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(54) N-TERMINAL TRUNCATED INSULIN (52) U.S. Cl.
ANALOGUES CPC

- (71) Applicant: CASE WESTERN RESERVE

INIVERSITY Cleveland OH (11S) (58) Field of Classification Search UNIVERSITY, Cleveland, OH (US)
- (72) Inventor: Michael A. Weiss, Moreland Hills, OH (US) (56) References Cited
- (73) Assignee: CASE WESTERN RESERVE FOREIGN PATENT DOCUMENTS UNIVERSITY, Cleveland, OH (US)
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- (86). PCT No.: PCT/US2O14/O12615 $\frac{1}{2}$ \$ 371 (c)(1), $\frac{1}{2}$ and $\frac{1}{2}$ are set of by examiner (2) Date: **Jul. 22, 2015** Primary Examiner — Gyan Chandra
- (87) PCT Pub. No.: **WO2014/116753** LLP
 $PCT Puh$ Dete: **Inl. 31, 2014** (57) PCT Pub. Date: Jul. 31, 2014 (57) ABSTRACT
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(2013.01)
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(21) Appl. No.: 14/762,712 Schwartz, Gerald and Katsoyannis, Panayotis G., "Synthesis of Des(tetrapeptide B1-4) and Des(pentapeptide B1-5) Human Insulins. Two Biologically Active Analogues'. American Chemical Society, 1978, pp.

EP14742940 Search Report dated Feb. 15, 2017.

(74) Attorney, Agent, or Firm — Hahn Loeser & Parks

An insulin analogue contains a foreshortened B-chain poly **Prior Publication Data** peptide lacking residues B1-B3 and optionally contains an (65) Prior Publication Data
additional substitution in the C-terminal B23-B30 segment
US 2015/0353621 A1 Dec. 10, 2015 of the B-chain. The insulin analogue lacking residues B1-B3 Dec. 10, 2015 of the B-chain. The insulin analogue lacking residues B1-B3 may contain substitutions at B28 and/or B29 that confer rapid action and optionally a non-standard substitution at Related U.S. Application Data B24. The analogue may be an analogue of a mammalian insulin, such as human insulin. A nucleic acid encoding such (60) Provisional application No. 61/755,020, filed on Jan. an insulin analogue is also provided. A method of treating a patient comprises administering a physiologically effective amount of the insulin analogue or a physiologically accept (51) Int. Cl. able salt thereof to a patient. A method of semi-synthesis is provided using an unprotected octapeptide by means of modification of an endogenous tryptic site by non-standard amino-acid substitutions.

A61K 38/00 (2006.01) 10 Claims, 7 Drawing Sheets

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N-TERMINAL TRUNCATED INSULIN ANALOGUES

CROSS-REFERENCE TO RELATED APPLICATION

This application claims priority to and the benefit of PCT/US2014/012615 filed Jan. 22, 2014, which claims the benefit of U.S. Provisional Application Ser. No. 61/755,020 incorporated by reference in their entirety. filed Jan. 22, 2013, the contents of which are hereby 10

STATEMENT REGARDING FEDERALLY SPONSORED RESEARCH OR DEVELOPMENT

This invention was made with government support awarded by the National Institutes of Health under grant numbers DK040949 and DK074176. The U.S. government has certain rights to the invention.

BACKGROUND OF THE INVENTION

This invention relates to polypeptide hormone analogues that exhibits enhanced pharmaceutical properties. Such as more rapid pharmacokinetics and/or augmented resistance 25 to thermal fibrillation above room temperature. More par ticularly, this invention relates to insulin analogues that are modified by deletion of residues B1-B3 in combination with other modifications of the A-chain or B-chain. Such non standard sequences may optionally contain standard amino- 30 acid substitutions at other sites in the A or B chains of an

insulin analogue.
The engineering of truncated and/or non-standard proteins, including therapeutic agents and vaccines, may have broad medical and societal benefits. An example of a medi-35 cal benefit would be optimization of the pharmacokinetic properties of a protein. An example of a further societal benefit would be the engineering of proteins more refractory than standard proteins with respect to degradation at or above room temperature for use in regions of the developing 40 world where electricity and refrigeration are not consistently available. An example of a therapeutic protein is provided by insulin. Analogues of insulin containing non-standard amino-acid Substitutions may in principle exhibit Superior properties with respect to pharmacokinetics or resistance to 45 thermal degradation. The challenge posed by the pharma cokinetics of insulin absorption following subcutaneous
injection affects the ability of patients to achieve tight glycemic control and constrains the safety and performance degradation is deepened by the pending epidemic of diabetes mellitus in Africa and Asia. These issues are often coupled following subcutaneous injection usually worsen the resistance of insulin to chemical and/or physical degradation. 55 Because fibrillation poses the major route of degradation above room temperature, the design of fibrillation-resistant formulations may enhance the safety and efficacy of insulin replacement therapy in Such challenged regions. The present invention pertains to the use of a truncated B-chain lacking 60 residues B1-B3 in combination with standard or non-stan dard substitutions elsewhere in the A-chain or B-chain to modify and improve distinct properties of insulin. During the past decade specific chemical modifications to the insu lin molecule have been described that selectively modify 65 one or another particular property of the protein to facilitate an application of interest. Whereas at the beginning of the of insulin pumps. The challenge posed by its physical 50

recombinant DNA era (1980) wild-type human insulin was envisaged as being optimal for use in diverse therapeutic contexts, the broad clinical use of insulin analogues in the past decade suggests that a suite of analogs, each tailored to address a specific unmet need, would provide significant medical and societal benefits.

15 Administration of insulin has long been established as a treatment for diabetes mellitus. Insulin is a small globular protein that plays a central role in metabolism in vertebrates. Insulin contains two chains, an A chain, containing 21 residues, and a B chain containing 30 residues. The hormone is stored in the pancreatic β -cell as a Zn^2 -stabilized hexamer, but functions as a Zn^{2+} -free monomer in the bloodstream. Insulin is the product of a single-chain precursor, proinsulin, in which a connecting region (35 residues) links the C-terminal residue of B chain (residue B30) to the N-terminal residue of the A chain (FIG. 1A). A variety of evidence indicates that it consists of an insulin-like core and disordered connecting peptide (FIG. 1B). Formation of three specific disulfide bridges (A6-A11, A7-B7, and A20-B19; FIGS. 1A and 1B) is thought to be coupled to oxidative folding of proinsulin in the rough endoplasmic reticulum (ER). Proinsulin assembles to form soluble Zn^{2+} -coordinated hexamers shortly after export from ER to the Golgi apparatus. Endoproteolytic digestion and conversion to insu lin occurs in immature secretory granules followed by morphological condensation. Crystalline arrays of zinc insulin hexamers within mature storage granules have been visualized by electron microscopy (EM). The sequence of insulin is shown in schematic form in FIG. 1C. Individual residues are indicated by the identity of the amino acid (typically using a standard three-letter code), the chain and sequence position (typically as a superscript).

