in SEQ ID NO: 15, 16, 17, 18 or 19, and a light chain variable region that has the amino acid sequence as shown in SEQ ID NO: 24.

In some specific embodiments of the present disclosure, for the monoclonal antibody,

(XIII) its heavy chain variable region has the amino acid sequence as shown in SEQ ID NO: 16, and its light chain variable region has the amino acid sequence as shown in SEQ ID NO: 21; or

(XIV) its heavy chain variable region has the amino acid sequence as shown in SEQ ID NO: 16, and its light chain variable region has the amino acid sequence as shown in SEQ ID NO: 22; or

(XV) its heavy chain variable region has the amino acid sequence as shown in SEQ ID NO: 18, and its light chain variable region has the amino acid sequence as shown in SEQ ID NO: 21; or

(XVI) its heavy chain variable region has the amino acid sequence as shown in SEQ ID NO: 19, and its light chain variable region has the amino acid sequence as shown in SEQ ID NO: 20; or

(XVII) its heavy chain variable region has the amino acid sequence as shown in SEQ ID NO: 19, and its light chain variable region has the amino acid sequence as shown in SEQ ID NO: 21; or

(XVIII) its heavy chain variable region has the amino acid sequence as shown in SEQ ID NO: 19, and its light chain variable region has the amino acid sequence as shown in SEQ ID NO: 22.

In the present disclosure, the sequence that has at least 97% sequence homology is an amino acid sequence obtained by substitution, deletion or addition of one or more amino acids on the basis of the original sequence, wherein the more amino acids refer to 2, 3, 4 or 5 amino acids.

In some specific embodiments of the present disclosure, the monoclonal antibody further comprises a constant region, in which the monoclonal antibody has a heavy chain constant region that is any one of human IgG1, IgG2, IgG3, or IgG4; and the monoclonal antibody has a light chain constant region that is of κ type or λ type.

In some specific embodiments of the present disclosure, the anti-A β monoclonal antibody provided by the present disclosure has a heavy chain constant region that is human IgG1, and a light chain constant region that is a constant region of human κ chain.

The humanized anti-A β monoclonal antibody provided by the present disclosure can bind to human A β ; in some embodiments, the affinity between the antibody and its target is characterized

by Ka (association constant), Kd (dissociation constant), and KD (equilibrium dissociation solution); and the KD value of the antibody provided by the present disclosure is not higher than 36.3 nM. The humanized anti-A β monoclonal antibody provided by the present disclosure can inhibit the polymerization of A β monomer, protect nerve cells from the toxicity of A β , and has a certain effect on improving cognitive learning and memory ability in Alzheimer's dementia model mice.

The present disclosure also provides nucleotides encoding the monoclonal antibody.

The present disclosure provides a nucleotide sequence encoding the heavy chain of the monoclonal antibody.

The present disclosure provides a nucleotide sequence encoding the light chain of the monoclonal antibody.

The present disclosure provides a nucleotide sequence encoding the heavy chain variable region of the monoclonal antibody.

Therein, the nucleotide sequence encoding the heavy chain variable region of the monoclonal antibody is shown in SEQ ID NOs: 25 to 29 or is a complementary sequence of SEQ ID NOs: 25 to 29.

In some specific embodiments of the present disclosure, the nucleotide has a nucleotide sequence that is obtained from the nucleotide sequence as shown in any one of SEQ ID NOs: 25 to 29 via substitution, deletion or addition of one or more nucleotides and has the same or similar function as the nucleotide sequence as shown in any one of SEQ ID NOs: 25 to 29.

In some specific embodiments of the present disclosure, for the nucleotide sequence that is obtained from the nucleotide sequence as shown in any one of SEQ ID NOs: 25 to 29 via substitution, deletion or addition of one or more nucleotides, the more nucleotides refer to 2, 3, 4 or 5 nucleotides.

The present disclosure provides a nucleotide sequence encoding the light chain variable region of the monoclonal antibody.

Therein, the nucleotide sequence encoding the light chain variable region of the monoclonal antibody is shown in SEQ ID NOs: 30 to 34, or is a complementary sequence of SEQ ID NOs: 30 to 34.

In some specific embodiments of the present disclosure, the nucleotide has a nucleotide sequence that is obtained from the nucleotide sequence as shown in any one of SEQ ID NOs: 30 to 34 via substitution, deletion or addition of one or more nucleotides and has the same or similar function as the nucleotide sequence as shown in any one of SEQ ID NOs: 30 to 34.

In some specific embodiments of the present disclosure, for the nucleotide sequence that is obtained from the nucleotide sequence as shown in any one of SEQ ID NOs: 30 to 34 via substitution, deletion or addition of one or more nucleotides, the more nucleotides refer to 2, 3, 4 or 5 nucleotides.

The expression vector provided by the present disclosure comprises nucleotides encoding the anti-A β monoclonal antibody.

The present disclosure also provides a host cell that is transformed or transfected with the expression vector.

The preparation method of the anti-A β monoclonal antibody of the present disclosure comprises: culturing the host cell and inducing the expression of the anti-A β monoclonal antibody.

The present disclosure also provides a conjugate, comprising the monoclonal antibody that is chemically or biologically labeled.

The chemical label is an isotope, immunotoxin and/or chemical drug.

The biological label is a biotin, avidin or enzyme label.

The enzyme label is preferably horseradish peroxidase or alkaline phosphatase.

The immunotoxin is preferably aflatoxin, diphtheria toxin, pseudomonas aeruginosa exotoxin, ricin, abrin, mistletoe lectin, volkensin toxin, PAP, saporin, gelonin or luffin.

The present disclosure also provides a coupling product that is prepared by coupling the monoclonal antibody or its conjugate with a solid medium or a semi-solid medium.

The solid medium or non-solid medium is selected from colloidal gold, polystyrene plates or beads.

The present disclosure also provides use of the monoclonal antibody, the conjugate and/or the coupling product in the manufacture of an agent for combating cognitive impairment, an agent for treating Alzheimer's disease, an agent for inhibiting the progression of Alzheimer's disease, an agent for inhibiting the formation of senile plaques, an agent for inhibiting $A\beta$ accumulation, an agent for combating neurotoxicity, an agent for inhibiting the formation of $A\beta$ amyloid fibrils, and/or an agent for combating synaptic toxicity.

The present disclosure also provides use of the humanized anti-A β monoclonal antibody, the conjugate and/or the coupling product in the manufacture of a medicament for the prevention and treatment of a disease;

the disease comprises amyloidosis, which is a disease and abnormality associated with amyloid protein, the amyloidosis comprises secondary amyloidosis and age-related amyloidosis, and the disease includes, but is not limited to, neurological disease such as Alzheimer's disease.

The present disclosure also provides a medicament, comprising the humanized anti-A β monoclonal antibody, its conjugate and/or coupling product.

The present disclosure also provides a method for the prevention and/or treatment of a disease, comprising administering the medicament of the present disclosure; the disease and disorder comprises amyloidosis, which is a disease and abnormality associated with amyloid protein, the amyloidosis comprises secondary amyloidosis and age-related amyloidosis, and the disease including but not limited to neurological disease such as Alzheimer's disease.

The humanized anti-A β monoclonal antibody provided by the present disclosure can inhibit the polymerization of A β monomers, protect nerve cells from the toxicity of A β , have a certain effect on improving cognitive learning and memory ability in Alzheimer's dementia model mice, and can be used for the treatment and diagnosis of a disease and disorder associated with amyloidosis, such as Alzheimer's disease.

The present disclosure also provides use of the humanized anti-A β monoclonal antibody, the conjugate and/or the coupling product in the manufacture of a product for detecting A β expression.

Experiments show that the humanized anti-A β monoclonal antibody provided by the present disclosure can bind to A β monomer. Therefore, the humanized anti-A β monoclonal antibody provided by the present disclosure can be used for the detection of A β monomer.

The present disclosure also provides a kit, comprising the humanized anti-A β monoclonal antibody, its conjugate and/or coupling product.

The kit for detecting $A\beta$ monomer or polymer mixture provided by the present disclosure further comprises a coating buffer, a washing solution, a blocking solution and/or a color developing solution.

The coating buffer is a carbonate buffer.

The washing solution comprises PBS, Tween, sodium chloride, potassium chloride, disodium hydrogen phosphate, and dipotassium hydrogen phosphate.

The blocking solution comprises PBS and BSA.

The color developing solution comprises TMB solution, substrate buffer solution and stop solution.

The substrate buffer comprises citric acid and disodium hydrogen phosphate.

The stop solution is an aqueous hydrogen peroxide solution.

The kit for detecting a cell with surface expression of $A\beta$ further comprises PBS, goat antimouse IgG Fc and TITC secondary antibody.

The present disclosure also provides a method for diagnosing a disease, comprising using the kit provided by the present disclosure to detect the expression of $A\beta$, and determining whether there is a disease based on the expression quantity of $A\beta$; the disease comprises amyloidosis, which is a disease and abnormality associated with amyloid protein, the amyloidosis comprises secondary amyloidosis and age-related amyloidosis, and the disease includes but is not limited to neurological disease such as Alzheimer's disease.

In some specific embodiments of the present disclosure, the standard for determining whether there is a disease based on the expression quantity of A β is: 600 to 1000 pg/ml for normal people, 200 to 450 pg/ml for AD patients, and the required detection sensitivity is < 20 pg/ml.

Unless otherwise defined, all scientific and technological terms used herein have the same meaning as understood by those of ordinary skill in the art. For definitions and terms in this field, professionals can refer to Current Protocols in Molecular Biology (Ausubel). The abbreviation for amino acid residue is the standard 3-letter and/or 1-letter code used in the art for referring to one of the 20 commonly used L-amino acids.

"Antibody" refers to a protein composed of one or more polypeptides that can specifically bind to an antigen. One form of antibody constitutes the basic structural unit of an antibody. This form is a tetramer, which is composed of two pairs of identical antibody chains, each of which has a light chain and a heavy chain. In each pair of antibody chains, the variable regions of the light chain and the heavy chain are jointly responsible for binding the antigen, while the constant regions are responsible for the effector function of the antibody.

The "variable region" of an antibody heavy or light chain is a N-terminal mature region of the chain. Currently known antibody types include κ and λ light chains, as well as α , γ (IgG1, IgG2, IgG3, IgG4), δ , ϵ , and μ heavy chains or their other type equivalents. The full-length immunoglobulin "light chain" (approximately 25 kDa or approximately 214 amino acids) comprises a variable region formed by approximately 110 amino acids at the NH₂-terminus, and a κ or λ constant region at the COOH-terminus. The full-length immunoglobulin "heavy chain" (approximately 50 kDa or approximately 446 amino acids) also comprises a variable region (approximately 116 amino acids) and one of the heavy chain constant regions, such as γ (approximately 330 amino acids).

"Antibody" comprises any isotype antibody or immunoglobulin, or antibody fragment that retains specific binding to the antigen, including but not limited to Fab, Fv, scFv and Fd fragments,

chimeric antibodies, humanized antibodies, single chain antibodies, and fusion proteins comprising an antigen-binding portion of antibody and a non-antibody protein. The antibody can be labeled and detected. For example, it can be labeled and detected by radioisotopes, enzymes, fluorescent proteins, biotin and so on that can produce detectable substances. The antibody can also be bound to a solid carrier, including but not limited to polystyrene plates or beads.

