(12) STANDARD PATENT (11) Application No. **AU 2003210125 B2**
(19) AUSTRALIAN PATENT OFFICE

(54) Title
Use of polysaccharide derivatives as anti-infective substances

(51) International Patent Classification(s)
A61K 8/72 (2006.01)
A01N 33/12 (2006.01)
A01N 63/00 (2006.01)
A61K 8/73 (2006.01)
A61K 31/715 (2006.01)
A61K 31/716 (2006.01)
A61K 31/717 (2006.01)
A61K 31/718 (2006.01)
A61K 31/719 (2006.01)
A61K 31/721 (2006.01)
A61P 31/04 (2006.01)
A61P 31/12 (2006.01)
C08B 31/12 (2006.01)

(21) Application No: **2003210125** (22) Date of Filing: **2003.01.10**

(87) WIPO No: **WO03/057227**

(30) Priority Data

(31) Number	(32) Date	(33) Country
102 00 717.9	**2002.01.10**	**DE**

(43) Publication Date: **2003.07.24**
(43) Publication Journal Date: **2003.09.04**
(44) Accepted Journal Date: **2008.01.24**

(71) Applicant(s)
Dow Wolff Cellulosics GmbH

(72) Inventor(s)
Heinze, Thomas;Mollmann, Ute;Dahse, Hans-Martin;Schmidtke, Michaela;Haack, Vera;Hartl, Albert

(74) Agent / Attorney
Davies Collison Cave, 255 Elizabeth Street, Sydney, NSW, 2000

(56) Related Art
US 6306835
WO 2000/026447
JP 04-057969

(12) NACH DEM VERTRAG ÜBER DIE INTERNATIONALE ZUSAMMENARBEIT AUF DEM GEBIET DES PATENTWESENS (PCT) VERÖFFENTLICHTE INTERNATIONALE ANMELDUNG

(19) Weltorganisation für geistiges Eigentum
Internationales Büro

(43) Internationales Veröffentlichungsdatum
17. Juli 2003 (17.07.2003)

PCT

(10) Internationale Veröffentlichungsnummer
WO 03/057227 A1

(51) Internationale Patentklassifikation[7]: A61K 31/715, 31/716, 31/717, 31/718, 31/719, 31/721, A61P 31/04, 31/12

(21) Internationales Aktenzeichen: PCT/DE03/00095

(22) Internationales Anmeldedatum:
10. Januar 2003 (10.01.2003)

(25) Einreichungssprache: Deutsch

(26) Veröffentlichungssprache: Deutsch

(30) Angaben zur Priorität:
102 00 717.9 10. Januar 2002 (10.01.2002) DE

(71) Anmelder *(für alle Bestimmungsstaaten mit Ausnahme von US)*: **HANS-KNÖLL-INSTITUT FÜR NATURSTOFF-FORSCHUNG E.V.** [DE/DE]; Beutenbergstrasse 11, 07745 Jena (DE). **FRIEDRICH-SCHILLER- UNIVERSITÄT JENA** [DE/DE]; Fürstengraben 1, 07743 Jena (DE).

(72) Erfinder; und
(75) Erfinder/Anmelder *(nur für US)*: **HAACK, Vera** [DE/DE]; Lutherstrasse 111, 07743 Jena (DE). **HEINZE, Thomas** [DE/DE]; Kösener Strasse 13, 07743 Jena (DE). **SCHMIDTKE, Michaela** [DE/DE]; Fröbelstieg 6a, 07743 Jena (DE). **MÖLLMANN, Ute** [DE/DE]; Schlendorfer Oberweg 17, 07749 Jena (DE). **DAHSE, Hans-Martin** [DE/DE]; Meyerstrasse 13a, 99423 Weimar (DE). **HÄRTL, Albert** [DE/DE]; Oskar-Zachau-Strasse 10, 07743 Jena (DE).

(81) Bestimmungsstaaten *(national)*: AE, AU, BA, BG, BR, CA, CN, CU, CZ, DZ, EE, GE, HR, HU, ID, IL, IN, IS, JP, KR, LT, MA, MK, MX, NO, NZ, PL, PT, RO, RU, SD, SG, SK, UA, US, YU, ZA.

(84) Bestimmungsstaaten *(regional)*: europäisches Patent (AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HU, IE, IT, LU, MC, NL, PT, SE, SI, SK, TR).

Veröffentlicht:
— *mit internationalem Recherchenbericht*
— *vor Ablauf der für Änderungen der Ansprüche geltenden Frist; Veröffentlichung wird wiederholt, falls Änderungen eintreffen*

Zur Erklärung der Zweibuchstaben-Codes und der anderen Abkürzungen wird auf die Erklärungen ("Guidance Notes on Codes and Abbreviations") am Anfang jeder regulären Ausgabe der PCT-Gazette verwiesen.