The pharmacokinetic features of insulin absorption after subcutaneous injection have been found to correlate with the rate of disassembly of the insulin hexamer. Although not wishing the present invention to be constrained by theory, modifications to the insulin molecule that lead to accelerated disassembly of the insulin hexamer are thought to promote more rapid absorption of insulin monomers and dimers from
the subcutaneous depot into the bloodstream. A major goal of insulin replacement therapy in patients with diabetes mellitus is tight control of the blood glucose concentration
to prevent its excursion above or below the normal range characteristic of healthy human subjects. Excursions below the normal range are associated with immediate adrenergic or neuroglycopenic symptoms, which in severe episodes lead to convulsions, coma, and death. Excursions above the normal range are associated with increased long-term risk of microvascular disease, including retinapathy, blindness, and renal failure. Because the pharmacokinetics of absorption of wild-type human insulin following subcutaneous injection is often too slow and too prolonged relative to the physiological requirements of post-prandial metabolic homeostasis, considerable efforts have been expended during the past 20 absorption with pharmacodynamic effects that are more rapid in onset and less prolonged in duration. Examples of such rapid-acting analogues known in the art are $[Lys^{B28}]$, Pro B^{29}]-insulin (KP-insulin, the active component of Huma-log®), [AspB28]-insulin (Novalog®), and [Lys B3 , Glu B29]insulin (Apidra®). Although widely used in clinical practice, these analogues exhibit two principal limitations. First, although their pharmacokinetic and pharmacodynamic pro files are more rapid than those of wild-type insulin, they are not rapid enough in many patients to optimize glycemic control or enable the safe and effective use of algorithm

based insulin pumps (closed-loop systems). Second, the amino-acid Substitutions in these analogues impair the ther modynamic stability of insulin and exacerbate its suscepti bility to fibrillation above room temperature. Thus, the safety, efficacy, and real-world convenience of these prod ucts have been limited by a trade-off between accelerated absorption and accelerated degradation.

Fibrillation, which is a serious concern in the manufac ture, storage and use of insulin and insulin analogues for the treatment of diabetes mellitus, is enhanced with higher 10 temperature, lower pH, agitation, or the presence of urea, guanidine, ethanol co-solvent, or hydrophobic Surfaces. Cur rent US drug regulations demand that insulin be discarded if fibrillation occurs at a level of one percent or more. Because fibrillation is enhanced at higher temperatures, patients with 15 diabetes mellitus optimally must keep insulin refrigerated prior to use. Fibrillation of insulin or an insulin analogue can be a particular concern for Such patients utilizing an external insulin pump, in which Small amounts of insulin or insulin analogue are injected into the patient's body at regular intervals. In Such a usage, the insulin or insulin analogue is not kept refrigerated within the pump apparatus, and fibril lation of insulin can result in blockage of the catheter used
to inject insulin or insulin analogue into the body, potentially levels or even dangerous hyperglycemia. At least one recent report has indicated that insulin Lispro (KP-insulin, an analogue in which residues B28 and B29 are interchanged relative to their positions in wild-type human insulin; trade name Humalog®) may be particularly susceptible to fibril- 30 lation and resulting obstruction of insulin pump catheters. Insulin exhibits an increase in degradation rate of 10-fold or more for each 10° C. increment in temperature above 25° C.; accordingly, guidelines call for storage at temperatures $\leq 30^{\circ}$ C. and preferably with refrigeration.

The present theory of protein fibrillation posits that the mechanism of fibrillation proceeds via a partially folded intermediate state, which in turn aggregates to form an amyloidogenic nucleus. In this theory, it is possible that or may not stabilize the partially folded intermediate state and may or may not increase (or decrease) the free-energy barrier between the native state and the intermediate state. Therefore, the current theory indicates that the tendency of increase or decrease the risk of fibrillation is highly unpre dictable.

Initial steps in the formation of an amyloidogenic nucleus may involve non-native hydrophobic interactions between nonpolar side chains that are either exposed on the Surface 50 of native insulin or are transiently exposed due to partial protein unfolding. Residues B1, B2, and B3 (Phe^{$B1$}-Val^{$B2$}-Asn^{$B3$}) exhibit significant exposure in crystal structures of wild-type insulin and substantial variability in their posi tioning $(FIO. ZA)$. Residues $B1-B3$ are also less well 55 ordered in NMR studies of engineered insulin monomers in solution as indicated by ¹H-NMR resonance line widths, $¹³C-NMR$ chemical shifts, and paucity of long-range inter-</sup> residue nuclear Overhauser effects (NOEs) (FIG. 2B). Despite the conservation of these residues among mamma- 60 lian insulin sequences, studies of synthetic analogs has shown that residues B1-B5 may be deleted from the B-chain without significant loss of binding to the insulin receptor and without significant loss of biological activity. Studies of the folding and secretion of wild-type and mutant proinsulins 65 have nonetheless shown that residues B1-B5, singly and in combination, contribute to the efficiency and fidelity of

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disulfide pairing in the endoplasmic reticulum (Liu, M., Hua, Q. X., Hu, S. Q., Jia, W., Yang, Y., Saith, S. E., Whittaker, J., Aryan, P. & Weiss, M.A. (2010)). Deciphering the hidden informational content of protein sequences: fold ability of proinsulin hinges on a flexible arm that is dispens able in the mature hormone. J. Biol. Chem. 285, 30989 31001). In particular, diverse polar or charged amino-acid substitutions at position B1 have been found to impair the cellular folding and secretion of proinsulin in a human cell line in culture. Residue Phe^{B1} may therefore be regarded as a "hydrophobic anchor" in the folding process. The B1-B5 arm of the B-chain represents a key folding element within the pancreatic beta-cell.

There is a need, therefore for an insulin analogue that displays increased resistance to fibrillation above room temperature while retaining rapid hexamer disassembly and while exhibiting at least a portion of the activity of the corresponding wild-type insulin and maintaining at least a portion of its chemical and/or physical stability.

SUMMARY OF THE INVENTION

to inject insulin or insulin analogue into the body, potentially
resulting in unpredictable fluctuations in blood glucose 25 provide insulin analogues that provide augmented resistance amino-acid substitutions that stabilize the native state may 40 Asn^{oo} thus contain a foreshortened B-chain (2*t* residues). 35 through the action of trypsin on a precursor that contains Lys It is, therefore, an aspect of the present invention to to fibrillation above room temperature while retaining rapid hexamer disassembly and hence accelerated absorption fol lowing subcutaneous injection. The present invention addresses previous limitations for fast-acting insulin ana logues, namely, that they are more susceptible to fibrillation than wild-type insulin. The claimed invention exploits the dispensability of residues B1-B3 once disulfide pairing and protein folding have been achieved in the manufacturing process. Removal of residues B1-B3 is accomplished or Arg at position B3 in the place of the wild-type residue Asn^{B_3}. An example of such a precursor is the analog insulin glulisine, the active component of the product Apidra (Sanofi-Aventis). Analogs lacking residues Phe^{$B1$}-Val^{$B2$}-The foreshortened B-chain confers resistance to fibrillation above room temperature while enabling native-like binding to the insulin receptor.