"Humanized antibody" refers to an antibody that contains a CDR region derived from a non-human antibody, and other parts of the antibody molecule are derived from one (or several) human antibodies. Moreover, in order to retain binding affinity, some residues of the framework (referred to as FR) segment can be modified.

The "monoclonal antibody" refers to a preparation of antibody molecules with a single molecular composition. The monoclonal antibody composition shows a single binding specificity and affinity for a specific epitope.

The medicament comprises at least one functional ingredient, and further comprises a pharmaceutically acceptable carrier. Preferably, the pharmaceutically acceptable carrier is water, buffered aqueous solution, isotonic salt solution such as PBS (phosphate buffered saline), glucose, mannitol, dextrose, lactose, starch, magnesium stearate, cellulose, magnesium carbonate, 0.3% glycerin, hyaluronic acid, ethanol or polyalkylene glycols such as polypropylene glycol, triglycerides, etc. The type of pharmaceutically acceptable carrier used depends in particular on whether the composition according to the present disclosure is formulated for oral, nasal, intradermal, subcutaneous, intramuscular or intravenous administration. The composition according to the present disclosure may comprise a wetting agent, an emulsifier or a buffer substance as an additive.

As used herein, "CDR region" or "CDR" refers to a hypervariable region of the heavy chain and the light chain of immunoglobulin, as defined by Kabat et al. (Kabat et al., Sequences of proteins of immunological interest, 5th Ed., US Department of Health and Human Services, NIH, 1991, and later versions). There are three heavy chain CDRs and three light chain CDRs. Depending on the situation, the term CDR or CDRs as used herein is used to indicate one of these regions, or several or even all of these regions, which contain most of the amino acid residues responsible for binding by the affinity of the antibody to the antigen or its recognition epitope.

The present disclosure provides a method for antibody humanization modification, in which a reasonable antibody humanization design is carried out by referring to multi-template to perform FR transplantation, thereby obtaining a humanized antibody with an affinity equivalent to that of a murine antibody.

The preparation method of the humanized anti-A β monoclonal antibody provided by the present disclosure comprises:

Step 1: preparing mouse-derived hybridoma, obtaining an antibody sequence through 5'RACE;

Step 2: antibody humanization, in which sequence alignment is performed on the NCBI tool to complete the humanization modification, and the modified antibodies are screened.

Specifically, the method comprises as follows.

The method for preparing the humanized anti-A β monoclonal antibody comprises: using murine antibody 066-4.26.14 as a template, performing PCR amplification to obtain a heavy chain variable region gene VH and a light chain variable region gene VL of the antibody, and translating them into amino acid sequences, and then aligning the amino acid sequences with the human antibody sequences in the NCBI database, and selecting 5 human antibody sequences with the highest similarity to the VH and the VL of the variable regions as reference templates for the humanization modification, determining the CDR regions of the murine antibody 066-4.26.14, leaving the CDR regions unchanged, transplanting the FR regions from the 5 reference templates of the above VH and VL respectively to 066-4.26.14 to obtain humanized sequences, which are codon optimized and then separately subjected to the construction of expression vectors for transient transfection; transferring the expression vectors to 293E cells for expression to obtain humanized antibodies that specifically bind to A β ; subjecting the humanized antibodies to affinity determination, antigen binding EC50 value determination, A β polymerization inhibition test and cytotoxicity protection test, and finally obtaining the humanized A β antibodies.

The humanized anti-A β monoclonal antibody provided by the present disclosure can inhibit the polymerization of A β monomers, protect nerve cells from the toxicity of A β , have a certain effect on improving the cognitive learning and memory ability in Alzheimer's dementia model mice, and can be used for the treatment and diagnosis of amyloidosis-associated diseases and disorders, such as Alzheimer's disease.

Brief Description of the Drawings

In order to explain the examples of the present disclosure or the technical solutions in the prior art more clearly, the following will briefly describe the drawings that need to be used in the description of the examples or the prior art.

Fig. 1 shows the SDS-PAGE and WB detection results of $A\beta$ monomer and polymer mixture; in which, lane M: protein molecular weight marker; lane 1: $A\beta$ monomer; lane 2: $A\beta$ polymer mixture; Fig. 1(A): SDS-PAGE detection results of $A\beta$ monomer and polymer mixture; Fig. 1(B): WB detection results of $A\beta$ monomer and polymer mixture;

Fig. 2 shows the SDS-PAGE detection results of the purified positive antibodies; in which, lane M: protein molecular weight marker; lane 1: 066-P01 (non-reducing); lane 2: 066-P01 (reducing); lane 3: 066-P02 (non-reducing); lane 4: 066-P02 (reducing); Fig. 2(A): SDS-PAGE detection results of the purified positive antibody 066-P01; Fig. 2(B): SDS-PAGE detection results of the purified positive antibody 066-P02;

Fig. 3 shows the detection results of anti-A β monoclonal antibodies in inhibiting A β polymerization; the abscissa represents different sample groups, the ordinate represents relative fluorescence intensity, and the anti-A β monoclonal antibodies such as 066-4.22.1, 066-4.26.14, 066-5.4.1 all can inhibit A β polymerization; in which, Fig. 3(A) shows the detection results of the sample groups IgG, anti-A β monoclonal antibodies 066-P01, 066-5.4.1, 066-6.1.1, 066-6.1.3, 066-6.2.1, 066-6.7.2 in inhibiting A β polymerization, respectively; Fig. 3(B) shows the detection results of the sample groups PBS, IgG, anti-A β monoclonal antibodies 066-4.21.13, 066-4.26.14, 066-4.6.8, 066-4.22.1, 066-4.18.2, 066-P01 in inhibiting A β polymerization, respectively;

Fig. 4 shows the activity detection results of anti-A β monoclonal antibodies in promoting macrophage phagocytosis of A β ; the abscissa represents different sample groups, the ordinate represents fluorescence intensity, and the anti-A β monoclonal antibodies 066-5.4.1, 066-7.17.2 have the activities of promoting macrophages phagocytosis of A β ;

Fig. 5 shows the protective activity detection results of anti-Aβ monoclonal antibodies against cytotoxicity; the abscissa represents different sample groups, the ordinate represents relative value of LDH release, the antibodies 066-4.26.14, 066-5.4.1, 066-6.1.1, 066-6.1.3, 066-6.2.1, 066-6.7.2, 066-7.17.2 all have protective effect against cytotoxicity, and their protective effect is equivalent to that of 066-P02; in which, Fig. 5(A) shows the protective activity detection results of sample groups Vehicle, IgG, anti-Aβ monoclonal antibodies 066-4.21.13, 066-4.17.28, 066-4.6.8, 066-4.22.1, 066-4.26.14, 066-4.18.2, 066-P02 against cytotoxicity, respectively; Fig. 5(B) shows the protective activity detection results of sample groups Vehicle, IgG, anti-Aβ monoclonal antibodies 066-6.2.1, 066-7.17.2, 066-5.4.1, 066-6.1.1, 066-6.1.3, 066-6.7.2, 066-P02 against cytotoxicity, respectively;

Fig. 6 shows the detection results of the Morris water maze experiment; in which the abscissa represents time after treatment (Days 1, 2, 3, 4 and 5), and the ordinate represents time to find hidden platform under water surface (unit: Sec);

Fig. 7 shows the total RNA agarose gel electrophoresis detection results; in which lane M: DL2000 molecular weight marker; lane 1: total RNA of 066-4.26.14;

Fig. 8 shows the agarose gel electrophoresis detection results of the heavy chain variable region and light chain variable region PCR products of the candidate antibody; lane M: DL2000

molecular weight marker; lane 1: PCR product of 066-4.26.14 heavy chain variable region; lane 2: PCR product of 066-4.26.14 light chain variable region; in which, Fig. 8(A) shows the PCR results of 066-4.26.14 heavy chain variable region; Fig. 8(B) shows the PCR results of 066-4.26.14 light chain variable region;

Fig. 9 shows the SDS-PAGE detection results of the purified mouse-human chimeric antibody 066-4.26.14-chAb; lane M: protein molecular weight marker; lane 1: mouse-human chimeric antibody 066-4.26.14-chAb (non-reducing); lane 2: mouse-human chimeric antibody 066-4.26.14-chAb (reducing); Fig. 9(A) shows the non-reducing electrophoresis results of mouse-human chimeric antibody 066-4.26.14-chAb; Fig. 9(B) shows the reducing electrophoresis results of mouse-human chimeric antibody 066-4.26.14-chAb;

Fig. 10 shows the SDS-PAGE detection results of the purified humanized candidate antibodies; lane M: protein molecular weight marker; in Fig. 10(A), lanes 1 to 6 are humanized antibodies 066-4.26.14H2L2, 066-4.26.14H2L3, 066-4.26.14H4L2, 066-4.26.14H5L1, 066-4.26.14H5L2, 066-4.26.14H5L3 (non-reducing), respectively; in Fig. 10(B), lanes 1 to 6 are humanized antibodies 066-4.26.14H2L2, 066-4.26.14H2L3, 066-4.26.14H4L2, 066-4.26.14H5L1, 066-4.26.14H5L2, 066-4.26.14H5L3 (reducing), respectively;

Fig. 11 shows the detection results of humanized $A\beta$ antibodies in inhibiting $A\beta$ polymerization; the abscissa represents different sample groups, and the ordinate represents relative fluorescence intensity; the humanized antibodies of 066-4.26.14 all can inhibit $A\beta$ polymerization;

Fig. 12 shows the protective activity detection results of humanized A β antibodies against cytotoxicity; the abscissa represents different sample groups, and the ordinate represents relative value of LDH release, the humanized candidate antibodies of 066-4.26.14 all have protective effect against cytotoxicity, and the protective effect is equivalent to that of 066-P02, in which 066-4.26.14H5L2 shows the best performance.

Specific Models for Carrying Out the present disclosure

The present disclosure discloses a humanized anti-A β monoclonal antibody and use thereof, and those skilled in the art can fulfill them by learning the contents of the present disclosure and appropriately improving the process parameters. In particular, it should be pointed out that all similar substitutions and modifications are obvious to those skilled in the art, and they are all deemed to be included in the present disclosure. The methods and use of the present disclosure have been described through the preferred examples, and it is obvious that those skilled in the art can make changes or appropriate alternations and combinations to the methods and use described

herein without departing from the content, spirit and scope of the present disclosure, so as to achieve and apply the technology of the present disclosure.

The humanized anti-A β monoclonal antibodies provided by the present disclosure and the raw materials and reagents used in the use were all commercially available.

