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## Yoshino et al.

## (54) POLYPEPTIDE, SCAFFOLD COMPOSITION, COMPOSITION FOR CARTILAGE TISSUE RESTORATION, COMPOSITION FOR CARTILAGE CELL CULTURE, AND COMPOSITION FOR PROMOTING GLYCOSAMINOGLYCAN PRODUCTION

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See application file for complete search history.

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## (57) ABSTRACT

A polypeptide having an amino acid sequence in which the number of RGD sequences contained per molecular weight of 10 kDa is not less than 0.30; the number of GFPGER sequences contained per molecular weight of 10 kDa is not less than 0.15; and the number of GVMGFP sequences contained per molecular weight of 10 kDa is less than 0.30; is provided. A scaffold composition, a composition for repairing a cartilage tissue, a composition for culturing cartilage cells, and a composition for promoting gly-cosaminoglycan production, which compositions contain the above polypeptide, are also provided.

### 12 Claims, 1 Drawing Sheet

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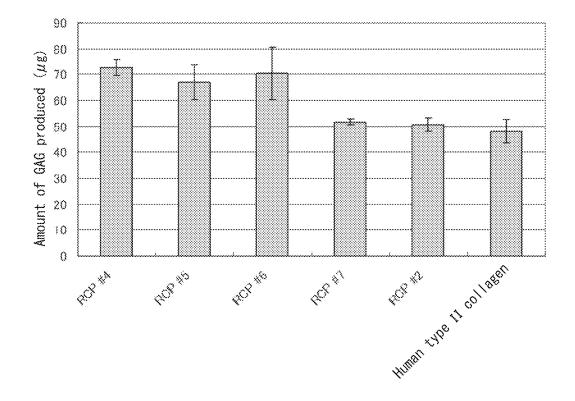
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## POLYPEPTIDE, SCAFFOLD COMPOSITION, **COMPOSITION FOR CARTILAGE TISSUE RESTORATION, COMPOSITION FOR** CARTILAGE CELL CULTURE, AND **COMPOSITION FOR PROMOTING GLYCOSAMINOGLYCAN PRODUCTION**

## CROSS-REFERENCE TO RELATED APPLICATIONS

This application is a continuation application of International Application No. PCT/JP2013/075946, filed Sep. 25, 2013, the disclosure of which is incorporated herein by reference in its entirety. Further, this application claims priority from Japanese Patent Application No. 2012-213110 15 filed on Sep. 26, 2012, the disclosure of which is incorporated herein by reference in its entirety.

## TECHNICAL FIELD

The present invention relates to a polypeptide, scaffold composition, composition for cartilage tissue restoration, composition for cartilage cell culture, and composition for promoting glycosaminoglycan production.

#### BACKGROUND ART

Currently, practical use of regenerative medicine, in which attempts are made to regenerate a body tissue or organ whose function is deteriorated or impaired, is being pro- 30 moted. Regenerative medicine is a new medical technology in which a body tissue that cannot be recovered by the self-healing ability is reconstructed using three factors, that is, cells, scaffolds and growth factors, such that the tissue has a morphology and/or function similar to those of the original 35 tissue.

In the field of regenerative medicine, collagen or gelatin, which has high biocompatibility, is used in some cases for the purpose of, for example, helping tissue repair or regeneration by cells. In particular, collagen or gelatin is some- 40 which the number of RGD sequences contained per molecutimes used for regeneration of a tissue having a threedimensional structure such as bone or skin, and, for the purpose of achieving better tissue regeneration, various modifications are being made for collagen and gelatin.

Cartilage, for example, articular cartilage, is a tissue 45 composed of a very small amount (about 2%) of cartilage cells together with an extracellular matrix, and the extracellular matrix is known to contain about 70% water, about 20% collagen and about 10% proteoglycan. The proteoglycan in the extracellular matrix is a glycoprotein containing 50 a polysaccharide called glycosaminoglycan (GAG) in an amount of about 95%, and about 5% protein. In a cartilage, cartilage cells are supported by being surrounded by collagen or proteoglycan produced by the cartilage cells themselves. In particular, glycosaminoglycan is thought to be a 55 having an isoelectric point (pI) of not more than 6.0. substance playing a role in keeping water in the cartilage matrix and involved in suppression of deterioration of, or in repair of, cartilage. Thus, studies are being carried out to develop a scaffold material for cartilage cells, which scaffold material allows favorable matrix production by the cartilage 60 ID NO:1, 2, or 3; cells.

As a scaffold material for cartilage cells, natural form of type II collagen is conventionally used.

Japanese National-Phase Publication (JP-A) No. 2007-528699 discloses a cell support coated with an RGD- 65 enriched gelatin-like protein with enhanced cell binding capacity, and describes that such a cell support can be used

for skin grafting, wound healing, or enhancement of the growth (regeneration) of bone or cartilage.

WO 2008/133196 discloses a recombinant gelatin having an RGD sequence as a cell adhesion signal, and describes that such a gelatin can be used as a cell-adhesive matrix. WO 2008/133196 also describes that, in cases of cell therapy, a cell-adhesive matrix material that can be used as a scaffold for cells is generally preferred, and that, in cases of cartilage regeneration, a high-strength matrix is desirable.

#### SUMMARY OF INVENTION

## Technical Problem

As described above, the GAG in the extracellular matrix is a matrix substance significantly involved in the metabolism of cartilage cells. However, natural form of type II collagen currently used shows only insufficient promotion of 20 production of the extracellular matrix. Scaffold materials which promote matrix production by cartilage cells have not been conventionally known so far. Moreover, compositions for cartilag tissues restoration or compositions for cartilage cell culture, which can promote repair of cartilage tissues from the viewpoint of extracellular-matrix production, or 25 compositions which can favorably promote cellular production of glycosaminoglycan among the extracellular matrix, have not been provided so far.

Accordingly, the invention aims to provide a scaffold composition excellent in promotion of extracellular-matrix production by cartilage cells, a composition for cartilage tissue restoration, a composition for cartilage cell culture, and a composition for promoting glycosaminoglycan production, and a material therefor.

#### Solution to Problem

The invention is as follows.

[1] A polypeptide having an amino acid sequence in lar weight of 10 kDa is not less than 0.30; the number of GFPGER (SEQ ID NO:12) sequences contained per molecular weight of 10 kDa is not less than 0.15; and the number of GVMGFP (SEQ ID NO:13) sequences contained per molecular weight of 10 kDa is less than 0.30.

[2] The polypeptide according to [1], wherein the number of amino acid residues in the full-length sequence is from 300 to 1400.

[3] The polypeptide according to [1] or [2], having an identity of not less than 85% to an amino acid sequence of natural form of human type II collagen.

[4] The polypeptide according to any one of [1] to [3], having a molecular weight of from 30 kDa to 80 kDa.

[5] The polypeptide according to any one of [1] to [4],

[6] The polypeptide according to any one of [1] to [5], which is a recombinant peptide.

[7] A polypeptide which is

(A) a polypeptide having the amino acid sequence of SEQ

(B) a polypeptide having the same amino acid sequence as the amino acid sequence of SEQ ID NO:1, 2, or 3 except that one or several amino acids are deleted, substituted and/or added, which polypeptide has a capacity to promote glycosaminoglycan production; or

(C) a polypeptide having an amino acid sequence having a sequence identity of not less than 80% to the amino acid

sequence of SEQ ID NO:1, 2, or 3, which polypeptide has a capacity to promote GAG production.

[8] A polypeptide having an amino acid sequence having a sequence identity of not less than 90% to the amino acid sequence of SEQ ID NO:1, 2, or 3, which polypeptide has 5 a capacity to promote glycosaminoglycan production.

[9] A polypeptide having an amino acid sequence having a sequence identity of not less than 95% to the amino acid sequence of SEQ ID NO:1, 2, or 3, which polypeptide has a capacity to promote glycosaminoglycan production.

[10] A scaffold composition comprising the polypeptide according to any one of [1] to [9].

[11] A composition for cartilage tissue testoration, comprising the polypeptide according to any one of [1] to [9].

[12] A composition for cartilage cell culture, comprising 15 the polypeptide according to any one of [1] to [9].