a given amino-acid substitution in the insulin molecule to 45 B-chain by N-terminal removal of residues B1-B3 protects The essence of this invention is that foreshortening of the the protein from fibrillation above room temperature and may be accomplished following the efficient folding and disulfide pairing of a precursor polypeptide containing resi dues Phe B_1 , Val^{B2}, and either Lys or Arg at position B3 to introduce a new tryptic site. The tryptic product des- $[B1 -$ B3]-des-B23-B30]-insulin analogue provides in turn a precursor for the trypsin-mediated semi-synthesis of insulin analogs containing one or more substitutions or non-standard modifications in the C-terminal B23-B30 segment; critically, trypsin-mediated semi-synthesis "repairs" the C-terminal deletion of residues B23-B30 but does not restore the B1-B3 segment. This order of steps (i.e., folding prior to tryptic digestion) circumvents the folding defect otherwise introduced by removal of residues B1-B3 prior to disulfide pairing. The starting material may be a mature insulin analogue containing Lys or Arg at position B3, an insulin analogue containing Lys or Arg at position B3 and an additional substitution at A8, a proinsulin analogue containing Lys or Arg at position B3, proinsulin analogue contain ing Lys or Arg at position B3 and an additional substitution at A8, or corresponding single-chain biosynthetic precursor containing Lys or Arg at position B3 and whose post-folding

tryptic digestion yields a des-[B1-B3]-des-B23-B30]-insulin analogue amenable to semi-synthesis to provide a des-[B1-B3]-insulin analogue.

In general, the present invention provides an insulin analogue comprising a foreshortened B-chain polypeptide that lacks residues B1-B3 in combination with one or more additional substitutions in the A- or B-chain. Unless stated otherwise, the locations of amino acids should be under stood to be the location in comparison to wild type insulin. Therefore in a des- $[B1-B3]$ insulin analogue, the seventh amino acid in the B chain would still be denoted as being at position B10, for example. In one example, the foreshort ened B-chain polypeptide incorporates an Ornithine (Orn), Norleucine (Nle), or Glutamic acid (Glu) at position B29. In contains Lysine (Lys) at position B28 and Proline (Pro) at position B29. In another embodiment the foreshortened B-chain polypeptide contains Aspartic acid (Asp) at position B28 and either Nle or Orn at position B29. In yet another embodiment, the insulin analogue is a mammalian insulin 20 human proinsulin (SEQ ID NO: 1) including the A- and analogue, such as an analogue of human insulin. In one particular set of embodiments, the B-chain polypeptide comprises an amino-acid sequence, prior to trypsin treatment, of SEQ ID NO: 4 and polypeptides having three or fewer additional amino-acid substitutions thereof. In another 25 particular set of embodiments, after digestion and semi synthesis with trypsin, the resulting polypeptides may com prise a sequence selected from the group consisting of SEQ ID NOS: 5-7. In yet another particular set of embodiments. ID NOS: 5-7. In yet another particular set of embodiments, designated single-chain insulin analogues, the B-chain poly 30 peptide is part of a single extended polypeptide of length 51-86 that comprises an amino-acid sequence provided in SEQ ID NO: 9, and polypeptides having three or fewer additional amino-acid Substitutions thereof. 10 another embodiment the foreshortened B-chain polypeptide 15

In addition or in the alternative, the insulin analogue may 35 contain a non-standard amino-acid Substitution at position 24 of the B-chain. In one particular example, the non standard amino acid at B24 is Cyclohexanylalanine (Cha). In another particular example, the non-standard amino acid at B24 contains a halogenated aromatic ring, such as penta- 40 fluoro-Phe, 2-F-Phe, 2-C1-Phe, 2-Br-Phe, 4-F-Phe, 4-Cl Phe, or 4-Br-Phe where "2" designates the a single halogenic substitution at the ortho position of the aromatic ring of Phe and where "4" designates the para position of the aromatic ring of Phe. In addition or in the alternative, the insulin 45 analogue may contain an amino-acid Substitution at position A8 whereby the β -branched side chain in wild-type insulin (Thr⁴⁸) is substituted by a non- β -branched polar or charged side chain, such as Arg, Gln, Glu, His, or Lys.

Also provided is a nucleic acid encoding an insulin 50 analogue comprising a B-chain polypeptide that incorpo rates a non-standard amino acid at position B24 or B29 or both. In one example, the non-standard amino acid is encoded by a stop codon, such as the nucleic acid sequence TAG. An expression vector may comprise Such a nucleic 55 acid and a host cell may contain Such an expression vector.

Also provided is a nucleic acid encoding an insulin analogue comprising an A-chain polypeptide that incorpo rates a lysine (LyS) or Arginine (Arg) Substitution at position rise to a glutamate substitution at position B29 or an amino acid other than Thr at position A8. An expression vector may comprise Such a nucleic acid and a host cell may contain such an expression vector. A3 and optionally, other substitutions such as those giving 60

The method comprises administering a physiologically effective amount of an insulin analogue or a physiologically acceptable salt thereof to the patient, wherein the insulin analogue or a physiologically acceptable salt thereof con tains a foreshortened B-chain polypeptide lacking residues B1-B3 in combination of one or more additional substitu tions at sites B24, B28, B29, or A8 as described above. In one embodiment, the foreshortened B-chain in the insulin analogue administered to a patient contains Glu at position B29. In still another embodiment, the insulin analogue is a mammalian insulin analogue, such as an analogue of human insulin. In one particular set of embodiments, the B-chain polypeptide comprises an amino-acid sequence selected from the group consisting of SEQ ID NOS: 5-7 and poly peptides having three or fewer additional amino-acid Sub stitutions thereof.

BRIEF DESCRIPTION OF THE SEVERAL VIEWS OF THE DRAWINGS

FIG. 1A is a schematic representation of the sequence of B-chains and the connecting region shown with flanking dibasic cleavage sites (filled circles) and C-peptide (open circles).

FIG. 1B is a structural model of proinsulin, consisting of an insulin-like moiety and a disordered connecting peptide (dashed line).

FIG. 1C is a schematic representation of the sequence of human insulin (SEQ ID NOS: 2 and 3) indicating the position of residue B24 in the B-chain.

FIGS. 2A and 2B provide molecular models showing the potential conformational variability of the N-terminal arm of graphic T-state protomers (FIG. 2A) and ensemble of NMRderived solution structures (FIG. 2B). The arms are in each case shown as sticks against a space-filling model of one representative structure. Gray scale code is shown at upper right of FIG. 2A. Structures in FIG. 2A were obtained from PDB depositions 4INS, 1APH, 1BPH, 1CPH, 1DPH, 1G7A, 1TRZ, 1TYL, 1TYM and 1ZNI; structures in FIG. 2B were obtained from PDB depositions 1TYL and 1G7A.

FIG. 3 is a pair of graphs showing the results of receptor binding studies of insulin analogues. In FIG. 2A, relative activities for the B isoform of the insulin receptor (IR-B) are determined by competitive binding assay in which receptorbound ¹²⁵I-labeled human insulin is displaced by increasing concentrations of human insulin (\blacksquare) or its analogues: Orn^{B29}-insulin (\triangle) and des-[B1-B3]-Orn^{B29}-insulin ($\vec{\bm{\nabla}}$). In FIG. 2B, relative activities for the Type I IGF receptor (IGF-1R) are determined by competitive binding assay in which receptor-bound 125 I-labeled IGF-I is displaced by increasing concentrations of human insulin (\blacksquare) or its analogues: Orn^{$B29$}-insulin (\blacktriangle) and des-[B1-B3]-Orn^{$B29$}-insulin (∇).