The present disclosure is further illustrated in conjunction with the following examples:

Example 1: Preparation of AB antigen and positive control antibody

Preparation of A β monomer and polymer mixture

 $A\beta_{1-42}$, $A\beta_{1-16}$, and $A\beta_{14-29}$ polypeptides were synthesized by Ji'er Biochemical (Shanghai) Co., Ltd. The amino acid sequence of $A\beta_{1-42}$ polypeptide was: DAEFRHDSGYEVHHQKLVFFAEDVGSNKGAIIGLMVGGVVIA (SEQ ID: 35), the amino acid sequence of $A\beta_{1-16}$ polypeptide was: DAEFRHDSGYEVHHQK (SEQ ID: 36), and the amino acid sequence of $A\beta_{14-29}$ polypeptide was: HQKLVFFAEDVGSNKGA (SEQ ID: 37).

Preparation method of $A\beta_{1-42}$ monomer (abbreviated as $A\beta$ monomer): 1 ml of hexafluoro isopropanol (HFIP) was added to 1 mg of $A\beta_{1-42}$ polypeptide dry powder, subjected to vortex and shaking for 1 min, and sonicated in a water bath for 1-5 min until the dissolution was completed; placed in 37°C, 200 rpm shaking incubator and shaken for 1.5 hours; a vacuum rotary dryer was used to volatilize hexafluoro isopropanol; 192 μ l of anhydrous dimethyl sulfoxide (DMSO) was added to the dried $A\beta_{1-42}$ polypeptide to dissolve the polypeptide, then added with 27 μ l of 20× PBS solution, 54 μ l of 2% SDS, 267 μ l of ddH₂O, mixed well, subpackaged in small amounts, stored in a refrigerator at -80°C, which was the $A\beta_{1-42}$ monomer; and the detection thereof was performed by SDS-PAGE and WB (hybridization detection was performed by using the positive antibody 066-P02 that specifically recognizes $A\beta_{1-42}$) (see Fig. 1).

Preparation method of $A\beta_{1-42}$ polymer mixture (abbreviated as $A\beta$ polymer mixture): 1 ml of hexafluoro isopropanol (HFIP) was added to 1 mg of $A\beta_{1-42}$ polypeptide dry powder, subjected to vortex and shaking for 1 min, sonicated in a water bath for 1-5 min until the dissolution was completed; placed in 37°C, 200 rpm shaking incubator and shaken for 1.5 h; a vacuum rotary dryer was used to volatilize hexafluoro isopropanol; 192 μ l of DMSO was added to the dry $A\beta_{1-42}$ polypeptide to dissolve the polypeptide, then added with 27 μ l of 20× PBS solution, 54 μ l of 2% SDS, 267 μ l of ddH₂O, mixed well, and placed in a 37°C water bath for 18-24 h; added with 1.62 ml of ddH₂O, mixed well, and placed in a 37°C water bath for 18-24 h; transferred into PBS by using 10 KDa ultrafiltration tube for buffer replacement, subpackaged in small amounts, stored in a refrigerator at -80°C, which was the $A\beta_{1-42}$ polymer mixture; and the detection thereof was performed by SDS-PAGE and WB (hybridization detection was performed using the positive antibody 066-P02 that specifically recognizes $A\beta_{1-42}$) (see Fig. 1).

2. Construction of positive control antibody expression vector

pGS003-hIgG1CH and pGS003-hIgKCL were separately selected as the expression vectors for constructing the heavy chain and the light chain of anti-human Aβ-positive antibodies (066-P01: Solanezumab, Eli lily; 066-P02: Aducanumab, Biogen); after the codon optimization of the amino acid sequences of the positive antibody variable regions, the positive antibody VH and VL genes were separately cloned into pGS003-hIgG1CH and pGS003-hIgKCL using restriction enzyme digestion method to obtain transient transfection expression vectors pGS003-066-P01VH-hIgG1CH, pGS003-066-P01VL-hIgKCL, pGS003-066-P02VH-hIgG1CH and pGS003-066-P02VL-hIgKCL of the heavy chain and the light chain of the positive antibody . The amino acid sequence of the heavy chain variable region of the positive antibody 066-P01 was as follows (SEQ ID: 38):

EVQLVESGGGLVQPGGSLRLSCAASGFTFSRYSMSWVRQAPGKGLELVAQINSVGN STYYPDTVKGRFTISRDNAKNTLYLQMNSLRAEDTAVYYCASGDYWGQGTLVTVSS

The amino acid sequence of the light chain variable region of the positive antibody 066-P01 was as follows (SEQ ID: 39):

DVVMTQSPLSLPVTLGQPASISCRSSQSLIYSDGNAYLHWFLQKPGQSPRLLIYKVSN RFSGVPDRFSGSGSGTDFTLKISRVEAEDVGVYYCSQSTHVPWTFGQGTKVEIK

The amino acid sequence of the heavy chain variable region of the positive antibody 066-P02 was as follows (SEQ ID: 40):

QVQLVESGGGVVQPGRSLRLSCAASGFAFSSYGMHWVRQAPGKGLEWVAVIWFD GTKKYYTDSVKGRFTISRDNSKNTLYLQMNTLRAEDTAVYYCARDRGIGARRGPYYM DVWGKGTTVTVSS

The amino acid sequence of the light chain variable region of the positive antibody 066-P02 was as follows (SEQ ID: 41):

DIQMTQSPSSLSASVGDRVTITCRASQSISSYLNWYQQKPGKAPKLLIYAASSLQSGV PSRFSGSGSGTDFTLTISSLQPEDFATYYCQQSYSTPLTFGGGTKVEIK

3. Expression by transient transfection

pGS003-066-P01VH-hIgG1CH and pGS003-066-P01VL-hIgKCL;

pGS003-066-P02VH-hIgG1CH and pGS003-066-P02VL-hIgKCL were transiently expressed.

FreeStyleTM 293E cells were used for expression by transient transfection in Freestyle medium. Twenty-four hours before transfection, 30 ml of 293E cells were inoculated at 0.5×10⁶ cells/ml in a 125 ml conical flask, and cultured on a shaker at 130 rpm in a 37°C, 5% CO₂ incubator. During

transfection, 60 µl of 293E Fectin was firstly taken and added to 1 ml of Opti-MEM, mixed well, and incubated at room temperature for 5 minutes; meanwhile, Total 30 µg plasmid DNA of transient transfection expression vectors (recombinant vectors) was dissolved in 1 ml of Opti-MEM. Then, the plasmid DNA and 293E Fectin were mixed thoroughly, with a total volume of 2 ml, incubated at room temperature for 15 minutes, and then all the mixture was added to the cell culture wells, mixed, and incubated on a shaker in a 37°C, 5% CO₂ incubator at 130 rpm for 7 days. The culture broth was centrifuged at a high speed and the supernatant was taken and subjected to vacuum filtration with a microporous membrane.

4. Purification of protein

According to the operating method provided by the manufacturer, Protein A column (protein purification liquid chromatography system/AKTA Purifier 10, GE) and nickel column were used for purification to obtain purified positive antibodies 066-P01 and 066-P02. As shown in Fig. 2.

Example 2: Preparation of anti-Aß monoclonal hybridoma

Immunization of BALB/c mice

 $A\beta_{1-42}$ polypeptide antigen and Freund's complete adjuvant were vortexed and mixed according to their doses, after emulsification was completed, first immunization was performed to 6-week-old BALB/c female mice. Each mouse was injected intraperitoneally with 200 µg of antigen, in total 3 groups of mice were immunized, 5 mice in each group. Two weeks after the first immunization, the mice were given second intraperitoneal immunization, in which Freund's incomplete adjuvant was used, while the dose of immune antigen was the same as the first immunization. After that, the mice were immunized intraperitoneally twice a month, and the adjuvant and antigen doses were the same as the second immunization.

After the first immunization, a small amount of blood was collected from mouse orbit and serum titer was tested every six weeks. After the serum titer reached 1:200000 or above by the indirect ELISA method, the mice used for fusion were subjected to booster immunization.

Preparation of myeloma cells for fusion

Myeloma cells P3X63Ag8.653 used for fusion were resuscitated three weeks in advance, cultured in DMEM medium containing 1×8-azaguanine and 10% fetal bovine serum for two weeks, and cultured with DMEM containing 10% fetal bovine serum before one week of fusion, in which the density of P3X63Ag8.653 was maintained at 70% to 80% until the day of fusion.

Cell fusion and HA screening

Obtaining and preparation of spleen cells: 2 mice after booster immunization were taken, sacrificed after collection of immune serum, and soaked in 75% alcohol for 2-3 minutes. The skin

and peritoneum on the abdomen side of the immunized mice were cut to expose spleen. The spleen was obtained by removing the surrounding tissues with scissor tip, ground with a grinding rod, and filtered through a cell sieve to prepare a single cell suspension. The supernatant was discarded after centrifugation.

Treatment before cell fusion: P3X63Ag8.653 in the culture flask was collected, centrifuged at 1000 rpm/5 min, then the supernatant was discarded, the cells were resuspended, and the live myeloma cells were counted. The spleen cell suspension was centrifuged to discard the supernatant, added with ACK lysate, incubated and centrifuged to discard supernatant to remove red blood cells, resuspended in DMED, and the viable spleen cells were counted.

Cell fusion: The cells were mixed at the ratio of spleen cells:P3X63Ag8.653 = 1:2, centrifuged at 2000 rpm/5 min to discard the supernatant, shaken to disperse cell pellets, added with 1 mg/ml Pronase at $400 \,\mu\text{l}/1\times10^8$ spleen cells; after incubating for 15 seconds, 10 ml of fetal bovine serum was added to stop the reaction, electroporation solution (ECF) was supplemented to 50 ml, centrifuged at 2500 rpm for 5 minutes to discard the supernatant, resuspended with ECF and the viable cells were counted, and the spleen cell density was adjusted to $2\times10^6/\text{ml}$. The cell suspension with the well-adjusted density was added to an electrofusion tank, and an electroporator was run for cell fusion. After the fusion, the cell suspension was transferred from the fusion tank to $1/2 \, \text{HA}$ medium, allowed to stand for 3 hours and then cell plating was carried out.

HA medium selection: AT selection medium containing 1/2 HA, 1× penicillin-streptomycin, 20% fetal bovine serum and 80% DMEM medium was prepared. The mouse hybridoma cells were resuspended in the above 1/2 HA selection medium and mixed well. The cell suspension was added to a 96-well cell culture plate at 200 μl/well, 1×10⁶ spleen cells/plate, placed in a cell incubator and cultured at 37°C. After 1 week of culture, the 1/2 HA medium was used for the first renewing of the medium, and the culture was continued in 37°C cell incubator. After 3 days of culture, the 1/2 HA medium was used for the second renewing of the medium.

Screening of positive cell lines

Two weeks after the fusion, the cell supernatant was taken and subject to ELISA experiment to detect the binding of the cell supernatant to human $A\beta_{1-42}$, and after the cells with positive ELISA result were screened out, the second ELISA experiment was retested. The cell supernatant with positive retested results was taken for subcloning and expansion culture.

Expansion culture

The cell lines with positive ELISA test result were transferred from the 96 well-plate to a 24 well-plate and cultured, after the cells grew all over the plate, and they were transferred to a 25 cm² culture flask and cultured.

