

(12) United States Patent

Ikuta et al.

(54) HUMAN ANTI-HUMAN INFLUENZA VIRUS ANTIBODY

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Related U.S. Application Data

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Jun. 19, 2009	(JP) .	 2009-146832

(51) Int. Cl.

C07K 16/10	(2006.01)
A61K 39/00	(2006.01)

- (52) U.S. Cl. CPC C07K 16/1018 (2013.01); A61K 2039/505 (2013.01); C07K 2317/14 (2013.01); C07K 2317/21 (2013.01); C07K 2317/34 (2013.01); C07K 2317/76 (2013.01)
- (58) Field of Classification Search None See application file for complete search history.

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(57)ABSTRACT

Provided is a human antibody having a neutralization activity against a human influenza virus. More specifically, provided is a human antibody which recognizes a highly conserved region in a human influenza A virus subtype H3N2 or a human influenza B virus and has a neutralization activity against the virus. The human antibody is a human anti-human influenza virus antibody, which has a neutralization activity against a human influenza A virus subtype H3N2 and binds to a hemagglutinin HA1 region of the human influenza A virus subtype H3N2, or which has a neutralization activity against a human influenza B virus, and includes, as a base sequence of a DNA encoding a variable region of the antibody, a sequence set forth in any one of SEQ ID NOS: 5 to 12.

12 Claims, 18 Drawing Sheets

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LOCATIONS OF PEPTIDE CHAINS RECOGNIZED BY B-1 AND D-1 MONOCLONAL ANTIBODIES IN CONFORMATIONAL STRUCTURE OF HA1 REGION OF HUMAN INFLUENZA A VIRUS SUBTYPE H3N2.



Figure 16



DAYS AFTER INFECTION



MADMINISTRATION OF ANTIBODY OR PBS DIADMINISTRATION OF INFLUENZA VIRUS EACH PLOT DENOTES INDIVIDUAL WEIGHT CHANGE

Figure 18



DAYS AFTER INFECTION





HUMAN ANTI-HUMAN INFLUENZA VIRUS ANTIBODY

TECHNICAL FIELD

The present invention relates to a human anti-human influenza virus antibody, which has a neutralization activity against a human influenza A virus subtype H3N2 and binds to a hemagglutinin HA1 region of the human influenza A virus subtype H3N2, or which has a neutralization activity against a human influenza B virus.

The present application is a continuation application of U.S. patent application Ser. No. 13/141,998, filed Jun. 24, 2011, which is a National Stage Application of PCT/JP2009/007159, filed Dec. 24, 2009, which claims priority of Japanese Patent Application No. 2008-330425, filed Dec. 25, 2008, and Japanese Patent Application No. 2009-146832, filed Jun. 19, 2009, which are incorporated herein by reference.

BACKGROUND ART

Influenza viruses belong to the Orthomyxoviridae family and are classified into three genera of types A, B, and C, 25 which are referred to as influenza A virus, influenza B virus, and influenza C virus, respectively. In general, the influenza virus often refers to the types A and B in particular. Differences among types A, B, and C are based on differences in antigenicity of an M1 protein and an NP protein among 30 proteins which form viral particles. Further, even though the influenza virus is classified into the same types A and B, each of the types is further classified into several subtypes or strains due to a difference in antigenicity of hemagglutinin (hereinafter, also referred to as "HA" simply) which is a 35 molecule on the surface of an envelope or a difference in antigenicity of neuraminidase (NA). Thus, for example, the influenza A virus is further classified into subtypes H1N1, H2N2, H3N2, and the like. The human influenza A virus periodically mutates HA and NA. Thus, vaccination corre- 40 sponding to the conventional subtype often cannot exert its expected effect.

HA in the influenza A virus is formed of a head region and a stem region which are different in structure, the head region includes a receptor-binding site for the virus to bind 45 to a target cell and is involved in hemagglutination activity of HA, and the stem region includes a fusion peptide required for membrane fusion between the envelope of the virus and an endosome membrane of a cell and is involved in fusion activity (Non Patent Literature 1). Most of anti-HA 50 antibodies, which recognize each of the influenza A virus subtypes H1N1 and H2N2, recognize the head region of HA. However, this region is most frequently mutated. Thus, these antibodies do not react with the subtypes of the human influenza A virus in common, and often lose their recogni-55 tion abilities along with an antigenic change of HA in the virus.

Patent Literature 1 and Non Patent Literature 2 disclose that a polypeptide was synthesized from the amino acid sequence of the stem region of HA from one type of the 60 influenza A virus subtype H3N2 and an antibody against this polypeptide was acquired. However, a viral neutralization activity was weak in such antibody (Patent Literature 1), and the polypeptide itself used as an antigen exhibited no reactivity with rabbit anti-virus serum obtained by immu-65 nization with the subtype H3N2, which was also problematic in antigenicity (Non Patent Literature 2).

If an antibody which is common to the subtypes of the influenza virus, recognizes an antigenic site which is hardly mutated in, for example, the HA or NA molecule, and has a neutralization activity against the influenza virus can be obtained, this can be utilized for diagnosing, preventing, and treating a disease caused by an infection with the virus, and the antigenic site itself becomes useful as a vaccine. There is disclosed an antibody which has a viral neutralization activity against influenza A virus subtypes H1N1 and H2N2 and exhibits no neutralization activity against the subtype H3N2 (Patent Literatures 2 and 3). There is also disclosed an antibody which recognizes a specific polypeptide sequence in the stem region of the influenza A virus subtype H3N2 and does not recognize the subtypes H1N1 and H2N2 (Patent Literature 4). There is also disclosed a human Fab antibody which neutralizes the influenza A virus subtype H3N2 (Patent Literature 5 and Non Patent Literature 3).

In the influenza viruses, the human influenza A virus causes worldwide epidemics and brings many deaths (Patent Literatures 2 to 4). The influenza A virus subtype H3N2 is a subtype which caused worldwide epidemics in the past, and there is a report that a strain resistant to a medicament such as amantadine having an anti-influenza virus action has been increasing in recent years (New York Times, Jan. 15, 2006). However, no report is available for an antibody which effectively exhibits a neutralization activity against a region highly conserved for about 20 years in the influenza A virus subtype H3N2 and in the influenza B virus.

CITATION LIST

Patent Literature

- [PTL 1] JP 59-501714 A
- [PTL 2] JP 06-100594 A
- [PTL 3] JP 07-265077 A
- [PTL 4] JP 07-304799 A
- [PTL 5] JP 2006-254777 A

Non Patent Literature

[NPL 1] Rev. Biochem., 56, 365-394 (1987)

[NPL 2] Cell, 28, 477-487 (1982)

[NPL 3] Microbiol. Immunol., 52, 162-170 (2008)

SUMMARY OF INVENTION

Technical Problem

It is an object of the present invention to provide a human antibody having a neutralization activity against a human influenza virus. More particularly, it is the object of the present invention to provide a human antibody which recognizes a highly conserved region in a human influenza A virus subtype H3N2 or a human influenza B virus and has a neutralization activity against the virus.

Solution to Problem

As a result of an extensive study for solving the abovementioned problems, the inventors of the present invention have achieved the production of a human anti-human influenza virus antibody by making a hybridoma from a peripheral blood mononuclear cell collected from a healthy donor vaccinated with an influenza vaccine and a cell capable of fusing with a human-derived lymphocyte with high efficiency, and selecting a cell producing an antibody which has a binding activity to an influenza virus-derived protein. The inventors have obtained an antibody of the present invention by selecting a human antibody which recognizes a highly conserved region particularly in a human influenza A virus subtype H3N2 or a human influenza B virus and has a 5 neutralization activity against the virus, from the obtained antibodies.

That is, the present invention includes the following. 1. A human anti-human influenza virus antibody, which has a neutralization activity against a human influenza A virus 10 subtype H3N2 and binds to a hemagglutinin HA1 region of the human influenza A virus subtype H3N2, or which has a neutralization activity against a human influenza B virus. 2. A human anti-human influenza virus antibody according to the item 1, in which the human anti-human influenza virus 15 antibody is free of a neutralization activity against human influenza A virus subtypes H1 and H2.

3. A human anti-human influenza virus antibody according to the item 1, in which the human anti-human influenza virus antibody, which has a neutralization activity against a human 20 influenza A virus subtype H3N2, has a neutralization activity against at least an A/Hiroshima/52/05 strain, and in which the human anti-human influenza virus antibody, which has a neutralization activity against a human influenza B virus, has a neutralization activity against at least a B/Malaysia/ 25 2506/04 strain.

4. A human anti-human influenza virus antibody according to any one of the items 1 to 3, in which an epitope recognized by the antibody includes a region including an amino acid sequence at positions 173 to 181 and/or a region 30 7. A human anti-human influenza virus antibody, including including an amino acid sequence at positions 227 to 239 counting from an N-terminus of an amino acid sequence which forms the hemagglutinin HA1 region of the human influenza A virus subtype H3N2.

5. A human anti-human influenza virus antibody according 35 to the item 4, in which the epitope includes an amino acid

sequence including an amino acid sequence set forth in SEQ ID NO: 1 or 2, or any of amino acid sequences having substitutions, deletions, additions, or insertions of one or two amino acids in the amino acid sequence:

1) NFDKLYIWG; and	(SEQ ID NO: 1)
2)	
KFDKLYIWG.	(SEQ ID NO: 2)

6. A human anti-human influenza virus antibody according to the item 4, in which the epitope includes an amino acid sequence including an amino acid sequence set forth in SEQ ID NO: 3 or 4, or any of amino acid sequences having substitutions, deletions, additions, or insertions of one or two amino acids in the amino acid sequence:

1) SSRISIYWTIVKP; and	(SEQ ID NO: 3)
2)	(CEO ID NO. 4)
PSRISIYWTIVKP.	(SEQ ID NO: 4)

a base sequence of a DNA encoding a variable region of the antibody, the base sequence being selected from a base sequence set forth in any one of SEQ ID NOS: 5 to 12, or including any one of base sequences having substitutions, deletions, additions, or insertions of one or more nucleotides in the base sequence:

(SEQ ID NO: 5) CTGTGCAGGCTCTGGATTCACGTTTAGTACTTACGCCATGACCTGGGTCCGCCAGGCTCCAG GACAGGGGCTGGAGTGGGTCTCCTCTATTAGCGGTAGTGGTGAAATTTCCTATTACGCAGAC TCCGTGAAGGGCCTGTTCACCATCTCCAGGGACAATTCCAAGGACACAGTGTTTCTGCAAAT GACCAGCCTGAGAGCCGAAGACACGGCCGTATATTACTGTGCGAAATCCGACGTTTGGGAGG GTTATCGACCCTCAAAAGATGCTCTTCATATGTGGGGGCCAAGGGACAATGGTCACCGTCTCT TCA 2) (SEQ ID NO: 6) GACGTCCAGATGACTCAGTCTCCATCCTCCCTGTCTGCATCTGTGGGAGACAGAGTCACCAT CACTTGTCGGGCAAGTCAGAGCGTGAGCAATTATGTGAATTGGTATCAACAGAAGCCAGGGA GAGCCCCTAGGCTCCTCATCTCTAGTGCGTCCAATTTGTGGGCTGGGGTCCCGCCAAGGTTC AGTGGCCGTGGAGAAGAGACAGACTTCACTCTCACCATCACCAGTCTGCAACCTGAAGATTC TGCAGTTTACTACTGTCAACAGAGGTTACAGTGACCTTCTCAGTTTCGGCGGAGGGACCAAGG TGGAGATCAAA;

3)

1)

(SEO ID NO: 7) CAGGTGCAGCTGGTGCAATCTGGGTCTGAGTTGAAGAAGCCTGGGGCCTCAGTGAAGGTTTC

CTGCAAGGCTTCTGGATACACCTTCACCTCTTATTCTATATATTGGGTGCGACAGGCCCCTG

- continued GACAAGGGCTTGAGTGGATGGATGGATGGATGGATCAACACCAACACTGGGAACCCAAGGTATGCCCAG GGCTTCACAGGACGGTTTGTCTTCTCCTTCGACACCTCTGTCAGCACGGCATATCTGGAGAT CAGCAGCCTAAAGGCTGAGGACACTGCCGTGTATTACTGTGCGAGGAAGCGGAGGAGAGGAGGAGATTACGATA TTTTGACTGGTTATTATTATTACTTTGACTACTGGGGCCAGGGAACCCTGGTCACCGTCTCC TCA:

4)

(SEQ ID NO: 8)

CAGACTGTGGTGACCCAGGAGCCATCGTTCTCAGTGTCCCCTGGAGGGACAGTCACACTCAC TTGTGGCTTGAGCTCTGGCTCAGTCTCTCCTAGTTACTACGCCAGCTGGTACCAGCAGACCC CAGGCCAGGCTCCACGCACGCTCATCTACAACACAAACACTCGCTCCTCTGGGGTCCCTGAT CGCTTCTCTGGCTCCTTCCTTGGGAGCGACGCTGCCTCACCATCACGGGGGCCCAGGCAGA TGATGAGTCTGATTATTTCTGTGTGCCTGTATATGCCTAGTGGCGATTGGGTTTTCGGCCGAG GGACCAAGCTGACCGTCCTAGGT;

5)

(SEQ ID NO: 9)

CAGGTGCAGTTGCAGGAGTCGGGCCCAGGACTGGTGAAGCCTTTACAGACCCTGTCCCTCAC CTGCGTTGTCTCTGGTGACTCCATCAGCAGGGGGGGGTGGTTACTACTGGAGTTGGGTCCGCCAGC CCCCAGAGAGGGGCCTGGAGTGGATTGGGGACATCTATCACAGTGGGGAGTACCAACTACAAC CCGGCCCTCAAGAGTCGAACTACCATCTCAGTAGAGACGGTCCAAGAAACCAGTTCTCCCCTGCA GCTGAACTCTGTGACCGCCGCAGACACGGCCGTGTATTACTGTGCCAGAGAGACCTCCACCTG ACTACAGTGACTACAAGGTTGGGAAGGGTTATTTTGACTACTGGGGCCAGGGAGCCCTGGTC ACCGTCTCCTCA;

6)

(SEQ ID NO: 10)

7)

(SEQ ID NO: 11) CAGGTGAAGTTGGTGCAGTCTGGCGGAGGGCGCAGTCCAGCCTGGGAGGTCCCTGAGACTCTC CTGTGAAGGCGTTGAGTGGATTCGACTTCACTGTGTATGACATCCACTGGGTCCGCCAGGCTCCAG GCAAGGGGCCTGAGTGGGCGCATCTATTTGGCATAACGGAGGAAAAGCATATTATGCGGAC TCCGTGAAGGGCCGATTCACCGTGTCCAGAGAACAATCCCCAGAAGACAGTGTATCTGCAAAT GAGTGGCCTGAGACCCGAGGACACGGCTACATATTACTGTGCGAGAGAGTTTCCTTTCATGG GCATCTATGACTACGGCATGGACGCCTGGGGCCAAGGGACCACGGTCACCGTCGCCTCA; and

8)

(SEQ ID NO: 12) CAGTCTGTGCTGGCTCAGCCACCCTCAGCGTCTGGGACCCCCGGGCAGAGGGGTCATCATCTC TTGTTCTGGAACCTCCTCCAACATCGGCGGTAATTCTGTCAACTGGTACCAGCAGCCCCCAG GGGCGGCCCCGAGACTCCTCATCTATACTACCGATCAGCGACCCTCAGGGGGTCCCTGACCGA TTCTCTGGCTCCCAAGTCTGGCACCTCTGCCTCCCTGGCCATCAGTGGGGCTCCCAATCTGAGGA 1)

8

-continued

TGAGGCTGATTATTACTGTGAAGTTTGGGATGACAGCCTGACTCGTCCGGTGTTCGGCCGGAG

GGACCAAGTTGACCGTCCTACGT .

8. A human anti-human influenza virus antibody according to the item 7, in which an antigen of the human anti-human influenza virus antibody is a human influenza virus A/Hiroshima/52/05 strain or B/Malaysia/2506/04 strain.

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9. A human anti-human influenza virus antibody according 10 to the item 7, in which the antibody having a variable region encoded by DNA set forth in any one of SEQ ID NOS: 5 to 8 among the DNAs each encoding a variable region of the antibody includes an antibody having a neutralization activity against a human influenza A virus subtype H3N2, and in 15 which the antibody having a variable region encoded by DNA set forth in any one of SEQ ID NOS: 9 to 12 includes a neutralization activity against a human influenza virus antibody according to the item 8, in which the DNA set forth in any one of SEQ

ID NOS: 5, 7, 9 and 11 among the DNAs each encoding a variable region of the antibody includes a DNA encoding a heavy chain variable region, and the DNA set forth in any one of SEQ ID NOS: 6, 8, 10, and 12 includes a DNA encoding a light chain variable region.

- 11. A human anti-human influenza virus antibody according to any one of the items 1 to 11, in which the antibody includes an intact antibody.
- 12. A DNA, which encodes a variable region of the human anti-human influenza virus antibody according to any one of the items 1 to 11, the DNA including a polynucleotide which is selected from a base sequence set forth in any one of SEQ ID NOS: 5 to 12 or which includes any one of base sequences having substitutions, deletions, additions, or insertions of one or more nucleotides in the base sequence:

3)

TGGAGATCAAA;

(SEQ ID NO: 7)

CAGGTGCAGCTGGTGCAATCTGGGTCTGAGTTGAAGAAGCCTGGGGCCTCAGTGAAGGTTTC CTGCAAGGCTTCTGGATACACCTTCACCTCTTATTCTATATATTGGGTGCGACAGGCCCCTG GACAAGGGCTTGAGTGGATGGATGGATCAACACCAACACTGGGAACCCCAAGCTATGCCCAG GGCTTCACAGGACGGTTTGTCTTCTCCTTCGACACCTCTGTCAGCACGGCATATCTGGAGAT CAGCAGCCTAAAGGCTGAGGACACTGCCGTGTATTACTGTGCGAGAGAGGGAGATTACGATA TTTTGACTGGTTATTATTATTATTACTTTGACTACTGGGGCCAGGGAACCCTGGTCACCGTCTCC TCA;

TGCAGTTTACTACTGTCAACAGAGTTACAGTGACCTTCTCAGTTTCGGCGGAGGGACCAAGG

4)

- continued TGATGAGTCTGATTATTTCTGTGTGCGTGTATATGCCTAGTGGCGATTGGGTTTTCGGCGGAG GGACCAAGCTGACCGTCCTAGGT;

5)

(SEQ ID NO: 9)

CAGGTGCAGTTGCAGGAGTCGGGCCCAGGACTGGTGAAGCCTTTACAGACCCTGTCCCTCAC CTGCGTTGTCTCTGGTGACTCCATCAGCAGGGGGGGGTGGTTACTACTGGAGTTGGGTCCGCCAGC CCCCAGAGAGGGGCCTGGAGTGGATTGGGGACATCTATCACAGTGGGAGTACCAACTACAAC CCGGCCCTCAAGAGTCGAACTACCATCTCAGTAGAGACGTCCAAGAAACCAGTTCTCCCTGCA GCTGAACTCTGTGACCGCCGCAGACACGGCCGTGTATTACTGTGCCAGAGAGCCTCCACCTG ACTACAGTGACTACAAGGTTGGGAAGGGTTATTTTGACTACTGGGGCCAGGGAGCCCTGGTC ACCGTCTCCTCA;

6)

(SEQ ID NO: 10)

7)

(SEQ ID NO: 11) CAGGTGAAGTTGGTGCAGTCTGGCGGAGGCGCAGTCCAGCCTGGGAGGCCCTGAGACTCTC CTGTGAGGCGTCTGGATTCGACTTCACTGTGTATGACATCCACTGGGTCCGCCAGGCTCCAG GCAAGGGGCCTGAGTGGGCGCATCTATTTGGCGAAGAGAAAAGCATATTATGCGGAC TCCGTGAAGGGCCGATTCACCGTGTCCAGAGAACAATCCCCAGAAGACAGTGTATCTGCCAAAT GAGTGGCCTGAGACCCGAGGACACGGCTACATATTACTGTGCGAAGAGAGTTTCCTTTCATGG GCATCTATGACTACGGCATGGACGCCTGGGGCCAAGGGACCACGGTCACCGTCGCCTCA; and

13. A composition, including the human anti-human influenza virus antibody according to any one of the items 1 to 11. 55

Advantageous Effects of Invention

The human anti-human influenza virus antibody of the present invention has a neutralization activity against a ⁶⁰ highly conserved region in each of the human influenza A virus subtype H3N2 and the human influenza B virus.

Specifically, in the antibodies of the present invention, the antibody against the human influenza A virus subtype H3N2 has a neutralization activity against at least a viral strain of 65 an A/Hiroshima/52/05 strain (isolated in 2005), and the antibody against the human influenza B virus has a neutral-

ization activity against at least a viral strain of a B/Malaysia/ 2506/04 strain (isolated in 2004). Further, the antibodies also have neutralization activities against influenza virus vaccine strains in various generations including: various viral strains from the human influenza A virus subtype H3N2, such as an A/Aichi/2/68 strain (isolated in 1968), an A/Guizhou/54/89 strain (isolated in 1989), an A/Wyoming/3/03 strain (isolated in 2003), an A/New York/55/04 strain (isolated in 2004), and an A/Hiroshima/52/05 strain (isolated in 2005); and various viral strains from the human influenza B virus, such as a B/Victoria/2/87 strain (isolated in 1987), a B/Malaysia/ 2506/04 strain (isolated in 2004), a B/Mie/1/93 strain (isolated in 1993), and a B/Shanghai/261/02 strain (isolated in 2002), respectively.

25

50

Further, when administered prophylactically or therapeutically in vivo, the antibody against the human influenza A virus subtype H3N2 of the present invention exhibits effects on a survival rate and a weight loss in at least an infection with the influenza virus A/Guizhou/54/89xA/PR/8/34 ⁵ (H3N2) strain.

That is, the antibody against the human influenza A virus subtype H3N2 of the present invention has an activity against a region which has been conserved for about 20 years, and also includes one having an activity against a region which has been conserved for 40 years or more. The antibody against the human influenza B virus has an activity against a region which has been conserved for 20 years or more. Meanwhile, an HI (hemagglutination inhibition) activity in the antibody of the present invention is equal to or less than the detection limit.

BRIEF DESCRIPTION OF DRAWINGS

FIG. 1 are images showing the results of confirming reactivity of a culture supernatant of each hybridoma producing an A-1, A-2, B-1, B-2, B-3, C-1, D-1, D-2, E-1, or E-2 monoclonal antibody against HA of each influenza virus by western blotting assay (Experimental Example 1-2).

FIG. 2 are graphs showing the results of confirming a neutralization activity of each hybridoma culture supernatant against influenza virus vaccine strains in various generations (Experimental Example 1-4).

FIG. **3** is a view showing a sequence of a heavy chain 30 variable region of the B-1 monoclonal antibody (Experimental Example 1-5).

FIG. **4** is a view showing a sequence of a light chain variable region of the B-1 monoclonal antibody (Experimental Example 1-5).

FIG. **5** is a view showing a sequence of a heavy chain variable region of the D-1 monoclonal antibody (Experimental Example 1-5).

FIG. **6** is a view showing a sequence of a light chain variable region of the D-1 monoclonal antibody (Experi- 40 mental Example 1-5).

FIG. 7 is a view showing a sequence of a heavy chain variable region of the E-2 monoclonal antibody (Experimental Example 1-5).

FIG. **8** is a view showing a sequence of a light chain 45 variable region of the E-2 monoclonal antibody (Experimental Example 1-5).

FIG. **9** is a view showing a sequence of a heavy chain variable region of the B-3 monoclonal antibody (Experimental Example 1-5).