WO 03/057227 A1

(54) **Title:** USE OF POLYSACCHARIDE DERIVATIVES AS ANTI-INFECTIVE SUBSTANCES

(54) **Bezeichnung**: VERWENDUNG VON POLYSACCHARID-DERIVATEN ALS ANTIINFEKTIVE SUBSTANZEN

(57) **Abstract:** The invention relates to polysaccharides such as polyglucans and chemically or enzymatically partially hydrolyzed starches that are substituted by quaternary ammonium groups, which are bound via linkers and have a degree of substitution ranging from 0.4 to 3.0. These polysaccharides are suited for use as anti-infective agents or for treating infectious diseases caused by viruses and bacteria.

(57) **Zusammenfassung:** Polysaccharide, wie Polyglucane und chemisch oder enzymatisch partiell hydrolysierte Stärken, die mit über Linker gebundenen quaternären Ammoniumgruppen mit einem Substitutionsgrad von 0,4 bis 3,0 substituiert sind, sind als antiinfektive Mittel bzw. für die Behandlung von Infektionserkrankungen, hervorgerufen durch Viren und Bakterien, geeignet.

Use of polysaccharide derivatives as antiinfective substances

The invention relates to the use of substances based on oligosaccharides and polysaccharides as antiinfective agents, such as antibacterial agents and antiviral agents. These antiinfective agents can be employed, for example, as preservatives in cosmetic and pharmaceutical formulations, as biologically active compounds in drug preparations, for the biocidal finishing of surfaces, of textiles and of packaging materials for, for example, foodstuffs or products which are used in medicine, biology and pharmacy, and for wound protection for use in the cosmetic and pharmaceutical industries, in agriculture and in the foodstuffs and feedstuffs industries.

It is known that infections with bacterial pathogens are increasing globally and that antibacterial resistance is a general health problem in this connection. A worldwide increase in tuberculosis infections due to mycobacterial strains which, over the course of time, has become resistant to the usual therapeutic agents (B.R. Bloom, J.L. Murray, Science **257**, 1992, 1055), and the treatment of infections due to multiresistant staphylococci (M. Kresken, Bundesgesundheitsblatt [Federal Health Newspaper] **38**, 1996, 170), make it necessary to design new active substances. Alternative active compounds, possessing new mechanisms of action, in particular for counteracting antibiotic resistance and for controlling bacterial infections when there is intolerance towards existing active compounds, are urgently required.

While a significant advance in controlling life-threatening infections was achieved with the development of highly selective nucleoside and nucleotide virustatic agents, such as acyclovir, penciclovir, ganciclovir, sorivudine and cidofovir for herpes viruses, these therapeutic agents all have the same principle of action. They inhibit the viral DNA polymerase. Another disadvantage of these compounds is that they also interfere with the DNA metabolism of the infected cell and therefore harbor the risk of inducing mutagenic, teratogenic and oncogenic effects (Wutzler, P. Thust, R. Antiv. Res. **49**, 2001, 55). Furthermore, when nucleoside and nucleotide virustatic agents have been used over long periods, resistance to these medicaments has been shown to develop both in infected cell cultures and in immunosuppressed patients (Andrei, G. et al. Antimicrob. Agents Chemother., 1995, **39**, 1632; Pavic, I. et al.

Antimicrob. Agents Chemother., 1997, **39**, 2686). For this reason, it is necessary to additionally develop novel highly active antiviral prophylactic and therapeutic agents which have another mechanism of action.

The large group of biologically active substances includes quaternary ammonium compounds. They are able to destroy microorganisms such as bacteria and fungi. Low molecular weight quaternary ammonium salts are used as disinfectants or biocidal coating materials (J. Controlled Release **50**, 1998, 145). A typical problem associated with the low molecular weight compounds is inadequate bioavailability which is caused, for example, by a variety of transport and breakdown processes. Polymeric quaternary amine-functionalized materials can be synthesized from commercial quaternary exchange resins, by the graft polymerization of polyurethanes with polybutadiene hydroxytelecheles or from polysiloxanes possessing primary alcohol functions in the side chain. These biocidal polymers usually have high production costs and are frequently toxic since they contain residues of the toxic monomers (Trends in Polymer Science **4**, 1996, 364). In addition to this, the polymers may accumulate in an undesirable and dangerous manner in the body since they are not biologically degradable. Furthermore, synthetic polymers which contain cationic functions are used as dispersions for preserving wood (US 5,049,383). Disadvantages of the synthetic polymers which contain cationic functions are the high costs of preparing them, their toxicity (contamination with residual monomer) and their stability towards biological degradation.