Subcloning by limiting dilution method

The positive cell lines were mixed well by beating and pipetting, and a small amount thereof was pipetted to count the viable cells. About 200 cells were pipetted and added to 80 ml of complete medium and mixed well, and plated on 4 plates. In addition, about 400 cells were pipetted and added to 80 ml of complete medium and mixed well, and plated on 4 plates. In addition, about 1000 cells were pipetted and added to 20 ml of complete medium and mixed well, and plated on 1 plate. A total of 9 plates were plated at 3 different cell densities, respectively 0.5 cells/well, 1 cell/well, and 10 cells/well. The 96-well plates were plated in a 37°C, 5% CO₂ incubator for culture.

Clone detection and expansion culture

The supernatants of the monoclonal cell wells were taken for ELISA to detect the binding of the cloned antibody to the full length of $A\beta_{1-42}$ as well as the N-terminal, C-terminal and middle peptide fragments of $A\beta_{1-42}$, respectively.

Coating: Streptavidin was diluted with CBS (pH 9.6) to 1 μ g/ml, added to 96-well microtiter plate, 50μ l per well, incubated overnight at 2-8°C.

Blocking: After washing the plate once with PBST, it was blocked with 1% BSA, 200 µl per well, and incubated for 1 hour at room temperature.

Antigen: After washing the plate three times with PBST, the biotinylated $A\beta_{1-42}$, $A\beta_{1-16}$, and $A\beta_{14-29}$ polypeptides were taken respectively, diluted with PBS (pH 7.2) to 1 μ g/ml, and added to enzyme-labeled 96 well-plate, 50 μ l per well, and incubated for 1 hour at room temperature.

Addition of primary antibody: After washing the plate three times with PBST, mouse candidate antibody was added, 50 µl/well, and incubated at room temperature for 2 hours.

Addition of secondary antibody: After washing the plate three times with PBST, anti-mouse IgG Fc-HRP antibody in 1:5000 diluent was added, 50 μ l/well, and incubated for 1 hour at room temperature.

Color development: After washing the plate six times with PBST, TMB color development solution was added, 50 µl per well, and developed in the dark for 10 minutes at room temperature.

Stop: a stop solution was directly added to stop the reaction, 50 µl per well.

Detection: After stopping the reaction, the microtiter plate was immediately placed into a microplate reader, the OD value was measured at 450 nm, and the original data was stored.

Data processing: The raw data were input into the software SoftMax Pro 6.2.1 for data processing. See Table 1 for specific data. The results showed that the 12 murine candidate antibodies contained three different antigen binding epitopes, namely N-terminal ($A\beta_{1-16}$), C-

terminal ($A\beta_{30-42}$), and middle ($A\beta_{14-29}$) peptide fragments, in which, the antigen binding epitope of 066-4.26.14 was grouped into $A\beta_{1-42}$ C-terminal peptide fragment (because 066-4.26.14 could bind to the full length of $A\beta_{1-42}$, but did not bind to $A\beta_{1-16}$, $A\beta_{14-29}$, it was deduced that it bound to $A\beta_{30-42}$ region).

The cell lines with positive ELISA result were transferred from the 96 well-plate to a 24 well-plate for culture, and the cells grew over the plate, they were transferred to a 25 cm² culture flask and cultured.

Table 1: Grouping detection results of murine candidate antibody antigen binding epitopes

Antibody name	Full length Aβ ₁₋₄₂	Aβ ₁₋₁₆ (N-terminal peptide fragment)	Aβ ₁₄₋₂₉ (middle peptide fragment)	Epitope
066-4.6.8	1.2944	0.0561	0.5115	Middle peptide fragment
066-4.17.28	1.4540	0.0690	1.1238	Middle peptide fragment
066-4.18.2	1.0925	1.2210	0.0848	N-terminal peptide fragment
066-4.21.13	1.3647	0.056	1.1457	Middle peptide fragment
066-4.22.1	1.2517	0.0773	1.0401	Middle peptide fragment
066-4.26.14	1.9312	0.0602	0.0674	C-terminal peptide fragment
066-5.4.1	1.2483	0.0540	1.1566	Middle peptide fragment
066-6.1.1	1.3752	0.1001	0.1180	C-terminal peptide fragment
066-6.1.3	1.3665	0.0618	0.0803	C-terminal peptide fragment
066-6.2.1	1.3085	0.0975	0.1003	C-terminal peptide fragment
066-6.7.2	1.4707	1.6608	0.3058	N-terminal peptide fragment
066-7.17.2	1.0833	0.9197	0.0612	N-terminal peptide fragment

Identification of subtypes

Goat anti-mouse IgG1, IgG2a, IgG2b, IgG2c, IgG3, IgM and IgGA were coated, $50 \text{ng}/100 \mu\text{l/well}$, 4°C overnight, blocked with BSA at room temperature, the cell supernatant to be tested was added, incubated at room temperature for 2 hours, added with enzyme-labeled secondary antibody goat anti-mouse IgG, κ , λ , after color development, stopping, and 450 nm reading, it was judged that the tested cell line was subtypes IgG1, κ or IgG2a, κ or IgG2b, κ . The results are shown in Table 2, in which for the antibody 066-4.26.14, its heavy chain constant region

was murine IgG2a, and its light chain constant region was the constant region of the murine κ chain.

Table 2: Detection results of mouse candidate subtypes

Antibody name	Subtype
066-4.6.8	IgG1, Kappa
066-4.17.28	IgG1, Kappa
066-4.18.2	IgG2b, Kappa
066-4.21.13	IgG1, Kappa
066-4.22.1	IgG1, Kappa
066-4.26.14	IgG2a, Kappa
066-5.4.1	IgG1, Kappa
066-6.1.1	IgG2a, Kappa
066-6.1.3	IgG2a, Kappa
066-6.2.1	IgG2a, Kappa
066-6.7.2	IgG2a, Kappa
066-7.17.2	IgG1, Kappa

Cell cryopreservation

Preparation of cryopreservation solution: 90% fetal bovine serum, 10% DMSO.

The cells in the culture flask were resuspended; after the cell counting, the cells were centrifuged at 1000 rpm/min for 5 min, the supernatant was discarded, and the suspension was beaten by pipetting with fetal bovine serum containing 10% DMSO, stored at 5×10^6 cells/tube in a cryopreservation box at -80° C overnight, and transferred into liquid nitrogen on the next day.

Preservation of monoclonal hybridoma gene

Positive monoclonal cell lines were collected, added with TRizol to lyse the cells and extract RNA, which was reverse-transcribed into cDNA, and stored at -80°C.

Preparation of antibodies by in vitro culture method

The prepared hybridoma cell lines were resuscitated by a method as follows. The hybridoma cell lines were resuscitated in a DMEM medium containing 10% fetal bovine serum and 1% penicillin streptomycin, and cultured in a vial; after the cell confluence was about 90%, passage expansion was performed, the expansion was performed until the cell culture supernatant in total was about 200 ml, then the supernatant was collected, centrifuged and filtered for purification.

Example 3: Detection of anti-Aß monoclonal antibody in inhibiting Aß polymerization

8.2% DMSO/DPBS solution (DMSO: sigma; DPBS: Hyclone) was used to dissolve Aβ dry powder to 1 mg/ml, the Aβ solution was diluted with DPBS to 33 μg/ml, the anti-Aβ monoclonal antibodies 066-4.6.8, 066-4.18.2, 066-4.22.1, 066-4.26.14, 066-5.4.1, 066-6.1.1, 066-6.1.3, 066-6.2.1, 066-6.7.2 were diluted to 450 µg/ml (IC100), and ThT (sigma) was diluted with ultrapure water to 20 µM. 50 µl of antibody diluent was taken and added to a 96-well black plate (corning), then added with 50 µl of Aβ diluent, finally added with 100 µl of ThT, incubated for 24 hours at room temperature in the dark, and the fluorescence intensity (Ex/Em =440/485) was detected with a multifunctional microplate reader. The abscissa represented different sample groups, and the ordinate represented relative fluorescence intensity. The results are shown in Fig. 3. In Fig. 3(A), when the relative fluorescence intensity of the IgG group was 1.0, the relative fluorescence intensity of the anti-Aβ monoclonal antibody 066-5.4.1 group was 0.59; in Fig. 3(B), when the relative fluorescence intensity of the PBS group was 1.0, the relative fluorescence intensity of the anti-Aß monoclonal antibody 066-4.22.1 group was 0.44, and the relative fluorescence intensity of the anti-Aβ monoclonal antibody 066-4.26.14 group was 0.46; it could be seen that the anti-Aβ monoclonal antibodies such as 066-4.22.1, 066-4.26.14 and 066-5.4.1 all could inhibit Aβ polymerization.

Example 4: Detection of activity of anti-A β monoclonal antibody in promoting macrophage phagocytosis of A β

Mouse primary peritoneal macrophages that were in good condition after 3 days of adherent culture were digested with 0.25% trypsin and counted. The cell density was adjusted to 2×10⁵/ml with DMEM medium (Gibco) containing 10% fetal bovine serum and the cells were inoculated on a 96-well cell culture plate, 100 μl/well; the anti-Aβ monoclonal antibodies 066-4.6.8, 066-4.17.28, 066-4.18.2, 066-4.21.13, 066-4.22.1, 066-4.26.14, 066-5.4.1, 066-6.1.1, 066-6.1.3, 066-6.2.1, 066-6.7.2, 066-7.17.2 were diluted with DMEM medium containing 1% fetal bovine serum to 20 μg/ml and used as working solutions, Aβ was diluted to 240 μg/ml, and ThT (sigma) was diluted to 20 µM with ultrapure water. The culture medium in the culture plate was discarded, 50 µl of antibody diluent was first added, then added with 50 µl of Aβ diluent, multiple wells were set; incubation was performed in a 37°C, 5% CO₂ incubator for 6 hours; 50 µl of supernatant was taken and add to a 96-well black plate, then added with 50 µl of ThT, and the fluorescence intensity (Ex/Em=440/485) was detected with a multifunctional microplate reader. The abscissa represented different sample groups, and the ordinate represented fluorescence intensity. The results are shown in Fig. 4. Among them, the fluorescence intensity of the anti-Aβ monoclonal antibody 066-5.4.1 group was 650,000, and the fluorescence intensity of the anti-A\beta monoclonal antibody 066-7.17.2 group was 600,000. It could be seen that the anti-Aβ monoclonal antibodies 066-5.4.1, 066-7.17.2 had the activity of promoting the phagocytosis of Aβ by macrophages.