[13] A composition for promoting glycosaminoglycan production, comprising the polypeptide according to any one of [1] to [9].

[9] in production of a scaffold composition.

[15] Use of the polypeptide according to any one of [1] to [9] in production of a composition for cartilage tissue restoration.

[16] Use of the polypeptide according to any one of [1] to  $^{25}$ [9] in production of a composition for cartilage cell culture.

[17] Use of the polypeptide according to any one of [1] to [9] in production of a composition for promoting glycosaminoglycan production

30 [18] A method for restoration of cartilage or regeneration of cartilage, comprising administering the composition for cartilage tissue restoration according to [11] to a damaged area of cartilage.

### Advantageous Effects of Invention

By the invention, a scaffold composition excellent in promotion of extracellular-matrix production by cartilage cells, a composition for cartilage tissue restoration, a composition for cartilage cell culture, and a composition for  ${\rm ^{40}}$ promoting glycosaminoglycan production, and a material therefor can be provided.

## BRIEF DESCRIPTION OF DRAWINGS

FIG. 1 is a graph showing the results of evaluation of the GAG production-promoting capacity of each polypeptide in Examples and Comparative Examples.

### DESCRIPTION OF EMBODIMENTS

The polypeptide of the invention is a polypeptide having an amino acid sequence in which the number of RGD sequences contained per molecular weight of 10 kDa is not less than 0.30; the number of GFPGER (SEQ ID NO:12) 55 sequences contained per molecular weight of 10 kDa is not less than 0.15; and the number of GVMGFP (SEQ ID NO:13) sequences contained per molecular weight of 10 kDa is less than 0.30.

In the invention, according to the above constitution, 60 production of an extracellular matrix, especially glycosaminoglycan (which may be hereinafter referred to as GAG), by cartilage cells is promoted when the cartilage cells are in contact with the polypeptide according to the invention. 65

That is, in order to promote production of GAG more efficiently than natural form of type II collagen, not less than 4

the predetermined numbers of RGD sequences and GFPGER (SEQ ID NO:12) sequences need to be present. In addition, the number of GVMGFP (SEQ ID NO:13) sequences needs to be 0, or not more than 0.30 per molecular weight of 10 kDa in the full-length polypeptide. In the invention, GAG production by cartilage cells is promoted by satisfaction of the conditions of the numbers of RGD sequences, GFPGER (SEQ ID NO:12) sequences, and GVMGFP (SEQ ID NO:13) sequences contained. It can be 10 assumed that GAG may be present in a large amount in the vicinity of cartilage cells after contacting with the polypeptide according to the invention, and that excellent proliferation and growth of the cartilage cells may also be obtained thereby. However, the invention is not bound by these theories.

The polypeptide according to the invention may be hereinafter referred to as "specific polypeptide".

The invention is described below.

In the present description, the term "step" means not only [14] Use of the polypeptide according to any one of [1] to 20 an independent step, but also a step which cannot be clearly distinguished from other steps, as long as an expected object of the step can be achieved therewith.

> In the present description, a numerical range indicated using "to" means the range in which the values described before and after "to" are included as the minimum value and the maximum value, respectively.

> In the present description, the amount of each component in a composition means, in cases in which plural substances corresponding to the component are present in the composition, the total amount of the plural substances present in the composition, unless otherwise specified.

In the invention, each amino acid residue in an amino acid sequence may be represented by the single-letter code (for example, "G" represents a glycine residue) or three-letter 35 code (for example, "Gly" represents a glycine residue), which are well known in the art.

In the invention, "%" as used in relation to the amino acid sequence of a polypeptide is based on the number of amino acid (or imino acid) residues, unless otherwise specified.

In the present description, the meaning of an expression such as "corresponding amino acid residue" as used for a specific amino acid residue in an amino acid sequence is as follows: when 2 or more amino acid sequences to be compared are aligned by a method well known in the art in consideration of insertions, deletions, and substitutions such that the number of identical amino acid residues becomes maximum, the amino acid residue, in another amino acid sequence, at the same position as the position of a specific amino acid residue in the amino acid sequence as a reference 50 is the "corresponding amino acid residue".

In the invention, the "identity" between the amino acid sequences of two polypeptides to be compared means the value calculated by the following equation. Comparison of plural polypeptides (alignment) is carried out by an ordinary method such that the number of identical amino acid residues is maximum.

In judgment of the identity between recombinant peptides, each of the two polypeptides to be compared is separated into arbitrary fragments each having not less than 10 amino acid residues, and the correspondence of the fragments derived from one polypeptide to the fragments derived from the other polypeptide is determined such that the identity becomes maximum. The amino acid sequence is then compared between the corresponding fragments, to determine the identity as a whole. In a case in which repeated sequences (sequences each having not less than 10 amino acid residues) are contained, the second and later

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repeats are excluded before the determination of the identity (%) between the corresponding portions.

Identity (%)=[(Number of identical amino acid residues)/(Alignment length)]×100

## [Specific Polypeptide]

The specific polypeptide according to the invention has an amino acid sequence in which the number of RGD sequences contained per molecular weight of 10 kDa is not less than 0.30; the number of GFPGER (SEQ ID NO:12) 10 sequences contained per molecular weight of 10 kDa is not less than 0.15; and the number of GVMGFP (SEQ ID NO:13) sequences contained per molecular weight of 10 kDa is less than 0.30.

Since the specific polypeptide has an amino acid sequence 15 containing the predetermined numbers of RGD sequences, GFPGER (SEQ ID NO:12) sequences, and GVMGFP (SEQ ID NO:13) sequences, the polypeptide can work as a favorable scaffold that promotes production of the matrix by cartilage cells.

The RGD sequence is known as an integrin-binding site or a sequence (motif) having a cell adhesion function. The number of RGD sequences contained in the specific polypeptide is not less than 0.30 per molecular weight of the specific polypeptide of 10 kDa. In cases in which the number 25 is less than 0.30, the matrix production by cartilage cells cannot be sufficiently promoted. The number of RGD sequences contained in the specific polypeptide may also be not less than 0.35, or may be not less than 0.40. Although the upper limit of the number of RGD sequences contained in 30 the specific polypeptide varies depending on the total length of the specific polypeptide, the number is, for example, preferably not more than 2.0, more preferably not more than 1.0, still more preferably not more than 0.5 per 10 kDa.

In cases in which plural RGD sequences are contained in 35 the specific polypeptide, the number of amino acid residues between the RGD sequences is preferably from 0 to 100, more preferably from 25 to 60, although the number varies depending on the total length of the particular polypeptide. The RGD sequences are preferably unevenly distributed in 40 the specific polypeptide such that the number of amino acid residues therebetween falls within the above ranges.

The GFPGER (SEQ ID NO:12) sequence is known as an  $\alpha 2\beta 1$  integrin-binding site or a sequence having a cell adhesion function. The number of GFPGER (SEQ ID 45 NO:12) sequences contained in the specific polypeptide is not less than 0.15 per molecular weight of the specific polypeptide of 10 kDa. In cases in which the number is less than 0.15, the matrix production by cartilage cells cannot be sufficiently promoted. The number of GFPGER (SEQ ID 50 NO:12) sequences contained in the specific polypeptide may also be not less than 0.20, or may be not less than 0.30. Although the upper limit of the number of GFPGER (SEQ ID NO:12) sequences contained in the specific polypeptide varies depending on the total length of the specific polypep- 55 tide, the number is, for example, preferably not more than 1.0, more preferably not more than 0.5 per 10 kDa.

"P" (proline residue) in the GFPGER (SEQ ID NO:12) sequences may also be an oxyproline residue.