Polysaccharide derivatives possessing quaternary ammonium functions are known and have thus far been used, in particular, as surface improvement additives for the paper and textile industries and as consistency regulators in cosmetics, in connection with which they only have a low degree of substitution (DS) of < 0.2. To date, nothing is known about their biological effects. On the other hand, starch ethers which contain long alkyl chains (C_8-C_{22}) and are bonded to the starch by way of silyl ether groups are reported to have antiinfective effects, especially antibacterial effects (JP 05295002). The low chemical stability of the alkyl silyl ethers of polysaccharides leads to an uncontrolled release of functional groups simply as a result of the effect of atmospheric moisture and consequently to a reduction in, or to the loss of, the biological activity (D. Klemm et al., Comprehensive Cellulose Chemistry, Wiley-

VCH, 1998). In addition to this, low molecular weight silyl compounds are toxic. In addition, publications refer to cellulose fibers and chitosan derivatives which possess antibacterial activity (W. H. Daly, M. M. Guerrini, Polym. Mat. Sci. Eng. 79, 1998, 220). As a natural cationic polysaccharide, chitosan is the one which is most frequently described and is used as a fungicidal agent in cosmetics (T. Tashiro, Macromol. Mater. Eng. 286, 2001, 63, K.C. Gupta, M.N.V.R. Kumar, J.M.S.-Rev. Macromol. Chem. Phys. C40, 2000, 273). Disadvantages of these polysaccharides are that they are frequently contaminated with other biogenic substances, that they are expensive as a result of the elaborate isolation and purification methods required and their inherent structure, with the ammonium groups being exclusively located on the polymer backbone. In addition to this, it is not possible to control their distribution and the content is limited to a degree of substitution of 1. Superabsorbers composed of cationically modified and crosslinked polysaccharides such as cellulose (EP 0 582 624 B1) have also been described.

While there are statements in the literature to the effect that the biological activity results from the presence of the quaternary ammonium functions, it is reported, on the other hand, that, as can be shown, typical compounds possessing cationic tetra¬alkylnitrogen groups, such as polyquaternium 10, do not possess any bioactivity (W.A. Daly, M.M. Guerrini, D. Culberson, J. Macossay, in: Science and Technology of Polymers and Advanced Materials, Plenum Press 1998, 493). It is in no way possible to conclude from the results which are available in the literature whether the structures, and if so which, are in fact biologically active.

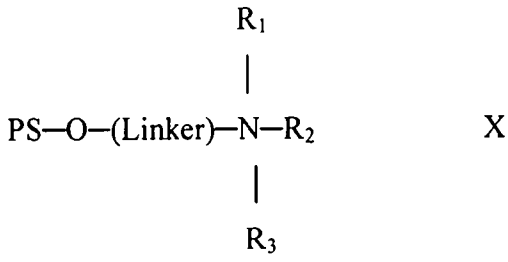
The invention is based on the aim of finding polymers which have a novel anti-infective activity; the substances should have a powerful antiinfective effect over a broad spectrum, should make it possible to effectively combat antibiotic resistance in association with bacterial infections, should offer new possibilities for treating viral infections and should be well tolerated, biologically degradable, nontoxic and easy to prepare.

According to the invention, the aim is achieved by providing polysaccharide derivatives in which etherification reactions have been used to introduce cationic functions with a degree

of substitution (DS) in the range from 0.4 to 3.0, in particular alkylammonium groups with a DS in the range from 0.6 to 1.8, into polysaccharides by way of spacer groups. Because of their biological degradability and nontoxicity, polysaccharides are particularly suitable.

The invention consequently relates to the use of polysaccharides, which are substituted, with a degree of substitution of from 0.4 to 3.0, by quaternary ammonium groups which are bonded by way of linkers, as antiinfective agents and/or for treating infectious diseases.

As now claimed, according to one aspect, the present invention provides use of alpha-glycosidically linked starch polysaccharide derivatives of the formula (I),



X

wherein PS is a polysaccharide residue based on starch,
X is an anion,
R_1 is hydrogen, C_{1-4}-alkyl or benzyl or substituted benzyl,
R_2 and R_3 independently are from C_{1-4}-alkyl or benzyl or substituted benzyl, and
Linker is a C_{2-4}-alkylene group optionally substituted by hydroxyl,
the degree of quaternary ammonium group substitution bound to Linker is from 0.4 to 3.0, for treating infectious diseases.

The compounds which are used in accordance with the invention exhibit a high degree of biological activity and surprisingly inhibit the growth of pathogenic bacteria, such as staphylococci and mycobacteria, at minimal inhibitory concentrations in the range of 5-60 mg/l, and also inhibit the replication of herpes viruses and influenza viruses in a range of 3-50 mg/l. Because of these properties, the compounds can be used for producing drugs for preventing and controlling bacterial and viral infections. They can be used both on their own and in combination with known therapeutic agents or physiologically tolerated auxiliary and carrier substances.