FIG. **10** is a view showing a sequence of a light chain variable region of the B-3 monoclonal antibody (Experimental Example 1-5).

FIG. **11** are images showing the results of confirming cells infected with an influenza virus vaccine strain and stained 55 with a supernatant of each hybridoma producing the A-2, B-1, B-2, or D-1 monoclonal antibody (Experimental Example 1-6).

FIG. **12** are images showing the results of confirming cells infected with the influenza virus vaccine strain and stained 60 with a supernatant of each hybridoma producing the B-3 or E-2 monoclonal antibody (Experimental Example 1-7).

FIG. **13** is a view showing sequences of peptide chains (epitopes) recognized by the B-1 and D-1 monoclonal antibodies, respectively, in a hemagglutinin HA1 region of 65 each influenza virus vaccine strain from a human influenza A virus subtype H3N2 (Experimental Example 1-8).

FIG. **14** is a view showing locations of the peptide chains recognized by the B-1 and D-1 monoclonal antibodies, respectively, in a conformational structure of the hemagglutinin HA1 region of the human influenza A virus subtype H3N2 (Experimental Example 1-8).

FIG. **15** is a view showing a comparison between the peptide chains recognized by the B-1 and D-1 monoclonal antibodies, respectively, and epitopes in a database (Example 1-8).

FIG. **16** is a graph showing survival rates when each of the B-1 and D-1 monoclonal antibodies is prophylactically administered to five mice (Experimental Example 2-1).

FIG. **17** are graphs showing body weight changes when each of the B-1 and D-1 monoclonal antibodies is prophylactically administered to five mice. Each plot shows the body weight change of each mouse (Experimental Example 2-1).

FIG. **18** is a graph showing survival rates when each of the B-1 and D-1 monoclonal antibodies is therapeutically ²⁰ administered to five mice (Example 2-2).

FIG. **19** are graphs showing body weight changes when each of the B-1 and -1 monoclonal antibodies is therapeutically administered to five mice. Each plot shows the body weight change of each mouse (Experimental Example 2-2).

DESCRIPTION OF EMBODIMENTS

The present invention relates to a human anti-human influenza virus antibody having the following properties (a) to (c):

(a) having a neutralization activity against a viral A/Hiroshima/52/05 strain (isolated in 2005) from the human influenza A virus subtype H3N2, or a neutralization activity against a viral B/Malaysia/2506/04 strain (isolated in 2004) from the human influenza B virus;

(b) having an HI (hemagglutination inhibition) activity equal to or less than the detection limit; and

(c) having no neutralization activity against human influenza A virus subtypes H1N1 and H2N2.

The antibody of the present invention against the human influenza A virus subtype H3N2 has a neutralization activity against at least the A/Hiroshima/52/05 strain (isolated in 2005), and further, has neutralization activities against influenza virus vaccine strains in various generations including various viral strains from the human influenza A virus subtype H3N2, such as an A/Aichi/2/68 strain (isolated in 1968), an A/Guizhou/54/89 strain (isolated in 1989), an A/Wyoming/3/03 strain (isolated in 2003), and an A/New York/55/04 strain (isolated in 2004). Further, the antibody of the present invention against the human influenza B virus has a neutralization activity against at least the B/Malaysia/ 2506/04 strain (isolated in 2004), and further has neutralization activities against influenza virus vaccine strains in various generations including various virus strains derived from the human influenza B virus, such as a B/Victoria/2/87 strain (isolated in 1987), a B/Mie/1/93 strain (isolated in 1993), and a B/Shanghai/261/02 strain (isolated in 2002).

Among the antibodies of the present invention each having the above-mentioned properties, the antibody against the human influenza A virus subtype H3N2 recognizes a region including an amino acid sequence at positions 173 to 181 and/or a region including an amino acid sequence at positions 227 to 239 counting from the N-terminus of an amino acid sequence which forms the hemagglutinin HA1 region of the human influenza A virus. Here, the amino acid sequences which form the human influenza A virus subtype H3N2 are disclosed in GenBank Accession No. EU501660 for the A/Hiroshima/52/05 strain, in GenBank Accession No. D49963 for the A/Guizhou/54/89 strain, in GenBank Accession No. AY531033 for the A/Wyoming/3/03 strain, and in GenBank Accession No. EU501486 for the A/New York/55/04 strain.

The 227th amino acid counting from the N-terminus in the amino acid sequence which forms the hemagglutinin HA1 region of the human influenza A virus is serine (S) or proline (P), but such difference has no effect on the neutral-10ization activity. The amino acid at that position is also S or Pin the literature (Karoline et al Virology J. 2008, 5, 40). The variant at position 173 in the sequence is asparagine (N) or lysine (K), is K except in the A/Aichi strain isolated in 1968, and is K or glutamic acid (E) in the literature. Further, the 15 229th and 230th amino acids are different and are arginine (R) or glycine (G), and isoleucine (I) or valine (V), respectively, in the literature (Underwood, Mol. Immunol., 1987, 21, 7), and the 238th and 239th amino acids are different and are K or N, and P or R, respectively, in the sequences 20 registered in GenBank.

In the light of the foregoing, a peptide chain (epitope) according to the region including an amino acid sequence at positions 173 to 181 is specifically an amino acid sequence including the amino acid sequence set forth in SEQ ID NO: 1 or 2, or is formed of an amino acid sequence in which one or two amino acids may be substituted, deleted, added or

introduced in that sequence. A peptide chain (epitope) according to the region including an amino acid sequence at positions 227 to 239 is specifically an amino acid sequence including the amino acid sequence set forth in SEQ ID NO: 3 or 4, or is formed of an amino acid sequence in which one or two amino acids may be substituted, deleted, added, or introduced in that sequence.

1)	(950	тп	NO.	1)
N	FDKLYIWG	(SEQ	тр	110:	1)
2)		TD	NO	21
к	FDKLYIWG	(SEQ	ID	NO:	2)
3)		TD	NO	21
S	SRISIYWTIVKP	(SEQ	ID	NO:	3)
4)		TD	NO	4.)
P	SRISIYWTIVKP	(SEQ	тD	100 :	4)

Base sequences of DNAs encoding the variable region of the antibody of the present invention having the abovementioned properties are specifically shown below.

 ²⁵ A-1. Human Antibody (B-1) Against Human Influenza A Virus Subtype H3N2

Light chain variable region sequence:

(SEQ ID NO: 6) GACGTCCAGATGACTCAGTCTCCATCCTCCCTGCATCTGCGGAGACAGAGGCCACCAT

CACTTGTCGGGCAAGTCAGAGCGTGAGCAATTATGTGAATTGGTATCAACAGAAGCCAGGGA GAGCCCCTAGGCTCCTCATCTCTAGTGCGTCCAATTTGTGGGGCTGGGGTCCCGCCAAGGTTC AGTGGCCGTGGAGAAGAGACAGACTTCACTCTCACCATCACCAGTCTGCAACCTGAAGATTC TGCAGTTTACTACTGTCAACAGAGTTACAGTGACCTTCTCAGTTTCGGCGGAGGGACCAAGG TGGAGATCAAA.

A-2. Human Antibody (D-1) Against Human Influenza A Virus Subtype H3N2

Heavy chain variable region sequence:

(SEQ ID NO: 7) CAGGTGCAGCTGGTGCAATCTGGGTCTGAGTTGAAGAAGCCTGGGGCCTCAGTGAAGGTTTC CTGCAAGGCTTCTGGATACACCTTCACCTCTTATTCTATATATTGGGTGCGACAGGCCCCTG GACAAGGGCTTGAGTGGATGGATGGATCAACACCAACACTGGGAACCCAAGCTATGCCCAG GGCTTCACAGGACGGTTTGTCTTCTCCTTCGACACCTCTGTCAGCACGGCATATCTGGAGAT CAGCAGCCTAAAGGCTGAGGACACTGCCGTGTATTACTGTGCGAGAGAGGGAGATTACGATA

-continued TTTTGACTGGTTATTATTATTACTTTGACTACTGGGGGCCAGGGAACCCTGGTCACCGTCTCC TCA.

Light chain variable region sequence: (SEQ ID NO: 8) ${\tt CAGACTGTGGTGACCCAGGAGCCATCGTTCTCAGTGTCCCCTGGAGGGACAGTCACACTCAC}$ ${\tt TTGTGGCTTGAGCTCTGGCTCAGTCTCCTCCTAGTTACTACGCCAGCTGGTACCAGCAGACCC}$ ${\tt CAGGCCAGGCTCCACGCACGCTCATCTACAACACAAAACACTCGCTCCTCTGGGGTCCCTGAT}$ CGCTTCTCTGGCTCCTTCCTTGGGAGCGACGCTGCCCTCACCATCACGGGGGCCCAGGCAGA TGATGAGTCTGATTATTTCTGTGTGCTGTATATGCCTAGTGGCGATTGGGTTTTCGGCGGAG GGACCAAGCTGACCGTCCTAGGT.

B-1. Human Antibody (E-2) Against Human Influenza B Virus

Heavy chain variable region sequence:

CAGGTGCAGTTGCAGGAGTCGGGCCCAGGACTGGTGAAGCCTTTACAGACCCTGTCCCTCAC CTGCGTTGTCTCTGGTGACTCCATCAGCAGGGGGGGGTGGTTACTACTGGAGTTGGGTCCGCCAGC CCCCAGAGAGGGGCCTGGAGTGGATTGGGGACATCTATCACAGTGGGAGTACCAACTACAAC CCGGCCCTCAAGAGTCGAACTACCATCTCAGTAGAGACGTCCAAGAACCAGTTCTCCCTGCA GCTGAACTCTGTGACCGCCGCAGACACGGCCGTGTATTACTGTGCCAGAGAGCCTCCACCTG ACTACAGTGACTACAAGGTTGGGAAGGGTTATTTTGACTACTGGGGCCAGGGAGCCCTGGTC ACCGTCTCCTCA .

Light chain variable region sequence:

(SEO ID NO: 9)

(SEQ ID NO: 10) ${\tt GAAATTGTGTTGGCACAGTCTCCAGCCACCCTGTCTTTGTCTCCAGGGGAAAGAGCCACCCT}$ ${\tt CTCCTGCAGGGCCAGTGAGACCGTTGACACCTACTTAGCCTGGTACCAACAGAAACCTGGCC$ AGTGGCAGTGGGTCTGGGACAGACTTCACTCTCACCATCAGCGGCCTAGAGCCTGAAGATTT TGGAGATTAAA

B-2. Human Antibody (B-3) Against Human Influenza B⁴⁵ Virus

Heavy chain variable region sequence:

(SEO ID NO: 11)

CAGGTGAAGTTGGTGCAGTCTGGCGGAGGCGCAGTCCAGCCTGGGAGGTCCCTGAGACTCTC ${\tt CTGTGAGGCGTCTGGATTCGACTTCACTGTGTATGACATCCACTGGGTCCGCCAGGCTCCAG$ GCAAGGGGCTTGAGTGGGTGGCATCTATTTGGCATAACGGAGGAAAAGCATATTATGCGGAC TCCGTGAAGGGCCGATTCACCGTGTCCAGAGACAATCCCCAGAAGACAGTGTATCTGCAAAT GAGTGGCCTGAGACCCGAGGACACGGCTACATATTACTGTGCGAGAGAGTTTCCTTTCATGG ${\tt GCATCTATGACTACGGCATGGACGCCTGGGGGCCAAGGGACCACGGTCACCGTCGCCTCA}\,.$ Light chain variable region sequence: (SEO ID NO: 12) ${\tt CAGTCTGTGCTGGCTCAGCCACCCTCAGCGTCTGGGACCCCCGGGCAGAGGGTCATCATCTC}$ TTGTTCTGGAACCTCCTCCAACATCGGCGGTAATTCTGTCAACTGGTACCAGCACCCCCAG

GGGCGGCCCCGAGACTCCTCATCTATACTACCGATCAGCGACCCTCAGGGGTCCCTGACCGA

-continued

TTCTCTGGCTCCAAGTCTGGCACCTCTGCCTCCCTGGCCATCAGTGGGCTCCAATCTGAGGA

TGAGGCTGATTATTACTGTGAAGTTTGGGATGACAGCCTGACTCGTCCGGTGTTCGGCGGAG

GGACCAAGTTGACCGTCCTACGT.