The GVMGFP (SEQ ID NO:13) sequence is commonly 60 found among fibrous collagen, and known as a recognition site of DDR-2 (Discoidin domain receptor-2). The GVMGFP (SEQ ID NO:13) sequence is also known to be involved in the proliferation of cells. The number of GVMGFP (SEQ ID NO:13) sequences contained in the 65 specific polypeptide is less than 0.30 per molecular weight of the specific polypeptide of 10 kDa. In cases in which the

number is not less than 0.30, the matrix production by cartilage cells cannot be sufficiently promoted. The number of GVMGFP (SEQ ID NO:13) sequences, if present, contained in the specific polypeptide may also be not more than 0.28, or may be not more than 0.25 per molecular weight of the particular polypeptide of 10 kDa. In terms of the lower limit of the number of GVMGFP (SEQ ID NO:13) sequences contained in the specific polypeptide, the number of the sequences may be, for example, not less than 0.2, or may be zero, per molecular weight of the specific polypeptide of 10 kDa.

From the viewpoint of promotion of the matrix production, the ratio of the number of RGD sequences contained to the total number of GFPGER (SEQ ID NO:12) sequences and GVMGFP (SEQ ID NO:13) sequences contained, that is, [number of RGD sequences contained/(total number of GFPGER (SEQ ID NO:12) sequences and GVMGFP (SEQ ID NO:13) sequences contained)] is preferably from 0.8 to 1.2, more preferably 1.

The positional relationship among the RGD sequences. GFPGER (SEQ ID NO:12) sequences, and GVMGFP (SEQ ID NO:13) sequences in the entire polypeptide is not limited as long as the ratios of these sequences present in the polypeptide satisfy the predetermined conditions described above. For example, a GVMGFP (SEQ ID NO:13) sequence may be placed either in the N-terminal side or C-terminal side of a GFPGER (SEQ ID NO:12) sequence. In cases in which plural RGD sequences are present, all of the RGD sequences may be placed between a GVMGFP (SEQ ID NO:13) sequence and the C-terminus of the polypeptide. In cases in which plural GFPGER (SEQ ID NO:12) sequences are present, all of the RGD sequences may be placed between the GFPGER (SEQ ID NO:12) sequence most close to the N-terminus and the GFPGER (SEQ ID NO:12) sequence most close to the C-terminus. Alternatively, at least one RGD sequence may be placed either in the N-terminal side of the GFPGER (SEQ ID NO:12) sequence most close to the N-terminus or in the C-terminal side of the GFPGER (SEQ ID NO:12) sequence most close to the C-terminus.

The specific polypeptide may contain, in addition to the RGD sequence(s), GFPGER (SEQ ID NO:12) sequence(s), and/or GVMGFP (SEQ ID NO:13) sequence(s), one or more other known sequences (motifs).

For example, the specific polypeptide may have repeats of a sequence(s) represented by Gly-X-Y. In cases in which plural Gly-X-Y sequences are present, the plural Gly-X-Y sequences may be either the same or different. In Glv-X-Y, Gly represents a glycine residue, and each of X and Y represents an arbitrary amino acid residue other than a glycine residue. A large number of imino acid residues, that is, proline residues and/or oxyproline residues, are preferably contained as X and Y. The ratio of the imino acid residues contained in the entire specific polypeptide is preferably from 10% to 45%. The ratio of Gly-X-Y contained in the entire specific polypeptide is preferably not less than 80%, more preferably not less than 95%, still more preferably not less than 99%.

The specific polypeptide may also contain one or more other cell adhesion signals from the viewpoint of biocompatibility. Examples of such cell adhesion signals include sequences such as the LDV sequence, REDV (SEQ ID NO:14)sequence, YIGSR (SEQ ID NO:15) sequence, PDSGR (SEQ ID NO:16) sequence, RYVVLPR (SEQ ID NO:17) sequence, LGTIPG (SEQ ID NO:18) sequence, RNIAEIIKDI (SEQ ID NO:19) sequence, IKVAV (SEQ ID NO:20) sequence, LRE sequence, DGEA (SEQ ID NO:21) sequence, and HAV sequence. Preferred examples of the cell

adhesion signals include YIGSR (SEQ ID NO:15) sequence, PDSGR (SEQ ID NO:16) sequence, LGTIPG (SEQ ID NO:18) sequence, IKVAV (SEQ ID NO:20)sequence, and HAV sequence. These other cell adhesion signals may be used singly, or in combination of two or more kinds thereof. 5

The number of amino acid residues in the entire specific polypeptide is not limited as long as the 3 kinds of sequences described above are contained at the predetermined ratios. The number of amino acid residues in the entire particular polypeptide is preferably from 300 to 1400, more preferably 10 from 400 to 1000, still more preferably from 500 to 800. In cases in which the number of amino acid residues is not less than 300, the effect of promoting the matrix production of cartilage cells tends to be more securely exerted, and, in cases in which the number of amino acid residues is not 15 more than 1400, solubility of the polypeptide in water is not largely deteriorated, and the polypeptide tends to have excellent handling properties.

The molecular weight of the specific polypeptide is preferably from 30 kDa to 80 kDa, more preferably from 40 kDa 20 to 70 kDa. With a molecular weight of not less than 30 kDa, the effect of promoting the matrix production of cartilage cells tends to be more securely exerted, and, with a molecular weight of not more than 80 kDa, solubility of the polypeptide in water is not largely deteriorated, and the 25 polypeptide tends to have excellent handling properties. In the invention, the molecular weight of the specific polypeptide is a value measured by electrospray ionization mass spectrometry (ESI-MS) (Q-TOF PREMIER, manufactured by Waters Corporation) according to an ordinary method.

As long as the specific polypeptide has an amino acid sequence containing the predetermined numbers of RGD sequences, GFPGER sequences, and GVBMGFP sequences, the amino acid sequence of the remaining part is not limited. From the viewpoint of, for example, promotion of proliferation of cartilage cells, the identity to the amino acid sequence of natural form of collagen is preferably not less than 85%, more preferably not less than 90%, still more preferably not less than 98%.

Examples of the natural from of collagen to be used as the standard of identity include type I, type II, type III, type IV, and type V. From the viewpoint of promotion of cartilage matrix production, the identity to the amino acid sequence of natural from of human type II collagen may be preferably not less than 85%, more preferably not less than 90%, still more preferably not less than 98%.

Preferred examples of the origin of the natural from of collagen to be used as the standard of identity include human, horse, pig, mouse and rat. The origin of the natural from of collagen is more preferably human.

The natural from of collagen to be used as the standard of identity is more preferably native human type II collagen. A known example of the sequence of natural from of human type II collagen is the following amino acid sequence of SEQ ID NO:4. The amino acid sequence of natural from of human type II collagen is shown in Table 1. In Table 1, RGD, GFPGER (SEQ ID NO:12), and GVMGFP (SEQ ID NO:13) sequences are indicated in bold.