The antiinfective compounds can be prepared for use as solutions or suspensions in pharmaceutically acceptable media for topical administration, for parenteral administration, by way of intravenous, subcutaneous or intramuscular injections, or for intranasal administration, and as tablets, capsules or suppositories. The compounds can be employed in doses of 0.1-1000 mg/kg of bodyweight.

Polysaccharides, preferably polyglucans such as cellulose, lichenan, pullulan and dextran, and particularly preferably starches, such as native starches of different provenance, for example potato starch, wheat starch, corn starch and rice starch, and starches which have been partially hydrolyzed chemically or enzymically, such as solamyl, amylose, amylopectin and waxy corn starch, and also starches, such as the hylon types, which have been obtained from genetically modified plants, are used for

preparing the active compounds according to the invention. The content of amylose in the starches, like that of amylopectin, can in each case be from 0 to 100%, preferably from 30 to 70%. The molecular weights of suitable polysaccharides are in the range of 10^3 - 10^7 g/mol (cf. tab. 1). The anhydroglucose unit repeating units (AGU) can be linked to each other by way of α(1-4), α(1-6), α(1-3), β(1-4) and β(1-3) bonds or combinations thereof, such as α(1-4) and α(1-6), as shown diagrammatically in fig. 1, or α(1-6), α(1-3) and α(1-4), and contain side chains which are differing lengths and which are linked in different ways. In addition to this, other functional groups, such as phosphate ester functions, can also be present, for example in the case of natural potato starch.

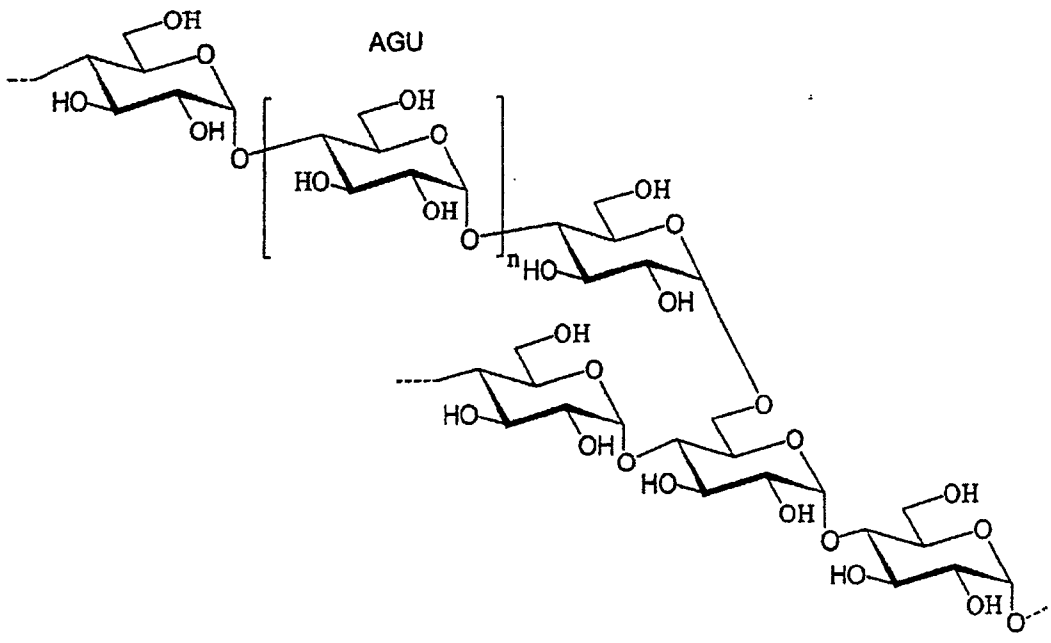


Fig. 1 Example of the structure of the polysaccharides which can be employed.

Tab. 1:

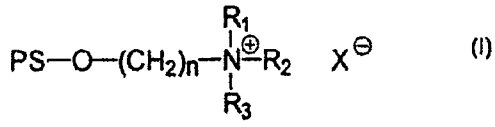
Starch material	Amylose content (%)	Molecular weight (GPC[1]) ($g \cdot mol^{-1}$)
hylon VII (H)	70	$9 \cdot 10^6$
potato starch (P, Emsland)	28	$40 \cdot 10^6$
corn starch (C)	28	$76 \cdot 10^6$
wheat starch (W)	26	$65 \cdot 10^6$
waxy corn starch (WC)	1	$51 \cdot 10^6$
solamyl (S)	28	9700

[1] determined in DMSO

The extent to which the hydroxyl groups have been transformed is described by the mean degree of substitution (DS). This mean value indicates, without any differentiation, the number of functionalized hydroxyl groups and is accordingly, in the case of the abovementioned polysaccharides, by definition in the range from 0 to 3. The DS of cationic groups in the polysaccharide derivatives of the invention having an antiinfective effect is between 0.4 and 3.0, preferably between 0.6 and 1.8. If, during derivatizations, functional groups are introduced which themselves contain reactive groups, e.g. hydroxyl groups as a result of the etherification of polysaccharides with epoxides, these latter groups can likewise react, resulting in the formation of longer side chains.