(\overrightarrow{V}).
FIG. 4 is a graph showing the dose-response of des-[B1insulin (Lys^{B3}, Glu^{B29}) (\blacklozenge). The graph shows the rate of decrease of blood sugar levels in the first hour after admin istration versus the dose of insulin in nanomoles per 300 g rat.

DETAILED DESCRIPTION OF THE INVENTION

65 logue that provides resistance to fibrillation above room The present invention is directed toward an insulin ana temperature where the analogue retains rapid hexamer dis assembly and where the analogue then maintains at least a

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portion of biological activity of the corresponding unmodi fied insulin or insulin analogue.

The present invention pertains to removal of residues B1-B3 following the folding and disulfide pairing of a precursor molecule to improve the properties of the insulin 5 analogue with respect to the lag time prior to onset of fibrillation. In one instance the removal of residues B1-B3 is combined with one or more amino-acid Substitutions or modifications in the B23-B30 segment and in particular at positions B24, B28 and/or B29. 10

In one embodiment, the present invention provides an insulin analogue that provides more rapid hexamer disas sembly by substitution of phenylalanine at position B24 by a non-standard amino acid. Such as Cyclohexanylalanine (Cha) or a derivative of Phenylalanine containing a halogen 15 atom (F, Cl, or Br) at the 4 position of the aromatic ring. In another particular embodiment the des-[B1-B3] analogue is enhanced with respect to thermodynamic stability by a substitution of Phe $^{\hat{\beta}24}$ by a derivative containing a halogen atom (F, Cl, or Br) at the 2 position of the aromatic ring. In yet another embodiment the des-[B1-B3] analogue contains substitutions at positions B28 and/or B29 to confer more rapid rates of disassembly of the hexamer, examples known in the art are Asp^{$B28$} (as in Novolog®), Lys^{$B28$}-Pro $B29$ (as in Humalog®), and Glu^{B29} (as in Apidra®). The present inven- 25 tion thus provides des-[B1-B3] forms of these analogues. The present invention is not limited, however, to human insulin and its analogues. It is also envisioned that these substitutions may also be made in animal insulins such as porcine, bovine, equine, and canine insulins, by way of 30 non-limiting examples. In addition or in the alternative, the insulin analogue of the present invention may contain a non-standard amino-acid substitution at position 29 of the B chain, which is lysine (LyS) in wild-type insulin. In one particular example, the non-standard amino acid at B29 is 35 ornithine (Orn); the non-standard amino acid at B29 could also be norleucine (Nle). Removal of residues B1-B3 may also be combined with an A-chain substitution by a non beta-branched side chain at position A8, singly or in com bination with the above modifications at positions B24, B28, 40
or B29.

It has also been discovered that des-[B1-B3]- Orn^{B29} insulin confers protection from fibrillation relative to wild type insulin or Orn^{B29} -insulin without impairment of binding to the insulin receptor.

Furthermore, in view of the similarity between human and animal insulins, and use in the past of animal insulins in human patients with diabetes mellitus, it is also envisioned that other minor modifications in the sequence of insulin may be introduced, especially those Substitutions considered 50 "conservative." For example, additional substitutions of amino acids may be made within groups of amino acids with invention. These include the neutral hydrophobic amino acids: Alanine (Ala or A), Valine (Val or V), Leucine (Leu 55 or L), Isoleucine (Ile or I), Proline (Pro or P), Tryptophan (Trp or W), Phenylalanine (Phe or F) and Methionine (Met or M). Likewise, the neutral polar amino acids may be substituted for each other within their group of Glycine (Gly or G), Serine (Ser or S), Threonine (Thr or T), Tyrosine (Tyr $60₁$ or Y), Cysteine (Cys or C), Glutamine (Glu or Q), and Asparagine (ASn or N). Basic amino acids are considered to include Lysine (Lys or K), Arginine (Arg or R) and Histidine (His or H). Acidic amino acids are Aspartic acid (Asp or D) and Giluamic acid (Gilu or E). Unless noted otherwise or δ 5 wherever obvious from the context, the amino acids noted herein should be considered to be L-amino acids. Standard

amino acids may also be substituted by non-standard amino acids belong to the same chemical class. By way of non limiting example, the basic side chain Lys may be replaced
by basic amino acids of shorter side-chain length (Ornithine, Diamino butyric acid, or Diaminopropionic acid). Lys may also be replaced by the neutral aliphatic isostere Norleucine (Nle), which may in turn be substituted by analogues con taining shorter aliphatic side chains (Aminobutyric acid or Aminopropionic acid).

In one example, the insulin analogue of the present invention contains three or fewer conservative substitutions other than the removal of residues B1-B3 of the present invention.

As used in this specification and the claims, various amino acids in insulin or an insulin analogue may be noted by the amino-acid residue in question, followed by the position of the amino acid, optionally in superscript. The position of the amino acid in question includes the A- or B chain of insulin where the substitution is located. Thus, Phe^{B1} denotes a phenylalanine as the first amino acid of the B chain of wild-type insulin; Val^{B2} the second amino acid of the B-chain of wild-type insulin; and so forth until Thr^{B30} at the C-terminal position. On removal of residues B1-B3 (desig nated as des-[B1-B3]), this numbering is retained such that Glu^{B4} would be the first amino acid of the des-[B1-B3] B-chain, Phe^{$B24$} is the 21^{st} amino acid of the des-[B1-B3] B-chain, and so forth.

Although not wishing to be constrained by theory, the present invention envisions that residues B1-B3 defines a nonpolar arm that engages in non-native intermolecular interactions in the mechanism of formation of an amyloido genic nucleus, and so its removal delays or prevents on the onset of fibrillation above room temperature. Substitution of the wild-type B3 residue (Gln) by a basic side chain (Lys or Arg) confers a novel tryptic site in the B-chain of a precursor insulin analogue or in the B-domain of a precursor single chain polypeptide. Because this invention specifies the tryp tic removal of residues B1-B3 occurs only after folding and native disulfide pairing of a precursor polypeptide have been accomplished, the critical role of residues B1-B3 in these processes is retained. Further, because this arm is not required for receptor binding or biological activity, it may be removed from the mature analogue product intended as a treatment of diabetes mellitus by subcutaneous injection or by continuous infusion via an external or internal pump.

We have found that the B1-B3 segment may readily be removed from insulin glulisine ([Lys^{\tilde{B}^3}, Glu^{B29}]-insulin; the active component of Apidra®) by trypsin digestion to yield des-[B1-B3]-des-B23-B30]-insulin. The latter species was found to provide a starting material for the semi-synthesis of des- $[B1-B3]$ -Orn^{$B29$}-insulin by trypsin-mediated semi-synthesis in the presence of an excess of the octapeptide GFFYTPOT (SEQ ID NO:10) where O designates Ornithine (Orn). It is also envisioned that the N-terminal B-chain deletion of the present invention may be made in any of a number of existing insulin analogues. For example, the B1-B3 segment may be removed from insulin lispro ([Lys^{B28}, Pro^{B29}]-insulin, herein abbreviated KP-insulin), insulin aspart (Asp^{B28}-insulin) via analogues of these products or corresponding single-chain precursors containing Lys or Arg at B3. These analogues are described in U.S. Pat. Nos. 5,149,777 and 5,474,978. These analogues are each known as fast-acting insulins.