Example 5: Detection of protective activity of anti-A β monoclonal antibody against cytotoxicity

Logarithmic growth phase SHSY5Y cells were digested with 0.25% trypsin, counted, adjusted with EMEM medium (ATCC) containing 10% fetal calf serum to a cell density of 3×10⁴/ml, inoculated on a 96-well cell culture plate, 100 µl/well; the anti-Aß monoclonal antibodies 066-4.6.8, 066-4.17.28, 066-4.18.2, 066-4.21.13, 066-4.22.1, 066-4.26.14, 066-5.4.1, 066-6.1.1, 066-6.1.3, 066-6.2.1, 066-6.7.2, 066-7.17.2 were diluted with EMEM medium containing 1% fetal bovine serum to 200 μg/ml (IC100), and used as working solution, Aβ was diluted to 240 μg/ml. The culture medium in the culture plate was discarded, 50 µl of antibody diluent was added, then added with 50 µl of Aβ diluent, multiple wells were set; incubation was performed in a 37°C, 5% CO₂ incubator for 48 hours; 50 ul of the supernatant was taken and added to a new 96 well-plate, then added with 50 µl of LDH assay buffer, reacted in the dark at room temperature for 30 minutes, added with 50 µl of stop solution, and the absorbance value was measured with a multifunctional microplate reader. The abscissa represented different sample groups, and the ordinate represented relative value of LDH release. The results are shown in Fig. 5. Among them, when the relative value of LDH release of the Vehicle group was 1.0, the relative fluorescence intensity of the anti-Aβ monoclonal antibody 066-4.26.14 group was 1.2, the relative value of LDH release of the anti-Aβ monoclonal antibody 066-5.4.1 group was 1.37, the relative value of LDH release of the anti-Aβ monoclonal antibody 066-6.1.1 group was 1.43, the relative value of LDH release of the anti-Aß monoclonal antibody 066-6.1.3 group was 1.35, the relative value of LDH release of the anti-Aβ monoclonal antibody 066-6.2.1 group was 1.34, the relative value of LDH release of the anti-Aβ monoclonal antibody 066-6.7.2 group was 1.44, the relative value of LDH release of the positive control antibody 066-P02 group was 1.26 (A) and 1.53 (B). It could be seen that the antibodies 066-4.26.14, 066-5.4.1, 066-6.1.1, 066-6.1.3, 066-6.2.1, 066-6.7.2, 066-7.17.2 all had protective effect against cytotoxicity, and the protective effect was equivalent to that of 066-P02.

Example 6: Morris water maze experiment

1. Experimental method and steps:

Experimental animals 3×Tg mice were purchased from Beijing Weitong Lihua Experimental Animal Technology Co., Ltd. and raised by the Experimental Animal Center of Medical College of Jilin University. The grouping situation was as follows:

according to the different drugs to be injected, they were divided into antibody 066-4.26.14 treatment group, antibody 066-5.4.1 treatment group, antibody 066-7.17.2 treatment group, 3×Tg blank control group, wild-type PBS injection group, positive antibody 066- P02 control group, 8 animals in each group.

By referring to the literature method (Nabeshima, 2007), 6-month-old male 3×Tg mice (500 g/mouse) were intraperitoneally injected with monoclonal antibodies 066-4.26.14, 066-5.4.1, 066-7.17.2, once per week, continuously injected for 10 weeks, and Morris water maze test was performed 8 weeks after injection.

Morris water maze test steps:

- (1) The specially designed water maze was mainly composed of a cylindrical pool and a movable platform. The pool had a height of 45 cm and a diameter of 100 cm, the platform had a diameter of 9 cm and an adjustable height from 15 to 40 cm, and digital camera was mounted above the pool and connected to a computer.
- (2) Clean water was filled into the pool in advance. The walls and bottom of the pool were all black. White pigment for food was added to the pool water to prevent mice from seeing the platform under the water surface. The water depth was 30 cm and the water surface was 1 cm higher than the platform.
- (3) The water temperature was controlled at 19±1°C, and except the quadrant where the platform was located, other quadrants on the pool were marked with points for entering water. On the sidewalls corresponding to each quadrant, markers of different shapes were adhered. The position of the platform was unchanged during the experiment.
- (4) Each test was carried out in a soundproof room, and the positions of laboratory objects such as the pool, light sources, and cages remained unchanged.
- (5) In the 8th week, training was started on the 3rd day after the administration. The experiment lasted for 5 days (water maze-hidden platform test), 4 times a day. When the mouse entered the water, it faced the wall of the pool and was gently put into the water. Five training sessions (experiments) a day were conducted randomly in areas other than the quadrant where the platform was located, and the first two training sessions in the first two days of the experiment were performed as exercises. If the mouse found the platform within 60 seconds, it was allowed to stay on the platform for 15 seconds. If the mouse could not find the platform within 60 seconds (the latent period was recorded as 60 seconds), the experimenter would guide it to the platform and stay on the platform for 15 seconds. The average of four latent periods of the mouse was taken as the daily performance of the mouse.

2. Experimental results (see Fig. 6):

On the second day of Hidden platform test, the time to find the hidden platform under the water (escape latent period) was significantly shortened for all antibody administration groups compared with the 3×Tg blank group, which was statistically significant. On day 3, the anti-Aβ

monoclonal antibody 066-7.17.2 group took 18 s for escape latent period, the anti-A β monoclonal antibody 066-4.26.14 group took 27 s for escape latent period, the anti-A β monoclonal antibody 066-5.4.1 group took 30 s for escape latent period, and the blank control group took 39 s for escape latent period, in which the 066-7.17.2 and 066-4.26.14 administration groups had a significantly shorter escape latent period as compared with the 3×Tg blank group, and there was statistical significance. It could be seen that the anti-A β monoclonal antibodies 066-7.17.2 and 066-4.26.14 had a certain effect on improving the cognitive learning and memory ability in Alzheimer's dementia model mice.

Example 7: Monoclonal antibody gene sequencing and chimeric antibody preparation

1. Monoclonal antibody gene sequencing

After immunization, fusion and monoclonalization, based on the experimental results of binding epitope, detection of inhibiting A β polymerization, detection of protective activity against cytotoxicity, Morris water maze, etc., the 066-4.26.14 monoclonal antibody cell line was selected for total RNA extraction which was reverse-transcribed into cDNA, and then the cDNA was used as a template for PCR amplification of the heavy chain variable region and light chain variable region of the antibody.

The TRIzol reagent kit (15596-026) of Invitrogen was used, and the total RNA was extracted from the 066-4.26.14 monoclonal antibody cell line according to its instructions. The results are shown in Fig. 7.

The 5'RACE FULL kit (D315) of Takara was then used, the total RNA was reverse-transcribed into the first strand cDNA using the random primers in the kit, and then the PCR amplification of heavy chain was performed using the constant region primer mIgGR (5'-CTCAGGGAARTARCCYTTGAC-3', SEQ ID NO: 42) and the RACE primer in the kit, and the PCR amplification of light chain was performed using the constant region primer mIgKR (5'-TCACTGCCATCAATCTTCCAC-3', SEQ ID NO: 43) and the RACE primer in the kit. The results are shown in Fig. 8.

The PCR fragments were recovered by the agarose gel recovery kit and subjected to TA cloning, and then single clones were picked up for colony PCR. The colony PCR primers were M13F (5'-TGTAAAACGACGGCCAGT-3', SEQ ID NO: 44) and M13R (5'-CAGGAAACAGCTATGACC-3', SEQ ID NO: 45). Part of the samples selected from the correct strains upon the identification were sent to Invitrogen for sequencing. It was finally determined that the nucleotide sequence of the heavy chain variable region was SEQ ID NO: 46, the nucleotide sequence of the light chain variable region was SEQ ID NO: 47, the amino acid sequence of the

heavy chain variable region was SEQ ID NO: 48, and the amino acid sequence of the light chain variable region was SEQ ID NO: 49, see Table 3.

Table 3: Specific sequences of heavy chain variable region and light chain variable region of 066-4.26.14 antibody

	Nucleotide	sequence	Amino acid sequence	
Antibody	Heavy chain variable region	Light chain variable region	Heavy chain variable region	Light chain variable region
066-4.26.14	SEQ ID NO:46	SEQ ID NO:47	SEQ ID NO:48	SEQ ID NO:49

The nucleotide sequence of the heavy chain variable region of the 066-4.26.14 antibody was as follows (SEQ ID NO: 46):

GAAGTGAAGCTGGTGGAGTCTGGGGGGAGGCTTAGTGAAGCCTGGAGGGTCCCT
GAAACTCTCCTGTGCAGCCTCTGGATTCACTTTCAGAAGTTATGCCATGTCTTGGGTT
CGCCAGACTCCAGAGAAGAGGCTGGAGTGGGTCGCATCCATTAGTACTACTAGTAA
CACCTACTATCCAGACAGTGTGAAGGGCCGATTCACCACCTCCAGAGATAACGCCA
GGAACATCGTGTACCTGCAAATGAGCAGTCTGAGGTCTGACGACACGGCCATGTAT
TACTGTGGAAGAGGCGTGATTACGAACCAGGCCTGGTTTGCTTACTGGGGCCAAGG
GACTCTGGTCACTGTCTCTGCA

The nucleotide sequence of the light chain variable region of the 066-4.26.14 antibody was as follows (SEQ ID NO: 47):

GATATTGTGCTAACTCAGTCTCCAGCCACCCTGTCTGTGACTCCAGGAGATAGCG
TCAGTCTTTCCTGCAGGGCCAGCCAAAGTATTAGCAACAACCTACACTGGTATCAGC
AAAAATCACATGAGTCTCCAAGGCTTCTCATCAAGTATGCTTCCCAGTCCATCTCTG
GGATCCCCTCCAGGTTCAGTGGCAGTGGATCAGGGACAGATTTCACTCTCAGTGTCA
ACAATGTGGGGACTGAAGATTTTGGAATGTATTTCTGTCAACAGAGTAACAGCTGGC
CGCTCACGTTCGGTGCTGGGACCAAGCTGGAGCTGAAA

The amino acid sequence of the heavy chain variable region of the antibody 066-4.26.14 was as follows (SEQ ID NO: 48):

EVKLVESGGGLVKPGGSLKLSCAASGFTFRSYAMSWVRQTPEKRLEWVASISTTSN TYYPDSVKGRFTTSRDNARNIVYLQMSSLRSDDTAMYYCGRGVITNQAWFAYWGQGT LVTVSA

The amino acid sequence of the light chain variable region of the antibody 066-4.26.14 was as follows (SEQ ID NO: 49):

DIVLTQSPATLSVTPGDSVSLSCRASQSISNNLHWYQQKSHESPRLLIKYASQSISGIP SRFSGSGSGTDFTLSVNNVGTEDFGMYFCQQSNSWPLTFGAGTKLELK

2. 066-4.26.14 mouse-human chimeric antibody expression vector construction

The pGS003-hIgG1CH and pGS003-hIgKCL were selected as the expression vectors for constructing the heavy chain and the light chain of the anti-human Aβ mouse-human chimeric antibody, respectively. Using the synthesized 066-4.26.14 mouse antibody sequence as a template, the VH and the VL mouse antibody genes were PCR amplified and cloned into pGS003-hIgG1CH and pGS003-hIgKCL using restriction enzyme digestion and ligation methods to obtain the transient transfection expression vectors pGS003-066-4.26.14-chAbVH-hIgG1CH and pGS003-066-4.26.14-chAbVH-hIgG1CH of the mouse-human chimeric antibody.