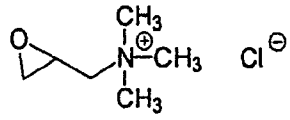
The polysaccharide derivatives which can be used in accordance with the invention are known or can be obtained in a manner known per se, in particular by etherifying polysaccharides with reactive compounds, either thereby directly forming quaternary ammonium compounds of the general formula (I) (PS: polysaccharide residue, only one substituent shown) or with the quaternization taking place after the etherification reaction. In formula (I), R_1, R_2 and R_3 are preferably, independently of each other, alkyl having 1-4 C atoms or benzyl or substituted benzyl (examples of substituents are 1 to 3 alkyl, halogen, alkoxy, carbamoyl, alkoxycarbonyl, cyano and dialkylamino), R_1 is also hydrogen, X is an anion (e.g. halide, hydroxide, sulfate,

hydrogen sulfate and another anion of inorganic and carboxylic acids), and n can be 2-4.

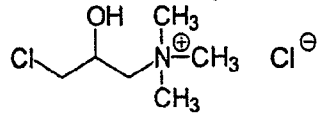


(I)

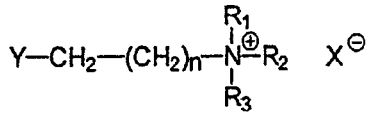
Quaternary cationizing reagents which are preferably used are 2,3-epoxypropyltrimethylammonium chloride (QUAB®151, Degussa AG, formula II) or 3-chloro-2-hydroxypropyltrimethylammonium chloride (QUAB®188, Degussa AG, formula III). Reagents of the general formula IV, in which Y = Cl or Br and n = 1-3, can also be used for etherifying the polysaccharides.



(II)



(III)



(IV)

Accordingly, the linker by way of which the quaternary ammonium groups are bonded to the polysaccharides is C_2-C_4-alkylene which is optionally substituted by hydroxyl.

The etherification for preparing the biologically active polysaccharide derivatives can be carried out in different ways and takes place in a manner which is known per se, with high contents of the cationic groups being achieved. Both suspensions of the polymers in an alcohol and sodium hydroxide solution and water (heterogeneous method), with alcohols such as methanol, isopropanol and, preferably ethanol, or aqueous alkali metal hydroxide solutions, preferably sodium hydroxide solution/

water with a transition from the heterogeneous to the homogeneous system, and also homogeneous solutions of the polymers in dipolar aprotic solvents such as dimethyl sulfoxide or dimethylacetamide in the presence of lithium chloride or other solvents as reaction media, are suitable. The time required for the reaction with the cationizing reagent is between 1 and 48 h, preferably from 3 to 24 h, and the temperature is between 30 and 130°C, preferably from 40 to 80°C. In addition, the degree of substitution of the products can be determined, and varied within wide limits, by the molar equivalents of the etherifying agent which are employed. In addition to this, multistep reactions for obtaining the polysaccharide derivatives are also suitable, with a product which is already cationized or amino-functionalized being once again reacted under the abovementioned conditions. Customary methods of polymer chemistry are used to work up the reaction products, with the low molecular weight byproducts and reagent residues being separated off by means of dialysis or washing processes or reprecipitating from water in organic solvents.

The degrees of substitution are calculated using nitrogen values determined by elemental analysis, in accordance with the following defining formula:

$$DS_N = \frac{162.15 \cdot \%N}{1401 - 151.64 \cdot \%N}$$

In addition to this, the counterions, such as chloride, which are present in the compounds, and the NMR spectra, are suitable for determining the DS values.

The following implementation examples are intended to explain the invention in more detail but without restricting it in any way.

Implementation examples:

1. Preparing the compounds by conducting a heterogeneous reaction in ethanol/ sodium hydroxide solution/water

20 g of polysaccharide (nature of the polysaccharide, see the following table 2) are suspended in 80 ml of ethanol. A solution of 10.85 g of NaOH in 28 ml of water and

80 ml of ethanol, and also a solution of 0.246 mol of QUAB®188 (69% aqueous solution), are added dropwise to this suspension. The reaction mixture is stirred at 60°C for 6 h. The product is neutralized with 0.1 N HCl, dialyzed and freeze-dried. The yield is 95% (based on the DS achieved).