The amino-acid sequence of human proinsulin is pro vided, for comparative purposes, as SEQ ID NO: 1.

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The amino-acid sequence of a precursor B chain of human insulin may be modified with a substitution of a Lysine (Lys) or Arginine (Arg) at position B3 and optionally Glutamate (Glu) at position B29. An example of such a sequence is $50₁$ provided as SEQ ID NO: 4.

Phe-Tyr-Thr-Pro-Xaa₂-Thr [Xaa₁ is Arg or Lys; Xaa₂ is Lys or Glu]

Following folding and trypsin digestion of a precursor analogue or precursor single-chain insulin analog containing Arg or Lys at position B3 and following trypsin mediated semi-synthesis with a modified octapeptide, the resulting des-(B1-B3) modification may optionally be combined with non-standard substitutions at other positions such as B24 as provided in SEQ ID NOS: 5-7. 65

SEQ ID NO: 9 Phe-Val-Xaa₁-Gln-His-Leu-Cys-Gly-Ser-His-Leu-Val-

substitu-

Glu-Ala-Leu-Tyr-Leu-Val-Cys-Gly-Glu-Arg-Gly-Xaa

substitutions in a single-chain insulin analogue or proinsulin analogue having the amino-acid sequence represented by

Phe-Tyr-Thr-Xaa₃-Xaa₄-Thr-Xaa₅-Gly-Ile-Val-Xaa₆-

Gln-Cys-Cys-Xaa₇-Ser-Ile-Cys-Ser-Leu-Tyr-Gln-Leu-

Glu-Asn-Tyr-Cys-Asn;

SEQ ID NO: 9,

wherein Xaa₁ is Arg or Lys; wherein Xaa₂ is Phe, Cha, penta-fluoro-Phe, 2F-Phe, 2-Cl-Phe, 2-Br-Phe, 4-F-Phe, 4-Cl-Phe, or 4-Br-Phe; wherein Xaa₃ is Pro, Lys, or Asp; wherein Xaa₄ is Lys or Pro; wherein Xaa₆ is His or Glu; wherein Xaa_7 is His, Lys, Arg, Glu, Gln or Thr; wherein $Xaa₅$ is 0-34 of any amino acid or a break in the amino-acid chain such that the C-terminal residue is Thr; and wherein $Xaa₆$ is Glu, His, Arg or Lys.

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Trypsin-mediated semisynthesis employs synthetic octa peptides of SEQ ID NO: 10-15, which contain key receptorbinding determinants of intact insulin and share the property of not containing tryptic digestion sites.

Phe SEQ ID NO: 14_{20}

Gly-Xaa₁-Phe-Tyr-Thr-Pro-Orn-Thr [Wherein Xaa₁ is Cha, penta-fluoro-Phe, 2F-Phe, 2-Cl-Phe, 2-Br-Phe, 4F-Phe, 4-Cl-Phe, or 4-Br-Phe

SEQ ID NO: 15 $_{25}$ $\operatorname{Gly-Kaa_1-Phe-Tyr-Thr-Pro-Glu-Thr}$ [Wherein Xaa₁ is Cha, penta-fluoro-Phe, 2F-Phe, 2-Cl-Phe, 2-Br-Phe, 4F-Phe, 4-Cl-Phe, or 4-Br-Phe

> SEQ ID NO: 16 30

> > 35

Gly-Xaa $_1$ -Phe-Tyr-Thr-Asp-Xaa $_2$ -Thr [Wherein Xaa₁ is Cha, penta-fluoro-Phe, 2F-Phe, $2-Cl-Phe$, $2-Br-Phe$, $4F-Phe$, $4-Cl-Phe$, or $4-Br-Phe$; and wherein Xaa , is Ornithine, Diaminobutyric acid, Diaminoproprionic acid, Norleucine, Aminobutric acid, or Aminoproprionic acid

Analogues of insulin lacking residues B1-B3 were pre pared using insulin glulisine (SEQ ID NO:2 and SEQ ID NO:4, Lys 53 , Glu^{B29}) as starting material. Complete trypsin digestion of this analogue yielded des- $[B1-B3]$ -des-B23- ₄₀ Baselines were approximated by pre- and post-transition B30]-insulin, which was purified by reverse-phase highperformance liquid chromatography (HPLC). From this fragment, which contains the three native disulfide bridges of wild-type insulin, modified B23-B30 peptide segments fied by high-performance liquid chromatography (Mirmira, R. G., and Tager, H. S., 1989. J. Biol. Chem. 264: 6349-6354.) This protocol employs (i) a synthetic octapeptide representing residues (N)-GFFYTPOT or (N)-GFFYTPET tively) and (ii) truncated analogue des-tripeptide [B1-B3]des-octapeptide [B23-B30]-insulin or, in the case of Glu⁴⁸-
insulin analogues, Glu^{48} -des-tripeptide [B1-B3]-des- Glu^{48} -des-tripeptide [B1-B3]-desoctapeptide [B23-B30]-insulin. Trypsin does not cleave after Orn. In brief, des-octapeptide (15 mg) and octapeptide (15 55 mg) were dissolved in a mixture of dimethylacetamide/1.4 butandiol/0.2 M Tris acetate (pH 8) containing 10 mM calcium acetate and 1 mMethylene diamine tetra-acetic acid (EDTA) $(35:35:30, v/v, 0.4 \text{ mL})$. The final pH was adjusted to 7.0 with $10 \mu L$ of N-methylmorpholine. The solution was 60 cooled to 12° C., and 1.5 mg of TPCK-trypsin was added and incubated for 2 days at 12°C. An additional 1.5 mg of trypsin was added after 24 hr. The reaction was acidified with 0.1% trifluoroacetic acid and purified by preparative reverse-phase HPLC (C4). Mass spectrometry using matrix- 65 assisted laser desorption/ionization time-of-flight (MALDI TOF; Applied Biosystems, Foster City, Calif.) in each case were attached by trypsin-catalyzed semi-synthesis and puri- 45 (substitutions underlined; SEQ ID NOS: 10 and 11, respec- 50

gave expected values (not shown). The general protocol for solid-phase synthesis is as described (Merrifield et al., 1982. Biochemistry 21: 5020-5031). 9-fluoren-9-yl-methoxy-car bonyl (F-moc)-protected phenylalanine analogues were pur chased from Chem-Impex International (Wood Dale, Ill.).