The amino acid sequence of the heavy chain of the 066-4.26.14 mouse-human chimeric antibody was as follows (SEQ ID NO: 50):

EVKLVESGGGLVKPGGSLKLSCAASGFTFRSYAMSWVRQTPEKRLEWVASISTTSN
TYYPDSVKGRFTTSRDNARNIVYLQMSSLRSDDTAMYYCGRGVITNQAWFAYWGQGT
LVTVSAASTKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFP
AVLQSSGLYSLSSVVTVPSSSLGTQTYICNVNHKPSNTKVDKRVEPKSCDKTHTCPPCPA
PELLGGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAKTK
PREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQPREPQV
YTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFLYS
KLTVDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPG

The amino acid sequence of the light chain of the 066-4.26.14 mouse-human chimeric antibody was as follows (SEQ ID NO: 51):

DIVLTQSPATLSVTPGDSVSLSCRASQSISNNLHWYQQKSHESPRLLIKYASQSISGIP SRFSGSGSGTDFTLSVNNVGTEDFGMYFCQQSNSWPLTFGAGTKLELKRTVAAPSVFIFP PSDEQLKSGTASVVCLLNNFYPREAKVQWKVDNALQSGNSQESVTEQDSKDSTYSLSST LTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGEC

3. Expression by transient transfection

pGS003-066-4.26.14-chAbVH-hIgG1CH and pGS003-066-4.26.14-chAbVL-hIgKCL were subjected to transient expression.

FreeStyleTM 293E cells were used and subjected to transient transfection expression in Freestyle medium. Twenty-four hours before transfection, 30 ml of 293E cells were inoculated at 0.5×10⁶ cells/ml in a 125 ml conical flask, and cultured on a shaker at 130 rpm in a 37°C, 5% CO₂ incubator. During transfection, 60 μl of 293E Fectin was first taken and added to 1 ml of Opti-MEM, mixed well, and incubated at room temperature for 5 minutes; meanwhile, the total plasmid DNA of the recombinant vector in an amount of 30 μg was dissolved in 1 ml of Opti-MEM. Then,

the plasmid DNA and 293E Fectin were mixed thoroughly, with a total volume of 2 ml, incubated at room temperature for 15 minutes, and then the whole mixture was added to the cell culture wells, mixed, and incubated in a 37°C, 5% CO₂ incubator on a shaker at 130 rpm for 7 days. The culture broth was centrifuged at a high speed and the supernatant was taken for vacuum filtration with a microporous membrane.

4. Purification of protein

According to the operating method provided by the manufacturer, Protein A column (Protein Purification Liquid Chromatography System/AKTA Purifier 10, GE) and nickel column were used for purification to obtain the purified mouse-human chimeric antibody 066-4.26.14-chAb, as shown in Fig. 9.

Example 8: Humanization of antibodies

The mouse antibody 066-4.26.14 was selected for humanization. The humanization process comprised mainly human template search and reshaping.

The main goal of humanization was the FR sequence in the variable region. Using the amino acid sequences of the mouse antibody 066-4.26.14 VH and VL as templates, sequences alignment were performed on the NCBI website, and 5 humanized reference sequences were found, which were used as reference templates for the humanization of antibody FR regions to design the humanized sequences.

The specific sequences of the CDR regions are shown in Table 4, and the sequences of the humanized antibodies after reshaping are shown in Table 5.

Table 4: Sequences of CDR regions of 066-4.26.14 antibody

	Antibody	CDR1 sequence	CDR2 sequence	CDR3 sequence
	066-4.26.14	SYAMS	SISTTSNTYYPDSVKG	GVITNQAWFAY
	H chain	(SEQ ID NO:1)	(SEQ ID NO:2)	(SEQ ID NO:3)
•	066-4.26.14	RASQSISNNLH	YASQSIS	QQSNSWPLT
	L chain	(SEQ ID NO:4)	(SEQ ID NO:5)	(SEQ ID NO:6)

Table 5: Humanized sequences of 066-4.26.14 antibody

	Humanized sequence
06 6- 4.2 6.1 4 H1	EVQLVESGGGLVKPGGSLRLSCAASGFTFRSYAMSWVRQAPGKGLEWVASISTT SNTYYPDSVKGRFTTSRDNAKNSLYLQMNSLRAEDTAVYYCGRGVITNQAWFA YWGQGTLVTVSS(SEQ ID NO:15)
06 6-	EVQLVESGGGLVKPGGSLRLSCAASGFTFRSYAMSWVRQTPEKRLEWVASISTT SNTYYPDSVKGRFTTSRDNAKNSVYLQMSSLRAEDTAVYYCGRGVITNQAWFA
4.2	YWGQGTTVTVSS(SEQ ID NO:16)

	Humanized sequence
6.1	
4 H2	
06	
6-	EVOLVOCCA EVIZIDOECI VICCY COCYCED CVA MCWA/DOMDOVOLEWA A CICT
4.2	EVQLVQSGAEVKKPGESLKISCKGSGYSFRSYAMSWVRQMPGKGLEWVASIST TSNTYYPDSVKGRVTTSRDKSISTAYLQWSSLKASDTAMYYCGRGVITNQAWF
6.1	AYWGQGTLVTVSS(SEQ ID NO:17)
H3	
06	
6-	EVQLVESGGGLVQPGGSLRLSCAASGFTFRSYAMSWVRQAPGKGLEWVASISTT
4.2 6.1	SNTYYPDSVKGRFTTSRDNAKNSVYLQMSSLRDEDTAMYYCGRGVITNQAWFA
4	YWGQGILVTVSS(SEQ ID NO:18)
H4	
06 6-	
4.2	EVQLVESGGGLVQPGGSLRLSCVASGFTFRSYAMSWVRQAPGKGLEWVASISTT
6.1	SNTYYPDSVKGRFTTSRDNSKNTVYLQMSSLRAEDTAVYYCGRGVITNQAWFA YWGQGTLVTVSS(SEQ ID NO:19)
4	1 WOQOTEV I VSS(SEQ ID NO.19)
H5 06	
6-	
4.2	DIVLTQSPATLSVSPGERVTLSCRASQSISNNLHWYQQKSGQAPRLLIKYASQSIS GIPSRFSGSGSGTDFTLTISSLQSEDFAVYFCQQSNSWPLTFGGGTQVEIK(SEQ ID
6.1	NO:20)
L1	
06	
6-	DIVLTQSPATLSVSPGERATLSCRASQSISNNLHWYQQKPGQAPRLLIKYASQSIS
4.2 6.1	GIPARFSGSGSGTDFTLTISSLQSEDFAVYFCQQSNSWPLTFGGGTKVEIK(SEQ ID
4	NO:21)
L2	
06 6-	
4.2	DIVLTQSPDFQSVTPKEKVTISCRASQSISNNLHWYQQKPDQSPKLLIKYASQSIS
6.1	GIPSRFSGSGSGTDFTLTINSLEAEDAAAYFCQQSNSWPLTFGPGTKVEIK(SEQ ID NO:22)
4 L3	,
06	
6-	EIVLTQSPGTLSLSPGERATLSCRASQSISNNLHWYQQKPGQAPRLLIKYASQSIS
4.2	GIPDRFSGSGSGTDFTLTISRLEPEDFAVYYCQQSNSWPLTFGGGTKVEIK(SEQ ID
6.1	NO:23)
L4	
06	
6-	EIVLTQSPDFQSVTPKEKVTITCRASQSISNNLHWYQQKPDQSPKLLIKYASQSIS
4.2 6.1	GVPSRFSGSGSGTDFTLTINSLEAEDAATYYCQQSNSWPLTFGQGTKVEIK(SEQ
4	ID NO:24)
L5	

066-4.26.14H1 (SEQ ID NO: 25)

>066-4.26.14H2 (SEQ ID NO: 26)

>066-4.26.14H3 (SEQ ID NO: 27)

>066-4.26.14H4 (SEQ ID NO: 28)

>066-4.26.14L1 (SEQ ID NO: 30)

 ttctagattcagcggcagcggaagcggcacagatttcaccctgaccatcagcagcctgcagagcgaggacttcgccgtctacttctgccagcaggagcaactcttggcccctgacctttggcggaggcacccaggtggagatcaag

>066-4.26.14L2 (SEQ ID NO: 31)

>066-4.26.14L3 (SEQ ID NO: 32)

>066-4.26.14L4 (SEQ ID NO: 33)

>066-4.26.14L5 (SEQ ID NO: 34)

gagategtgetgaccagageceagaetteeagtgacceceaaggagaaggteaceateacttgeagggecagecagagea teagcaacaacetgeattggtaccageagaagecegaceagagececaagetgetgateaagtaegceagecagtetateageggagtg cettetagatteageggaageggaageggaacagattteaceetgaccateaacageetggaggcagaggaagegcagecacetatattge cageagagaaactettggeceetgacetteggacagggeaceaaggtggagateaag

Example 9: Preparation of anti-Aß humanized full-length antibody

1. Construction of expression vectors for transient transfection of full-length antibody

The pGS003-hIgG1CH and pGS003-hIgKCL were selected as the expression vectors for constructing the heavy chain and the light chain of the anti- Aβ humanized full-length antibody, respectively. Codon optimization was performed on the 066-4.26.14 humanized antibody sequence. After PCR amplification, the heavy chain was digested with HindIII and NheI, and the light chain was digested with HindIII and NarI, and then 5 VH and 5 VL antibody genes were cloned into pGS003-hIgG1CH and pGS003-hIgKCL, respectively, as shown in Table 6. After sequencing to identify the correct insertion of antibody gene, the recombinant expression vector was transformed into *E. coli* TOP10F', and a single colony was picked and inoculated in LB medium containing 100 μg/ml ampicillin, and cultured with shaking at 37°C for 16 hours. The

plasmids were extracted using endotoxin-free large-scale extraction kit of Zymo Research, and finally the plasmids were dissolved in 1 ml of ultrapure water, and the plasmids concentration and $OD_{260/280}$ were measured with a spectrophotometer. The plasmids DNA with $OD_{260/280}$ between 1.8 and 1.9 were of a relatively high purity.

Table 6: List of expression vectors for transient transfection of the heavy chain and the light chain

Name of Heavy chain expression vector	Name of Light chain expression vector
H1	L1
H2	L2
Н3	L3
H4	L4
Н5	L5

2. Transfection, expression and detection in mammalian cells 293E

The above 5 heavy chain expression vectors and 5 light chain expression vectors of 066-4.26.14 were combined in pairs (a total of 25 combinations), and then the transient transfection expression in 2 ml 293E system was evaluated, and the expression levels and ELISA values of the 25 combinations were evaluated. The results are shown in Table 7. Among them, 6 full-length antibodies were preferably selected, which were 066-4.26.14-H2L2, 066-4.26.14-H2L3, 066-4.26.14-H5L1, 066-4.26.14-H5L2, 066-4.26.14-H5L3, respectively.