Elemental analysis: N 3.51%

Mean degree of substitution (DS_N) = 0.66

Tab. 2: Heterogeneous cationization

	Reagent			Degree of substitution
Polysaccharide	Type	Molar ratio AGU : Reagent	Product	
Hylon	QUAB®188	1 : 3	H 1 (466)	0.50
Hylon	QUAB®188	1 : 2	H 2 (KS005)	0.66
Amioca	QUAB®188	1 : 3	WC 1 (505)	0.14
Potato starch	QUAB®188	1 : 3	P 1[1] (491)	0.34
Potato starch	QUAB®188	1 : 3	P 2[1] (469)	0.58
Wheat starch	QUAB®188	1 : 3.25	W 1 (527)	0.99
Wheat starch	QUAB®188	1 : 2	W 3 (571)	0.61
Wheat starch	[2]	1 : 5	W 4	0.72
Wheat starch	[2]	1 : 10	W 5	0.81

[1] different NaOH concentration

[2] reagent: $Cl\text{-}CH_2\text{-}CH_2\text{-}N(C_2H_5)H_2Cl$

2. Conversion of polysaccharides in aqueous sodium hydroxide solution

20 g of polysaccharide (nature of polysaccharide, see following table 3) are suspended in a sodium hydroxide solution (0.5 g of NaOH in 100 ml of water). The mixture is stirred at 60°C for one hour. 0.123 mol of QUAB®151 or, in the case of the degraded starch solamyl, QUAB®188, is added dropwise at this temperature. The

reaction mixture is stirred at 60°C for 6 h. After cooling down to room temperature, the mixture is neutralized with 0.1 N HCl; it is then dialyzed and the product freeze-dried.

The yield is 98% (based on the DS achieved).

Elemental analysis: N 3.34%

Mean degree of substitution (DS_N) = 0.60

Tab. 3: Heterogeneous/homogeneous cationization

	Reagent			Degree of substitution
Polysaccharide	Type	Molar ratio AGU : Reagent	Product	
Hylon	QUAB®151	1 : 1	H 3 (477)	0.40
Hylon	QUAB®151	1 : 2	H 4 (KS006)	0.92
Amioca	QUAB®151	1 : 0.5	WC 2 (503)	0.38
Amioca	QUAB®151	1 : 1	WC 3 (501)	0.60
Amioca	QUAB®151	1 : 2	WC 4 (502)	0.92
Corn starch	QUAB®151	1 : 0.5	C 1 (517)	0.35
Corn starch	QUAB®151	1 : 1	C 2[1] (516)	0.55
Corn starch	QUAB®151	1 : 1	C 3[1] (519)	0.72
Corn starch	QUAB®151	1 : 2	C 4 (515)	1.03
Potato	QUAB®151	1 : 3	P 3 (KS 016)	0.69
Potato	QUAB®151	1 : 2	P 4 (KS 013)	1.05
Wheat starch	QUAB®151	1 : 0.5	W 4 (520)	0.39
Solamyl	QUAB®188	1 : 2	S 1[1] (554)	0.68
Solamyl	QUAB®188	1 : 3	S 2 (555)	0.76
Solamyl	QUAB®188	1 : 2	S 3[1] (568)	0.80

[1] different NaOH concentration

3. Conducting a homogeneous reaction in dimethyl sulfoxide (DMSO)

15 g of polysaccharide (nature of the polysaccharide, see following table 4) are suspended in dimethyl sulfoxide (DMSO) at room temperature and the suspension is heated to 80°C, in connection with which the polysaccharide dissolves. The solution is cooled down to room temperature and 0.5 g of NaOH, dissolved in 20 ml of water, is added. 0.0925 mol of QUAB®151 is then added dropwise, while stirring. The reaction mixture is stirred at 60°C for 24 h. After it has been cooled down to room temperature, the mixture is neutralized with 0.1 N HCl and then dialyzed and freeze-dried.

The yield is 99% (based on the DS achieved).

Elemental analysis: N 3.22%

Mean degree of substitution (DS_N) = 0.57

Tab. 4: Homogeneous cationization

	Reagent			Degree of substitution
Polysaccharide	Type	Molar ratio AGU : Reagent	Product	
Hylon	QUAB®151	1 : 3	H 5 (436)	0.55
Potato starch	QUAB®151	1 : 1	P 5 (KS 9)	0.42
Amioca	QUAB®151	1 : 1	WC 5 (504)	0.57
Wheat starch	QUAB®151	1 : 1	W 5 (531)	0.41
Corn starch	QUAB®151	1 : 1	C 5 (532)	0.40
Solamyl	QUAB®151	1 : 1	S 4 (588)	0.60

4. Cationization in several steps

5 g of cationized polysaccharide (nature of the polysaccharide, see following table 5) are suspended in a sodium hydroxide solution (0.5 g of NaOH in 100 ml of water). The mixture is stirred at 60°C for one hour. 0.2 mol of QUAB®151 is added

dropwise at this temperature. The reaction mixture is stirred at 60°C for 6 h. After it has been cooled down to room temperature, the mixture is neutralized with 0.1 N HCl and then dialyzed and freeze-dried.