The antibody of the present invention is not particularly limited as long as the antibody has the above-mentioned properties. For example, the base sequence encoding the 10 variable region of the human antibody against the human influenza A virus subtype H3N2 is not limited to the base sequence set forth in any one of SEQ ID NOS: 5 to 8 described above, and may be a sequence in which one or more nucleotides are substituted, deleted, added or intro- 15 duced in the above-mentioned sequence as long as the sequence has a neutralization ability against the human influenza A virus subtype H3N2. In an example of the substitution of one or more nucleotides, the base sequence may be varied due to, for example, codon degeneracy but 20 has the neutralization ability. Preferably, the antibody having a neutralization ability against the human influenza A virus subtype H3N2 further recognizes any one of the peptide chains (epitopes) described above. Further, the base sequence encoding the variable region of the human anti- 25 body against the human influenza B virus is also not limited to the base sequence set forth in any one of SEQ ID NOS: 9 to 12 described above, and may be a sequence in which one or more nucleotides are substituted, deleted, added or introduced in the above-mentioned sequence as long as the 30 sequence has a neutralization ability against the human influenza B virus. In an example of the substitution of one or more nucleotides, the base sequence may be varied different due to, for example, codon degeneracy but has the neutralization ability.

A monoclonal antibody obtained by a cell fusion method is typically derived from an immunized animal species, e.g., a mouse. A mouse antibody cannot exert its expected effect in a human because when administered to the human, the mouse antibody can be metabolized as foreign matter and 40 example, a so-called human combinatorial antibody library thus the half-life of the mouse antibody is relatively short in the human. Further, a human anti-mouse antibody (HAMA) generated against the administered mouse antibody elicits an immune response such as serum disease or other allergy which is inconvenient and dangerous for a patient. Thus, 45 medical and therapeutic values of the monoclonal antibody derived from another animal species are limited in a human. Therefore, when the monoclonal antibody is administered to a human as a pharmaceutical or the like, a human antibody is strongly desired.

A known method or any method to be developed in the future may be employed for a method of producing the human antibody. However, there is no standardized method capable of being universally applied to any human antibody, and various efforts are required for producing a human 55 antibody which exhibits a sufficient binding activity and neutralization activity against a certain antigen. For example, reference can be made to Sato, K. et al, Cancer Res., 53, 851-856, 1993 and JP 2008-161198 A. The type of the human antibody is not particularly limited, and may be 60 an Fab type or an intact type. The intact type antibody is desirable in order to effectively exert an antibody activity. The intact type antibody is not particularly limited, and can be an antibody in which a complementarity determining region (CDR) of the antibody is derived from an original 65 animal species and a constant region (C region) is derived from an appropriate human. In such chimera antibody, the

variable region including CDR of the antibody derived from an immunized animal is generally linked to the constant region of the human antibody. The chimera antibody can be easily constructed by genetic modification technology. A humanized antibody obtained by CDR grafting in which CDR of the antibody from the immunized animal species is grafted in the variable region of the human antibody may also be employed. The human antibody may have the human antibody having CDR derived from a human, and CDR derived from the immunized animal species in terms of antigenicity because amino acid substitutions frequently occur in the CDR region.

The human anti-human influenza virus antibody of the present invention having the above-mentioned properties (a) to (c) can be produced specifically by the following method. Peripheral blood mononuclear cells corresponding to 10 mL of blood are collected from a healthy donor 2 to 4 weeks after being vaccinated with an influenza vaccine. Hybridomas are made simply and efficiently by fusing the peripheral blood mononuclear cells with partner cells derived from a human, e.g., SPYMEG cells (manufactured by MBL) capable of highly efficient cell fusion with lymphocytes derived from a human by a method such as a polyethylene glycol method. A cell producing the antibody which has a binding activity to a protein derived from the influenza virus is selected from the hybridomas by an ELISA method in which a purified protein such as HA derived from the 35 influenza virus is immobilized or a staining method using a cell infected with the influenza virus as an antigen, thereby enabling to produce the human anti-human influenza virus antibody.

As another method of producing the human antibody, for in which human antibody fragments are presented on the surface of Escherichia coli phage is constructed, and the desired human antibody can be obtained by screening of the antibodies with biopanning. In this case, the desired antibody can be screened without immunological work in an animal.

The present invention also encompasses a composition including the antibody of the present invention. Further, when used for medical usages, the composition of the present invention may include an effective amount of one or more kinds of human anti-human influenza virus antibodies of the present invention and further a pharmaceutically acceptable carrier. In the present invention, a pharmaceutically acceptable salt is exemplified by the following salts.

There are given as base addition salts: alkali metal salts such as a sodium salt and a potassium salt; alkaline earth metal salts such as a calcium salt and a magnesium salt; for example, an ammonium salt; aliphatic amine salts such as a trimethylamine salt, a triethylamine salt, a dicyclohexylamine salt, an ethanolamine salt, a diethanolamine salt, a triethanolamine salt, and a procaine salt; aralkylamine salts such as an N,N-dibenzylethylenediamine salt; heterocyclic aromatic amine salts such as a pyridine salt, a picoline salt, a quinoline salt, and an isoquinoline salt; quaternary ammonium salts such as a tetramethylammonium salt, a tetraethylammonium salt, a benzyltrimethylammonium salt, a benzyltriethylammonium salt, a benzyltributylammonium salt, a

methyltrioctylammonium salt, and a tetrabutylammonium salt; basic amino acid salts such as an arginine salt and a lysine salt; and the like.

There are given as acid addition salts: inorganic acid salts such as a hydrochloric acid salt, a sulfuric acid salt, a nitric acid salt, a phosphoric acid salt, a carbonic acid salt, a hydrogen carbonate salt, and a perchloric acid salt; organic acid salts such as an acetic acid salt, a propionic acid salt, a lactic acid salt, a maleic acid salt, a fumaric acid salt, a tartaric acid salt, a malic acid salt, a citric acid salt, and an ascorbic acid salt; sulfonic acid salts such as a methanesulfonic acid salt, an isethionic acid salt, a benzenesulfonic acid salt, and a p-toluenesulfonic acid salt; acidic amino acid salts such as an aspartic acid salt and a glutamic acid salt; and the like.

Such composition can be orally or parenterally administered as a pharmaceutical composition. When administered orally, the human anti-human influenza virus antibody of the present invention can be used as any one of dosage forms 20 such as: solids such as tablets, powders, granules, and capsules; aqueous solutions; oil suspensions; or solutions such as syrups or elixirs. When administered parenterally, the human anti-human influenza virus antibody of the present invention can be used as an aqueous or oil suspension for 25 injection or a nasal solution. Commonly used excipients, binders, lubricants, aqueous solvents, oil solvents, emulsifiers, suspending agents, preservatives, stabilizers, and the like can be optionally used in its preparation.

EXAMPLES

To help understanding of the present invention, the present invention is specifically described below with reference to Examples and Experimental Examples, but it goes without saying that the present invention is not limited thereto.

Example 1

Preparation of Human Anti-Human Influenza Virus Antibody

In this example, a description is made of the preparation of a monoclonal antibody using a hybridoma.

1) Preparation of Viruses

Human influenza virus vaccine strains, i.e., A/New Caledonia/20/99 strain as A virus subtype H1N1, A/Hiroshima/ 52/05 strain as A virus subtype H3N2, and B/Malaysia/2506/ 04 strain as the human influenza B virus, given from 50 National Institute of Infectious Diseases were each used to infect MDCK (canine kidney epithelial cell line) cells, which were then cultured at 37° C. in the presence of trypsin for 2 to 3 days, and then the virus was collected. 55

2) Purification of Influenza Virus HA Antigen

An influenza virus HA antigen was purified by a method well-known to those skilled in the art. The above-mentioned influenza virus vaccine strains were each inoculated to an incubated chicken egg, which was then cultured at 33 to 35° C. for 2 days and subsequently left stand overnight at 4° C., 60 and then an infected allantoic fluid was collected. The fluid was then concentrated by ultrafiltration and the like, and viral particles were purified by a sucrose density gradient centrifugation method. That is, the fluid was ultracentrifuged at 35,000 rpm in 0 to 60% sucrose density gradient, and a 65 fraction around 40% of the sucrose density gradient was collected. This concentrated viral fraction was treated with

ether followed by adding formalin, and further purified by the sucrose density gradient centrifugation method to obtain the influenza HA antigen.

3) Preparation of Hybridoma

10 mL of peripheral blood were collected from a healthy donor 2 to 4 weeks after being vaccinated with an influenza vaccine prepared from a seasonal influenza vaccine in 2006/2007, specifically, an A/Hiroshima/52/05 strain as the human influenza A virus subtype H3N2 or a B/Malaysia/ 2506/04 strain as the human influenza B virus. A mononuclear cell fraction was collected using Ficoll Paque Plus (manufactured by GE Healthcare) to use as a cell fraction for producing hybridomas. The mononuclear cell fraction was washed with serum-free DMEM before cell fusion to obtain mononuclear cells as cells for preparing hybridomas. SPYMEG cells (manufactured by MBL) as hybrid myeloma cells of murine myeloma cells and human megakaryoblasts were used as partner cells for making hybridomas. The SPYMEG cells were cultured in a DMEM medium supplemented with 10% fetal bovine serum for 2 days after passage, and the resultant was washed with serum-free DMEM before cell fusion.

Subsequently, the mononuclear cells obtained in the foregoing and the SPYMEG cells were mixed at a ratio of 1:5 to 1:10, and centrifuged to remove a supernatant. The precipitated cell mass was loosened sufficiently, subsequently 0.6 mL of a 50% polyethylene glycol 1500-PBS solution was slowly added thereto with stirring over 1 minute, and then 10 mL of serum-free DMEM was slowly 30 added thereto over 2 minutes. Further, 10 mL of serum-free DMEM were added followed by adding 1 mL of fetal bovine serum to complete the cell fusion. Subsequently, the resultant was centrifuged followed by removal of a supernatant, and the cells were washed with 20 mL of serum-free DMEM. Finally, the cells were gradually loosened, 120 mL of a hypoxanthine-aminopterin-thymidine (HAT) medium {HAT and additives for a human hybridoma medium, such as BM condimed (manufactured by Roche), were added to DMEM supplemented with 15% fetal bovine serum} were 40 added thereto, and the cells were gradually suspended using a measuring pipette.

4) Cloning of Hybridoma

The cell suspension of the above-mentioned section 3) was dispensed in six 96-well microplates for culture, and 45 cultured at 37° C. in an incubator containing 5% CO₂ for 10 to 14 days. A half amount of the HAT medium was changed every 3 to 4 days during this period. Subsequently, an aliquot of the culture supernatant was taken, and subjected to screening of the hybridomas.

The purified HA antigen (1 µg/well) prepared in the above-mentioned section 2) was immobilized on a 96-well microplate for ELISA, and further blocked with a PBS-0.1% Tween (TBS-T) solution of 5% defatted dry milk. Subsequently, 50 µL of the culture supernatant from the abovementioned cultured cell suspension were added to each well of the microplate for ELISA, and reacted at 37° C. for 30 minutes to form a primary immune complex by the purified HA antigen and the anti-HA antigen antibody (human antihuman influenza virus antibody) on the solid phase in the well. The human anti-human influenza virus antibody in the culture supernatant was detected by reacting the primary immune complex with a peroxidase-labeled goat anti-antibody followed by peroxidase reaction with color development.