The above protocol was also employed to prepare des tripeptide $[B1-B3]$ analogues of human insulin containing Ornithine or Glutamic acid at position B29. The method of preparation of these analogues exploits non-standard amino acid substitutions at position 29 to eliminate the tryptic site ordinarily present within the C-terminal octapeptide of the B chain (i.e., between Lys^{$B29$} and Thr^{$B30$}) while maintaining a Proline at position 28. Pro B^{28} contributes to the stability of the dimer interface within the insulin hexamer, and so this method of preparation provides near-isosteric models of wild-type insulin in which other modifications may conve niently be incorporated without the need for cumbersome side-chain protection.

Circular dichroism (CD) spectra were obtained at 4°C. and/or 25°C. using an Aviv spectropolarimeter (Weiss et al., Biochemistry 39: 15429-15440). Samples contained ca. 25 uM DKP-insulin or analogues in 50 mM potassium phos phate (pH 7.4); samples were diluted to 5 μ M for guanidineinduced denaturation studies at 25° C. To extract free energies of unfolding, denaturation transitions were fitted by non-linear least squares to a two-state model as described by Sosnick et al., Methods Enzymol. 317: 393-409. In brief, CD data $\mu(x)$, where x indicates the concentration of denaturant, were fitted by a nonlinear least-squares program according tO

$$
\theta(x) = \frac{\theta_A + \theta_B e^{\left(-\Delta C_{H_2O}^2 - mx\right)/RT}}{1 + e^{\left(\Delta C_{H_2O}^2 - mx\right)/RT}}
$$

where x is the concentration of guanidine and where θ_A and v_B are baseline values in the native and unfolded states. Baselines were approximated by pre- and post-transition lines $\theta_A(x)=\theta_A^{H_2O}+m_Ax$ and $\theta_B(x)=\theta_B^{H_2O}+m_Bx$. The m values obtained in fitting the variant unfolding transitions are lower than the m value obtained in fitting the wild-type unfolding curve. To test whether this difference and apparent change in ΔG_u result from an inability to measure the CD signal from the fully unfolded state, simulations were performed in which the data were extrapolated to plateau CD values at higher concentrations of guanidine; essentially identical estimates of ΔG_u and m were obtained.

Relative activity is defined as the ratio of the hormone receptor dissociation constants of analogue to wild-type human insulin, as measured by a competitive displacement assay using 125 human insulin. Microtiter strip plates (Nunc Maxisorb) were incubated overnight at 4° C. with AU5 IgG (100 µl/well of 40 mg/ml in phosphate-buffered saline). Binding data were analyzed by a two-site sequential model. Data were corrected for nonspecific binding (amount of radioactivity remaining membrane associated in the pres ence of $1 \mu M$ human insulin. In all assays the percentage of tracer bound in the absence of competing ligand was less than 15% to avoid ligand-depletion artifacts. Representative data are provided in FIG. 3A; corresponding assays conducted with the Type I IGF receptor (IGF-1R) are shown in FIG. 3B.

The far-ultraviolet circular dichroism (CD) spectrum of the des-tripeptide $[B1-B3]$ analogue are similar to those of the parent analogues. Free energies of unfolding (ΔG_{μ}) at

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25° C. were estimated based on a two-state model as extrapolated to zero denaturant concentration. Lag time indicates time (in days) required for initiation of protein fibrillation on gentle agitation at 30° C. in zinc-free phos phate-buffered saline (pH 7.4).

The baseline thermodynamic stability of wild-type insulin and Orn^{B29} -insulin, as inferred from a two-state model of denaturation at 25° C., are indistinguishable: 3.5 ± 0.1 kcal/ mole and 3.5 ± 0.1 kcal/mole, respectively. CD-detected guanidine denaturation studies indicate that deletion of residues B1-B3 is associated with a small decrement in thermodynamic stability in the context of Orn^{B29} -insulin: ΔG_{ν} 3.2±0.1 kcal/mole, implying that $\Delta \Delta G_{\nu}$ 0.3±0.2 kcal/ mole. Nonetheless, the physical stability of the des-[B1-B3] analogue was found to be greater than that of wild-type insulin or Orn B29 -insulin as evaluated in triplicate during incubation in 300 μ M phosphate-buffered saline (PBS) at pH 7.4 at 30° C. under gentle agitation. The samples were observed for 10 days or until signs of precipitation or frosting of the glass vial were observed. Whereas the three tubes of wild-type insulin and three tubes of Orn^{B29} -insulin became cloudy in less than 4 days, the three tubes of des-[B1-B3]-Orn^{$B29$}-insulin stayed clear for at least 7 days. des-[B1-B3]-Orn^{o29}-insulin stayed clear for at least 7 days.
Similar comparison of fibrillation lag times between des-[B1-B3]-Glu^{2^{29}}-insulin and [Lys^{2^{19}}, Glu 2^{19}]-insulin (the active component of Apidra®) showed that removal of residues B1-B3 lengthened the lag time by at least a factor of two. 25

Dissociation constants (K_d) were determined as described by Whittaker and Whittaker (2005. *J. Biol. Chem.* 280: 20932-20936), by a competitive displacement assay using 125 I-Tyr⁴¹⁴-insulin (kindly provided by Novo-Nordisk) and the purified and solubilized insulin receptor (isoform B or A) in a microtiter plate antibody capture assay with minor modification; transfected receptors were tagged at their C-terminus by a triple repeat of the FLAG epitope (DYKDDDDK) and microtiter plates were coated by anti FLAG M2 monoclonal antibody (Sigma). The percentage of tracer bound in the absence of competing ligand was less than 15% to avoid ligand-depletion artifacts. Binding data were analyzed by non-linear regression using a heterologous competition model (Wang, 1995, FEBS Lett. 360: 111-114) to obtain dissociation constants. Results are provided in Table 1; dissociation constants are provided in units of nanomolar. Interestingly, removal of residues B1-B3 appears to enhance receptor binding by at least twofold. 30 35 40 45

TABLE 1.

Binding of Insulin Analogues to Insulin Receptor and IGF Receptor			
Protein	IR-B binding	IGF-1R binding	
insulin Orn^{B29} -insulin des -[B-B3]- $OmB29$ -insulin	$0.061 \pm 0.010 \text{ nM}$ 0.118 ± 0.018 nM 0.035 ± 0.006 nM	$6.4 \pm 1.0 \text{ nM}$ 7.5 ± 1.2 nM 3.2 ± 0.5 nM	

DR-B, B isoform of the insulin receptor; IGF-1R, Type 1 IGF receptor

Referring to FIG. 4, Glulisine-insulin (Apidra®, SEQ ID NOS:2 and 4, \blacktriangledown) and des-[B1-3] glulisine insulin (SEQ ID 60 NOS:2 and 5; \Box) were purified by HPLC, lyophilized to powder, and dissolved in insulin diluent. Male Lewis rats were injected subcutaneously at time–0 with either 0.875, 3.5 or 8.75 mM of glulisine or des-[B1-3 glulisine in 100 μ] of diluent; the higher dose is at the plateau of the wild-type 65 insulin dose-response curve whereas the lower dose corre sponds to about 40% maximal initial rate of glucose dis

posal. Injection of diluent alone was performed as a negative control. 6 rats were studied in each group. Blood was obtained from clipped tip of the tail at time 0 and at successive intervals up to 120 min. Blood glucose was measured using a Hypoguard Advance Micro-Draw meter. Blood glucose concentrations were observed to decrease. Any differences in initial rate or duration were not statisti cally significant.