TABLE 1

Col	lagen II hu	man alpha I	(1487 a.a.	) (SEQ ID N	O: 4)
MIRLGAPQTL	VLLTLLVAAV	LRCQGQDVQE	AGSCVQDGQR	YNDKDVWKPE	PCRICVCDTG
TVLCDDIICE	DVKDCLSPEI	PFGECCPICP	TDLATASGQP	GPKGQKGEPG	DIKDIVGPKG
PPGPQGPAGE	QGP <b>rgdrgd</b> K	GEKGAPGPRP	RDGEPGTPGN	PGPPGPPGPP	GPPGLGGNFA
AQMAGGFDEK	AGGAQLGVMQ	GPMGPMGPRG	PPGPAGAPGP	QGFQGNPGEP	GEPGVSGPMG
PRGPPGPPGK	PGDDGEAGKP	GKAGERGPPG	PQGARGFPGT	PGLPGVKGHR	GYPGLDGAKG
EAGAPGVKGE	SGSPGENGSP	GPMGPRGLPG	ERGRTGPAGA	AGARGNDGQP	GPAGPPGPVG
PAGGPGFPGA	PGAKGEAGPT	GARGPEGAQG	PRGEPGTPGS	PGPAGASGNP	GTDGIPGAKG
SAGAPGIAGA	PGFPGPRGPP	GPQGATGPLG	KPGQTGEPGI	AGFKGEQGPK	GEPGPAGPQG
APGPAGEEGK	RGARGEPGGV	GPIGPPGERG	APGNRGFPGQ	DGLAGPKGAP	GERGPSGLAG
PKGANGDPGR	PGEPGLPGAR	GLTGRPGDAG	PQGKVGPSGA	PGEDGRPGPP	gpqgargqp <b>g</b>
<b>VMGFP</b> GPKGA	NGEPGKAGEK	GLPGAPGLRG	LPGKDGETGA	AGPPGPAGPA	GERGEQGAPG
PSGFQGLPGP	PGPPGEGGKP	GDQGVPGEAG	APGLVGPRGE	R <b>gfpger</b> gsp	GAQGLQGPRG
LPGTPGTDGP	KGASGPAGPP	GAQGPPGLQG	MPGERGAAGI	agpkgd <u><b>rgd</b></u> V	GEKGPEGAPG
KDGGRGLTGP	IGPPGPAGAN	GEKGEVGPPG	PAGSAGARGA	PGERGETGPP	GPAGFAGPPG
ADGQPGAKGE	QGEAGQKGDA	GAPGPQGPSG	APGPQGPTGV	TGPKGARGAQ	GPPGATGFPG
AAGRVGPPGS	NGNPGPPGPP	GPSGKDGPKG	A <u><b>RGD</b></u> SGPPGR	AGEPGLQGPA	GPPGEKGEPG
DDGPSGAEGP	PGPQGLAGQR	GIVGLPGQRG	ERGFPGLPGP	SGEPGKQGAP	GASGDRGPPG
PVGPPGLTGP	AGEPGREGSP	GADGPPGRDG	AAGVKGDRGE	TGAVGAPGAP	GPPGSPGPAG
PTGKQGDRGE	AGAQGPMGPS	GPAGARGIQG	PQGP <u>RGD</u> KGE	AGEPGERGLK	GHRGFTGLQG
LPGPPGPSGD	QGASGPAGPS	GPRGPPGPVG	PSGKDGANGI	PGPIGPPGPR	GRSGETGPAG

TABLE	1	-continued

Collagen II human alpha I (1487 a.a.) (SEQ ID NO: 4)
PPGNPGPPGP PGPPGPGIDM SAFAGLGPRE KGPDPLQYMR ADQAAGGLRQ HDAEVDATLK
SLNNQIESIR SPEGSRKNPA RTCRDLKLCH PEWKSGDYWI DPNQGCTLDA MKVFCNMETG
ETCVYPNPAN VPKKNWWSSK SKEKKHIWFG ETINGGFHFS YGDDNLAPNT ANVQMTFLRL
LSTEGSQNIT YHCKNSIAYL DEAAGNLKKA LLIQGSNDVE IRAEGNSRFT YTALKDGCTK
HTGKWGKTVI EYRSQKTSRL PIIDIAPMDI GGPEQEFGVD IGPVCFL

The isoelectric point (pI) of the specific polypeptide is not limited, and may be, for example, not more than 10.0. The <sup>15</sup> kDa, and a pI of from 5.0 to 10.0; isoelectric point is preferably not more than 9.2, more preferably not more than 7.0, still more preferably not more than 6.0 from the viewpoint of promotion of proliferation of cartilage cells. In terms of the lower limit of the isoelectric 20 point, the isoelectric point may be, for example, not less than 5.0. The pI of the polypeptide may be adjusted by an ordinary method. For example, the pI can be lowered by increasing the content of neutral amino acid residues (for example, glycine residues and alanine residues) and/or 25 acidic amino acid residues (glutamic acid residues and aspartic acid residues), or by decreasing the content of basic amino acid residues (lysine residues, arginine residues and histidine residues), among the amino acid residues in the amino acid sequence of the polypeptide. In the invention, the pI of the specific polypeptide is a value measured by isoelectric focusing according to an ordinary method.

From the viewpoint of antigenicity of the specific polypeptide, each of a serine residue(s) and/or threonine 35 residue(s) is preferably substituted by other amino acid residue. An example of the other amino acid residue for substitution of a serine residue or threonine residue is a lysine residue. For example, use of a lysine residue instead of a serine residue or threenine residue leads to introduction  $_{40}$ of an amino group to the specific polypeptide, which then results in an increased number of cross-linking points. As a result, the polypeptide tends to be more stable and less likely to be decomposed, achieving better properties for formulation. 45

The specific polypeptide is preferably a recombinant polypeptide from the viewpoints of reduction of antigenicity, mass production, safety, and the like. In the present description, the "recombinant peptide" means a polypeptide artificially prepared by a gene recombinant technology using 50 E. coli, yeast, cultured cells, or the like as a host.

The solubility of the specific polypeptide in water is preferably not less than 2% by mass from the viewpoint of properties for formulation. The solubility in water in the invention means the solubility in water under normal pressure at 25° C.

From the viewpoint of the capacity to promote matrix production in cartilage cells, examples of the specific polypeptide include the following:

(1) a polypeptide having an amino acid sequence in which the number of RGD sequences contained per molecular weight of 10 kDa is not less than 0.30; the number of GFPGER (SEQ ID NO:12) sequences contained per molecular weight of 10 kDa is not less than 0.15; and the 65 number of GVMGFP (SEQ ID NO:13) sequences contained per molecular weight of 10 kDa is less than 0.30; which

polypeptide has a molecular weight of from 30 kDa to 80

(2) a polypeptide having an amino acid sequence composed of from 300 to 1400 amino acid residues in which the number of RGD sequences contained per molecular weight of 10 kDa is not less than 0.30; the number of GFPGER (SEQ ID NO:12) sequences contained per molecular weight of 10 kDa is not less than 0.15; and the number of GVMGFP (SEQ ID NO:13) sequences contained per molecular weight of 10 kDa is less than 0.30; which polypeptide has a pI of from 5.0 to 10.0;

(3) a polypeptide having an amino acid sequence in which the number of RGD sequences contained per molecular weight of 10 kDa is not less than 0.30; the number of GFPGER (SEQ ID NO:12) sequences contained per molecular weight of 10 kDa is not less than 0.15; and no GVMGFP (SEQ ID NO:13) sequence is contained; which polypeptide has a molecular weight of from 30 kDa to 80 kDa, and a pI of from 5.0 to 10.0;

(4) a polypeptide having an amino acid sequence in which the number of RGD sequences contained per molecular weight of 10 kDa is not less than 0.35; the number of GFPGER (SEQ ID NO:12) sequences contained per molecular weight of 10 kDa is not less than 0.20; and the number of GVMGFP (SEQ ID NO:13) sequences contained per molecular weight of 10 kDa is less than 0.30; which polypeptide has a molecular weight of from 40 kDa to 70 kDa, and a pI of from 5.0 to 10.0; and

(5) a polypeptide having an amino acid sequence composed of from 300 to 1400 amino acid residues in which the number of RGD sequences contained per molecular weight of 10 kDa is not less than 0.35; the number of GFPGER (SEQ ID NO:12)sequences contained per molecular weight of 10 kDa is not less than 0.20; and the number of GVMGFP (SEQ ID NO:13) sequences contained per molecular weight of 10 kDa is less than 0.30; which polypeptide has a pI of from 5.0 to 10.0.