Cells contained in each well in the microplate for culture, in which cells were confirmed to produce the antibody and proliferated, were taken out, and limiting dilution for the

cells was performed three times, and target cells were cloned by the same method as in the foregoing. The cloned hybridoma strains were designated as R1D8, K4E7, and G4G11

5) Purification of Antibody

Each hybridoma strain was finally cultured in serum-free medium by reducing the content of the fetal bovine serum in the medium from 10% to 2%. 100 mL of each hybridoma culture supernatant obtained by culturing in the serum-free medium for 3 to 7 days were centrifuged at 2,000 rpm for 10⁻¹ minutes, and the resulting supernatant was filtrated with a filter of 0.45 μ m to remove solid contents. The filtrate was purified by 1 mL of 6% agarose gel having immobilized thereon Protein G (HiTrap Protein G HPTM manufactured by GE Healthcare). The monoclonal antibodies produced by 1 the hybridoma strains R1D8, K4E7, and G4G11 were designated as B-1, D-1, and E-2, respectively. Likewise, monoclonal antibodies such as A-1, A-2, B-2, B-3, C-1, D-2, and E-1 were purified from other hybridoma supernatants.

Experimental Example 1-1

Characterization of Culture Supernatant of Each Hybridoma

1) Neutralization Activity

A neutralization activity of each hybridoma culture supernatant against each influenza virus was measured in accordance with Arch. Virol., 86, 129-135 (1985), Microbiol. Immunol., 29, 327-335 (1985).

MDCK cells were dispensed at 2×10⁴ cells/well in a 96-well microplate (for measuring a neutralization activity), and cultured overnight at 37° C. An 8-fold diluted solution of each antibody culture supernatant from which non-specific inhibitors had been removed by RDE treatment and a 35 viral solution obtained from the following influenza virus vaccine strain, each of which had been prepared so that a focus forming unit/well was 100, were mixed in equal amounts, and incubated at 37° C. for 1 hour. Subsequently, 30 μ L of this mixture were dispensed in each well of the 40 microplate, to which the cultured MDCK cells had been added, and incubated at 37° C. for 30 minutes. Subsequently, the solution in each well was removed, each well was washed with PBS, and fetal bovine serum-free MEM was added. The microplate was incubated at 37° C. for 6 to 10 45 hours. Then, the added solution was removed, and the cells were treated with absolute ethanol at room temperature for 10 minutes to fix the cells. Each well was dried, and the cells were stained by the same enzyme antibody staining method as the staining test. After being stained, the cells were 50 washed with tap water, and after being dried, the number of stained foci was counted under an optical microscope. The neutralization activity was represented by a focus reduction rate. The results were shown in Table 1.

Human influenza virus vaccine strain:

A. Influenza A: A/New Calcdonia/20/99 strain (H1), A/Hiroshima/52/05 strain (H3N2)

B. Influenza B: B/Malaysia/2506/04 strain

2) Hemagglutination Inhibition Activity (HI) of Antibody

 $25 \ \mu\text{L}$ of serial dilution (2 to 64 times) of the hybridoma ⁶⁰ culture supernatant treated with RDE were added to each well of a 96-well microplate (for measuring a hemagglutination inhibition activity (HI)), then were mixed with $25 \ \mu\text{L}$ of each of the viruses (8 HA units/50 μ L) used in the above-mentioned section 1), and reacted at room temperature for 30 minutes. Subsequently, 50 μ L of 0.75% guinea pig erythrocytes were added and mixed thoroughly to exam-

ine the effect of the antibody in the hybridoma culture supernatant on the hemagglutination activity of each virus. The results were shown in Table 1.

TABLE 1

	Human monoclonal antibodies against influenza virus					
10	HuMAb	ELISA ^a H1 H3 B	Staining of infected cells ^b H1 H3 B	Rec- ognized virus type	HI activity ^c H1/H3/B	Neutralization activity (% inhibition) ^c H1/H3/B
15	A-1 A-2 B-1 B-2 B-3 C-1 D-1	+ -+- -+- -+- +	+ -+- -+- -+- +	B AH3 AH3 AH3 B B AH3	<2 <2 <2 <2 <2 <2 <2 <2 <2 <2 <2 <2 <2 <	0 4.3 98.9 0 31.7 0 80.5
20	D-2 E-1 E-2	+ + +	+ + +	B AH1 B	<2 <2 <2 <2	16.7 0 37.8

^aAntigenic virus for ELISA: virus vaccine antigens of H1: A/New Caledonia/20/99 strain, H3: A/Hiroshima/52/05 strain, and B: B/Malaysia/2506/04 strain "Staining of infected cells: MDCX cells infected with H1: A/New Caledonia/20/99 strain, H3: A/Hiroshima/52/05 strain, or B: B/Malaysia/2506/04 strain and fixed with ethanol

were used. 'HI activity with neutralization activity: activity againist any one of H1: A/New Caledonia/ 20/99 strain, H3: A/Hiroshima/52/05 strain, and B: B/Malaysia/2506/04 strain recognized by the antibody was shown.

As a result of examining the neutralization activity and HI activity of the culture supernatant of the hybridoma producing each monoclonal antibody against each influenza virus, it was confirmed that the culture supernatant of the hybridoma producing the B-1 or D-1 monoclonal antibody had a high neutralization activity against the human influenza A virus subtype H3N2, and that the culture supernatant of the hybridoma producing the B-3 or E-2 monoclonal antibody had a slightly high neutralization activity against the human influenza B virus. However, the HI activity was equal to or less than the detection limit in all of the culture supernatants.

Experimental Example 1-2

Confirmation of Binding of Hybridoma Culture Supernatant to HA of Each Virus by Western Blotting

Western blotting assay of the culture supernatant of the hybridoma producing an A-1, A-2, B-1, B-2, B-3, C-1, D-1, D-2, E-1 or E-2 monoclonal antibody was performed for binding to HA of each virus. Plasma obtained from a patient with informed consent was used as a control. The purified HA antigen obtained in the section 2) of Example 1 was fractionated on SDS-PAGE, transferred onto a polyvinylidene fluoride (PVDF) membrane, and then blocked with 55 a TBS-T solution of 5% defatted milk. The membrane was incubated with undiluted hybridoma culture supernatant or the plasma diluted to 2,000 folds with PBS-T at room temperature for 1 hour to perform an antigen antibody reaction. The blotted membrane was washed several times with PBS-T, and subsequently incubated in a solution containing a peroxidase-conjugated anti-human IgG antibody at room temperature for 1 hour. Development was performed with an ECL detection kit (manufactured by Amersham Biosciences).

As a result, it was confirmed that the culture supernatants of the hybridomas producing the D-1 and B-1 monoclonal antibodies each had a binding activity to HA of the human

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influenza A virus subtype H3N2, and that the culture supernatants of the hybridomas producing the A-1 and C-1 monoclonal antibodies each had a binding activity to HA of the human influenza B virus (FIG. 1).

Experimental Example 1-3

Staining Activity of Hybridoma Culture Supernatants in Cells Infected with Various Influenza Virus Vaccine Strains

Staining activities in cells infected with the influenza virus vaccine strains in various generations shown below was examined for the supernatant of the hybridoma producing the A-2, B-1, B-2, D-1 as well as A-1, B-3, C-1, D-2, or 15 E-2 monoclonal antibody. The staining test was performed in accordance with the method described in J. Clin. Microbiol., 28, 1308-1313 (1990).

MDCK cells infected with the human influenza A virus subtype H3N2 strain or B virus strain shown below in a 20 96-well microplate were washed with PBS (pH 7.4), and then fixed with absolute ethanol at room temperature for 10 minutes, in the same manner as in the case of measuring the neutralization activity in the section 1) of Experimental Example 1. Each hybridoma culture supernatant containing 25 the monoclonal antibody was serially diluted at 4-fold dilutions. These were reacted sequentially with a 500-fold dilution of rabbit anti-human IgG serum (manufactured by Jackson), a 500-fold dilution of goat anti-rabbit IgG serum (manufactured by Cappel), and a 10,000-fold dilution of a peroxidase-rabbit anti-peroxidase complex (manufactured 30 by Cappel) each for 40 minutes, and washed with PBS. Finally, a peroxidase reaction was performed using a PBS solution of 0.01% H2O2 and 0.3 mg/mL of 3,3'-diaminobenzidine tetrahydrochloride in accordance with the Graham-Karnovsky method in J. Histochem. Cytochem., 14, 291-302 35 (1966), and the stained cells were observed under an ordinary optical microscope.

A. Influenza a Virus Subtype H3N2 Strains
Ai/68: A/Aichi/2/68 strain (isolated in 1968)
Gz/89: A/Guizhou/54/89 strain (isolated in 1989)
Wy/03: A/Wyoming/3/03 strain (isolated in 2003)
NY/04: A/New York/55/04 strain (isolated in 2004)
Hi/05: A/Hiroshima/52/05 strain (isolated in 2005)
B. Human Influenza B Virus Strains
Vi/87: B/Victoria/2/87 strain (isolated in 1987)
Ma/04: B/Malaysia/2506/04 strain (isolated in 2004)
Mi/93: B/Mie/1/93 strain (isolated in 1993)
Sh/02: B/Shanghai/261/02 strain (isolated in 2002)

TABLE 2

	Stai infec	ining activ cted with	vity of hyb influenza v	oridoma c vaccine st	ulture s rains in	upernata various	nts in cel generatio	ls ons	
			A H3N2			Vic	B toria- oup	E Yama gro	gata- up
HnMAb	Ai/68	Gz/89	Wy/03	NY/04	Hi/05	Vi/87	Ma/04	Mi/93	Sh/02
A-2	>128	>128	>128	>128	>128				
B-1	128	>128	>128	>128	>128				
B-2	<2	>128	>128	128	>128				
D-1	<2	>128	>128	>128	>128				
A-1						>128	>128	>128	>128
B-3						32	8	32	32
C-1						128	128	>128	>128
D-2						<2	<2	<2	2
E-2						>128	128	128	>128

The results in Table 2 confirmed that the culture supernatant of the hybridoma producing the A-2 or B-1 monoclonal antibody exhibited a high staining activity for the human influenza A virus subtype H3N2 strains, and it was also confirmed that the culture supernatant of the hybridoma producing the A-1, C-1, or E-2 monoclonal antibody exhibited a high staining activity for the human influenza B virus. These confirmed that these monoclonal antibodies had staining activities for the strains of the influenza A virus subtype H3N2 or the influenza B virus, even for those conserved for about 40 years or for 20 years or more.

Experimental Example 1-4

Neutralization Activity of Hybridoma Culture Supernatants Against Various Influenza Virus Vaccine Strains

Neutralization activities against cells infected with the influenza virus vaccine strains in various generations shown in Experimental Example 3 was examined for the supernatant of the hybridoma producing the B-1, D-1 as well as E-2, or B-3 monoclonal antibody. The neutralization activity was measured in accordance with the techniques in Experimental Example 1.

As a result, the culture supernatant of the hybridoma producing the B-1 monoclonal antibody exhibited neutralization activities against the various viral strains after A/Aichi/2/68 strain (isolated in 1968), and the culture supernatant of the hybridoma producing the D-1 monoclonal antibody exhibited neutralization activities against the various viral strains after A/Guizhou/54/89 strain (isolated in 1989) of the various viral strains derived from the human influenza A virus subtype 3H2N. The culture supernatant of the hybridoma producing the E-2 or B-3 monoclonal antibody exhibited neutralization activities against the various influenza B virus strains after B/Victoria/2/87 strain (isolated in 1987). These confirmed that these monoclonal antibodies exhibited neutralization activities against the viral strains from the influenza A virus subtype H3N2 or the influenza B virus, even for those conserved for about 40 40 years or for 20 years or more (FIG. 2).

Experimental Example 1-5

Base Sequence of Variable Region of Each Human Anti-Human Influenza Virus Antibody

From the results of the above-mentioned experiment, it was thought that the B-1, D-1, as well as E-2, or B-3

monoclonal antibody (human anti-human influenza virus antibody) could recognize a region conserved for 40 years or about 20 years in the human influenza A virus subtype H3N2 or the human influenza B virus. Thus, the base sequences encoding the variable regions of these monoclonal antibod-⁵ ies were examined in this experimental example.