The yield is 95% (based on the DS achieved).

Elemental analysis: N 5.11%

Mean degree of substitution (DS_N) = 1.32

Tab. 5: Cationization in several steps

Cationized starch		Reagent			Degree of substit.
Poly-saccharide	Initial DS	Type	AGU : Reag. molar ratio	Product	DS_N
Hylon	0.40	QUAB®151	1 : 2	H 6 (479)	0.90
Hylon	0.80	QUAB®151	1 : 1.5	H 7 (498)	1.10
Potato	0.77	QUAB®151	1 : 2	P 6 (506)	1.18
Potato	0.42	QUAB®151	1 : 10	P 7 (597)	1.32
Amioca	0.60	QUAB®151	1 : 10	WC 6 (598)	1.25
Wheat	0.41	QUAB®151	1 : 10	W 6 (603)	1.16
Wheat	0.88	QUAB®151	1 : 10	W 7 (602)	1.41
Corn	0.55	QUAB®151	1 : 10	C 6 (600)	1.13
Potato	1.32	QUAB®151	1 : 15	P 8	1.80
Corn	1.13	QUAB®151	1 : 10	C 7	1.65
Solamyl	0.80	QUAB®151	1 : 2	S 5 (610)	0.91

5. Determining the antibacterial/antimicrobial activity of the compounds against Gram-positive and Gram-negative bacteria and against yeasts.

The antibacterial activity of the compounds, as directed against Staphylococcus aureus SG 511, S. aureus 134/93 (multiresistant) and Mycobacterium vaccae IMET 10670, was tested by means of determining the minimum inhibitory concentrations

(MHCs) in a microbroth dilution test in Müller-Hinton broth (DIFCO) in accordance with the NCCLS guidelines [National Committee for Clinical Laboratory Standards: Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically; 4th ed.; Villanova, Ed.; Approved standard Document M7-A4. NCCLS, (1997)]. The results are shown in table 6.

Tab. 6: Antibacterial activity

Sample	Degree of substitution DS_N	MHC [mg/l]		
		Staphylococcus aureus		Mycobacterium vaccae
		SG 511	134/93	IMET10670
H 4 KS006	0.92	15.6	15.6	7.8
H 7 498	1.10	15.6	15.6	7.8
WC 4 502	0.92	31.25	62.5	15.6
C 3 519	0.72	15.6	62.5	7.8
C 4 515	1.03	31.25	62.5	15.6
P 3 016	0.69	31.25	62.5	7.8
P 4 013	1.05	15.6	125	3.9
W 2 567	0.57	62.5	62.5	15.6
W 3 571	0.61	31.25	31.25	7.8
W 1527	0.99	31.25	31.25	15.6
S 1 554	0.68	31.25	62.6	15.6
S 2 555	0.76	15.6	31.25	7.8
S 3 568	0.80	15.6	15.6	7.8

6. Determining the antiviral effect in regard to herpes simplex virus type 1

Prior to the antiviral investigations, the 50% cytotoxic concentration (CC_{50}) in Green monkey kidney (GMK) cells was determined in order to be able to exclude nonspecific substance effects. To do this, continuous GMK cell lawns in microtiter

plates are inoculated with the appropriate substance dilution series (factor 2) (Schmidtke et al.; J. Virol. Meth. **95**, 2001, 133). After a 72-hour incubation, the cells are stained with crystal violet/methanol. After the dye has been leached out, the optical densities of the individual wells are measured (550/630 nm) in a Dynatech plate photometer and compared with the mean value for 6 untreated cell controls, which value is taken to be 100%. The CC_{50} is the substance concentration at the point where the extinction curve of the dilution series intersects with the 50% line of the mean value for the control. The antiviral effect of the compounds in regard to HSV-1 was investigated in a cytopathic effect-inhibiting test (CPE-inhibiting test) performed in GMK cells, and the 50% inhibitory dose (IC_{50}) was determined (Schmidtke et al.; J. Virol. Meth. **95**, 2001, 133). The selection index was calculated as the ratio of CC_{50} to IC_{50} (table 7).

The starting compounds (QUAB reagents and unmodified starches) did not exhibit any antiviral effect (results not shown).