The specific polypeptide in the invention is preferably the polypeptide of SEQ ID NO: 1, 2 or 3 shown below, because of their high capacity to promote GAG production. In each sequence, RGD, GFPGER (SEQ ID NO:12), and GVMGFP (SEQ ID NO:13) sequences are indicated in bold. In SEQ ID NOs:1 to 3, each base corresponding to a serine residue or threonine residue in the amino acid sequence of natural form of human type II collagen is substituted by a glycine residue, alanine residue, lysine residue, or the like.

sequence	Number of residues	SEQ ID No.
GPQGARGQP <u>GVMGFP</u> GPKGANGEPGKAGEKGLPGAPGLRGLPGKDGEAGAAGPPGPAGPAGERGEQ GAPGPPGFQGLPGPPGPPGEGGKPGDQGVPGEAGAPGLVGPRGER <u>GFPGER</u> GAPGAQGLQGPRGLP GAPGPPGPAGANGEKGEVGPPGPAQAGPGLQGMPGERGAAGIAGPKGD <u>RGD</u> VGEKGPEGAPGKDGGRGLGG PIGPPGPAGANGEKGEVGPPGPAGAAGARGAPGERGEAGPPGPAGPAGPAGPAGPAGPAGACGQEAAGQ KGDAGAPGPQGPGGAPGPQGPAGPAGAAGRGAQGPPGAAGGPFGAAGRVGPPGLAQONGGPPGPPA GKOGPKGA <u>RGD</u> AGPPGRAGEPGLQGPAGPGEKGEPGDDGPFGAEGPPGPQGLAAGQRGIVGLPGQRG ERGFPGLPGPAGEPGKQGAPGAAGDRGPPGPVGPPGPAGEPGPGPGAAGGPGAAGPPGRAGAVKGD RGEAGAVGAPGAPGPPGAPGPAGPPGPQGDRGEAGAQQP	506	1
GPQGARGQP <b>GYMGFP</b> GPKGANGEPGKAGEKGLPGAPGLRGLPGKDGEAGAAGPPGPAGPAGERGEQ GAPGPPGFQCLPGPPGPPGEGGKPGDQGVPGEAGAPCLVGPRGER <b>GFPGER</b> GKPGAQGLQGPRGLP GAPGKDGPKGAAGPAGPPGAQGPPGLQGMPGERGAAGIAGPKGD <b>RGD</b> VGEKGPEGAPGKDGGRGLGG PIGPPGPAGANGEKGEVGPPGPAGAAGARGAPGERGEKGPPGPAGFAGPPGADQOPGAKGEQGEAGQ KGDAGAPGPQGPKGAPGPQGPAGVAGPKGARGAQGPPGAAGGPPGAAGRVGPPGPPGPA GKDGPKGA <b>RGD</b> AGPPGRAGEPGLQGPAGPAGPGEKGEPGDDGPPGAAGGPPGPAQGLAQQRCIVGLPQQRG ERGFPGLPGPKGEPGKQGAPGAKGDRGPPGPVGPPGPAGEGPGBGEGGPGADGPPGRDGAAGVKGD RGEKGAVGAPGAPGPAGPAGPPGPQGDRGEAGAQGP	506	2
MGFPGPKGANGEPGKAGEKGLPGAPGLRGLPGKDGEAGAAGPPGPAGPAGERGEQGAPGPGFQGLP GPPGPGEGGKPGDQGVPGEAGAPGLVGPRGER <b>GFPGER</b> GKPGAQGLQGPRGLPGAPGKDGPKGAA GPAGPPGAQGPPGLQGMPGERGAAGIAGPKGD <u>RGD</u> VGEKGPEGAPGKDGGRGLGGPIGPPGPAGAAG EKGEVGPPGPAGAAGARGAQGPEGAAGFPGAAGRVGPPGLQGNPGPPGPAGKDGPKGABGDA GPPGRAGEPGLQGPAGPGEKGEPGDDGPPGAEGPPGPQGLAGQRGIVGLPGQPGERGFPGLPGPKG EPGKQGAPGAKGDRGPFGPVGPPGLAGPAGEPGFQGLAGQPFGRDGAAGVKDRGERGAPGAPG PGPPGAPGPAGPPGPQGDRGEAGAQGPMGFPGPKGANGEPGKAGEKGLPGAPGLRGLPGKDGERGAVGAPKA GPPGPAGPGQQAPGGPAGPGFQGLPGPPGPQGEGGKPGDQGVPGEAGAPGLCGPRGERG <u>FPGER</u> GKPGAQGPAGPAGPAGPAGPAGPCGKDGERGAPGAPGF CKGAPGACGAPGACGPCGACGPFGFQGLPGPGPGPGFGCGCKPGDQGVPGEAGAPGLVGPRGERG <u>FPGER</u> GKPGAQGLQGPRGLPGAPGFCGCFGAQGPPGLQG	644	3

TABLE 2

The polypeptide of the invention is preferably (A) a polypeptide having the amino acid sequence of SEQ ID NO:1, 2, or 3; (B) a polypeptide having the same amino acid sequence as the amino acid sequence of SEQ ID NO:1, 2, or 3 except that one or several amino acids are deleted, 35substituted and/or added, which polypeptide has a capacity to promote GAG production; or (C) a polypeptide having an amino acid sequence with a sequence identity of not less than 80% to the amino acid sequence of SEQ ID NO:1, 2, or 3, which polypeptide has a capacity to promote GAG  $_{40}$ production. The polypeptide of (C) is more preferably a polypeptide having an amino acid sequence with a sequence identity of not less than 90% to the amino acid sequence of SEQ ID NO:1, 2, or 3, which polypeptide has a capacity to promote GAG production; still more preferably a polypep- 45 tide having an amino acid sequence with a sequence identity of not less than 95% to the amino acid sequence of SEQ ID NO:1, 2, or 3, which polypeptide has a capacity to promote GAG production.

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Further, the polypeptide of the invention is preferably 50 (A1) a polypeptide composed of the amino acid sequence of SEQ ID NO:1, 2, or 3; (B1) a polypeptide composed of the same amino acid sequence as the amino acid sequence of SEQ ID NO:1, 2, or 3 except that one or several amino acids are deleted, substituted and/or added, which polypeptide has 55 a capacity to promote GAG production; or (C1) a polypeptide composed of an amino acid sequence with a sequence identity of not less than 80% to the amino acid sequence of SEQ ID NO:1, 2, or 3, which polypeptide has a capacity to promote GAG production. The polypeptide of (C1) is more 60 preferably a polypeptide composed of an amino acid sequence with a sequence identity of not less than 90% to the amino acid sequence of SEQ ID NO:1, 2, or 3, which polypeptide has a capacity to promote GAG production; still more preferably a polypeptide composed of an amino acid sequence with a sequence identity of not less than 95% to the 65 amino acid sequence of SEQ ID NO:1, 2, or 3, which polypeptide has a capacity to promote GAG production.

In the amino acid sequence of each of the polypeptide of (B) and the polypeptide of (B1), 1 or several amino acid residues may be deleted, substituted and/or added. Although the number of the amino acid residues to be deleted, substituted and/or added varies depending on the total number of amino acid residues in the particular polypeptide, the number may be from 2 to 15, preferably from 2 to 5.

The specific polypeptide can be produced by a gene recombinant technology known to those skilled in the art. Examples of the method which may be used for producing the polypeptide include the methods described in EP 0926543 A1, EP 1014176 A2, U.S. Pat. No. 6,992,172, WO 01/34646, WO 2004/85473, and WO 2008/103041. More specifically, a gene encoding the amino acid sequence of the polypeptide of interest is obtained, and the gene is then incorporated into an expression vector to prepare a recombinant expression vector. The prepared recombinant expression vector is introduced into an appropriate host to prepare a transformant. By culturing the obtained transformant in an appropriate medium, the polypeptide of interest is produced. By recovering the produced polypeptide from the culture, the particular polypeptide according to the invention can be obtained.

The capacity to promote GAG production can be evaluated by bringing the polypeptide into contact with cartilage cells, and then measuring the GAG production after a predetermined period of time.

Specific examples of the evaluation method include the following method.

The subject polypeptide is dissolved in water for injection (for dissolving polypeptide) such that the polypeptide is contained in a predetermined amount, for example, 0 µg/ml, 0.2 µg/ml, or 20 µg/ml, to prepare sample liquids. To each well of a 24-well plate (24 WELL NON-TRATED PLATE, BD Company), 625 µl of each of the obtained sample liquids is placed. The samples are fixed in the wells by air-drying at 25° C. to provide a test plate.

To the test plate, cartilage cells derived from Japanese white rabbits are seeded at 20,000 cells/well, and culture is

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performed at 37° C. under 5% (v/v)  $\rm CO_2$ . The culture supernatant is collected at Hour 2, Day 1, Day 2, Day 3, and Day 7 for quantification of GAG in the culture supernatant.

The quantification of GAG is carried out using a "SUL-FATED GLYCOSAMINOGLYCAN QUANTIFICATION <sup>5</sup> KIT" (trade name, Seikagaku Biobusiness Corporation).