1) Extraction of Total RNA and RT-PCR Specific for Human Antibody (IgG) Sequence

The hybridoma obtained by cloning as described above 10 was homogenized in a nucleic acid extraction reagent (RNAisoTM, TakaraBio), and total RNA was purified in accordance with the protocol of the kit.

cDNA was synthesized using a part of the total RNA sample after purification and using the following RT primer ¹⁵ or dT primer designed specifically for the sequences of the H chain and L chain constant regions of a gene encoding a human antibody (IgG).

Human_IgGH_RT_Primer: TGGAGGGCACGGTCACCACGC Human_IgGL_RT_Primer: TTGTGACGGGCGAGCTCAGGC (SEQ ID NO: 14) 25

RACE PCR was further performed using the following PCR primers located upstream of the above-mentioned primers.

```
Human_IgGH_PCR_Primer:

AAGGTGTGCACGCCGCTGGTC

Human_IgGL (\kappa)_PCR_Primer:

(SEQ ID NO: 15)

GTGCTGCTGAGGGCTGTAGGTG

Human IgL (\lambda) PCR Primer 1:

(SEQ ID NO: 17)

CCAYTGTCTTCTCCACRGTRCTCYC

Human IgL (\lambda) PCR Primer 2:

(SEQ ID NO: 18)

TCAGAGGAGGRYGGGAACAGAGTG
```

2) Purified PCR Products

The resulting PCR product was electrophoresed on 3% agarose gel, and the PCR product was purified from the resulting band. The base sequence of DNA was analyzed in the purified PCR product. The analysis was carried out by AB13730 Sequencer (ABI) using BigDye® Terminators v3.1 Cycle Sequencing Kit (ABI). As a result, the sequences represented in SEQ ID NOS: 5 to 12 shown below were identified for the heavy chain variable region sequence and light chain variable region sequence.
 ²⁵ and the light chain variable region sequence, and translated amino acid sequences therefrom for the each monoclonal antibody were as shown in FIGS. 3 to 10 (SEQ ID NOS: 19 to 26).

A-1. Human Antibody (B-1) Against Human Influenza a Virus Subtype H3N2

Heavy chain variable region sequence:

Light chain variable region sequence:

(SEO ID NO: 6)

GACGTCCAGATGACTCAGTCTCCATCCTCCCTGTCTGCATCTGTGGGAGACAGAGTCACCAT CACTTGTCGGGCAAGTCAGAGCGTGAGCAATTATGTGAATTGGTATCAACAGAAGCCAGGGA GAGCCCCTAGGCTCCTCATCTCTAGTGCGTCCAATTTGTGGGCTGGGGGTCCCGCCAAGGTTC AGTGGCCGTGGAGAAGAGACAGACTTCACTCTCACCATCACCAGTCTGCAACCTGAAGATTC TGCAGTTTACTACTGTCAACAGAGTTACAGTGACCTTCTCAGTTTCGGCGGAGGGACCAAGG TGGAGATCAAA.

A-2. Human Antibody (D-1) Against Human Influenza a Virus Subtype H3N2

Heavy chain variable region sequence:

(SEQ ID NO: 7)

 ${\tt CAGGTGCAGCTGGTGCAATCTGGGTCTGAGTTGAAGAAGCCTGGGGCCTCAGTGAAGGTTTC}$

CTGCAAGGCTTCTGGATACACCTTCACCTCTTATTCTATATATTGGGTGCGACAGGCCCCTG

- continued GACAAGGGCTTGAGTGGATGGATGGATGGATGGATCAACACCAACACTGGGAACCCAAGCTATGCCCAG GGCTTCACAGGACGGTTTGTCTTCTCCTTCGACACCTCTGTCAGCACGGCATATCTGGAGAT CAGCAGCCTAAAGGCTGAGGACACTGCCGTGTATTACTGTGCGAGGAAGGGGAGATTACGATA TTTTGACTGGTTATTATTATTACTTTGACTACTGGGGCCAGGGAACCCTGGTCACCGTCTCC TCA.

Light chain variable region sequence: (SEQ ID NO: 8) CAGACTGTGGGTGACCCAGGAGCCATCGTTCTCAGTGTCCCCTGGAGGGACAGTCACACTCAC TTGTGGCTTGAGCTCTGGGTCAGTCTCTCCTAGTTACTACGCCAGGCGGGGCCCAGGCAGA CAGGCCAGGCTCCACGCACGCTCATCTACAACACAAACACTCGCTCCTCTGGGGGCCCAGGCAGA TGATGAGTCTGATTATTTCTGTGTGCTGTATATGCCTAGTGGCGATTGGGTTTTCGGCGGAG GGACCAAGCTGACCGTCCTGAGT.

B-1. Human Antibody (E-2) Against Human Influenza B Virus

Heavy chain variable region sequence:

(SEQ ID NO: 9) CAGGTGCAGTTGCAGGAGTCGGGCCCAGGACTGGTGAAGCCTTTACAGACCCTGTCCCTCAC CTGCGTTGTCTCTGGTGACTCCATCAGCAGGGGGGGGGTGGTTACTACTGGAGTTGGGGCCAGC CCCCAGAGAGGGGCCTGGAGTGGATTGGGGACATCTATCACAGTGGGAGTACCAACTACAAC CCCGGCCCTCAAGAGTCGAACTACCATCTCAGTAGAGACGTCCAAGAACCAGTTCTCCCCTGCA GCTGAACTCTGTGACCGCCGCAGACACGGCCGTGTATTACTGTGCCAGAGAGCCCTCGTCG ACTACAGTGACTACAAGGTTGGGAAGGGTTATTTTGACTACTGGGGCCAGGGAGCCCTGGTC ACCGTCTCCTCA.

Light chain variable region sequence:

B-2. Human Antibody (B-3) Against Human Influenza B 50 Virus

Heavy chain variable region sequence: (SEQ ID NO: 11) CAGGTGAAGTTGGTGCAGTCTGGCGGAGGCGCAGTCCAGCCTGGGAGGGTCCCTGAGACTCTC CTGTGAGGGCGTTGAGTGGGTGGCATCTATTGGCGATAACGGAGGGAAAAGCATATTATGCGGAC GCAAGGGGCCTGAGGGCGGATTCACCGTGTCCAGAGACAATCCCCCAGAAGACAGTGTATCTGCAAAT GAGTGGCCTGAGACCCGAGGACACGGCTACATATTACTGTGCGAGAGAGTTTCCTTTCATGG GCATCTATGACTACGGCATGGACGCCTGGGGCCAAGGGACCACGGTCACCGTCGCCTCA.

-continued Light chain variable region sequence:

(SEQ ID NO: 12) CAGTCTGTGCTGGCTCAGCCACCCTCAGCGTCTGGGACCCCCGGGCAGAGGGTCATCATCTC

TTGTTCTGGAACCTCCTCCAACATCGGCGGTAATTCTGTCAACTGGTACCAGCACCCCCCAG GGGCGGCCCCGAGACTCCTCATCTATACTACCGATCAGCGACCCTCAGGGGTCCCTGACCGA TTCTCTGGCTCCAAGTCTGGCACCTCTGCCTCCCTGGCCATCAGTGGGCTCCAATCTGAGGA TGAGGCTGATTATTACTGTGAAGTTTGGGATGACAGCCTGACTCGTCCGGTGTTCGGCGGAG GGACCAAGTTGACCGTCCTACGT

Experimental Example 1-6

Staining Activity of Hybridoma Culture Supernatants in Cells Infected with Various Influenza Virus Vaccine Strains

Staining activities in cells infected with A/Hiroshima/52/ 20 05 (H3N2) strain was examined for the supernatant of the hybridoma producing the A-2, B-1, B-2, or D-1 monoclonal antibody as well as C43 antibody and F49 antibody. The staining test was performed by an ordinary indirect fluorescence antibody method. Here, the C43 antibody used as a 25 control is an antibody against the human influenza A virus, particularly a murine monoclonal antibody against a nucleoprotein (NP). Further, the F49 antibody is an antibody against the human influenza A virus subtype H3N2, particularly a murine monoclonal antibody against HA. 30

MDCK cells infected with the human influenza A virus subtype H3N2, A/Hiroshima/52/05 strain were washed with PBS (pH 7.4) on an 8-well chamber slide and fixed with absolute ethanol at room temperature for 10 minutes, in the same manner as in the case of measuring the neutralization ³⁵ activity in the section 1) of Experimental Example 1. Each undiluted hybridoma culture supernatant containing the monoclonal antibody was sequentially reacted for 60 minutes with a 1000-fold dilution of fluorescein isothiocyanate (FITC)-labeled rabbit anti-human IgG antibody (manufac- 40 monoclonal antibodies in this example. tured by Jackson). The stained cells washed with PBS were observed under a fluorescence microscope.

As a result, the culture supernatant of the hybridoma producing the A-1, B-1, B-2, or D-1 monoclonal antibody exhibited the same staining pattern as that of the murine 45 monoclonal antibody F49 against HA of the human influenza A virus subtype H3N2 (FIG. 11). This led to the speculation that these monoclonal antibodies obtained using the purified HA antigen were antibodies against HA of the influenza A virus subtype H3N2.

Experimental Example 1-7

Staining Activity of Hybridoma Culture Supernatants in Cells Infected with Various Influenza Virus Vaccine Strains

Staining activities in cells infected with B/Malaysia/2506/ 04 strain was examined for the supernatant of the hybridoma producing the B-3 or E-2 monoclonal antibody as well as 60 9F3 antibody and 9E10 antibody. The staining test was performed by an ordinary indirect fluorescence antibody method. Here, the 9F3 antibody used as a control is an antibody against the human influenza B virus, particularly a murine monoclonal antibody against NP. Further, the 9E10 65 antibody is an antibody against the human influenza B virus, particularly a murine monoclonal antibody against HA.

MDCK cells infected with the human influenza B virus, 15 B/Malaysia/2506/04 strain were washed with PBS (pH 7.4) on an 8-well chamber slide and fixed with absolute ethanol at room temperature for 10 minutes, in the same manner as in the case of measuring the neutralization activity in the section 1) of Experimental Example 1. Each undiluted hybridoma culture supernatant containing the monoclonal antibody was sequentially reacted for 60 minutes with a 1000-fold dilution of FITC-labeled rabbit anti-human IgG antibody (manufactured by Jackson). The stained cells washed with PBS were observed under a fluorescence microscope.

As a result, the culture supernatant of the hybridoma producing the B-3 or E-2 monoclonal antibody exhibited the same staining pattern as that of the murine monoclonal antibody 9E10 against HA of the human influenza B virus (FIG. 12). This led to the speculation that these monoclonal antibodies obtained using the purified HA antigen were antibodies against HA of the human influenza B virus.

Experimental Example 1-8

Epitope Mapping

Epitope analysis was performed for the B-1 and D-1

A total of 166 sets of peptides were synthesized which were the peptides of 15 residues consecutively selected from 345 amino acid residues which formed the hemagglutinin HA1 region including a signal peptide portion in Hi/05: A/Hiroshima/52/05 strain (isolated in 2005) and which were adjusted so that 13 residues were overlapped. A peptide array in which each peptide from the above-mentioned 166 sets had been immobilized on a glass surface was prepared. The glass surface was blocked with a buffer for blocking (Super Block® TBS manufactured by Piace), and subsequently, each monoclonal antibody diluted to 10 µg/mL with the buffer for blocking was reacted. After incubation, the peptide array was washed three times with TBS containing 55 0.1% Tween 20, and Cy5-labeled anti-human IgG (H+ L) diluted to 1 µg/mL was reacted. After incubation, the peptide array was washed three times with the above-mentioned tris buffered saline (TBS) and washed thoroughly with a 3 mM citrate buffer (SSC), then dried, and the fluorescence was measured by a fluorescence scanner to detect the peptide capable of reacting with the monoclonal antibody. As a control, an experiment in which no monoclonal antibody was added and other manipulations were performed in the same manner was performed simultaneously. An antigen antibody reaction was performed using the B-1 or D-1 monoclonal antibody, and epitope analysis was performed by an overlap peptide scanning method. Specifically, the

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analysis was conducted in accordance with the protocol of RepliTope[™] Microarrays (JPT Peptide Technologies Gmbh Germany).