Table 7: Detection values of expression level and EC50 for small system transient transfection expression of 066-4.26.14 humanized full-length antibodies of 5×5 combination

No	Combination of heavy chain	Expression level	EC50 value of
No.	and light chain	(mg/L)	Aβ ₄₂ monomer
1	066-4.26.14H1L1	60	
2	066-4.26.14H1L2	61	
3	066-4.26.14H1L3	68.2	
4	066-4.26.14H1L4	32.1	
5	066-4.26.14H1L5	68.4	
6	066-4.26.14H2L1	89.7	0.02135
7	066-4.26.14H2L2	86.6	0.01291
8	066-4.26.14H2L3	51.8	0.01532
9	066-4.26.14H2L4	7.72	
10	066-4.26.14H2L5	6.14	
11	066-4.26.14H3L1	5.42	
12	066-4.26.14H3L2	0	
13	066-4.26.14H3L3	6.78	
14	066-4.26.14H3L4	3.88	
15	066-4.26.14H3L5	3.68	
16	066-4.26.14H4L1	69.1	0.04948
17	066-4.26.14H4L2	71.2	0.01685
18	066-4.26.14H4L3	25.3	0.1406
19	066-4.26.14H4L4	3.96	

No	Combination of heavy chain	Expression level	EC50 value of
No.	and light chain	(mg/L)	Aβ ₄₂ monomer
20	066-4.26.14H4L5	0	
21	066-4.26.14H5L1	118.5	0.004218
22	066-4.26.14H5L2	113.2	0.00772
23	066-4.26.14H5L3	103.3	0.01075
24	066-4.26.14H5L4	3.12	
25	066-4.26.14H5L5	0	

Note: "--" in the table means no combination.

293E was used for transient transfection and expression of 6 candidate antibodies in Freestyle medium. Twenty-four hours before transfection, 300 ml of 293E cells were inoculated at 0.5×10⁶ cells/ml in a 1L cell culture flask, and cultured in a 37°C, 5% CO₂ incubator with a shaker at 120 rpm. During transfection, 300 μl of 293 fectin was firstly taken and added to 5.7 ml of Opti-MEM, mixed well, and incubated at room temperature for 2 minutes; meanwhile, the expression plasmids for the heavy chain and the light chain in amount of 300 μg were diluted to 6 ml with Opti-MEM, respectively. The above-diluted transfection reagent and plasmid were mixed thoroughly, incubated at room temperature for 15 minutes, then the whole mixture was added to the cells, mixed well, and incubated in a 37°C, 5% CO₂ incubator with a shaker at 120 rpm for 7 days.

3. Purification and detection of antibodies

The cell culture medium was centrifuged at 2000 g for 20 min, the supernatant was collected, and the antibody expression level in the supernatant was detected by Octet. See Table 8.

Table 8: Detection of expression level of 6 candidate antibodies expressed by transient transfection in 300 ml

Antibody name	Heavy chain	Light chain	Expression level of
	sequence	sequence	transient transfection
			(mg/L)
066-4.26.14H2L2	066-4.26.14H2	066-4.26.14L2	146
066-4.26.14H2L3	066-4.26.14H2	066-4.26.14L3	56
066-4.26.14H4L2	066-4.26.14H4	066-4.26.14L2	101
066-4.26.14H5L1	066-4.26.14H5	066-4.26.14L1	164
066-4.26.14H5L2	066-4.26.14H5	066-4.26.14L2	128
066-4.26.14H5L3	066-4.26.14H5	066-4.26.14L3	135

The supernatant was filtered with a 0.22 µm filter, and then passed through a MabSelect SuRe affinity chromatography column (GE), eluted with 20 mM citrate-sodium citrate, pH 3.0, and the pH was adjusted to neutral with 1 M Tris base, and the solution was adjusted to an isotonic solution by adding with 10×PBS. The purified protein was detected by SDS-PAGE with 4-20% gradient gel (Nanjing Jinsirui Biotechnology Co., Ltd.). The results are shown in Fig. 10 below.

Example 10: Determination of EC50 value of humanized candidate antibody

Coating: The human A β_{42} monomer was diluted with CBS (pH 9.4) to 1 μ g/ml, added to 96-well microtiter plate, 50 μ l per well, and incubated overnight at 2-8°C.

Blocking: After washing the plate three times with PBST, 3% BSA was used for blocking, 200 µl per well, and incubated for 1 hour at 25°C.

Sample processing: The humanized candidate antibody and chimeric antibody were taken respectively, subjected to 2-fold gradient dilution using $10 \mu g/ml$ as the starting concentration (2^0 to 2^{-11}), $50 \mu l/well$, and incubated at 25° C for 1 h.

Addition of antibody: After washing the plate four times with PBST, anti-human IgG (H+L)-HRP antibody in 1:5000 diluent was added, 50 µl/well, and incubated at 25°C for 1 h.

Color development: After washing the plate four times, TMB color development solution was added, 50 µl per well, and developed in the dark for 3 minutes at room temperature.

Stop: The stop solution was directly added to stop the reaction, 50 µl per well.

Detection: After the reaction was stopped, the microtiter plate was immediately placed into a microplate reader to measure the OD value at 450 nm, and the original data were stored.

Data processing: The raw data were input into the software SoftMax Pro 6.2.1 for data processing. See Table 9 for the specific data. The results showed that the binding capability of the 6 humanized candidate antibodies to human Aβ was equivalent to that of the chimeric antibody.

Table 9: EC50 values of 6 candidate antibodies binding to antigen

Antibody name	EC50 value of Aβ ₄₂ monomer
066-4.26.14H2L2	0.0129
066-4.26.14H2L3	0.0153
066-4.26.14H4L2	0.0168
066-4.26.14H5L1	0.0042
066-4.26.14H5L2	0.0077
066-4.26.14H5L3	0.0108
066-4.26.14-chAb	0.0255

Example 11: Determination of KD value of humanized candidate antibody

Biacore-T200 detection was performed, ProteinA chip was used to capture candidate antibodies or positive antibodies, different concentrations of human $A\beta$ antigen were used to flow

through the chip, and the fitting analysis was performed based on the collected data. The antigen sample was subjected to 2-fold gradient dilution using HBS-EP+ Buffer to obtain solutions with gradient concentrations of 400 nmol/L, 200 nmol/L, 100 nmol/L, 50 nmol/L, 25 nmol/L, 12.5 nmol/L, 6.25 nmol/L, 3.125 nmol/L, 1.56 nmol/L, 0.78 nmol/L, 0 nmol/L. The sample of 25 nmol/L was used for repeat concentration detection. The detection conditions were: capture time: 30 s; antigen binding time: 120 s; dissociation time: 900 s; flow rate: 30 µl/min. And the regeneration conditions were: 20 mM NaOH solution, flow rate: 30 µl/min. The specific experimental results are shown in Table 10. It could be seen from the experimental results that, compared with the mouse-human chimeric antibody, the KD value of the humanized antibody could be equivalent to that of the mouse antibody.

Table 10: KD value detection of 6 candidate antibodies

Antibody name	Ka (1/Ms)	Kd (1/s)	KD (M)
066-4.26.14H2L2	1.92E+04	4.33E-04	2.25E-08
066-4.26.14H2L3	1.83E+04	4.72E-04	2.58E-08
066-4.26.14H4L2	1.28E+04	4.63E-04	3.63E-08
066-4.26.14H5L1	1.89E+04	4.69E-04	2.48E-08
066-4.26.14H5L2	1.34E+04	2.78E-04	2.08E-08
066-4.26.14H5L3	1.42E+04	4.60E-04	3.25E-08
066-4.26.14-chAb	2.65E+04	3.33E-04	1.26E-08

Note: E+04: $\times 10^4$; E-04: $\times 10^{-4}$; E-08: $\times 10^{-08}$.

Example 12: Detection of effect of humanized anti-AB antibody inhibiting AB polymerization

8.2% DMSO/DPBS solution (DMSO: sigma; DPBS: Hyclone) was used to dissolve Aβ dry powder to 1 mg/ml, DPBS was used to dilute the Aβ solution to 33 μg/ml, and the humanized candidate antibody 066-4.26.14 was diluted to 450 μg/ ml (IC100), and ThT (sigma) was diluted to 20 μM with ultrapure water. 50 μl of the candidate antibody diluent was taken and added to 96-well black plate (corning), then added with 50 μl of the Aβ diluent and finally added with 100 μl of ThT, incubated at room temperature for 24 hours in the dark, and measured with a mutifunctional microplate reader to determine fluorescence intensity (Ex/Em=440/485). The abscissa represented different sample groups, and the ordinate represented relative fluorescence intensity. The results are shown in Fig. 11. The relative fluorescence intensity of hlgG was 1.00, the relative fluorescence intensity of the 066-4.26.14-mAb group was 0.70, the relative fluorescence intensity of the 066-4.26.14-mAb group was 0.71, the relative fluorescence intensity of the 066-4.26.14H2L2 group was 0.62, the relative fluorescence intensity of the 066-4.26.14H4L2 group was 0.70, the relative fluorescence intensity of the 066-4.26.14H4L2 group was 0.70, the relative fluorescence intensity of the 066-4.26.14H5L1 group was 0.68, the relative

fluorescence intensity of the 066-4.26.14H5L2 group was 0.67, and the relative fluorescence intensity of the 066-4.26.14H5L3 group was 0.78. It could be seen that the humanized antibodies of 066-4.26.14 could inhibit A β polymerization.

Example 13: Detection of protective activity of humanized anti-Aβ antibody against cytotoxicity

Logarithmic growth phase SHSY5Y cells were digested with 0.25% trypsin, counted, adjusted with EMEM medium (ATCC) containing 10% fetal calf serum to have a cell density of 3×10^4 /ml, inoculated on a 96-well cell culture plate, 100 µl/well; the humanized candidate antibodies of 066-4.26.14 was diluted with EMEM medium containing 1% fetal bovine serum to 200 µg/ml (IC100) and used as working solutions, Aβ was diluted to 240 µg/ml. The culture medium in the culture plate was discarded, 50 µl of the candidate antibody diluent was firstly added, then added with 50 μl of the Aβ diluent, multiple wells were set; incubation was performed in a 37°C, 5% CO₂ incubator for 48 hours; 50 µl of the supernatant was taken and added to a new 96-well plate, then added with 50 µl of LDH assay buffer, reacted in the dark at room temperature for 30 minutes, added with 50 µl of stop solution, and measured with a multifunctional microplate reader to determine absorbance. The abscissa represented different sample groups, and the ordinate represented relative value of LDH release. The results are shown in Fig. 12. The relative value of LDH release of hIgG was 1.00, the relative value of LDH release of the 066-P02 group was 1.26, the relative value of LDH release of the 066-4.26.14-mAb group was 1.20, the relative value of LDH release of the 066-4.26.14-chAb group was 1.32, the relative value of LDH release of the 066-4.26.14H2L2 group was 1.41, the relative value of LDH release of the 066-4.26.14H2L3 group was 1.42, the relative value of LDH release of the 066-4.26.14H4L2 group was 1.30, the relative value of LDH release of the 066-4.26.14H5L1 group was 1.37, the relative value of LDH release of the 066-4.26.14H5L2 group was 1.26, and the relative value of LDH release of the 066-4.26.14H5L3 group was 1.41. It could be seen that the humanized candidate antibodies of 066-4.26.14 all had protective effect against cytotoxicity, and the protective effect was equivalent to that of 066-P02, in which the 066-4.26.14H5L2 showed the best performance.

The humanized anti-A β monoclonal antibody and its use provided by the present disclosure have been introduced in detail above. The principle and implementation of the present disclosure are illustrated with specific examples, while the description of the above examples is only used to help understand the method and the core idea of the present disclosure. It should be pointed out that for those skilled in the art, without departing from the principle of the present disclosure, several improvements and modifications can be made to the present disclosure, and these improvements and modifications also fall within the protection scope of the claims of the present disclosure.