Tab. 7: Antiviral activity

Sample	Degree of substitution DS_N	CC_{50} (µg/ml) in GMK cells	IC_{50} (µg/ml) against HSV-1	Selection index (CC_{50}/IC_{50})
H 3 477	0.40	> 200	8.54	> 23.42
H 1 466	0.50	> 200	5.11	> 39.14
H 2 005	0.66	> 200	7.07	> 28.29
WC 2 503	0.38	> 200	7.05	> 28.37
C 1 517	0.35	> 200	7.84	> 25.51
P 1 491	0.34	> 200	10.15	> 19.70
W 4 520	0.39	> 200	10.55	> 18.96
S 1 554	0.68	141.54	3.59	39.43

7. Determining the mode of action in a modified PRT

The investigations into the mechanism of action of the substance were carried out in a modified PRT using the acyclovir-sensitive and phosphonoformic acid-sensitive

HSV1 strain Kupka and taking the compound M 1 as an example (Schmidtke et al.; J. Virol. Meth. 95, 2001, 133). In the test, the substance was added at various concentrations:

1. only to cell-free virus (10^6 pfu/ml), which was then incubated with the compound at 37°C for 6 h and, after the substance had been diluted, incorporated in the PRT: no plaque reduction in the dose range up to 6.25-25 µg/ml (results not shown),
2. only to the agar: no plaque reduction in the dose range 6.25-25 µg/ml (results not shown),
3. only during the one-hour adsorption, at 4°C for 2 h (3.12-12.5 µg/ml), and
4. 1, 2 and 4 h before adding virus (3.12-12.5 gg/ml).

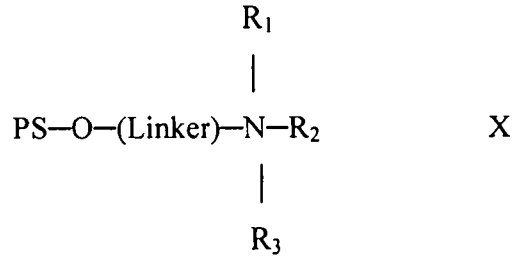
The results of the investigations into the mechanism of action show that the compounds according to the invention do not have a virucidal effect since it was not possible to observe any inactivation of cell-free virus. Instead, the prerequisite for inhibiting herpes virus replication is for the substance to be present before or during the adsorption of the virus to the test cells.

Throughout this specification and the claims which follow, unless the context requires otherwise, the word "comprise", and variations such as "comprises" or "comprising", will be understood to imply the inclusion of a stated integer or step or group of integers or steps.

The reference in this specification to any prior publication (or information derived from it), or to any matter which is known, is not, and should not be taken as an acknowledgment or admission or any form of suggestion that that prior publication (or information derived from it) or known matter forms part of the common general knowledge in the field of endeavour to which this specification relates.

The Claims defining the invention are as follows:

1. Use of alpha-glycosidically linked starch polysaccharide derivatives of the formula (I),



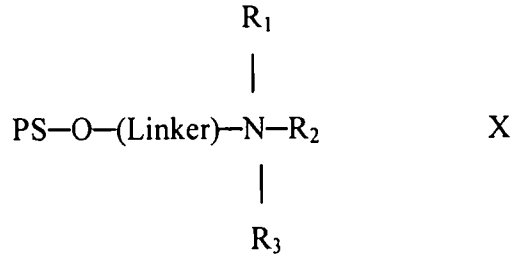
wherein PS is a polysaccharide residue based on starch,
X is an anion,
R_1 is hydrogen, C_{1-4}-alkyl or benzyl or substituted benzyl,
R_2 and R_3 independently are from C_{1-4}-alkyl or benzyl or substituted benzyl, and
Linker is a C_{2-4}-alkylene group optionally substituted by hydroxyl,
the degree of quaternary ammonium group substitution bound to Linker is from 0.4 to 3.0,
for treating infectious diseases.

2. The use of claim 1, wherein the degree of substitution is from 0.6 to 1.8.

3. The use of claim 1 wherein the starch is potato starch, wheat starch, corn starch, rice starch, starch which has been partially hydrolyzed chemically or enzymically, or starch which has been obtained from genetically modified plants.

2003210125 13 Dec 2007

4. Use of alpha-glycosidically linked starch polysaccharide derivatives of the formula (I),



X

wherein PS is a polysaccharide residue based on starch,

X is an anion,

R_1 is hydrogen, C_{1-4}-alkyl or benzyl or substituted benzyl,

R_2 and R_3 independently are from C_{1-4}-alkyl or benzyl or substituted benzyl, and

Linker is a C_{2-4}-alkylene group optionally substituted by hydroxyl,

the degree of quaternary ammonium group substitution bound to Linker is from 0.4 to 3.0,

for the production of a medication against infectious diseases.

5. Use of alpha-glycosidically linked starch polysaccharide derivatives according to claim 1 substantially as hereinbefore described with reference to the examples.