As a result, a neutralization activity was detected in the sites shown in FIG. 13. Two peptide chain portions of hemagglutinin HA1 which reacted with the B-1 or D-1 monoclonal antibody were conserved except one residue in an N terminal side in the five H3N2 viral strains. Thus, it was confirmed that the variation was limited in this region. The 227th amino acid counting from the N-terminus of the amino acid sequence which forms hemagglutinin HA1 of the 10human influenza A virus subtype H3N2 is serine (S) or proline (P), but such difference has no effect on the neutralization activity. The amino acid residue at this position is also S or P in the literature (Karoline et al Virology J., 5, 40 (2008)). The variant amino acid at position 173 is asparagine $_{15}$ (N) or lysine (K), that in the virus vaccine strains other than A/Aichi strain in 1968 is K, and that is K or glutamic acid (E) in the literature. Further, the amino acid residues at positions 229 and 230 are different and are arginine (R) or glycine (G) and isoleucine (I) or valine (V), respectively, in 20 the literature (Underwood, Mol. Immunol., 21, 7 (1987)). The amino acid residues at positions 238 and 239 are different and are K or N and P or R, respectively, in the sequences registered in GenBank.

The two peptide chain portions recognized by the B-1 and D-1 monoclonal antibodies are conformationally close, and ²⁵ thus these antibodies are estimated to conformationally recognize the highly conservative two chains and are anticipated to be highly resistant to mutation of the virus (FIG. 14). The epitope portions recognized by the B-1 and D-1 monoclonal antibodies are different from the epitope recognized by the antibody (F10 antibody, J. Sui et al. Nature structural & molecular biology (2009)) which widely neutralizes the publicly known influenza virus strains. Thus, the viral type which can be neutralized by these monoclonal antibodies is thought to be different.

Further, the epitopes were searched by epitope database (http://www.immuneepitope.org/doc//influenza/index.html) linked to NIAID (National Institute of Allergy and Infectious Disease). As a result, although there was a rabbit polyclonal antibody (No. 42, 63) against a wider range of epitopes including the sequences recognized by the B-1 and D-1 monoclonal antibodies concerning linear epitopes, this includes a portion having many mutations. Concerning conformational epitopes, there is a murine monoclonal antibody (No. 34) which recognizes the epitope formed of partially overlapped amino acid residues at positions 229 45 and 230, but this epitope is different from the epitopes corresponding to the portions recognized by the B-1 and D-1 monoclonal antibodies. Therefore, the epitopes recognized by the B-1 and D-1 monoclonal antibodies are thought to be novel (see FIG. 15).

In FIG. 15: the portions recognized by the B-1 and D-1 monoclonal antibodies are shown in italic type; horizontal lines denote the numbers and the epitope portions described in Table 6 or B cell epitopes in the above-mentioned database; stars denote the epitope portions of No. 34 in Table 55 9 describing the conformational antibody against influenza in the same database; and triangles denote the sites at which the amino acid had been substituted in the strains registered in the literature or Pub Med other than the above-mentioned 5 strains.

Example 2

Production of Human Anti-Human Influenza Virus Antibody

In this example, the culture of the hybridomas R1D8 and K4E7 cloned in Example 1 is described, and the production of the B-1 monoclonal antibody (hereinafter, referred to as B-1 antibody) and the D-1 monoclonal antibody (hereinafter, referred to as D-1 antibody) is described.

Each hybridoma was cultured on a large scale using a serum-free medium (Hybridoma-SFMTM; GIBCO) in an incubator containing 5% CO2 at 37° C. The culture supernatant was collected, an antibody molecule was adsorbed to a Protein G SepharoseTM (Protein G SepharoseTM 4 Fast Flow; GE healthcare) column, which was then washed twice with PBS, and the antibody molecule was eluted with a solution of 0.17 M glycine at pH 2.3. The eluted antibody molecule was dialyzed against PBS using a dialysis membrane (Spectra/Por® (fractioned molecules); 6 K to 8 K; Nippon Genetics), and collected. Final yields of R1D8 and K4E7 were 0.249 mg/L and 24.38 mg/L, respectively.

Experimental Example 2-1

Effects of Human Anti-Human Influenza Virus Antibody (Preventive Effects)

In this experimental example, a survival rate and an effect on body weight changes were identified when the B-1 antibody or the D-1 antibody obtained by purifying in Example 2 was prophylactically administered to mice.

The B-1 antibody or the D-1 antibody obtained by purifying in Example 2 was intraperitoneally administered at a dosage of 100 ug/mouse to 4-week old female Balb/c mice (5 in one group), and 1×10^5 FFU/mouse of an influenza virus A/Guizhou/54/89xA/PR/8/34 (H3N2) strain was nasally inoculated thereto after 24 hours. As a control group, 0.5 mL/mouse of PBS was administered instead of the monoclonal antibody. The A/Guizhou/54/89xA/PR/8/34 (H3N2) strain is a re-assortant virus obtained by replacing HA and NA in an A/PR/8/34 (H1N1) strain with those in an 35 A/Guizhou/54/89 (H3N2) strain.

The survival rates in the D-1 antibody-administered group and B-1 antibody-administered group were 40% and 100%, respectively, whereas the survival rate in the control group was 0%. In particular, a good effect was obtained with the B-1 antibody (FIG. 16). Further, concerning the body weight change after the infection with the virus, the body weight was slightly reduced in the B-1 antibody-administered group. Meanwhile, the body weight in the D-1 antibodyadministered group was reduced to a similar degree to that in the control group (FIG. 17). In FIG. 17, the body weight change in an individual mouse is shown in each plot.

Experimental Example 2-2

Effects of Human Anti-Human Influenza Virus Antibody (Therapeutic Effects)

In this experimental example, a survival rate and an effect on body weight changes were identified when the B-1 antibody or the D-1 antibody obtained by purifying in Example 2 was therapeutically administered to mice.

An influenza virus A/Guizhou/54/89xA/PR/8/34 (H3N2) strain at a dosage of 1×10⁵ FFU/mouse was nasally inoculated to 4-week old female Balb/c mice (5 in one group), and the B-1 antibody or the D-1 antibody obtained by purifying in Example 2 was intraperitoneally administered at a dosage of 100 µg/mouse after 24 hours. As a control group, 0.5 mL/mouse of PBS was administered instead of the monoclonal antibody.

The survival rates in the D-1 antibody-administered group and B-1 antibody-administered group were 40% and 60%, respectively, whereas the survival rate in the control group was 0% (FIG. 18). Further, concerning the body weight change after the infection with the virus, the body weight

was reduced rather slightly with the B-1 antibody as compared to the control group. Meanwhile, the body weight was reduced to a similar degree with the D-1 antibody to that in the control group (FIG. **19**). In FIG. **19**, the body weight change in an individual mouse is shown in each plot.

INDUSTRIAL APPLICABILITY

As described above in detail, in the antibodies of the present invention, the antibody against the human influenza 10 A virus subtype H3N2 has a neutralization activity against at least an A/Hiroshima/52/05 strain (isolated in 2005) viral strain, and the antibody against the human influenza B virus has a neutralization activity against at least a B/Malaysia/ 2506/04 strain (isolated in 2004) viral strain. Further, the 15 antibodies of the present invention also have neutralization activities against influenza virus vaccine strains in various generations including: various viral strains from the human influenza A virus subtype H3N2, such as an A/Aichi/2/68 strain (isolated in 1968), an A/Guizhou/54/89 strain (isolated 20 in 1989), an A/Wyoming/3/03 strain (isolated in 2003), and an A/New York/55/04 strain (isolated in 2004); and various viral strains from the human influenza B virus, such as a B/Victoria/2/87 strain (isolated in 1987), a B/Mie/1/93 strain (isolated in 1993), and a B/Shanghai/261/02 strain (isolated 25 in 2002).

That is, the antibody against the human influenza A virus subtype H3N2 of the present invention has an activity against a region which has been conserved for about 20 years, and also includes one having an activity against a region which has been conserved for 40 years or more. The antibody against the human influenza B virus also has an activity against a region which has been conserved for 20 years or more. Meanwhile, an HI (hemagglutination inhibition) activity in the antibody of the present invention is equal to or less than the detection limit.

Further, when administered prophylactically or therapeutically in vivo, the antibody against the human influenza A virus subtype H3N2 of the present invention exhibits effects on a survival rate and a weight loss in at least the infection with the influenza virus A/Guizhou/54/89xA/PR/8/34 (H3N2) strain.

Due to the above-mentioned properties, the human antihuman influenza virus antibody of the present invention has effects particularly on regions conserved in an influenza virus to be easily mutated. The influenza A virus subtype H3N2 is a subtype which caused a worldwide epidemic in the past, and strains resistant to medicaments such as amantadine having the anti-influenza virus action have been increasing in recent years. However, the composition including the human anti-human influenza virus antibody of the present invention is expected to have a therapeutic effect on influenza even when new influenza virus occurs and becomes epidemic in the future. The human anti-human influenza virus antibody of the present invention has a neutralization activity against the human influenza A virus subtype H3N2 or the human influenza B virus. Thus, the composition including at least one or more kinds of such antibodies is expected to be utilized as a therapeutic drug and a prophylactic drug for influenza.

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Thr															
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Tyr	Tyr	Ala 35	Ser	Trp	Tyr	Gln	Gln 40	Thr	Pro	Gly	Gln	Ala 45	Pro	Arg	Thr
Leu	Ile 50	Tyr	Asn	Thr	Asn	Thr 55	Arg	Ser	Ser	Gly	Val 60	Pro	Asp	Arg	Phe
Ser 65	Gly	Ser	Phe	Leu	Gly 70	Ser	Asp	Ala	Ala	Leu 75	Thr	Ile	Thr	Gly	Ala 80
Gln	Ala	Asp	Asp	Glu 85	Ser	Asp	Tyr	Phe	Cys 90	Val	Leu	Tyr	Met	Pro 95	Ser
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Pro	Lys	Ala 115	Ala	Pro	Ser	Val	Thr 120	Leu	Phe	Pro	Pro	Ser 125	Ser	Glu	Glu
Leu	Gln 130	Ala	Asn	Lys	Ala	Thr 135	Leu	Val	Суз	Leu	Ile 140	Ser	Asp	Phe	Tyr
Pro 145	Gly	Ala	Val	Thr	Val 150	Ala	Arg	Lys	Ala	Asp 155	Ser	Ser	Pro	Val	Lys 160
Ala	Gly	Val	Glu	Thr 165	Thr	Thr	Pro	Ser	Lys 170	Gln	Ser	Asn	Asn	Lys 175	Tyr
Ala	Ala	Ser	Ser 180	Tyr	Leu	Ser	Leu	Thr 185	Pro	Glu	Gln	Trp	Lys 190	Ser	His
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Gly	Tyr	Tyr 35	Trp	Ser	Trp	Val	Arg 40	Gln	Pro	Pro	Glu	Arg 45	Gly	Leu	Glu
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Ala Ser Ile Trp His Asn Gly Gly Lys Ala Tyr Tyr Ala Asp Ser Val Lys Gly Arg Phe Thr Val Ser Arg Asp Asn Pro Gln Lys Thr Val Tyr Leu Gln Met Ser Gly Leu Arg Pro Glu Asp Thr Ala Thr Tyr Tyr Cys Ala Arg Glu Phe Pro Phe Met Gly Ile Tyr Asp Tyr Gly Met Asp Ala Trp Gly Gln Gly Thr Thr Val Thr Val Ala Ser Ala Ser Thr Lys Gly Pro Ser Val Phe Pro Leu Ala Pro Ser Ser Lys Ser Thr Ser Gly Gly Thr Ala Ala Leu Gly Cys Leu Val Lys Asp Tyr Phe Pro Glu Pro Val Thr Val Ser Trp Asn Ser Gly Ala Leu Thr Ser Gly Val His Thr <210> SEO ID NO 34 <211> LENGTH: 210 <212> TYPE: PRT <213> ORGANISM: Homo sapiens <400> SEOUENCE: 34 Gln Ser Val Leu Ala Gln Pro Pro Ser Ala Ser Gly Thr Pro Gly Gln Arg Val Ile Ile Ser Cys Ser Gly Thr Ser Ser Asn Ile Gly Gly Asn Ser Val Asn Trp Tyr Gln His Pro Pro Gly Ala Ala Pro Arg Leu Leu Ile Tyr Thr Thr Asp Gln Arg Pro Ser Gly Val Pro Asp Arg Phe Ser Gly Ser Lys Ser Gly Thr Ser Ala Ser Leu Ala Ile Ser Gly Leu Gln Ser Glu Asp Glu Ala Asp Tyr Tyr Cys Glu Val Trp Asp Asp Ser Leu Thr Arg Pro Val Phe Gly Gly Gly Thr Lys Leu Thr Val Leu Arg Gln Pro Lys Ala Ala Pro Ser Val Thr Leu Phe Pro Pro Ser Ser Glu Glu Leu Gln Ala Asn Lys Ala Thr Leu Val Cys Leu Ile Ser Asp Phe Tyr Pro Gly Ala Val Thr Val Ala Trp Lys Ala Asp Ser Ser Pro Val Lys Ala Gly Val Glu Thr Thr Thr Pro Ser Lys Gln Ser Asn Asn Lys Tyr Ala Ala Ser Ser Tyr Leu Ser Leu Thr Pro Glu Gln Trp Lys Ser His Lys Ser Tyr Ser Cys Gln Val Thr His Glu Glu Ser Thr Val Glu Lys 2.05 Thr Met