What is claimed is:

- 1. An anti-A β humanized monoclonal antibody, wherein,
- (I) the amino acid sequences of three heavy chain CDR regions of the monoclonal antibody are the amino acid sequences set forth in SEQ ID NOs: 1, 2 and 3, respectively; and (II) the amino acid sequences of three light chain CDR regions of the monoclonal antibody are the amino acid sequences set forth in SEQ ID NOs: 4, 5 and 6, respectively;

or

(III) the amino acid sequences are the amino acid sequences obtained from the amino acids of (I) or (II) via substitution, deletion or addition of one or more amino acids, and are the amino acid sequences having the same function as the amino acid sequences of (I) or (II);

or

- (IV) the amino acid sequences are the amino acid sequences having at least 97% homology with the amino acid sequences of (I), (II) or (III).
 - 2. The monoclonal antibody according to claim 1, wherein,
- (V) the amino acid sequences of 4 heavy chain FR regions of the monoclonal antibody are the amino acid sequences set forth in SEQ ID NOs: 7, 8, 9 and 10, respectively; and (VI) the amino acid sequences of 4 light chain FR regions of the monoclonal antibody are the amino acid sequences set forth in SEQ ID NOs: 11, 12, 13 and 14, respectively;

or

(VII) the amino acid sequences are the amino acid sequences obtained from the amino acids of (V) or (VI) via substitution, deletion or addition of one or more amino acids, and are the amino acid sequences having the same function as the amino acid sequence of (V) or (VI);

or

- (VIII) the amino acid sequences are the amino acid sequences having at least 97% homology with the amino acid sequences of (V), (VI) or (VII).
 - 3. The monoclonal antibody according to claim 1, wherein,
- (IX) its heavy chain variable region has the amino acid sequence as shown in any one of SEQ ID NOs: 15 to 19; and (X) its light chain variable region has the amino acid sequence as shown in any one of SEQ ID NOs: 20 to 24;

or

(XI) its heavy chain variable region or its light chain variable region has the amino acid sequence obtained from the amino acids of (IX) or (X) via substitution, deletion or addition of one or more amino acids, and are the amino acid sequence having the same function as the amino acid sequence of (IX) or (X);

or

- (XII) its heavy chain variable region or its light chain variable region has the amino acid sequence having at least 97% homology with the amino acid sequence of (IX), (X) or (XI).
- 4. The monoclonal antibody according to claim 1 or 3, wherein the monoclonal antibody has an antigen binding epitope of A β_{30-42} .
 - 5. The monoclonal antibody according to claim 1 or 3, wherein,
- (XIII) its heavy chain variable region has the amino acid sequence as shown in SEQ ID NO: 16, and its light chain variable region has the amino acid sequence as shown in SEQ ID NO: 21; or
- (XIV) its heavy chain variable region has the amino acid sequence as shown in SEQ ID NO: 16, and its light chain variable region has the amino acid sequence as shown in SEQ ID NO: 22; or
- (XV) its heavy chain variable region has the amino acid sequence as shown in SEQ ID NO: 18, and its light chain variable region has the amino acid sequence as shown in SEQ ID NO: 21; or
- (XVI) its heavy chain variable region has the amino acid sequence as shown in SEQ ID NO: 19, and its light chain variable region has the amino acid sequence as shown in SEQ ID NO: 20; or
- (XVII) its heavy chain variable region has the amino acid sequence as shown in SEQ ID NO: 19, and its light chain variable region has the amino acid sequence as shown in SEQ ID NO: 21; or
- (XVIII) its heavy chain variable region has the amino acid sequence as shown in SEQ ID NO: 19, and its light chain variable region has the amino acid sequence as shown in SEQ ID NO: 22.
- 6. The monoclonal antibody according to any one of claims 1 to 5, wherein the more amino acids refer to 2, 3, 4, or 5 amino acids.
- 7. The monoclonal antibody according to any one of claims 1 to 6, further comprising a constant region, in which the monoclonal antibody has a heavy chain constant region that is any

one of human IgG1, IgG2, IgG3 or IgG4; and the monoclonal antibody has a light chain constant region that is of κ type or λ type.

- 8. Nucleotides encoding the monoclonal antibody according to any one of claims 1 to 7.
- 9. An expression vector, comprising nucleotides encoding the monoclonal antibody according to any one of claims 1 to 7.
- 10. A host, which is transformed or transfected with the expression vector according to claim 9.
- 11. A method for preparing the monoclonal antibody according to any one of claims 1 to 7, comprising: culturing the host cell according to claim 10, and inducing the expression of the humanized anti-A β monoclonal antibody.
- 12. A conjugate, characterized by comprising the monoclonal antibody according to any one of claims 1 to 7 that is chemically or biologically labeled.
- 13. A coupling product, which is prepared by coupling the monoclonal antibody according to any one of claims 1 to 7 or the conjugate according to claim 12 with a solid medium or a semi-solid medium.
- 14. Use of the monoclonal antibody according to any one of claims 1 to 7, the conjugate according to claim 12 and/or the coupling product according to claim 13 in the manufacture of an agent for combating cognitive impairment, an agent for treating Alzheimer's disease, an agent for inhibiting the progression of Alzheimer's disease, an agent for inhibiting the formation of senile plaques, an agent for inhibiting $A\beta$ accumulation, an agent for combating neurotoxicity, an agent for inhibiting the formation of $A\beta$ amyloid fibrils, and/or an agent for combating synaptic toxicity.
- 15. Use of the monoclonal antibody according to any one of claims 1 to 7, the conjugate according to claim 12 and/or the coupling product according to claim 13 in the manufacture of a medicament for preventing and treating a disease;

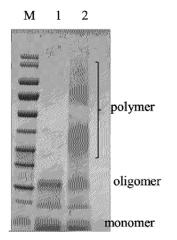
the disease comprises amyloidosis, the amyloidosis comprises secondary amyloidosis and agerelated amyloidosis, and the disease includes, but is not limited to, a neurological disease such as Alzheimer's disease.

- 16. A medicament, characterized by comprising the monoclonal antibody according to any one of claims 1 to 7, the conjugate according to claim 12 and/or the coupling product according to claim 13.
- 17. A method for the prevention and/or treatment of a disease, characterized by administering the medicament according to claim 16; wherein the disease comprises amyloidosis, the

amyloidosis comprises secondary amyloidosis and age-related amyloidosis, and the disease includes, but is not limited to, a neurological disease such as Alzheimer's disease.

- 18. Use of the monoclonal antibody according to any one of claims 1 to 7, the conjugate according to claim 12 and/or the coupling product according to claim 13 in the manufacture of products for detecting $A\beta$ expression.
- 19. A kit, comprising the monoclonal antibody according to any one of claims 1-7, the conjugate according to claim 12 and/or the coupling product according to claim 13.
- 20. A method for diagnosing a disease, characterized by detecting $A\beta$ expression by the kit according to claim 19, determining whether there is a disease according to the expression amount of $A\beta$;

wherein the disease comprises amyloidosis, the amyloidosis comprises secondary amyloidosis and age-related amyloidosis, and the disease includes, but is not limited to, a neurological disease such as Alzheimer's disease.





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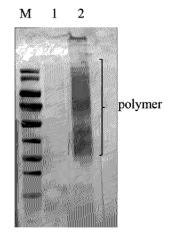
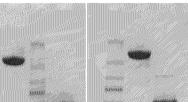


Fig. 1(B)

4

3

M



2

M

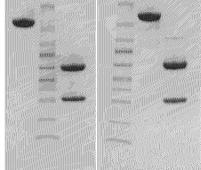


Fig. 2(A) Fig. 2(B)

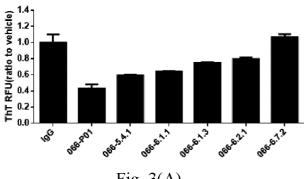


Fig. 3(A)

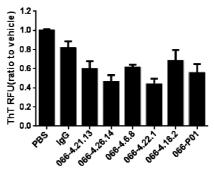
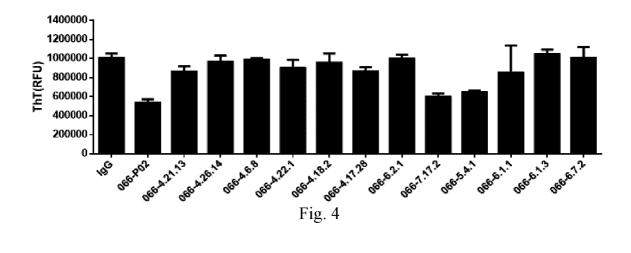
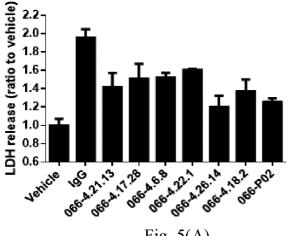
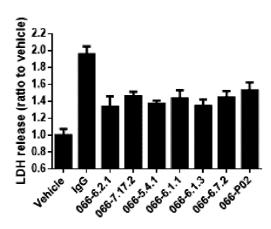


Fig. 3(B)







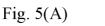


Fig. 5(B)

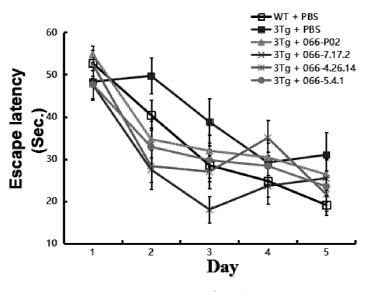


Fig. 6



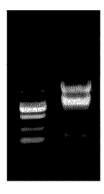
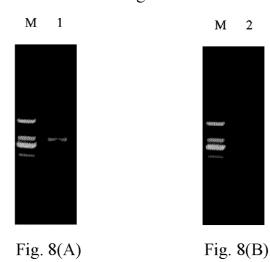


Fig. 7



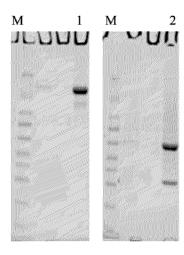


Fig. 9(A) Fig. 9(B)

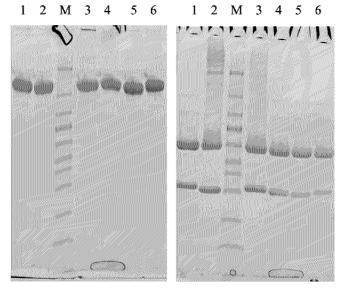
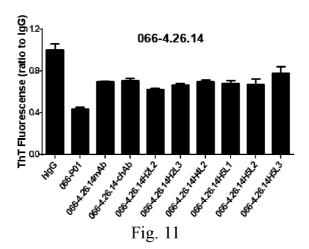


Fig. 10(A)

Fig. 10(B)



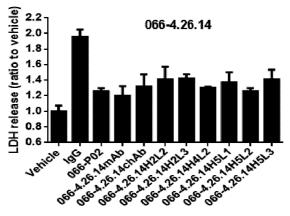


Fig. 12