A method for treating a patient comprises administering an insulin analogue containing a foreshortened des-tripep tide [B1-B3] B-chain as known in the art or described herein. It is another aspect of the present invention that des tripeptide [B1-B3]-des-octapeptide [B23-B30]-insulin may readily be obtained by tryptic digestion of a two-chain or single-chain precursor polypeptide following its folding to achieve native disulfide pairing. It is yet another aspect of the present invention that use of non-standard amino-acid substitutions enables a rapid and efficient method of preparation of insulin analogues by trypsin-mediated semi-synthesis using unprotected octapeptides.

In still another example, the insulin analogue is admin istered by an external or implantable insulin pump. An insulin analogue of the present invention may also contain other modifications, such as a tether between the C-terminus of the B-chain and the N-terminus of the A-chain as described more fully in co-pending U.S. patent application Ser. No. 12/419,169, the disclosure of which is incorporated by reference herein.

A pharmaceutical composition may comprise Such insulin analogues and which may optionally include zinc. Zinc ions may be included in Such a composition at a level of a molar ratio of between 2.2 and 3.0 per hexamer of the insulin analogue. In Such a formulation, the concentration of the insulin analogue would typically be between about 0.1 and about 3 mM; concentrations up to 3 mM may be used in the reservoir of an insulin pump. Modifications of meal-time insulin analogues may be formulated as described for (a) "regular" formulations of Humulin® (Eli Lilly and Co.), Humalog® (Eli Lilly and Co.), Novalin® (Novo-Nordisk), and Novalog ® (Novo-Nordisk) and other rapid-acting insulin formulations currently approved for human use, (b) "NPH" formulations of the above and other insulin analogues, and (c) mixtures of such formulations. Analogues of insulin lacking residues $B1-B3$ and containing Glu^{B29} may also be formulated in the absence of zinc ions as known in the art for the formulation of insulin glulisine.

50 a pharmaceutical composition may be used to treat a patient Excipients may include glycerol, glycine, arginine, Tris, other buffers and salts, and anti-microbial preservatives such as phenol and meta-cresol; the latter preservatives are known to enhance the stability of the insulin hexamer. Such having diabetes mellitus or other medical condition by administering a physiologically effective amount of the composition to the patient.

55 polypeptide encoding an insulin analogue containing a A nucleic acid comprising a sequence that encodes a sequence encoding at least a B-chain of insulin with Arg or Lys at position B3 with a non-standard amino acid at position B24 is also envisioned. The latter can be accom plished through the introduction of a stop codon (such as the amber codon, TAG) at position B24 in conjunction with a suppressor tRNA (an amber suppressor when an amber codon is used) and a corresponding tRNA synthetase, which incorporates a non-standard amino acid into a polypeptide in response to the stop codon, as previously described (Furter, 1998, Protein Sci. 7:419-426; Xie et al., 2005, Methods. 36: 227-238). The particular sequence may depend on the pre ferred codon usage of a species in which the nucleic-acid sequence will be introduced. The nucleic acid may also

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encode other modifications of wild-type insulin. The nucleic-acid sequence may encode a modified A- or B-chain sequence containing an unrelated substitution or extension elsewhere in the polypeptide or modified proinsulin ana logues. The nucleic acid may also be a portion of an 5 expression vector, and that vector may be inserted into a host cell such as a prokaryotic host cell like an E. coli cell line, or a eukaryotic cell line such as S. cereviciae or Pischia pastoris strain or cell line.

For example, it is envisioned that synthetic genes may be synthesized to direct the expression of a B-chain polypeptide in yeast Piscia pastoris and other microorganisms. The nucleotide sequence of a B-chain polypeptide utilizing a stop codon at position B24 for the purpose of incorporating a Cyclohexanylalanine or halogen derivative of Phe at that position may be either of the following or variants thereof:

(a) with Human Codon Preferences:

 $$\texttt{(SEQ}\texttt{ID}\texttt{NO}:17$$ TTGTGXXXCAACACCTGTGCGGCTCACACCTGGTGGAAGCTCTCTACC

TAGTGTGCGGGGAACGAGGCTAGTTCTACACACCCAAGACC wherein XXX is AGG, AGA, CGG or CGC (encoding Arg), or AAG or AAA (encoding Lys)

(b) with Pichia Codon Preferences

 $(\verb|SEQ ID NO: 18)| \label{eq:2}$ TTGTT**XXX**CAACATTTGTGGTTCTCATTTGGTTGAAGCTTTGTACT

TGGTTTGTGGTGAAAGAGGTTAGTTTTACACTCCAAAGACT [wherein **XXX** is AGA, CGU or AGG (encoding Arg) or
AAG or AAA (encoding Lys)]

Similarly, a full length proinsulin cDNA having human codon preferences, encoding Arg or Lys at position B3 and utilizing a stop codon at position B24 for the purpose of incorporating Cyclohexanylalanine or a halogenated deriva tive of Phe at that position may have the sequence of SEQ 35 ID NO: 19.

(SEQ ID NO: 19)
TTTGT**XXX**CC AACACCTGTG CGGCTCACAC CTGGTGGAAG

CTCTCTACCT AGTGTGCGGG GAACGAGGCT AGTTCTACAC ACCCAAGACC CGCCGGGAGG CAGAGGACCT GCAGGTGGGG CAGGTGGAGC TGGGCGGCGG CCCTGGTGCA. GGCAGCCTGC AGCCCTTGGC CCTGGAGGGG TCCCTGCAGA AGCGTGGCAT TGTGGAACAA TGCTGTACCA GCATCTGCTC CCTCTACCAG CTGGAGAACT ACTGCAACTA. G. wherein XXX is AGG, AGA, CGG or CGC (encoding Arg), or AAG or AAA (encoding Lys)

Likewise, a full-length human proinsulin cDNA utilizing a stop codon at position B24 for the purpose of incorporating a Cyclohexanylalanine or halogenated derivative of Phe at that position and having codons preferred by P . pastor is may 55 have the sequence of SEQ ID NO: 20.

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(SEQ ID NO: 20)
TTTGTTXXXC AACATTTGTG TGGTTCTCAT TTGGTTGAAG

10 CTTTGTACTT GGTTTGTGGT GAAAGAGGTT AGTTTTACAC TCCAAAGACT AGAAGAGAAG CTGAAGATTT GCAAGTTGGT CAAGTTGAAT TGGGTGGTGG TCCAGGTGCT GGTTCTTTGC AACCATTGGC TTTGGAAGGT TCTTTGCAAA AGAGAGGTAT TGTTGAACAA TGTTGTACTT CTATTTGTTC TTTGTACCAA

TTGGAAAACT ACTGTAACTA A or AAG or AAA (encoding Lys)] [wherein XXX is AGA, CGU or AGG (encoding Arg)

Other variants of these sequences, encoding the same polypeptide sequence, are possible, given the synonyms in the genetic code.