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Arg 65	Leu	Leu	Ile	Ser	Ser 70	Ala	Ser	Asn	Leu	Trp 75	Ala	Gly	Val	Pro	Pro 80
Arg	Phe	Ser	Gly	Arg 85	Gly	Glu	Glu	Thr	Asp 90	Phe	Thr	Leu	Thr	Ile 95	Thr
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Thr	Val	Ala	Ala	Pro	Ser	Val	Phe	Thr	Phe	Pro	Pro	Ser	Asp	Glu	Gln
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_	50	_	_	_	_	55	_	_	_	_	60	_	_	_	_	_
Gly 65	Leu	Glu	Trp	Ile	Gly 70	Asp	Ile	Tyr	His	Ser 75	Gly	Ser	Thr	Asn	Tyr 80	
Asn	Pro	Ala	Leu	Lys 85	Ser	Arg	Thr	Thr	Ile 90	Ser	Val	Glu	Thr	Ser 95	Lys	
Asn	Gln	Phe	Ser 100	Leu	Gln	Leu	Asn	Ser 105	Val	Thr	Ala	Ala	Asp 110	Thr	Ala	
Val	Tyr	Tyr 115	Сүз	Ala	Arg	Glu	Pro 120	Pro	Pro	Asp	Tyr	Ser 125	Asp	Tyr	Lys	
Val	Gly 130	Lys	Gly	Tyr	Phe	Asp 135	Tyr	Trp	Gly	Gln	Gly 140	Ala	Leu	Val	Thr	
Val 145	Ser	Ser	Ala	Ser	Thr 150	ГЛЗ	Gly	Pro	Ser	Val 155	Phe	Pro	Leu	Ala	Pro 160	
Ser	Ser	Lys	Ser	Thr 165	Ser	Gly	Gly	Thr	Ala 170	Ala	Leu	Gly	Сүз	Leu 175	Val	
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Leu	Ser	Pro 35	Gly	Glu	Arg	Ala	Thr 40	Leu	Ser	Сүз	Arg	Ala 45	Ser	Glu	Thr	
Val	Asp 50	Thr	Tyr	Leu	Ala	Trp 55	Tyr	Gln	Gln	Lys	Pro 60	Gly	Gln	Ala	Pro	
Arg 65	Leu	Leu	Ile	Asn	Asp 70	Ala	Ser	Lys	Arg	Ala 75	Thr	Gly	Ile	Pro	Ala 80	
Arg	Phe	Ser	Gly	Ser 85	Gly	Ser	Gly	Thr	Asp 90	Phe	Thr	Leu	Thr	Ile 95	Ser	
Gly	Leu	Glu	Pro 100	Glu	Asp	Phe	Ala	Val 105	Tyr	Trp	CÀa	Gln	Gln 110	His	Ser	
Asn	Trp	Pro 115	Pro	Thr	Phe	Gly	Gln 120	Gly	Ser	Arg	Leu	Glu 125	Ile	Lys	Arg	
Thr	Val 130	Ala	Ala	Pro	Ser	Val 135	Phe	Ile	Phe	Pro	Pro 140	Ser	Asp	Glu	Gln	
Leu 145	Lys	Ser	Gly	Thr	Ala 150	Ser	Val	Val	Cys	Leu 155	Leu	Asn	Asn	Phe	Tyr 160	
Pro	Arg	Glu	Ala	Lys 165	Val	Gln	Trp	Lys	Val 170	Asp	Asn	Ala	Leu	Gln 175	Ser	
Gly	Asn	Ser	Gln 180	Glu	Ser	Val	Thr	Glu 185	Gln	Asp	Ser	Lys	Asp 190	Ser		
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Arg Gl	ly	Leu 35	Thr	Met	Glu	Phe	Gly 40	Leu	Ile	Trp	Ile	Phe 45	Leu	Val	Ala
Leu Le 50	eu 0	Gly	Gly	Ala	Gln	Сув 55	Gln	Val	Lys	Leu	Val 60	Gln	Ser	Gly	Gly
Gly Al 65	la	Val	Gln	Pro	Gly 70	Arg	Ser	Leu	Arg	Leu 75	Ser	Суз	Glu	Ala	Ser 80
Gly Ph	he	Asp	Phe	Thr 85	Val	Tyr	Asp	Ile	His 90	Trp	Val	Arg	Gln	Ala 95	Pro
Gly Ly	Уs	Gly	Leu 100	Glu	Trp	Val	Ala	Ser 105	Ile	Trp	His	Asn	Gly 110	Gly	Lys
Ala Ty	yr	Tyr 115	Ala	Asp	Ser	Val	Lys 120	Gly	Arg	Phe	Thr	Val 125	Ser	Arg	Asp
Asn Pr 13	r0 30	Gln	Lys	Thr	Val	Tyr 135	Leu	Gln	Met	Ser	Gly 140	Leu	Arg	Pro	Glu
Asp Th	hr	Ala	Thr	Tyr	Tyr	Суз	Ala	Arg	Glu	Phe	Pro	Phe	Met	Gly	Ile
Tyr As	ab	Tyr	Gly	Met	Aab	Ala	Trp	Gly	Gln	Gly	Thr	Thr	Val	Thr	Val
Ala Se	er	Ala	Ser	165 Thr	Lys	Gly	Pro	Ser	170 Val	Phe	Pro	Leu	Ala	175 Pro	Ser
Ser Ly	Ув	Ser	180 Thr	Ser	Gly	Gly	Thr	185 Ala	Ala	Leu	Gly	Сув	190 Leu	Val	Lys
Asp Ty	yr	195 Phe	Pro	Glu	Pro	Val	200 Thr	Val	Ser	Trp	Asn	205 Ser	Gly	Ala	Leu
21 Thr Se	10 er	Glv	Val	His	Thr	215				-	220		-		
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Glu Ly	Уs	Thr 35	Gly	Leu	Arg	Thr	Leu 40	Ser	Ser	Met	Ala	Ser 45	Phe	Pro	Leu
Leu Le 50	eu 0	Thr	Leu	Leu	Thr	His 55	Суз	Ala	Gly	Ser	Trp 60	Ala	Gln	Ser	Val
Leu Al 65	la	Gln	Pro	Pro	Ser 70	Ala	Ser	Gly	Thr	Pro 75	Gly	Gln	Arg	Val	Ile 80
Ile Se	ər	Cys	Ser	Gly	Thr	Ser	Ser	Asn	Ile	Gly	Gly	Asn	Ser	Val or	Asn
Тгр Ту	yr	Gln	His	Pro	Pro	Gly	Ala	Ala	Pro	Arg	Leu	Leu	Ile	Tyr	Thr
Thr As	ab	Gln	100 Arg	Pro	Ser	Gly	Val	105 Pro	Asp	Arg	Phe	Ser	110 Gly	Ser	Lys
	-	115	2			4	120		-	5		125	-		-

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Ser	Gly 130	Thr	Ser	Ala	Ser	Leu 135	Ala	Ile	Ser	Gly	Leu 140	Gln	Ser	Glu	Asp	
Glu 145	Ala	Asp	Tyr	Tyr	Cys 150	Glu	Val	Trp	Asp	Asp 155	Ser	Leu	Thr	Arg	Pro 160	
Val	Phe	Gly	Gly	Gly 165	Thr	Lys	Leu	Thr	Val 170	Leu	Arg	Gln	Pro	Lys 175	Ala	
Ala	Pro	Ser	Val 180	Thr	Leu	Phe	Pro	Pro 185	Ser	Ser	Glu	Glu	Leu 190	Gln	Ala	
Asn	Lys	Ala 195	Thr	Leu	Val	Суз	Leu 200	Ile	Ser	Asp	Phe	Tyr 205	Pro	Gly	Ala	
Val	Thr 210	Val	Ala	Trp	Lys	Ala 215	Asp	Ser	Ser	Pro	Val 220	Lys	Ala	Gly	Val	

Glu Thr Thr Thr Pro Ser Lys Gln Ser Asn Asn Lys Tyr Ala Ala Ser 225 230 235 240 Ser Tyr Leu Ser Leu Thr Pro Glu Gln Trp Lys Ser His Lys Ser Tyr 245 250 255 Ser Cys Gln Val Thr His Glu Glu Ser Thr Val Glu Lys Thr Met 265 270

The invention claimed is:

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1. An isolated human anti-human influenza virus monoclonal antibody, comprising a variable region comprising:

- a heavy chain comprising the amino acid sequence of SEQ ID NO: 31 and a light chain comprising the amino 30 acid sequence of SEQ ID NO: 32; or
- a heavy chain comprising the amino acid sequence of SEQ ID NO: 33 and a light chain comprising the amino acid sequence of SEQ ID NO: 34.

2. The isolated human anti-human influenza virus mono- 35 clonal antibody according to claim 1, comprising a neutralization activity against a human influenza B virus.

3. The isolated human anti-human influenza virus monoclonal antibody according to claim 1, comprising a neutralization activity against at least a B/Malaysia/2506/04 strain.

40 4. The isolated human anti-human influenza virus monoclonal antibody according to claim 1, wherein an antigen of the isolated anti-human influenza virus monoclonal antibody is a human influenza virus B/Malaysia/2506/04 strain.

5. An isolated human anti-human influenza virus monoclonal antibody according to claim 1, wherein the isolated 45 antibody comprises an isolated intact antibody.

6. A composition comprising the isolated human antihuman influenza virus monoclonal antibody according to claim 1 and a pharmaceutically acceptable carrier.

7. A hybridoma that produces the isolated human antihuman influenza virus monoclonal antibody according to claim 1.

8. The isolated human anti-human influenza virus monoclonal antibody according to claim 1, wherein the isolated antibody is a humanized or chimeric antibody.

9. A composition comprising the isolated human antihuman influenza virus monoclonal antibody according to claim 1, and at least one preservative.

10. A composition comprising the isolated human antihuman influenza virus monoclonal antibody according to claim 1, and at least one stabilizer.

11. An isolated DNA encoding a variable region of an isolated human anti-human influenza virus monoclonal antibody comprising a neutralization activity against a human influenza B virus, the DNA comprising a base sequence set forth in any one of SEQ ID NOS: 9 to 12 or a sequence having substitutions of one or more nucleotides in the base sequence, and which encodes SEQ ID NOS: 31-34, respectively.

12. The isolated DNA according to claim 11, wherein the isolated DNA is a cDNA.

> * * *