In the quantification, the medium in the wells of the test plate is discarded, and washing is carried out once using 1 ml/well of phosphate buffered saline (PBS). To each well after washing, 150 µl of the protease liquid included the kit is added, and the liquid is then stirred using a plate shaker. Thereafter, treatment is carried out at 50° C. for 2 hours, and then at 100° C. for 10 minutes. To 50 µl of each sample, 50 µl of the reaction buffer II included in the kit is added, and the resulting mixture is mixed, followed by addition of 150 µl of a DMMB (dimethylmethylene blue) dye solution thereto. The same operations are carried out for GAG standard solutions. After 5 minutes of the reaction, the absorbance is measured at a wavelength of 530 nm using a 20 plate reader to perform quantification of GAG. The same operations are carried out for natural form of type II collagen. The amount of GAG in the case in which the subject polypeptide was used is compared with the amount of GAG in the case in which natural form of type II collagen was 25 used, and, when the amount of GAG in the case in which the subject polypeptide was used is larger than the amount of GAG in the case in which natural form of type II collagen was used, the subject polypeptide is evaluated as having a capacity to promote GAG production. The quantification of 30 GAG can also be carried out using a product equivalent to the above quantification kit, and examples of the equivalent product include the BLYSCAN GLYCOSAMINOGLYCAN ASSAY KIT (120 assays) (trade name, Biocolor Ltd., B1000). 35

[Scaffold Composition]

The scaffold composition according to the invention contains the specific polypeptide described above. As described above, the specific polypeptide contained in the scaffold composition can promote production of the matrix by cartilage cells when the polypeptide is brought into contact with the cartilage cells. Thus, the scaffold composition can promote the matrix production by cartilage cells.

The scaffold composition may contain, in addition to the specific polypeptide, one or more of other factors and the 45 like that are known to promote the matrix production. Examples of such other factors include basic fibroblast growth factor (bFGF), parathyroid hormone, transforming growth factor  $\beta$  (TGF $\beta$ ), insulin-like growth factor I (IGF-I), and insulin-like growth factor II (IGF-II). These other fac- 50 tors may be used singly, or in combination of two or more kinds thereof.

[Composition for Promoting GAG Production]

The composition for promoting GAG production according to the invention comprises the specific polypeptide 55 described above. As described above, the specific polypeptide can promote production of GAG by cells when the polypeptide is brought into contact with the cells. Thus, the specific polypeptide can be preferably employed as a composition for promoting GAG production for uses in which 60 promotion of GAG production is demanded.

Examples of the cells whose production of GAG is promoted by the composition for promoting GAG production according to the invention include cartilage cells, vascular endothelial cells, and corneal endothelial cells. The 65 composition for promoting GAG production is particularly preferably used for cartilage cells. 14

The composition for promoting GAG production may contain, in addition to the specific polypeptide, one or more of other factors and the like that are known to promote the matrix production. Examples of such other factors include bFGF, parathyroid hormone, TGF $\mu$ , IGF-I, and IGF-II. These other factors may be used singly, or in combination of two or more kinds thereof

[Composition for Cartilage Tissue Restoration and Composition for Cartilage Cell Culture]

As described above, the specific polypeptide, scaffold composition, and composition for promoting GAG production according to the invention promote production of a specific matrix by cells when the polypeptide or composition is brought into contact with the cells. Thus, the polypeptide and compositions can be applied to various uses. Examples of such uses include restoration or regeneration of damaged tissue, for example, damaged cartilage.

That is, the invention also includes a composition for cartilage tissue restoration and a composition for cartilage cell culture, containing the specific polypeptide. Examples of the cartilage in such a case include articular cartilage (in the knee, shoulder or hip joint), vertebral cartilage, auricular cartilage, and nasal septal cartilage. The composition for cartilage tissue restoration and the composition for cartilage cell culture, containing the specific polypeptide are particularly preferably used as compositions for restoration or regeneration of damaged cartilage in joints. Such use allows proliferation of cartilage cells and/or favorable repair of a cartilage tissue. The invention also includes a method for restoration or regeneration of damaged cartilage, comprising administering the composition for cartilage tissue restoration to the damaged area of cartilage.

[Other Uses]

The invention also includes uses of the specific matrixproducing polypeptide for production of a scaffold composition, composition for cartilage tissue restoration, composition for cartilage cell culture, or composition for promoting glycosaminoglycan production.

Further, the GAG production-promoting polypeptide, scaffold composition, and/or composition for promoting GAG production can be used for analyzing functions or properties of cells having a GAG production capacity, for example, cartilage cells, or for carrying out a test or study utilizing the functions or properties of these cells.

## EXAMPLES

The invention is described in detail by way of Examples. However, the invention is not limited to the Examples.

#### Examples 1 to 3

In order to produce GAG production-promoting polypeptides RCP #4 to RCP #6, which have the amino acid sequences of SEQ ID NO:1 to SEQ ID NO:3, polynucleotides (SEQ ID NO:5 to SEQ ID NO:7) having base sequences corresponding to the amino acid sequences of SEQ ID NO:1 to SEQ ID NO:3 were synthesized by an ordinary method. The obtained polynucleotides were amplified by polymerase chain reaction (PCR), and each of the resulting amplification products was introduced into pPICZ $\alpha$ A (Invitrogen), which is a plasmid containing the a-factor signal for protein secretion and the Zeocin resistance gene for selection, using an IN-FUSION HD CLON-ING KIT (Clontech Inc.).

*Pichia pastoris* cells were transformed with the obtained plasmid by electroporation, and transformed yeast strains were selected based on the resistance to an antibiotic Zeocin.

Polypeptides were produced based on the introduced polynucleotides according to the methods disclosed in EP- <sup>5</sup> A-0926543, EP-A-1014176, and WO 01/34646.

More specifically, the yeast strains obtained as described above were grown using the YNB (Yeast Nitrogen Base w/o amino acids) medium (BD Corporation), and then cultured in 3-L jar fermenters (B.E. Marubishi Co., Ltd.). More specifically, each yeast strain was first grown in a medium containing glycerol as a carbon source, and, from 1 hour before completion of the addition of glycerol, methanol was added as a carbon source to perform culture. After 96 hours of the culture, the culture supernatant was collected, and SDS-PAGE was carried out using the collected culture supernatant in order to confirm expression of the polypeptide of interest.

Culture supernatants for which expression of the polypeptides of interest could be confirmed were subjected to <sup>20</sup> purification with a cation-exchange chromatography CAPTO-S (trade name, GE Healthcare) and an anionexchange chromatography CAPTO-Q: (trade name, GE Healthcare) using an AKTA EXPLORER (trade name, GE Healthcare), to obtain polypeptides of interest RCP #4 to <sup>25</sup> RCP #6.

Properties of the polypeptides are show in Table 4. Each isoelectric point (pI) is a calculated value. The molecular weight was measured by ESI-MS (Q-TOF PREMIER, manufactured by Waters Corporation). The solubility of each polypeptide in water was not less than 2% by mass under normal pressure at 25° C.

In Table 4, "normal" as described for the amount of lysine means that each residue corresponding to a serine residue or threonine residue in the amino acid sequence of natural form<sup>35</sup> of human type II collagen is substituted by a glycine residue or alanine residue, and "high" means that each residue corresponding to a serine residue or threonine residue in the amino acid sequence of natural form of human type II collagen is substituted by a lysine residue.

Each identity indicates the identity to the amino acid sequence of natural form of human type II collagen. The symbol "\*" in Table 4 indicates that, in cases in which the polypeptide contained repeated sequences, the identity (%) was determined for the corresponding portions in the polypeptide sequence after exclusion of the repeated portion.

## Comparative Examples 1 to 4

As polypeptides for comparison, polypeptides RCP #7 and RCP #2, R-II collagen, and natural form of human type II collagen were prepared (Comparative Examples 1 to 4).

As shown in Table 3 and Table 4, the polypeptide RCP #7 (SEQ ID NO:8) has an amino acid sequence in which not less than 0.3 GVMGFP (SEQ ID NO:13) sequences are contained per 10 kDa. As shown in Table 3 and 4, the polypeptide RCP #2 (SEQ ID NO:9) has an amino acid sequence containing no GFPGER (SEQ ID NO:12)sequence. Each of the R-II collagen and the natural form of human type II collagen (SEQ ID NO:4) contains an amino acid sequence in which not more than 0.15 GFPGER (SEQ ID NO:12) sequences are contained per 10 kDa.