Based upon the foregoing disclosure, it should now be apparent that insulin analogues provided will carry out the objects set forth hereinabove. Namely, these insulin ana logues exhibit enhanced resistance to fibrillation while retaining rapid action due to hexamer disassembly and maintaining at least a fraction of the biological activity of wild-type insulin. It is, therefore, to be understood that any variations evident fall within the scope of the claimed invention and thus, the selection of specific component elements can be determined without departing from the spirit of the invention herein disclosed and described.

The following literature is cited to demonstrate that the testing and assay methods described herein would be under stood by one of ordinary skill in the art.

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Mirmira, R. G., and Tager, H. S. 1989. Role of the phenyl-
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- Sosnick, T. R., Fang, X., and Shelton, V. M. 2000. Appli cation of circular dichroism to study RNA folding tran sitions. Methods Enzymol. 317:393-409.
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	- Weiss, M.A., Hua, Q. X., Jia, W., Chu, Y.C., Wang, R. Y., and Katsoyannis, P. G. 2000. Hierarchiacal protein "un
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	- Whittaker, J., and Whittaker, L. 2005. Characterization of the functional insulin binding epitopes of the full length insulin receptor. J. Biol. Chem. 280: 20932-20936.
	- Xie, J. and Schultz, P. G. 2005. An expanding genetic code. Methods. 36: 227-238.

SEQUENCE LISTING

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 $<$ 220 > FEATURE: <221> NAME/KEY: MISC_FEATURE $<222>$ LOCATION: (2)..(2) <223> OTHER INFORMATION: Xaa is Cyclohexanylalanine, penta-fluoro-Phe, 2F-Phe, 2-Cl-Phe, 2-Br-Phe, 4F-Phe, 4-Cl-Phe, or 4-Br-Phe $<$ 220 > FEATURE: <221> NAME/KEY: MISC_FEATURE <222> LOCATION: (7) .. (7) <223> OTHER INFORMATION: Xaa is Glu, Lys, Ornithine, Diaminobutyric acid, Diaminoproprionic acid, Norleucine, Aminobutric acid, or Aminoproprionic acid <400> SEQUENCE: 16 Gly Xaa Phe Tyr Thr Asp Xaa Thr $\mathbf 1$ 5 <210> SEQ ID NO 17 $<$ 211> LENGTH: 90 $<$ 212> TYPE: DNA <213> ORGANISM: Homo sapiens $<$ 220 > FEATURE: <221> NAME/KEY: misc_feature $<222>$ LOCATION: (6)..(8) <223> OTHER INFORMATION: NNN is AGG, AGA, CGG, CGC, AAG or AAA $<$ 220 > FEATURE: <221> NAME/KEY: misc_feature $< 222 >$ LOCATION: (9)..(9) <223> OTHER INFORMATION: n is a, c, g, or t $<400>$ SEOUENCE: 17 tttgtgnnnc aacacctgtg cggctcacac ctggtggaag ctctctacct agtgtgcggg 60 gaacgagget agttetacae acccaagacc 90 $<$ 210> SEQ ID NO 18 $<$ 211> LENGTH: 90 $<$ 212> TYPE: DNA <213> ORGANISM: Homo sapiens $<$ 220 > FEATURE: <221> NAME/KEY: misc_feature $<222>$ LOCATION: (7) .. (9) <223> OTHER INFORMATION: NNN is AGA, CGU, AGG, AAG or AAA $<400>$ SEOUENCE: 18 tttgttnnnc aacatttgtg tggttctcat ttggttgaag ctttgtactt ggtttgtggt 60 gaaagaggtt agttttacac tccaaagact 90 $<$ 210> SEQ ID NO 19 < 211 > LENGTH: 261 $<$ 212> TYPE: DNA <213> ORGANISM: Homo sapiens $<$ 220 > FEATURE: <221> NAME/KEY: misc_feature $<222>$ LOCATION: (6)..(8) <223> OTHER INFORMATION: NNN is AGG, AGA, CGG, CGC, AAG or AAA $<400>$ SEQUENCE: 19 tttgtnnncc aacacctgtg cggctcacac ctggtggaag ctctctacct agtgtgcggg 60 gaacgagget agttetacae acceaagace egeegggagg eagaggaeet geaggtgggg 120 caggtggage tgggeggegg eeetggtgea ggeageetge ageeettgge eetggagggg 180 teeetgeaga agegtggeat tgtggaacaa tgetgtaeca geatetgete eetetaecag 240 ctggagaact actgcaacta g 261

<210> SEQ ID NO 20 < 211 > LENGTH: 260 $<$ 212> TYPE: DNA

What is claimed is:

analogue, the method comprising: providing a folded single-chain or two-chain insulin

- related precursor polypeptide containing Arg or Lys at position B3;
- adding trypsin to the folded single-chain or two-chain $_{25}$ insulin-related precursor polypeptide under conditions conducive to the proteolytic activity of trypsin to provide a reaction mixture;
- incubating the reaction mixture for sufficient time to yield a des-tripeptide $[B1-B3]$ -des-octapeptide $[B23-B30]$ - ₃₀ insulin intermediate product; adding a synthetic peptide in molar excess, wherein the
- synthetic peptide comprises residues corresponding to residues B23-B30 of insulin or an analogue thereof;
- matic activity of trypsin such that trypsin catalyzes formation of a peptide bond between ArgB22 of the des-tripeptide [B1-B3]-des-octapeptide [B23-B30]-insulin intermediate product and the synthetic peptide to yield the des-[B1-B3]insulin analogue. adding trypsin under conditions conducive to the enzy- $_{35}$

2. The method of claim 1, wherein the folded single-chain or two-chain insulin-related precursor polypeptide is pre pared by recombinant expression in a host cell.

1. A method for preparing a des-tripeptide [B1-B3] insulin 20 analogue has the amino acid sequence of SEQ ID NO: 5, 6, or 7. 3. The method of claim 2, wherein the des-[B1-B3]insulin

> 4. The method of claim 1, wherein the B23-B30 peptide segment has an amino acid sequence selected from SEQ ID NOS: 10-16.

> 5. The method of claim 1, wherein the des-[B1-B3]insulin analogue further comprises a B29 substitution.

> 6. The method of claim 5, wherein the substitution at B29 is selected from Glu^{B29}, Orn^{B29}, Diaminobutyric acid^{B29}, Diaminoproprionic acid^{B29}, Norleucine^{B29}, Aminobutric acid^{B29} , or Aminoprioprionic acid^{B29}

> 7. The method of claim 1, wherein the des-[B1-B3]insulin analogue further comprises a substitution at B24.

8. The method of claim 7, wherein the substitution at B24 is selected from Cha^{24}, penta-fluoro-Phe^{24}, 2F-Phe^{24}, 2-Cl-Phe^{B24}, 2-Br-Phe^{B24}, 4F-Phe^{B24}, 4-Cl-Phe^{B24}, or 4-Br- Phe^{B24} .

9. The method of claim 1, wherein the des-[B1-B3]insulin analogue further comprises a substitution at B10.

10. The method of claim 9, wherein the substitution at
 40 B10 is selected from a Asn^{B10} Pro^{B10} or Glu^{B10} substitu- $B10$ is selected from a Asp^{$B10$}, Pro $B10$, or Glu $B10$ substitution.