The polypeptides RCP #7 and RCP #2 were obtained in the similar manner as in Examples 1 to 3 except that the corresponding polynucleotides (SEQ ID NOs:10 and 11) were used.

The R-II collagen was obtained in the similar manner as in Examples 1 to 3 except that a polynucleotide having a base sequence corresponding to the amino acid sequence of SEQ ID NO:4 was provided. The serine residues and threonine residues were not substituted by other amino acid residues, and the identity to natural form of human type II collagen was 100%.

Properties of the polypeptides are shown in Table 3 and Table 4. In Table 4, "R-II" indicates the R-II collagen. The symbol "\*" in Table 4 indicates that, in cases in which the polypeptide contained repeated sequences, the identity (%) was determined for the corresponding portions in the polypeptide sequence after exclusion of the repeated portion. In Table 4, "Natural Form of type II collagen" means natural form of human type II collagen.

TABLE 3

	sequence	Number of residues	SEQ ID No.
RCP#7	GPQGARGQPGVMGPPGPKGANGEPGKAGEKGLPGAPGLRGLPGKDGEAGAAGPPGPAGPAGERGEQG APGPPGPQGLPGPPGPGEGGKPGDQGVPGEAGAPGLVGPRGERGFPGERGLPGAQGLQGPRGLPGA PGKDGPKGAAGPAGPPGAQGPPGLQGMPGERGAAGIAGPKGDRGDVGEKGPEGAPGKDGGRGLGGPI GPPGPAGANGEKGEVGPPGPAGAAGARGAPGERGEKGPPGPAGFAGPPGADGQPGAKGEQGEAQKG DAGAPGPQGPKGAPGPQGPAGVAGPKGARGAQGPPGAAGFPGAAGRVGPPGQLQGNPGPPGPPGAGK DGPKGARGDAGPPGRAGEPGLQGPAGPAGEKGEPGDDGPPGAEGPPGPQGLAQNPGPPGPAGAK GFPGLPGFKGEPGKQAPGAKGDRGPPGPVGPPGLAGFAGEPGREGGPGADGPPGRDGAAGVKGDRG EKGAVGAPGAPGPPGAPGPAGPPGPQGDRGEAGAQGPGPQGARGQPGVMGFPGPKGANGEPGKAGEK GLPGAPGLRGLPGKKDGEAGAAGPPGPAGPAGPAGERGEQGAPGPPGPQGLPGPPGEGGKPGDQGVPG EAGAPGLVGPRGERGFPGERGKPGAQGLQGPRGLPGAPGKDGPKGAAGPPGAQGPPGLQG	666	8
RCP#2	PGERGAAGIAGPKGDRGDVGEKGPEGAPGKDGGRGLGGPIGPPGPAGANGEKGEVGPPGPAGAAGAR GAPGERGEKGPPGPAGPAGPPGADGQPGAKGEQGEAGQKGDAGAPGPQGPKGAPGQQPAGVAGPKG ARGAQGPPGAAGFPGAAGRVGPPGLQGNPGPPGPPGPAGKDGPKGARGDAGPPGRAGEPGLQGPAGP PGEKGEPGDDGPPGAEGPPGPQGKAGQRGIVGLPGQRGERGFPGLPGPKGEPGKQGAPGAKGDRGPP GPVGPPGLAGPAGEPGREGGPGADGPPGRDGAAGVKGDRGEKGAVGAPGAPGPGPQG DRGEAGAQGPPGERGAAGIAGPKGDRGDVGEKGPEGAPGKDGGRGLGGPIGPPGPAGANGEKGEVGP PGPAGAAGARGAPGERGEKGPPGAPGFPGAPGADGQPGAKGEQGEAGQKGDAGAPGPQGPGG GPAGVAGPKGARGAQGPPGAAGPPGAAGPPGAQGNGDVGEKGPEGAPGPGPGPGPGAGANGEKGEVGP GPAGVAGPKGARGAQGPPGAAGPPGAAGPGPQGLAGQRGIVGLPGPGPGPAGANGEKGAPGPQ GPAGVAGPKGARGAQGPPGAAGPPGAAGPGPGPGQCLAGQRGIVGLPGQRGERGFPGLPGPKGEPGKQGA PGAKGDRGPPGPVGPPGLAGPAGEPGREGGPGADGPPGRDGAAGVKGDRGEKGAVGAPGAPGPPGPA GPAGPGPGQDRGEAGAQGP	690	9

<Evaluation>

The obtained polypeptides were evaluated as follows for their capacity to promote proliferation of cartilage cells, and their capacity to promote production of the extracellular matrix. Before the evaluation, test plates were prepared as 5 follows.

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(1) Preparation of Plates Coated with GAG Productionpromoting Polypeptide

Each of the polypeptides RCP #4 to #6, corresponding to Examples of the invention; and RCP #7, RCP #2, R-II collagen and natural form of human type II collagen, corresponding to Comparative Examples of the invention; was dissolved in a solution for dissolving RCP #4 to #7 and #2, and R-II collagen (water for injection), or in a solution for dissolving natural form of human type II collagen (acidic 15 solution prepared by adjusting the pH of distilled water to 3 with 1 M HCl) such that the polypeptide was contained at  $0.2 \,\mu\text{g/ml}$ ,  $2 \,\mu\text{g/ml}$ , or  $20 \,\mu\text{g/ml}$ , to prepare sample solutions. To each well of 24-well plates (24 well non-treated plate, BD Company), 625 ul of each of the obtained sample solutions 20 was placed. The samples were fixed in the wells by airdrying at 25° C. to prepare test plates.

(2) Evaluation of Proliferation of Cartilage cells For the evaluation of proliferation of cartilage cells, CHONDROCYTE CULTURE KIT (Code: CHC02) pur- 25 chased from Primary Cell Co., Ltd. was used.

To the test plates prepared as described above, cartilage cells derived from Japanese white rabbits, included in the kit, were seeded at 20,000 cells/well, and culture was performed at 37° C. under 5% (v/v)  $CO_2$ . For the culture, the 30 "differentiation medium" (RPMI1640, serum, ascorbic acid, etc.) included in the kit was used. Cartilage cells in each well were collected at Hour 2, Day 1, Day 2, Day 3, and Day 7 after the beginning of the culture, and the number of cartilage cells was quantified.

More specifically, the medium in the test plates was discarded, and washing was carried out once using 1 ml/well of PBS, followed by adding 150 µl of trypsin-EDTA to each well and leaving the plates to stand for 1 minute, thereby detaching the cells attached to the test plates. Into each well, 40 150 µl of the medium described above was added to prepare a cell suspension, and trypan blue was added thereto, followed by counting the number of live cells using a hemacytometer. The capacity to promote proliferation of cartilage cells was evaluated as follows based on the number 45 of obtained live cells. The results are shown in Table 4. In Table 4, "-" in the column showing the evaluation of proliferation of cartilage cells means that the evaluation was not carried out.

S: The number of cells was more than 125% with respect 50 to the number of cells obtained by the culture after addition of natural form of human type II collagen.

A: The number of cells was from more than 100% to 125% with respect to the number of cells obtained by the culture after addition of natural form of human type II collagen.

B: The number of cells was from more than 75% to 100% with respect to the number of cells obtained by the culture after addition of natural form of human type II collagen.

C: The number of cells was not more than 75% with respect to the number of cells obtained by the culture after addition of natural form of human type II collagen.

(3) Evaluation of Cartilage Matrix Production

In the similar manner as in the (2) described above, cartilage cells derived from Japanese white rabbits were cultured with each polypeptide in each test plate prepared in the (1) described above. GAG as a matrix was quantified at Hour 2, Day 1, Day 2, Day 3, and Day 7 after the beginning of the culture. The quantification of GAG was carried out using a "SULFATED GLYCOSAMINOGLYCAN QUAN-TIFICATION KIT" (Seikagaku Biobusiness Corporation).

More specifically, the medium in the wells of the test plates was discarded, and washing was carried out once using 1 ml/well of PBS, followed by adding 150 µl of the protease liquid included the kit to each well and stirring the liquid using a plate shaker. Subsequently, the reaction was allowed to proceed at 50° C. for 2 hours, and then at 100° C. for 10 minutes. To 50 µl of each sample, 50 µl of the reaction buffer II included in the kit was added, and the resulting mixture was mixed, followed by addition of 150 µl of a DMMB dye solution thereto. The same operations were carried out for the GAG standard solutions included in the kit. After 5 minutes of the reaction, the absorbance was measured at a wavelength of 530 nm using a plate reader (Sunrise (trade name) SUNRISE RAINBOW THERMO RC [model number], manufactured by TECAN Ltd.) to perform quantification of GAG. The results are shown in FIG. 1. The capacity to promote cartilage matrix production was evaluated as follows based on the amount of GAG. The results are shown in Table 4.

S: The amount of GAG produced was more than 125% with respect to the amount of GAG produced by the culture after addition of natural form of human type II collagen.

A: The amount of GAG produced was from more than 100% to 125% with respect to the amount of GAG produced by the culture after addition of natural form of human type II collagen.

B: The amount of GAG produced was from more than 75% to 100% with respect to the amount of GAG produced by the culture after addition of natural form of human type II collagen.

C: The amount of GAG produced was not more than 75% with respect to the amount of GAG produced by the culture after addition of natural form of human type II collagen

ΤA	BL	Æ	4

			Num-			Number of sequences contained           RGD(A)         GFPGER(B)         GVMGFP(									
	A-		ber of		Num- ber/	Num- ber/	Num- ber/	Num- ber/	Num- ber/	Num- ber/			Evalua	ation	-
	mount of ly- sine	pI	amino acid resi- dues	Molec- ular weight (kDa)	total length (se- quences)	10 kDa (se- quences)	total length (se- quences)	10 kDa (se- quences)	total length (se- quences)	10 kDa (se- quences)	[(B) +		Ma- trix produc- tion	Cell pro- lifer- ation	SEQ ID No.
RCP#4 RCP#5 RCP#6	normal high high	5.48 9.14 9.14	506 506 644	45.0 45.3 57.8	2.00 2.00 2.00	0.44 0.44 0.35	1.00 1.00 2.00	0.22 0.22 0.35	1.00 1.00 0.00	0.22 0.22 0.00	1.00 1.00 1.00	94.9 94.9 94.7*	S S S	A B B	1 2 3

						TABI	LE 4-con	tinued									
		Num-		RG	Nu D(A)	umber of se GFPGI		ontained GVMC	FP(C)								
	A-	ber A- of		A-			Num- ber/	Num- ber/	Num- ber/	Num- ber/	Num- ber/	Num- ber/			Evalua	ation	-
	mount of ly- sine	pI	amino acid resi- dues	Molec- ular weight (kDa)	total length (se- quences)	10 kDa (se- quences)	total length (se- quences)	10 kDa (se- quences)	total length (se- quences)	10 kDa (se- quences)	(A)/ [(B) + (C)]		Ma- trix produc- tion	Cell pro- lifer- ation	SEC ID No		
RCP#7	high	9.34	666	59.8	2.00	0.33	2.00	0.33	2.00	0.33	0.50	94.9*	А	В	8		
RCP#2	high	8.62	690	61.4	4.00	0.65	0	0.00	0.00	0.00		94.2*	Α	в	9		
R-II		9.27	1014	90.5	3.00	033	1.00	0.11	1.00	0.11	1.50	100	С		4		
Natural form of type II collagen	_	9.27	1014	90.5	3.00	0.33	1.00	0.11	1.00	0.11	1.50	100	А	Α	4		

As shown in Table 4 and FIG. 1, it was found that any of the polypeptides of the invention, RCP #4 to #6, promoted the GAG production significantly more efficiently than the natural form of human type II collagen. Moreover, any of the polypeptides of the invention, RCP #4 to #6, could promote the proliferation of cartilage cells equally to, or more efficiently than, the polypeptides of Comparative Examples 1 to 3.

Thus, the polypeptides RCP #4 to #6 were found to be scaffold compositions that are excellent in promotion of cartilage matrix production as well as in promotion of  $_{30}$  proliferation of cartilage cells.

It was also found that the cell proliferation capacity further increases when the pI is not more than 6.0 (see the result on RCP #4).

It was also found that, since the polypeptides RCP #4 to #6 were excellent in production of glycosaminoglycan and allowed proliferation of cartilage cells, these polypeptides

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Gly Glu Ala Gly Ala Pro Gly Leu Val Gly Pro Arg Gly Glu Arg Gly
          100
                             105
                                                   110
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can be used as compositions for cartilage tissue restoration, composition for cartilage cell culture, or composition for promoting glycosaminoglycan production

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Thus, the invention can provide a scaffold composition excellent in promotion of extracellular-matrix production by cartilage cells, a composition for cartilage tissue restoration, a composition for promoting glycosaminoglycan production, and a composition for cartilage cell culture, and a material therefor.

The disclosure of Japanese Patent Application No. 2012-213110, filed on Sep. 26, 2012, is hereby incorporated by reference in its entirety.

All the literatures, patent applications and technical standards described in the present description are hereby incorporated by reference to the same extent as in cases in which each literature, patent application or technical standard is concretely and individually described to be incorporated by reference.

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COILC	LIIUCU

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Asp Gly Glu Ala

The invention claimed is:

1. polypeptide which is

- (1) a polypeptide consisting of the amino acid sequence of SEQ ID NO: 1;
- (2) a polypeptide consisting of the amino acid sequence of SEQ ID NO: 1, except that 1 to 15 amino acids are <sup>35</sup> deleted, substituted and/or added, wherein said polypeptide has a capacity to promote glycosaminoglycan production;
- (3) a polypeptide consisting of the amino acid sequence of SEQ ID NO: 2;
- (4) a polypeptide consisting of the amino acid sequence of SEQ ID NO: 2, except that 1 to 15 amino acids are deleted, substituted and/or added, wherein said polypeptide has a capacity to promote glycosaminoglycan production;
- (5) a polypeptide consisting of the amino acid sequence of SEQ ID NO: 3; or
- (6) a polypeptide consisting of the amino acid sequence of SEQ ID NO: 3, except that 1 to 15 amino acids are deleted, substituted and/or added, wherein said poly-<sup>50</sup> peptide has a capacity to promote glycosaminoglycan production;
- wherein in the polypeptides of (1)-(6), the number of RGD sequences contained per molecular weight of 10 kDa is not less than 0.30, the number of GFPGER <sup>55</sup> sequences contained per molecular weight of 10 kDa is not less than 0.15, and the number of GVMGFP sequences contained per molecular weight of 10 kDa is less than 0.30.

2. A scaffold composition comprising the polypeptide according to claim 1.

**3**. A composition for cartilage tissue restoration, comprising the polypeptide according to claim **1**.

4. A composition for cartilage cell culture, comprising the polypeptide according to claim 1.

**5**. A composition for promoting glycosaminoglycan production, comprising the polypeptide according to claim **1**.

 A method for regeneration or restoration of cartilage comprising administering to a damage area of the cartilage
 the polypeptide according to claim 1.

7. A method for performing a cartilage cell culture comprising administering to said culture the polypeptide according to claim 1.

**8**. A method for promoting a cellular production of glycosaminoglycans in an extracellular matrix comprising administering to said matrix the polypeptide according to claim **1**.

**9**. The polypeptide according to claim **1**, having an isoelectric point (pI) of not more than 6.0.

10. The polypeptide according to claim 1, wherein the solubility of the polypeptide in water is at least 2% by mass.

11. The polypeptide according to claim 1, which consists of the amino acid sequence of SEQ ID NO:1, 2, or 3 except that 1 to 5 amino acids are deleted, substituted and/or added, wherein said polypeptide has a capacity to promote gly-cosaminoglycan production.

**12**. The polypeptide according to claim **1**, which consists of the amino acid sequence of SEQ ID NO:1, 2, or 3.

\* \* \* \* \*